SUMMARY

The immunization of rabbits with cells of mouse leukemia L1210 has been studied. The immune process was dissected into its γM and γG components. The main objective was the preparation of antisera with high cytolytic potencies.

A single i.v. injection of viable L1210 cells produced a classical immune response in rabbits, with γM antibody activity first detected on Day 4, maximal on Day 6, and diminishing thereafter; in contrast, γG activity was detected on Day 8, was maximal around Day 14, and diminished thereafter. Interestingly, additional immunizations gave new peaks of γM activity, each smaller than the previous one. As expected, γG activity rose sharply after secondary immunization and more gradually with further immunizations.

For production of highly potent cytolytic antisera, repeated i.v. injection of large numbers of viable leukemia cells was the most effective procedure tested. Immunization with homogenized or sonically disrupted cells was significantly inferior. Evidence for immune exhaustion after long-continued immunization was obtained.

INTRODUCTION

Attempts to treat leukemia by passive administration of heterologous antisera have been made both in experimental animals and in man (3-5, 7, 10, 12, 13, 15-17, 22-25, 29). With one exception (3), mild but significant therapeutic benefits have been obtained in experimental animals (7, 12, 16, 17, 22; 23, 25). While clinical trials with antisera to leukemia cells or with antilymphocyte globulin are currently proceeding at a number of laboratories (5, 15, 24, 29), it remains questionable whether such antisera have substantial therapeutic potentials in man, especially since antilymphocyte globulin possesses strong immunosuppressive properties (26).

We recently found that when a rabbit antiserum prepared against mouse leukemia cells is injected into mice carrying that leukemia there is a direct relationship above a certain threshold between the number of cytolytic potency units injected and therapeutic effectiveness (22). This finding suggested that passive therapy with antiserum can give positive results only when the antiserum has sufficiently high cytolytic strength and is administered in sufficient amount (22); similarly, Mohos and Kidd stressed the need for high cytolytic potency in antisera which are to be used for passive immunotherapy of tumors (17). Unfortunately, much of the more recent experience (9, 18, 30) in production of the closely related ALS is irrelevant, since ALS is raised to be effective for allograft prolongation in vivo, rather than to be strongly cytolytic, and there is poor correlation between these two properties (2, 8, 9).

The in vivo therapeutic potential of rabbit antisera to L1210 leukemia cells has already been demonstrated (16, 22, 23), and the in vitro specificity of such antisera to thymic, splenic, and leukemic lymphocytes has been investigated (1). The present study was therefore confined to attempts at production of highly cytolytic rabbit antisera to Leukemia L1210. For this purpose, immunization schedules that differed in amount of tissue, route of administration, and disruption of cells were studied; where pertinent, the immune response was dissected into its γG and γM antibody components.

MATERIALS AND METHODS

Mice and Leukemia. All mice were obtained from the Jackson Laboratory. The lymphocytic Leukemia L1210 (11) was transplanted i.p. in DBA/2 mice. Leukemia cells were harvested 6 days after i.p. transplantation of approximately 300,000 cells. Erythrocytes present were destroyed by hypotonic lysis; 1.5 volumes distilled water were added slowly with stirring at room temperature. After stirring further for 10 min, concentrated buffer was added to restore isotonicity. Cells were washed in isotonic Locke's buffer (20), resuspended in buffer, and standardized by hemacytometer count.

Cytolysis Assay System. A small scale assay system (21) was used. In outline, each assay tube contained 100,000 viable lymphocytes, suitably absorbed 10% complement (19), and various dilutions of antiserum. Tubes were incubated for 1 hr at 37° and placed in ice water. For each tube in turn, the supernatant was sucked off the cells that had settled, 0.02 ml of vital dye was added, and 200 cells were classified under the microscope as stained or unstained. Results, in percentage of stained cells, were plotted against final antiserum concentration (%), and the cytolytic titer was read at 50% stained cells, with a correction for stained cells in the control tube (19). The potency of an antiserum was expressed as 100 divided by the cytolytic titer obtained in this assay system (19, 21).

The abbreviation used is: ALS, antilymphocyte serum.

