Inhibition of Skin Tumor Promoter-caused Induction of Epidermal Ornithine Decarboxylase in SENCAR Mice by Polyphenolic Fraction Isolated from Green Tea and Its Individual Epicatechin Derivatives

Rajesh Agarwal,2 Santosh K. Katiyar, Syed I.A. Zaidi, and Hasan Mukhtar1

Department of Dermatology, Skin Diseases Research Center, University Hospitals of Cleveland, Case Western Reserve University, and Department of Veterans Affairs Medical Center, Cleveland, Ohio 44106

INTRODUCTION

Cancer chemoprevention is an important emerging area of research which in addition to providing a practical approach to identifying potentially useful inhibitors of cancer development, also affords opportunities for studying the mechanisms of carcinogenesis (Refs. 1–3 and references therein). For example, the involvement of cytochrome P-450 in the metabolic activation of polycyclic aromatic hydrocarbons leading to tumor initiation was made apparent by studies showing that inhibitors of aryl hydrocarbon hydroxylase, such as synthetic flavone, can inhibit TPA-induced epidermal ODC activity in a dose-dependent manner. The inhibitory effect of GTP was also dependent on the time of its application relative to TPA treatment. Maximum inhibitory effect was observed when GTP was applied 30 min prior to topical application of TPA. GTP application to animals also inhibited the induction of epidermal ODC activity caused by several structurally different mouse skin tumor promoters. In order to identify which of the specific epicatechin derivatives present in GTP is responsible for these inhibitory effects, they were isolated from GTP and evaluated for their inhibitory effects against TPA-caused induction of epidermal ODC activity. Among these, (−)-epigallocatechin-3-gallate (EGCG), which was the major constituent present in GTP by weight, exerted the maximum inhibition. EGCG also showed greater inhibitory effects against TPA-caused induction of epidermal ODC activity when compared with several other naturally occurring polyphenols. The results of this study suggest that GTP, specifically its epicatechin derivative EGCG, could provide anti-tumor-promoting effects against a wide spectrum of skin tumor promoters.

MATERIALS AND METHODS

Chemicals. TPA, mezerein, (−)-indolactam V, H2O2, anthralin, α-dodecanec, d-lornithine, nordihydroguaiaretic acid, quercetin, α-GA, β-GA, and EC were purchased from Sigma Chemical Co. (St. Louis, MO). Benzoyl peroxide, tert-butylperoxide, tannic acid, and ellagic acid were obtained from Aldrich Chemical Co. (Milwaukee, WI). Benzoyl peroxide was dissolved immediately before use in ethanol. TPA, mezerein, (−)-indolactam V, and H2O2 were dissolved in ethanol and stored at −20°C. None of the solvents was used in concentrations higher than 1% by volume. The abbreviations used are: GTP, polyphenolic fraction isolated from green tea; TPA, 12-O-tetradecanoylphorbol-13-acetate; H2O2, hydrogen peroxide; benzoyl peroxide, and EC, (−)-epicatechin; EGC, (−)-epicatechin-3-gallate; ODC, ornithine decarboxylase; α-GA and β-GA, 18α- and 18β-glycyrhizic acid.

Received 1/9/92; accepted 4/24/92.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

1Supported by USPHS Grants ES-1900 and P-30-AR-39750, American Institute for Cancer Research Grant 90A47, and research funds from the Department of Veterans Affairs.

2R. A. is a recipient of a Dermatology Foundation Research Grant Award.

3To whom requests for reprints should be addressed, at the Department of Veterans Affairs Medical Center, 10701 East Boulevard, Cleveland, OH 44106.
were purchased from Aldrich Chemical Co. (Milwaukee, WI). d,L-[³⁵]C]Ornithine (58 Ci/mmol) was from Amersham Searle (Chicago, IL). All other chemicals were obtained in the purest form commercially available.

Preparation of GTP. GTP from green tea leaves (a product of Korea; distributed by Hanni, Inc., Los Angeles, CA) was prepared by the method described earlier (19, 20) with some modifications. In brief, dried green tea leaves (100 g) were suspended and extracted twice with hot water (80°C) and thrice with 80% ethanol (700 ml each time) under nitrogen. The total combined extract (3.5 liters) was concentrated under vacuum. The residue obtained was dissolved in water (50 ml) and freeze-dried. The light-brown solid matter obtained was called GTP.

