Hormone-dependent Regulation of BRCA1 in Human Breast Cancer Cells

Jean M. Gudas,1 Hoang Nguyen,2 Tao Li,2 and Kenneth H. Cowan

Medicine Branch, Division of Cancer Treatment, Medical Breast Cancer Section, National Cancer Institute, Bethesda, Maryland 20892

Abstract

BRCA1 mRNA and protein levels are regulated by the steroid hormones estrogen and progesterone in human breast cancer cells. BRCA1 mRNA and protein levels were significantly decreased in estrogen-depleted MCF-7 and BT20T cells and increased again after stimulation with β-estradiol. The increase in BRCA1 expression upon stimulation with estrogen was not coordinated with the early induction of the estrogen-dependent p52 gene but closely paralleled the delayed increase in the S-phase dependent marker cyclin A. T47-D cells deprived of steroid hormones and subsequently stimulated with progesterone also showed a delayed increase in BRCA1 mRNA expression. However, no change in BRCA1 protein was detected in these cells. When considered together, the data suggest that steroid hormones may affect BRCA1 expression indirectly by altering the proliferative status of the cells rather than acting directly on DNA sequences in the BRCA1 gene itself.

Introduction

Breast cancer is the most common malignancy among women in North America, with the number of newly diagnosed cases increasing at a rate of about 1% a year (1). As one approach toward understanding the genetic basis of this disease, Miki et al. (2) have recently reported the cDNA sequence of a gene on chromosome 17q that that confers susceptibility to early onset familial breast and ovarian cancers. Analysis of the BRCA1 gene in familial and sporadic breast tumors has suggested that mutation is not a common event in sporadic cases of breast cancer; however, a recent report has indicated that somatic mutations in the BRCA1 gene may play a role in a subset of sporadic ovarian cancers (3). In addition to genetic factors, hormones of pituitary and ovarian origin, including estrogens, progesterins, and prolactin, all play a role in the control of normal and breast tumor cell proliferation (reviewed in Refs. 4 and 5). The response to polypeptide growth factor and steroid hormones is abrogated as normal mammary epithelial cells progress to gain a malignant phenotype (6). Since estrogens and progesterins can influence both the growth and differentiation of normal and breast cancer cells, we examined whether the breast cancer susceptibility gene BRCA1 was regulated in response to these hormonal signals.

Materials and Methods

Cells and Cell Culture. MCF-7 cells were obtained from Dr. R. Buick (University of Toronto, Toronto, Ontario, Canada). The BT20T cells were a gift from Dr. K. Keyomarsi (New York State Department of Health, Albany, NY), and the T47-D cells were purchased from American Type Culture Collection. All cells were cultured in IMEM2 (Biofluids, Inc.) supplemented with 10% HEPES, 2 mM glutamine, 0.1 mM nonessential amino acids, 10% fetal bovine serum, and 2 μg/ml insulin. The cells were routinely screened for Mycoplasma contamination and maintained at 37°C in an atmosphere of 6.0% CO2. To study the effects of hormones on BRCA1 expression, the respective cells were plated at approximately 2 × 106 cells/150-mm tissue culture dish into phenol red-free IMEM (GIBCO) supplemented with 5% charcoal-stripped calf serum and incubated for 5 or 6 days. At this time the cells were refed with phenol red-containing IMEM supplemented with 5% charcoal-stripped calf serum and 10 nm β-estradiol or 10 nm progesterone (Sigma).

RNA Isolation and Northern Blot Analysis. Total RNA was prepared and analyzed as described previously (7). RNA (20 μg) was electrophoresed on denaturing formaldehyde gels, transferred to Magna NT membranes, and cross-linked with UV. A combination of two probes was used for Northern blot analyses. An 1179-bp fragment corresponding to cDNA sequences from 1025 to 2222 of BRCA1 was amplified by PCR and subcloned into the EcoRI and Smal sites of pGem7. A 624-bp PCR product corresponding to cDNA sequences from 4388 to 5011 was generously provided by Dr. Roger Wiseman (NIEHS). The authenticity of the subcloned probes was verified by DNA sequence analysis using standard techniques (Sequenase; U. S. Biochemicals). The 3684 probe was an 800-bp Prl fragment (8), and the cyclin A probe contained the entire cDNA (generously provided by Dr. Tony Hunter). All probes were labeled with [α-32P]dCTP to a specific activity of approximately 1 × 109 cpmdμg DNA using a random primed labeling kit (Boehringer Mannheim). The amount of radioactivity hybridized with specific bands on the RNA filters was quantitated using a Molecular Dynamics PhosphorImager.