1 This investigation was supported by USPHS Research Grant CA-04469 from the National Cancer Institute.

2 Fellow in Surgery.

Received March 18, 1970; accepted September 14, 1970.

JANUARY 1971
Separation of γG and γM Antibodies. The 7 S and 19 S antibodies were separated by centrifugation for approximately 20 hr at 112,000 x g (28) in a horizontal rotor (SB-283) of a Model B-60 ultracentrifuge (International Equipment Co., Boston, Mass.). The rabbit antiserum to be separated was diluted with 4 parts 10% sucrose, and 1.0 ml was layered on 11.0 ml of a continuous gradient of 10 to 40% (w/v) sucrose in 0.9% NaCl solution. The gradient was formed from the bottom upward with a finger pump (Model 600-1200, Harvard Apparatus Co., Dover, Mass.). After centrifugation, the bottom of the tube was punctured with a 20-gauge hypodermic needle, and 0.5 ml fractions were collected with the finger pump. The fractions were tested for light absorption at 280 μμ in microcuvets (Chart 1) to determine the point of separation between γM and γG antibodies (28).

RESULTS

Immunization Experiment A. Three rabbits were each given i.v. injections of 30 million L1210 cells in a suspension of 0.9% NaCl solution. The same quantity of cells was injected again on Days 50, 87, 89, 99, 101, 106, and 108. The rabbits were bled at various times during this course of immunization. At each time point, equal volumes of sera from each of the 3 rabbits were pooled. Sera were separated into γG and γM fractions by ultracentrifugation in a continuous sucrose gradient (see “Materials and Methods”). These fractions contained all proteins present in the whole antiserum, except those contained in the single tube discarded at the minimum of absorbance (Chart 1). Each fraction could be tested directly for cytolytic antibody activity against L1210 cells without removing sucrose by dialysis.

Immunization Experiment B. Twenty-four rabbits were divided into 6 groups of 4, designated A to F, and immunized with L1210 cells by different procedures (Table 1). For the i.v. immunizations, each rabbit received the number of viable L1210 cells recorded in Footnote a of Table 1, or their equivalent in homogenized cells (3 min in a cooled semimicro container of a Waring Blender), or sonically disrupted cells (3 min at 70% power in a water-cooled Raytheon Model DF101 sonic oscillator).

The potencies of the resulting antisera were determined by immune cytolysis against L1210 cells. On Day 25, 1 week after the 2nd immunization with L1210 cells, the antibody response of rabbits in Group C, injected with homogenized cells, was much lower than that of other groups. However, after immunization with 6 further i.v. injections and/or 1 further i.m. injection of adjuvant-dispersed cells, the cytolytic potencies of sera from all groups of rabbits converged (Table 1).

The antibody activity of sera from rabbits given injections of leukemia cells incorporated in Freund-McDermott adjuvant (Groups E and F) was dissected into its γM and γG components. For rabbits in Group E, all the cytolytic antibody activity of serum taken on Day 6 was present in the γM fraction. For the serum of Day 18, 52% of the antibody activity was in the γM fraction, and the balance was in the γG fraction;
Antisera to Leukemia L1210

**Table 1**

<table>
<thead>
<tr>
<th>Group</th>
<th>No. of rabbits at end of experiment</th>
<th>State of L1210 cells injected</th>
<th>Route ( ^{a} )</th>
<th>Potencies(^{b} ) of pooled rabbit antisera for immune cytolysis of L1210 cells, after</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Immunization 1</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Day 6(^{c} )</td>
<td>Day 11(^{c} )</td>
</tr>
<tr>
<td>A</td>
<td>3</td>
<td>Whole cells, 1 dose/day i.v.</td>
<td>46</td>
<td>47</td>
</tr>
<tr>
<td>B</td>
<td>3</td>
<td>Whole cells, 3 divided doses(^{d} ) i.v.</td>
<td>47</td>
<td>64</td>
</tr>
<tr>
<td>C</td>
<td>4</td>
<td>Homogenized whole cells i.v.</td>
<td>8</td>
<td>11</td>
</tr>
<tr>
<td>D</td>
<td>3</td>
<td>Sonically disrupted whole cells i.v.</td>
<td>34</td>
<td>136</td>
</tr>
<tr>
<td>E</td>
<td>2</td>
<td>Freund-McDermott adjuvant only i.m.</td>
<td>30</td>
<td>78</td>
</tr>
<tr>
<td>F</td>
<td>4</td>
<td>Whole cells + Freund-McDermott adjuvant i.v. + m.p.</td>
<td>60</td>
<td>103</td>
</tr>
</tbody>
</table>