HPLC Analysis of GTP. The HPLC system comprised two Waters model 510 pumps and a model 680 automatic gradient controller. A Waters 4-μm Nova Pak C-18 column (3.9 mm x 15 cm) was used along with a sample loop of 2 ml capacity. The absorbance of the eluate was monitored at 280 nm using a Shimadzu SPD-6AV UV-visible spectrophotometer and recorded on a Shimadzu C-R3A Chromatopack integrator. The mobile phase was a mixture of water (solvent A) and methanol (solvent B). For HPLC analysis, GTP was dissolved in 10° aqueous methanol (2 mg/ml, w/v) and small aliquots (50–100 μl) were subjected to reverse-phase HPLC using a Waters model WISP 710B autoinjector. The gradient system used in terms of percentage of solvent B was: 0 min, 10%; 0–30 min, a linear increase to 30%; 30–45 min, 30% isocratic; and beyond 45 min, 100%. The individual polyphenolic constituents were eluted under these conditions at a uniform flow rate of 1 ml/min throughout the HPLC run. Identification of the individual epicatechin derivatives was based on the comparison of the retention times of unknown peaks to those of reference standards either obtained commercially (EC) or provided by Dr. R. Saijo (Vegetable and Ornamental Corp., Research Station, Japan) and Dr. Matsuaki (Food Research Laboratories, Mitsui Nourin Co., Japan) (EGC, EGCg, ECG).

Animals and Treatment. Six-week-old female SENCAR mice were obtained from the National Cancer Institute-Frederick Cancer Research Facility (Frederick, MD). SENCAR mice were used in the present study because they are the most studied mouse strain for the causation, mechanism, and chemoprevention of cutaneous carcinogenesis (Refs. 6 and 22 and references therein). Additionally, this mouse strain respond because they are the most studied mouse strain for the causation, mechanism, and chemoprevention of cutaneous carcinogenesis (Refs. 6 and 22 and references therein). Additionally, this mouse strain respond because they are the most studied mouse strain for the causation, mechanism, and chemoprevention of cutaneous carcinogenesis (Refs. 6 and 22 and references therein). Additionally, this mouse strain respond because they are the most studied mouse strain for the causation, mechanism, and chemoprevention of cutaneous carcinogenesis (Refs. 6 and 22 and references therein). Additionally, this mouse strain respond because they are the most studied mouse strain for the causation, mechanism, and chemoprevention of cutaneous carcinogenesis (Refs. 6 and 22 and references therein).

The animals were shaved with electric clippers, and Nair depilatory was applied 1 day prior to the experiments. Animals were divided into several groups of eight animals each. All treatments were done topically on the shaved areas, which later were used for cytosol preparation and ODC activity determination. In the control group, the animals received a single topical application of 0.2 ml acetone only. In experimental groups, the animals were treated either with a single topical application of known skin tumor promoters in 0.2 ml acetone only or with GTP or individual epicatechin derivatives isolated from GTP in 0.2 ml acetone 30 min prior to the skin application of tumor promoters in 0.2 ml acetone. The details of treatment protocols of various skin tumor promoters, GTP, and/or individual epicatechin derivatives in relation to their doses, time of application, and killing of animals after treatments are provided at appropriate places in "Results." The animals were killed by cervical dislocation, and the dorsal treated skin was removed and made free of connective tissue and fat. The epidermis was separated from the whole skin by brief heat treatment at 52°C for 30 s, and the epidermal 100,000 x g supernatant fraction was prepared as described earlier (23).

Assay of ODC Activity. ODC activity was determined utilizing 0.4 ml of epidermal 100,000 x g supernatant fraction per assay tube by measuring the release of ¹⁴C]CO₂ from the d,L-[¹⁴]C]Ornithine by the method of O'Brien et al. (24) as described by Verma et al. (25). The details of the assay procedure are described earlier (26). Enzyme activity is expressed as pmol CO₂ released/hr/mg protein. Protein concentration was determined by the method of Bradford (27).