Western Blot Analyses. Western blot analysis was performed as described previously (7). Protein lysate (100 μg/lane) was electrophoresed on SDS-polyacrylamide gels and transferred to nitrocellulose membranes. After blocking with Tris-buffered saline containing 8% dried milk and 0.1% Tween 20, filters were probed using antibodies to BRCA1 (2 μg/ml immunofinity-purified C20; Santa Cruz Biotechnology Inc.), cyclin A (4 μg/ml cyclin A antibody; Pharmingen), and actin (2 μg/ml actin antibody-1; Oncogene Science). Specific complexes were detected using a chemiluminescent technique according to the manufacturer’s recommendations (New England Nuclear). The X-ray films were captured with a Fotodyne digital image system and quantitated using the NIH Image Analysis software program.

Results

BRCA1 mRNA expression was examined in estrogen receptor-positive MCF-7 cells that were depleted of estrogens for 6 days and subsequently refed with phenol red-containing medium supplemented with charcoal-stripped calf serum and 10 nm β-estradiol for the times indicated (Fig. 1A). We have shown previously that steroid depletion results in a significant decrease in the growth of these MCF-7 cells (9). Northern blot analysis was performed using a combination of two probes that hybridize to distinct regions of the BRCA1 cDNA. Both of these probes recognize the major 8.1-kb transcript (designated M) while only the 624-bp 3’ probe hybridizes to the lower abundance 4.6-kb BRCA1 mRNA (designated m).4 BRCA1 mRNA levels were low in estrogen-deprived MCF-7 cells (time 0) and remained depressed for the first 8 h after feeding the cells with fresh medium containing β-estradiol. BRCA1 mRNA levels increased gradually 12 and 24 h after treatment. The kinetics of BRCA1 mRNA accumulation was strikingly similar to that observed for the S-phase-dependent marker cyclin A (Fig. 1A, second panel).

Received 7/27/95; accepted 8/27/95.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

1 To whom requests for reprints should be addressed, at Medicine Branch, Division of Cancer Treatment, National Cancer Institute, Building 10, Room 12N226, Bethesda, MD 20892.

2 These two authors contributed equally to this work.

3 The abbreviation used is: IMEM, improved MEM.

4 J. M. Gudas, T. Li., H. Nguyen, and H. Cowan, manuscript submitted for publication.
To compare the kinetics of BRCA1 mRNA accumulation with that of a gene containing a known estrogen-response element within its promoter, the same filters were stripped and reprobed for pS2 gene expression. pS2 mRNA levels were increased earlier (approximately 2–4 h after estrogen treatment) and rose to much higher levels than the proliferation-associated BRCA1 and cyclin A mRNAs. As a control for RNA loading and transfer, these and all subsequent filters were reprobed with the estrogen-independent marker 36B4 (10).

Using a peptide antibody that recognizes an epitope at the carboxyl terminus of the BRCA1 protein, we have demonstrated the presence of two BRCA1 proteins having approximate Mₜₛ 185,000 and 140,000, respectively, in most breast cancer cells. Examination of BRCA1 protein levels in samples that were treated and harvested in parallel revealed a decrease in estrogen-depleted MCF-7 cells (Fig. 1B). Digitization and quantitation of the X-ray films revealed an approximate 4-fold increase in the amount of the Mₜₛ 185,000 protein after estrogen treatment. Twenty-four h after estrogen addition, the amount of the Mₜₛ 185,000 BRCA1 protein was similar to that present in exponentially growing cells. In contrast, the Mₜₛ 140,000 BRCA1 protein did not increase until 24 h after estrogen stimulation. Thus, the rise in BRCA1 protein levels was coordinated with the rise in cyclin A protein levels (Fig. 1B, middle panel). The results observed were specifically due to the effects of estrogen, as MCF-7 cells depleted of estrogen and refiged with medium containing the antiestrogen tamoxifen showed no increase in either BRCA1 or cyclin A mRNA or protein levels (data not shown).

To determine whether the changes observed were specific for MCF-7 cells, the same experiments were performed using another estrogen-dependent cell line, BT20T. BRCA1 mRNA levels were significantly decreased in estrogen-depleted BT20T cells, but not to the same magnitude as observed in MCF-7 cells (Fig. 2A). Expression of both BRCA1 mRNAs increased approximately 8–12 h after re-stimulation with estrogen. The kinetics of BRCA1 mRNA accumulation was similar to that of the proliferation marker cyclin A. Both of these mRNAs were increased 12–24 h after estrogen stimulation. In contrast, the mRNA for the estrogen-inducible gene pS2 was maximally increased 2 h after refeeding with estrogen (Fig. 2A, third panel).

BRCA1 protein levels were lower in estrogen-depleted BT20T cells when compared with an exponentially growing population (Fig. 2B). The amounts of the Mₜₛ 140,000 and 185,000 BRCA1 proteins increased approximately 2.5-fold after estrogen stimulation for 24 h. Cyclin A protein levels were also decreased in the estrogen-depleted cells and did not increase significantly until 24 h after refeeding. When considered together, these data suggest that the increase in BRCA1 mRNA and protein levels after estrogen stimulation reflect a change in the proliferative status of the cells and may not be due to a direct effect of estrogen on DNA sequences in the BRCA1 gene.