\(^{a}\) Immunizations i.v., each with approximately 40 million L1210 cells, were given on Days 0 (initial injection), 19, 26, 28, 30, 33, 35, and 37. Injections i.m. of approximately 40 million L1210 cells incorporated in complete Freund-McDermott adjuvant (Groups E and F) were given on Days 0, 19, and 35. For multiportal injections (m.p.), the total dose of approximately 40 million L1210 cells incorporated in adjuvant was divided between i.m., s.c., i.p., and foot pad sites.

\(^{b}\) Mean of two determinations made on different days on sera pooled for each group.

\(^{c}\) After first immunization with L1210 cells.

\(^{d}\) The same dose of L1210 cells used for Group A was divided into 3 equal parts and injected separately, spaced over 24 hr.

---

**Table 2**

Rabbit immunization with L1210 leukemia cells

<table>
<thead>
<tr>
<th>Group</th>
<th>State of injected L1210 cells</th>
<th>No. of L1210 cells injected each time(^{a} )</th>
<th>Rabbits alive at end of experiment</th>
<th>Cytolytic potencies(^{b} ) of antisera</th>
</tr>
</thead>
<tbody>
<tr>
<td>G</td>
<td>Whole cells i.v.</td>
<td>20 million</td>
<td>a</td>
<td>2426</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>b</td>
<td>1440</td>
</tr>
<tr>
<td>H</td>
<td>Whole cells i.v.</td>
<td>60 million</td>
<td>c</td>
<td>5380</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>d</td>
<td>2280</td>
</tr>
<tr>
<td>I</td>
<td>Whole cells i.v.</td>
<td>180 million</td>
<td>f</td>
<td>7190</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>g</td>
<td>8210</td>
</tr>
<tr>
<td>J</td>
<td>Whole sonically disrupted cells</td>
<td>60 million</td>
<td>h</td>
<td>8720</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>i</td>
<td>2085</td>
</tr>
<tr>
<td>K</td>
<td>Supernatant after sonic disruption, centrifugation at 1000 x g</td>
<td>60 million</td>
<td>l</td>
<td>2655</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>m</td>
<td>2253</td>
</tr>
<tr>
<td>L</td>
<td>Homogenized, then treated as for K</td>
<td>60 million</td>
<td>n</td>
<td>1465</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>o</td>
<td>1020</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>p</td>
<td>1610</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>q</td>
<td>1403</td>
</tr>
</tbody>
</table>

\(^{a}\) All injections were given i.v. twice weekly for 3 weeks.

\(^{b}\) Mean of two separate determinations. Rabbits were bled 10 days after the last injection.
L) gave antisera that were significantly less potent ($p < 0.01$) than if the same number of viable cells was used (Group H).

All rabbits that survived were given 2 further i.v. injections of 140 million viable L1210 cells, and the resulting antisera were retested. When the results obtained (Table 3) were evaluated by the analysis of variance, the overall findings were nonsignificant. However, the progression of mean results obtained for the 6 groups of rabbits appeared to be geometric and suggested a logarithmic transformation of the data. Upon repetition of an analysis of variance following such transformation, the potencies of antisera from rabbits initially immunized with viable cells (Groups G, H, and I) were significantly greater (0.01 < $p < 0.025$) than for rabbits immunized with disrupted cells (Groups J, K, and L). These results further confirm the finding (Table 2) that viable cells are more immunogenic than disrupted cells.