Statistical Analysis. The statistical significance of the difference between TPA and GTP plus TPA groups, and TPA and naturally occurring polyphenol plus TPA were evaluated by Student’s t-test.

RESULTS

HPLC Analysis of GTP and Stability of Individual Epicatechin Derivatives. Under the procedure used for the preparation of GTP, the total yield of the light-brown solid matter was found to be 7.5% (w/w) of the dry green tea leaves used in the present study. As shown in Fig. 1, when subjected to HPLC, GTP separated into six sharp peaks, of which five were identified as ECG, caffeine, EGCg, ECG, and ECG. As shown in Table 1, epicatechin derivatives accounted for 68.5% (w/w) of the total GTP injected into the column, showing that the GTP preparation used in the present study was constituted maximally of epicatechin derivatives. Furthermore, in this GTP preparation, EGCg was the major constituent (42% of total GTP, w/w) and represented 3.15% (w/w) of the dry weight of green tea leaves (Table 1). As observed by HPLC analysis, the individual epicatechin derivatives present in GTP were highly stable at 4°C in the powder form for about 1 year (data not shown).

Inhibitory Effect of GTP on TPA-induced Epidermal ODC Activity. In order to determine the optimal effective dose and the 50% inhibitory dose of GTP against TPA-induced epidermal ODC activity in SENCAR mice, various groups of animals were treated topically with varying doses of GTP (0.5–24 mg GTP in 0.2 ml acetone/animal) 30 min prior to the topical application of TPA (5 μg in 0.2 ml acetone/animal). As shown in Fig. 2, pretreatment of animals with GTP resulted in a dose-dependent inhibition of TPA-induced epidermal ODC activity. At the highest dose of GTP (24 mg), as much as 90% inhibition was observed (Fig. 2). Similarly, at small doses GTP resulted in significant inhibition. The effective dose of GTP resulting in 50% inhibition of the levels of epidermal ODC activity induced by the topical application of 5 μg TPA was found to be 1.9 mg/animal (Fig. 2). GTP alone up to 24 mg in 0.2 ml acetone does not cause any induction of epidermal ODC activity (data not shown).

Based on the data shown in Fig. 2, a dose of 2 mg GTP, which produced a 55–60% inhibition of TPA-induced epidermal ODC activity, was selected for further experiments.

Fig. 1. Reverse-phase HPLC of GTP on a Waters 4-μm Nova Pak C-18 column. In a typical HPLC elution profile shown here, 200 μg GTP (2 mg/ml in 10° aqueous methanol, w/v) was subjected to HPLC analysis under the conditions described in "Materials and Methods." The identification of EGC, caffeine, EGCg, ECG, and ECG is based on the reference standards.

3583

Downloaded from cancerres.aacrjournals.org on April 14, 2017. © 1992 American Association for Cancer Research.
order to determine the time of application of GTP in relation to the topical application of TPA for its maximum inhibitory effect, 2 mg of GTP in 0.2 ml acetone were applied at various time intervals before or after the skin application of 5 μg TPA in 0.2 ml acetone. As shown in Fig. 3, the inhibitory effect of GTP was dependent on the time of its application in relation to TPA application. The maximum inhibition was observed when GTP was applied 30 min prior to TPA application. Comparable results were obtained when GTP was applied 15–25 min prior to TPA application (data not shown). Although not that profound, the topical application of GTP between 3 h before and 2 h after TPA application was also significantly effective in inhibiting the TPA-caused induction of epidermal ODC activity (Fig. 3).

The effect of the preapplication of GTP on TPA-induced epidermal ODC activity was also studied as a function of time after the TPA application. As shown in Fig. 4, consistent with published studies (28, 29), the maximum induction of epidermal ODC activity after the single topical application of TPA was observed at 6 h, which started declining after that period and was almost at the basal level by 14 h. When GTP (2 mg in 0.2 ml acetone) was applied 30 min prior to topical application of TPA, significant inhibition was observed at all the time points studied, in a pattern similar to that observed with TPA alone (Fig. 4).