T47-D breast carcinoma cells are dependent on progesterone and insulin for their growth in vitro (11). We examined BRCA1 mRNA expression in T47-D cells that were depleted of steroids and subsequently restimulated with medium containing progesterone (Fig. 3A).
The results show that BRCA1 mRNA was markedly decreased in steroid-depleted T47-D cells and increased significantly 8-12 h after progesterone stimulation. In these cells the increase in BRCA1 mRNA again correlated well with the respective increase in cyclin A mRNA levels (Fig. 3A, second panel). In contrast, the mRNA for a progesterone-regulated gene, fatty acid synthetase (12), was induced at earlier times (4 h after treatment), peaked by 12 h, and returned to basal levels 24 h after treatment.

Examination of BRCA1 protein levels in parallel cultures of T47-D cells revealed no change in the steroid-depleted versus progesterone-stimulated cells (Fig. 3B). However, analysis of cyclin A protein levels in the same extracts revealed a significant rise beginning 8-12 h after progesterone stimulation. Thus, in T47-D cells BRCA1 protein expression appears to be regulated independently of BRCA1 mRNA levels.

Discussion

In this study we show that BRCA1 mRNA levels are regulated in human breast cancer cells by the steroid hormones estrogen and progesterone. To determine whether this increase was correlated with a possible direct effect of estrogens or whether it was more closely associated with the proliferative effects of estrogens on breast cancer cells, the kinetics of BRCA1 mRNA accumulation was compared with that observed for pS2 and cyclin A mRNAs. pS2 was originally isolated as a gene that was specifically induced by estrogens in MCF-7 cells (13). Subsequent studies showed that the effects of estrogens were mediated by consensus estrogen-response elements within the promoter region of this gene (14). Cyclin A is one of the regulatory components of the cell division kinases (cyclin-dependent kinase 2). Because its level and activity is specifically increased just prior to S-phase entry (15, 16), we chose to use this gene as a marker for late G1-S-phase cells.

Our results demonstrated that BRCA1 mRNA and protein levels were regulated in concert in estrogen-deprived and restimulated MCF-7 and BT20T cells. The kinetics of BRCA1 induction was delayed when compared to the early rise in estrogen-regulated pS2 mRNA levels. In contrast, we observed a good correlation between the rise in BRCA1 mRNA and protein with that of the S-phase-regulated gene cyclin A. Therefore, the effects of estrogens on BRCA1 expression may be due to overall changes in the proliferative status of the cells as opposed to a direct effect of estrogen on DNA sequences in the BRCA1 gene itself.

The increased BRCA1 mRNA levels observed in T47-D cells may also reflect an indirect effect of progesterone on cell proliferation rather than a direct effect on DNA sequences in the BRCA1 gene itself. In these cells the kinetics of BRCA1 mRNA accumulation closely paralleled that of cyclin A rather than the increase in progesterone-regulated fatty acid synthetase mRNA levels. In contrast to estrogen-depleted and -stimulated cells, the level of BRCA1 protein was not decreased in progesterone-depleted T-47D cells. These results
suggest an uncoupling of mRNA and protein levels under certain metabolic conditions.

A similar uncoupling of BRCA1 mRNA and protein levels was detected in synchronized populations of immortalized MCF10 and 184B5 cells. In these two cell lines BRCA1 mRNA levels were tightly regulated during the cell cycle while BRCA1 protein levels remained constant. There are several possible explanations for discrepancies between mRNA and protein levels under different physiological conditions. BRCA1 proteins may have an increased half-life in T47-D cells. Alternatively, the low level of BRCA1 mRNA in the progesterone-depleted cells may be translationally regulated by other cellular proteins or antisense RNA transcripts. Precedents for both mechanisms of regulation exist for other genes (17). Interestingly, many developmentally regulated genes exhibit regulation at the level of mRNA translation (17).

The underlying mechanisms responsible for the changes in BRCA1 expression remain unknown. However, this report provides evidence to indicate that BRCA1 mRNA and protein levels can be regulated in response to steroid hormones. If the BRCA1 protein(s) functioned as a feedback regulator of growth in proliferating breast epithelial cells, women who are at increased risk of developing breast cancer due to nonsense or missense mutations in the BRCA1 gene may have an altered cellular response to hormonal stimuli. Future studies aimed at determining the role of this gene in the growth and/or differentiation of human breast epithelial cells should provide insights into novel approaches for the prevention and treatment of these cancers.

Acknowledgements

We thank Dr. Roger Wiseman for providing a BRCA1 cDNA probe and Dr. Jeffrey Struweing for supplying the PCR primers used in the genomic amplification. We are also grateful to Dr. Jeffrey Moscow for helpful comments on the manuscript.

References


Hormone-dependent Regulation of BRCA1 in Human Breast Cancer Cells

Jean M. Gudas, Hoang Nguyen, Tao Li, et al.


Updated version
Access the most recent version of this article at:
http://cancerres.aacrjournals.org/content/55/20/4561

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.