The 7 rabbits with the highest cytolytic potencies (Table 3) were further immunized on Days 5, 7, 19, 21, 33, 35, 47, 49, 61, 63, 75, 77, 89, 91, 117, and 119 after this bleeding. Between 40 and 282 million viable L1210 cells were injected i.v. each time. Approximately 40 to 50 ml of whole blood were taken from each rabbit by cardiac puncture on days 13, 18, 42, 47, 56, 69, 89, and 130. The sera of Rabbits a, d, f, g, h, j, and l, obtained on Day 130, had mean cytolytic potencies (determined in duplicate experiments) that were, respectively, 24, 24, 51, 56, 76, 57, and 35% of the potencies on Day 0 (Table 3); the mean reduction in cytolytic potency for all rabbits was 46%.

The above experiments suggested the presence of “immune exhaustion.” To check this further, the same rabbits were immunized with 150 ± 30 million viable L1210 cells, given i.v. on each of 6 occasions spaced evenly over 3 weeks, without intermediate bleedings. Sera were collected 7 days after the last injection or 158 days after harvest of the sera evaluated in Table 2. Compared to the latter, the cytolytic potencies of sera from the surviving Rabbits a, d, f, g, h, and j were changed by -64, -40, +70, -48, +6, and +14%, respectively (mean of duplicate experiments). Thus, following intensive hyperimmunization, antibody activity decreased in 3 rabbits, increased in 1 rabbit, and was substantially unchanged in 2 rabbits. Taken together with the data in the previous paragraph, these results substantiate a mild and individually variable degree of immune exhaustion in rabbits subjected to courses of long-continued immunizations with intermittent bleedings.

**DISCUSSION**

This study has illustrated that the immune response of rabbits given i.v. injections of mouse leukemia cells follows a classic pattern (27). There is an early $\gamma M$ response, followed quickly by a $\gamma G$ response, which shuts off $\gamma M$ production (28). In addition, a far less widely known facet of the immune response has been illustrated: repeated i.v. injections cause new but gradually diminishing peaks of $\gamma M$ antibody activity (Chart 2).

We previously found that a strong correlation exists between the *in vitro* cytolytic potency of heterologous anti-leukemia sera and their *in vivo* therapeutic effectiveness against mouse leukemias (22). Since the cytolytic activity of $\gamma M$ antibody produced by i.v. injection was highest after the very first immunization, and even then was only about 3 to 5% of the maximal cytolytic activity of $\gamma G$ antibody produced after hyperimmunization, it seems unlikely that rabbit $\gamma M$ antibody can be used successfully for *in vivo* therapy of mouse leukemias. Further, the distribution of $\gamma M$ and $\gamma G$ antibody activity was similar after i.m. immunization with leukemia cells incorporated in complete Freund-McDermott adjuvant. Hence, we know of no method of immunization which would produce a high level of $\gamma M$ cytolytic antibody activity in rabbits. Similarly, $\gamma G$, rather than $\gamma M$, appears to be the active antibody in ALS (14, 30).

The most effective procedure for preparation of ALS, as evaluated *in vivo* by skin allograft survival, is i.v. injection of viable lymphoid cells, plus foot pad injection of cells incorporated in complete Freund's adjuvant (30). In contrast, the *in vitro* leukagglutination titer of rabbit antisera was essentially unchanged when injection of adjuvant-suspended cells was omitted (30). Similarly, in the present study, we failed to obtain statistically significant evidence for increased cytolytic potency for sera from rabbits that received adjuvant-suspended cells in addition to their immunization by the i.v. route. With regard to future work, use of leukemic lymphocytes cross-linked with 1,5-difluoro-2,4-dinitrobenzene may produce further increases in the potency of the resulting antisera (6). Most desirable would be the chemical purification of leukemia-specific cell surface antigens for use in immunizations.

**ACKNOWLEDGMENTS**

Best thanks are due to Dr. Melvin Schwartz for his valuable help in the statistical evaluation of the data.
REFERENCES


Immunization Schedules for Potent Rabbit Antisera to Leukemia L1210

Chung-Ai H. Kim and Arnold E. Reif


Updated version
Access the most recent version of this article at:
http://cancerres.aacrjournals.org/content/31/1/7

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.