### Table 1. Qualitative and Quantitative Analysis of GTP in Relation to Epicatechin Derivatives by Reverse-Phase HPLC

<table>
<thead>
<tr>
<th>Percentage of Total GTP (w/w)</th>
<th>Percentage of Dry Green Tea Leaves (w/w)</th>
<th>Percentage of Total Epicatechin Derivatives (w/w)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total Epicatechin Derivatives</td>
<td>68.5</td>
<td>100</td>
</tr>
<tr>
<td>EGC</td>
<td>3.7</td>
<td>0.28</td>
</tr>
<tr>
<td>ECG</td>
<td>42.0</td>
<td>3.15</td>
</tr>
<tr>
<td>EC</td>
<td>4.0</td>
<td>0.3</td>
</tr>
<tr>
<td>ECG</td>
<td>18.8</td>
<td>1.41</td>
</tr>
</tbody>
</table>

*a GTP (100-200 μg) was subjected to reverse-phase HPLC on a Waters 4-μm Nova Pak C-18 column under the conditions detailed in "Materials and Methods." The quantitative data shown here in this table in relation to total epicatechin derivatives as well as individual epicatechin derivatives are based on the printout obtained by using a Shimadzu C-R3A chromatopak integrating recorder programmed for the area normalization method.

*b Calculations are based on the total percentage of GTP (7.5%, w/w) in dried green tea leaves.

*c Calculations are based on considering the total epicatechin derivatives as 100% GTP.

Fig. 2. Dose-dependent inhibition of TPA-induced epidermal ODC activity in SENCAR mice by GTP. Eight animals in each group were treated topically either with 0.2 ml acetone or with the indicated doses of GTP in 0.2 ml acetone 30 min prior to the application of 5 μg TPA in 0.2 ml acetone. Animals were killed 6 h after the TPA treatment by cervical dislocation, and the ODC activity was determined as described in "Materials and Methods." Each data point represents the mean ± SE of four individual values; the epidermis from two animals were pooled for each determination. The basal ODC activity in control animals receiving acetone alone was 98 ± 12 pmol CO₂ released/h/mg protein. The application of 5 μg TPA alone caused the induction of epidermal ODC activity to the level of 2046 ± 162 pmol CO₂ released/h/mg protein. For statistical analysis of the data, Student's t test was applied between TPA alone and GTP plus TPA groups. *P < 0.005 in case of TPA alone versus 1 to 24 mg GTP dosage plus TPA.
Inhibitory Effect of GTP on Epidermal ODC Induction Caused by Structurally Different Mouse Skin Tumor Promoters. In addition to its inhibitory effect against TPA-induced epidermal ODC, GTP was also evaluated for its inhibitory effects against epidermal ODC induced by several structurally different mouse skin tumor promoters. The doses of various tumor promoters used in the present study were based on the published studies by us as well as by others (Refs. 26, 29, and 30 and references therein). As shown by the data in Table 2, the topical application of GTP (2 mg in 0.2 ml acetone/animal) 30 min prior to the topical application of these tumor-promoting agents resulted in 45 to 61% inhibition of epidermal ODC induction caused by these agents; however, a maximal inhibitory effect was observed in the case of TPA, although it was comparable to that with H$_2$O$_2$ and benzoyl peroxide (Table 2).

Inhibitory Effects of Individual Epicatechin Derivatives Isolated from GTP on TPA-Induced Epidermal ODC. To identify which of the specific epicatechin derivatives present in GTP is involved in exerting the inhibitory effect against TPA-induced epidermal ODC activity, we evaluated the inhibitory effects of individual epicatechin derivatives isolated from GTP. For the isolation of individual epicatechin derivatives, GTP was subjected to HPLC under the conditions described in “Materials and Methods.” Each peak for EGC, EGCG, EC, and ECG was collected manually, lyophilized, and dissolved in acetone. As in the experiments utilizing GTP, the quantity of each of the epicatechin derivatives present in 2 mg GTP was applied topically 30 min prior to the application of TPA, and ODC activity was determined under similar experimental conditions. As shown by the data in Table 3, maximum inhibition (63%) was found to be associated with EGCG, the major constituent present in GTP (Table 1), followed by EGC (43%), ECG (38%), and EC (17%), as compared to 49% inhibition observed with GTP. Furthermore, in order to determine (a) if there is any loss of inhibitory effect of GTP during its fractionation on HPLC, and (b) that the inhibitory effect of GTP is only due to the epicatechin derivatives which constitute 68.5% of total GTP, all four epicatechin derivative fractions isolated from GTP (EGC, EGCG, EC, and ECG) were combined together in the same ratios in which they occur in unfractionated GTP, and this reconstituted GTP preparation was evaluated for its inhibitory effect against TPA-induced epidermal ODC activity. As shown by the data in Table 3, the reconstituted GTP preparation showed inhibition almost comparable to that observed with the unfractionated GTP preparation (51 and 49%, respectively).

