Advances in Brief

Vaccinations with Tumor Cells Genetically Engineered to Produce Different Cytokines: Effectivity not Superior to a Classical Adjuvant

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Abstract

The potential of tumor cells (J558L) engineered to produce one of 5 different cytokines (interleukin 2, interleukin 4, interleukin 7, tumor necrosis factor, or γ-interferon) to produce rise to systemic immunity protective against a contralateral challenge with the parental cells was analyzed. The rejection of all cytokine-producing cells appeared to induce some systemic response capable of mediating the rejection of low numbers of subsequently contralaterally injected cells, but the effect was much less obvious with higher cell numbers. The injection of any possible combination of two of the cytokine producers did not reveal any synergistic effects. The cytokine gene-transfected tumor cells were not superior to the parental cells admixed with the adjuvant Corynebacterium parvum with respect to their potential as immunogens to induce immunity.

Introduction

Attempts to vaccinate against cancer cells have not been rewarded with decisive success in clinical trials so far, even though considerable efforts have been taken to apply and improve various vaccination strategies to the benefit of cancer patients (1). The reasons for the apparent failure are not completely clear, especially since it was established on the basis of very promising results from experimental systems (2) and clinic trials (3) that cytokines (e.g. interleukin-2, -4, -7, tumor necrosis factor, interferon-γ, interferon-β, interferon-α) may induce or augment antitumor responses. However, the achievement of such antigen-specific antitumor responses is not straightforward and may depend on various factors (4). The cost of publication of this article was defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Results

The cell lines J558-IL-2, -IL-4, -IL-7, -TNF, and -IFN, which are all derived from parental J558L cells and produce the respective cytokine after gene transfer, were used in this study (Table 1). Due to secretion of the transfected cytokine their growth is strongly suppressed in vivo but not affected in vitro (15–18). We compared the effect of prior injection of any of the cytokine producers or irradiated parental cells on the tumor growth of a subsequent contralateral challenge with J558L cells. Also, we searched for synergism by mixing two cytokine producers in all possible combinations. Fig. 1 summarizes the results of 4 independent experiments: prior injection of irradiated cells did not lead to a significant change in tumor incidence when the animals were challenged with 2 × 10^6 after 2 weeks (Fig. 1d) or 4 × 10^6 after 4 weeks (Fig. 1, a and b), even if a slight delay in tumor growth was observed in comparison to animals which had never encountered this tumor before (not shown). In contrast, some of the mice which had been previously injected with cytokine producers were tumor free or only transiently developed a tumor. However, there was considerable variation in the protective effect induced by each cytokine in different experiments. No cytokine producer or any combination led to the reproducible protection of more than 2 of 5 animals, regardless of whether the challenge was introduced after 2 or after 4 weeks. Although in one experiment (Fig. 1b) both J558-IL-7 or the combination of J558-IL-4 and TNF resulted in statistically significant protection (P < 0.05, Fisher's exact test) in comparison to the control with irradiated cells, this was not observed in an independent parallel experiment under the same conditions (Fig. 1a), nor did the effect occur when animals were challenged after 2 weeks (Fig. 1, c and d). Thus, none of the cytokine producers or their permuted combination appeared to be strikingly superior in comparison to the others regarding the capacity to generate a systemic antitumor response. Given the variation of tumor incidence, however, more subtle differences would need much larger groups in each experiment to become statistically apparent.

Materials and Methods

Cell Lines. The mouse plasmacytoma cell line J558L is syngeneic in BALB/c mice and is a heavy chain loss variant of the plasmacytoma J558 (14). Culturing of J558L cells (15) and generation of all J558L sublines [J558-IL-2, -IL-4, -IL-7, tumor necrosis factor or interferon-γ (IFN-γ)] has been described; a more detailed characterization of the cell line is described elsewhere. Each subline produces the respective cytokine after gene transfer. The biological activities were determined by proliferation assays with CTLL-2 (for IL-2), CT-45 cells (for IL-4), and thymocytes (for IL-7); by cytotoxicity on L929 cells (for TNF); and by major histocompatibility complex class II expression on WEHI.13 cells (for IFN-γ) as described. One unit was defined as the reciprocal of dilution of conditioned medium required for 50% proliferation/cytotoxicity or in ng/ml as determined for IFN-γ by a commercially available enzyme-linked immunosorbent assay (1).
VACCINATION WITH CYTOKINE-PRODUCING TUMOR CELLS

Fig. 1. Systemic protection induced by injection of single-cytokine producers or combinations of cytokine-producing cells. Mice were immunized by injection of $4 \times 10^6$ cytokine-producing or irradiated parental J558L cells. When combinations of cytokines were tested, $2 \times 10^6$ of each cytokine producer were mixed before injection. The number of mice with tumor per Five mice in the experimentis given. N.D., not done; *, 10 mice were challenged.