Since one of the aims of the present study was to identify which of the epicatechin derivatives present in GTP is the most effective inhibitor of TPA-induced epidermal ODC activity, and since the data summarized in Table 3 are based on the quantity of each epicatechin derivative present in 2 mg GTP, we also evaluated the inhibitory effects of the topical application of equal quantities of individual epicatechin derivatives, on a molar basis, on TPA-induced epidermal ODC activity. As shown by the data in Table 4, the topical application of 4 μmol of EGC, EGCG, EC, or ECG in 0.2 ml acetone/animal 30 min prior to the topical application of 5 μg TPA in 0.2 ml acetone resulted in the significant inhibition of TPA-induced epidermal ODC activity, with a significant inhibition of 45% compared to TPA alone.

### Table 2: Inhibitory Effect of GTP on Epidermal ODC Induction Caused by Structurally Different Skin Tumor Promoters in SENCAR Mice

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose of tumor promoter</th>
<th>ODC activity</th>
<th>% Inhibition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acetone</td>
<td>77 ± 9</td>
<td></td>
<td></td>
</tr>
<tr>
<td>TPA</td>
<td>5 μg</td>
<td>2059 ± 160</td>
<td>61</td>
</tr>
<tr>
<td>GTP + TPA</td>
<td></td>
<td>795 ± 53%</td>
<td></td>
</tr>
<tr>
<td>Mezerein</td>
<td>5 μg</td>
<td>2872 ± 255</td>
<td>52</td>
</tr>
<tr>
<td>GTP + Mezerein</td>
<td></td>
<td>1378 ± 101</td>
<td></td>
</tr>
<tr>
<td>(−)Indolactam V</td>
<td></td>
<td>1140 ± 107</td>
<td>47</td>
</tr>
<tr>
<td>GTP + (−)Indolactam V</td>
<td></td>
<td>604 ± 38%</td>
<td></td>
</tr>
<tr>
<td>Anthratin</td>
<td>50 μg</td>
<td>498 ± 45</td>
<td>40</td>
</tr>
<tr>
<td>GTP + Anthratin</td>
<td></td>
<td>299 ± 25%</td>
<td></td>
</tr>
<tr>
<td>H$_2$O$_2$</td>
<td>5 mg</td>
<td>387 ± 31</td>
<td>58</td>
</tr>
<tr>
<td>GTP + H$_2$O$_2$</td>
<td></td>
<td>163 ± 21%</td>
<td></td>
</tr>
<tr>
<td>BPO$^\dagger$</td>
<td>20 mg</td>
<td>776 ± 49</td>
<td>59</td>
</tr>
<tr>
<td>GTP + BPO</td>
<td></td>
<td>318 ± 29%</td>
<td></td>
</tr>
<tr>
<td>t-BuPO$^\dagger$</td>
<td>20 mg</td>
<td>118 ± 12</td>
<td>52</td>
</tr>
<tr>
<td>GTP + t-BuPO</td>
<td></td>
<td>57 ± 10%</td>
<td></td>
</tr>
<tr>
<td>n-Dodecane</td>
<td>25 mg</td>
<td>739 ± 69</td>
<td>45</td>
</tr>
<tr>
<td>GTP + n-Dodecane</td>
<td></td>
<td>407 ± 35%</td>
<td></td>
</tr>
</tbody>
</table>

$^\dagger$ Eight animals in each group were treated topically either with 0.2 ml acetone or with 2 mg GTP in 0.2 ml acetone/animal 30 min prior to the skin application of indicated doses of tumor promoters in 0.2 ml acetone. In case of benzoyl peroxide and tert-butylperoxide, animals were killed 24 h after the treatment, whereas in rest of the cases animals were killed 6 h after the treatment by cervical dislocation. The ODC activity was determined in epidermal 100,000 × g supernatant fraction.

$^\ddagger$ Each value represents the mean ± SE of four individual values; the epidermis from two animals were pooled for each determination. For statistical analysis of the data, Student’s $t$ test was applied between tumor promoter and GTP plus tumor promoter groups.

$^\ddagger\ddagger$ Highly significant versus tumor promoter alone, $P < 0.0005$.

$^\ddagger\ddagger\ddagger$ Significant versus tumor promoter alone, $P < 0.005$.

$^\ddagger\ddagger\ddagger\ddagger$ Significant versus tumor promoter alone, $P < 0.01$.

BPO, benzoyl peroxide; t-BuPO, tert-butylperoxide.
ODC activity. However, even on a molar basis, the maximum inhibitory effect was associated with EGCG (Table 4).

Since in a number of studies from this laboratory (13, 26, 31) as well as by other investigators (Refs. 5, 29, and 32 and references therein), various naturally occurring polyphenols have been shown to be inhibitors of TPA-induced epidermal ODC activity and mouse skin tumor promotion, and since in the present study we found EGCG to be a potent inhibitor of TPA-induced epidermal ODC activity, we also evaluated the comparative inhibitory effects of various known naturally occurring polyphenols with EGCG. As shown by the data in Table 4, of all the polyphenols evaluated, EGCG showed the maximum inhibitory effect against TPA-induced epidermal ODC activity. At a dose of 4 µmol EGCG, an inhibition as high as 90% was observed, whereas EGC, EC, ECG, nordihydroguaiaretic acid, quercetin, tannic acid, ellagic acid, α-GA, and β-GA produced only 55–70% inhibition at the same molar dose (Table 4). However, application of any of the naturally occurring polyphenols alone does not induce epidermal ODC activity (data not shown).

DISCUSSION

Chemical carcinogenesis in murine skin is a stepwise process of at least three distinct stages: initiation; promotion; and progression (6, 22). Studies in murine skin have shown that even a lapse of up to 1 year between the application of initiator and the beginning of promoter treatment produces a tumor response similar to that observed when promotion is begun only 1 week after initiation, with the carcinogen administered as in a classical two-stage skin carcinogenesis protocol (6, 22). In addition, initiated cell populations in animals may persist throughout their lifetime but may not result in visible tumors in the absence of further treatment, indicating that initiation alone is not sufficient to elicit a tumor response. These studies have aroused an interest in defining the mechanism of tumor promotion and in finding new agents capable of affording protection against tumor promotion.

The morphological, biochemical, and physiological changes occurring in mouse skin as a result of the topical application of phorbol esters like TPA are reviewed extensively (Ref. 22 and references therein). The induction of ODC activity followed by an increase in the levels of polyamines, the induction of dark periplasia following multiple application, the ability to induce reactive oxygen species formation, and the interaction with and activation of specific receptor protein kinase C are the effects best studied in either mouse skin or keratinocytes following TPA treatment (Ref. 22 and references therein). Of all these effects, the induction of ODC activity is considered to be closely associated with, although not sufficient to produce the tumor-promoting activity of a variety of tumor promoters (6, 22). ODC is involved in polyamine biosynthesis, which plays an essential role in cell proliferation and differentiation (33). The importance of the induction of epidermal ODC in skin tumor promotion is evident from the fact that several inhibitors of ODC are capable of inhibiting tumor promotion in murine skin (5, 22, 29). For example, a variety of compounds such as n-propyl gallate, flavonoids, retinoids, and antioxidants which inhibit ODC induction have also been found to be effective inhibitors of TPA-mediated tumor promotion (5, 29, 34). As in to these reports, in the present study topical application of GTP prior to that of TPA resulted in the significant dose-
INHIBITION OF ODC BY EPICATECHINS IN GREEN TEA

dependent inhibition of TPA-induced epidermal ODC (Fig. 2). Since the effect of GTP application was more profound when it was applied prior to TPA as compared to the effect observed after TPA application (Fig. 3), it is reasonable to assume that GTP application inhibited the action of tumor promoters and/or the enzymatic pathway(s) which regulates the ODC induction rather than interacting directly with the enzyme. Furthermore, the inhibition of TPA-induced epidermal ODC by GTP at each time point studied (Fig. 4) suggests that GTP does not delay the peak of TPA-induced ODC.