Table 1 Cytokine-producing J558L sublines

<table>
<thead>
<tr>
<th>Cell line</th>
<th>Vector</th>
<th>Cytokine production</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>J558-IL2</td>
<td>pLTR-IL2</td>
<td>40 units/ml</td>
<td>footnote 4</td>
</tr>
<tr>
<td>J558-IL4</td>
<td>pXEP-IL4</td>
<td>75 units/ml</td>
<td>16</td>
</tr>
<tr>
<td>J558-IL7</td>
<td>pBA-IL7</td>
<td>50 units/ml</td>
<td>17</td>
</tr>
<tr>
<td>J558-TNF</td>
<td>pBA-TNF</td>
<td>17 units/ml</td>
<td>18</td>
</tr>
<tr>
<td>J558-IFN</td>
<td>pLTR-IFN</td>
<td>120 ng/ml</td>
<td>footnote 4</td>
</tr>
</tbody>
</table>

In view of the rather moderate systemic effects obtained by prior injection of cytokine producers, a comparison with the well-established adjuvant C. parvum (13) was performed. The growth of up to $10^6$ parental J558L cells when administered locally with an admixture of 100 µg C. parvum was completely inhibited, but when higher numbers of tumor cells were administered the effect of C. parvum became incomplete (Fig. 2a). When mice which had been given injections of either irradiated J558L cells or of the same number of irradiated cells mixed with C. parvum were contralaterally challenged 18 days later with $2 \times 10^6$ J558L cells, no effect could be observed (Fig. 2b). However, when mice which had survived the injection of $4 \times 10^6$ viable J558L cells mixed with C. parvum were challenged with $2 \times 10^6$ J558L cells 4 of 5 mice remained tumor free (Fig. 2b).

Finally, we wanted to directly compare the protective effects induced by C. parvum admixture and the cytokine-producing cells. Therefore, mice were injected with $4 \times 10^6$ cytokine-producing or parental J558L cells admixed with C. parvum. In order to be able to more sensitively assess differences in the strength of the systemic response, groups of mice were challenged with escalating cell doses. Fig. 3 shows that most mice develop a tumor after prior injection of the cytokine-producing cells, when challenged with $>1 \times 10^6$ tumor cells, whereas mice challenged with smaller cell numbers were obviously better protected. In contrast, prior injection of J558L cells mixed with C. parvum induced significantly better protection than irradiated cells ($P < 0.05$, Fisher’s exact test), even when animals were challenged with $2 \times 10^6$ or $4 \times 10^6$ J558L cells. The statistical analysis of contingency tables using the $\chi^2$ test revealed that the potencies of immunization with different cytokine producers did not differ from each other when animals challenged with all different doses were included in the analysis. However, the mixture of tumor cells and C. parvum was significantly superior ($P < 0.01$) to cytokine-producing tumor cells with respect to the generation of a systemic protective immune response.
VACCINATION WITH CYTOKINE-PRODUCING TUMOR CELLS

<table>
<thead>
<tr>
<th>challenge dose</th>
<th>Tumor incidence at challenge site in % (n=5-9)</th>
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<tbody>
<tr>
<td>J558L</td>
<td></td>
</tr>
<tr>
<td>4 x 10⁶</td>
<td></td>
</tr>
<tr>
<td>2 x 10⁶</td>
<td></td>
</tr>
<tr>
<td>1 x 10⁶</td>
<td></td>
</tr>
<tr>
<td>0.5 x 10⁶</td>
<td></td>
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<tr>
<td>0.25 x 10⁶</td>
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![Fig. 3. Comparison of the immunogenic potency of cytokine-producing J558L variants versus parental cells admixed with C. parvum](image)

Discussion

The potential to generate systemic immunity has been observed for tumor cells transplanted with any of the 5 cytokines dealt with in this study (5–12). Therefore, we were surprised to find our cytokine-producing cell lines to be immunogenic which were only slightly superior to irradiated cells not producing the cytokines. However, we do not think our data are necessarily in contrast to those of others with different tumor cell lines. The studies so far describing the generation of systemic immunity did not conclusively determine if this phenomenon was necessarily due to the secreted cytokines or might be explained solely by the exposure of viable tumor cells to the immune system during the primary rejection. Irradiated or mitomycin C-treated cells as controls were not included in those experiments (5–12). It should be noted, however, that in many cases well-characterized cell lines were used which are usually considered to be non-immunogenic. But even in this context the omission of irradiated cells and cytokines in the anti-tumor effect of interleukin 4. Science (Washington DC), 254: 713–716, 1991.

References

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