The results of the present study showing the inhibitory effect of GTP on epidermal ODC induction (Table 2) suggest that GTP may be of significant use in protecting against several tumor promoters with diversified mechanisms of action (Refs. 5, 6, 22, 29, 30, 35-42 and references therein). In fact, in our ongoing experiments, when assessed in an anti-tumor promotion protocol in 7,12-dimethylbenz[a]anthraenc-initiated SENCAR mouse skin, the pretreatment of animals topically with GTP before each skin application of TPA resulted in significant protection in terms of total number of tumors per mouse as well as percentage of mice with tumors when compared to non-GTP-pretreated animals. A preliminary report by Huang et al. (43), however, has also shown the inhibitory effects of GTP against TPA-caused epidermal ODC induction and skin tumor promotion in CD-1 mice.

In recent years, we and others have shown that green tea leaves are rich in epicatechin derivatives (17, 18), specifically EGCG (21, 44), and suggested that the antimutagenic (15, 16, 49-51), and antibacterial (52, 53) activities of water extracts of green tea and/or GTP may primarily be due to these epicatechin derivatives. In the present study when analyzed on HPLC, GTP separated into six sharp peaks, of which four were identified as EGC, EGCG, EC, and ECG, the epicatechin derivatives (Fig. 1), accounting for 68.5% of total GTP (Table 1). When these epicatechin derivatives were assayed individually against the TPA-caused induction of epidermal ODC activity, both on the basis of the quantity of each epicatechin derivative present in GTP (Table 3) and on a molar basis (Table 4), EGCG was found to be the most effective inhibitor and constitutes the major part of the GTP (42%, w/w; Table 1). Similarly, when compared with other naturally occurring polyphenols with known inhibitory effects against TPA-induced epidermal ODC and skin tumor promotion (Refs. 5, 13, 26, 29, 31, 32 and references therein), EGCG was most effective when tested at an equimolar dosage (Table 4). Based on the chemical structure of EGCG, it seems reasonable that the gallyl and gallate moieties at positions 2 and 3, respectively, in EGCG make it highly active against various inhibitory effects. Finally, since the inhibitory effect observed with reconstituted GTP (Table 3), obtained by mixing only the four epicatechin derivative fractions and not the unidentified peak and the caffeine fractions (Fig. 1), were comparable with that of unfractionated GTP, it seems reasonable to conclude that the inhibitory effects of GTP observed in the present study may be due to the epicatechin derivatives present therein.

Chemoprevention is a means of cancer control in which disease occurrence is prevented by the administration of one of a combination of several chemical agents (2, 3, 54-58). One of the exciting findings in this area of research is the identification of a large number of compounds with great structural diversity, which have shown promise in several animal bioassay systems (2, 3, 32, 54-58). One of the most impressive findings in this field is the presence of these compounds in the human diet, which makes it possible that simple changes in dietary habits could significantly lower cancer risk (2, 54-58). The results of present study, in conjunction with prior publications from our laboratory and others, suggest that GTP may prove to be a useful chemopreventive agent against some forms of human cancers induced by environmental agents and that more detailed studies on EGCG could lead to new strategies for recommending changes in dietary habits and for developing other foods with desired cancer chemopreventive activities.

REFERENCES


Downloaded from cancerres.aacrjournals.org on April 14, 2017. © 1992 American Association for Cancer Research.


Inhibition of Skin Tumor Promoter-caused Induction of Epidermal Ornithine Decarboxylase in SENCAR Mice by Polyphenolic Fraction Isolated from Green Tea and Its Individual Epicatechin Derivatives


Updated version Access the most recent version of this article at:
http://cancerres.aacrjournals.org/content/52/13/3582