Estrogenic Effects of Genistein on the Growth of Estrogen Receptor-positive Human Breast Cancer (MCF-7) Cells in Vitro and in Vivo


Departments of Food Science and Human Nutrition (C-Y. H., R. C. S., W. G. H.) and Physiology (S. Z. H.), Michigan State University, East Lansing, Michigan 48824, and Department of Food Science and Human Nutrition, University of Illinois, Urbana, Illinois 61801 (W. G. H.)

ABSTRACT

Genistein, found in soy products, is a phytochemical with several biological activities. In the current study, our research focused on the estrogenic and proliferation-inducing activity of genistein. We have demonstrated that genistein enhanced the proliferation of estrogen-dependent human breast cancer (MCF-7) cells in vitro at concentrations as low as 20 nM, with a concentration of 100 nM achieving proliferative effects similar to those of 1 nM estradiol. Expression of the estrogen-responsive gene pS2 was also induced in MCF-7 cells in response to treatment with a concentration of genistein as low as 1 µM. At higher concentrations (above 20 µM), genistein inhibits MCF-7 cell growth. In vivo, we have shown that dietary treatment with genistein (750 ppm) for 5 days enhanced mammary gland growth in 28-day-old ovariectomized athymic mice, indicating that genistein acts as an estrogen in normal mammary tissue. To evaluate whether the estrogenic effects observed in vitro with MCF-7 cells could be reproduced in vivo, MCF-7 cells were implanted s.c. in ovariectomized athymic mice, and the growth of the estrogen-dependent tumors was measured weekly. Negative control animals received the American Institute of Nutrition (AIN)-93G diet, the positive control group received a new s.c. estradiol (2 mg) pellet plus the AIN-93G diet, and the third group received genistein at 750 ppm in the AIN-93G diet. Tumors were larger in the genistein (750 ppm)-treated group than they were in the negative control group, demonstrating that dietary genistein was able to enhance the growth of MCF-7 cell tumors in vivo. Increased uterine weights were also observed in the genistein-treated groups. In summary, genistein can act as an estrogen agonist in vivo and in vitro, resulting in the proliferation of cultured human breast cancer cells (MCF-7) and the induction of pS2 gene expression. Here we present new information that dietary genistein stimulates mammary gland growth and enhances the growth of MCF-7 cell tumors in ovariectomized athymic mice.

INTRODUCTION

The phytoestrogen genistein is present at high concentrations in plant foods such as soybeans. Phytoestrogens are known to reduce the reproductive performance of sheep grazing on subterranean clover, rabbits fed soybean hay, captive cheetahs fed diets containing soybean protein, and desert quail feeding on desert brush (1, 2). Additionally, a decrease in reproductive performance was observed in female rats fed either a soybean-based diet or a diet supplemented with genistein (3). These observations have been attributed to an estrogenic effect produced by dietary phytoestrogens. In vitro, genistein at low concentrations (200 nM) stimulates the growth of cultured human breast cancer (MCF-7) cells (4) and enhances the expression of the estrogen-responsive pS2 gene (5). These effects can be inhibited by tamoxifen (5), further confirming that these effects are mediated via the estrogen receptor. Conversely, high concentrations of genistein (above 20 µM) inhibit the growth of estrogen-dependent human breast cancer cells in vitro. Genistein blocks the cell cycle at G2-M phase, and this inhibitory effect is likely mediated via inhibition of tyrosine phosphorylation (6).

It is well documented that estrogens act to stimulate the growth of estrogen-dependent tumors in vivo. However, there are numerous reports that suggest that estrogen and estrogen agonists, such as genistein, can act as chemopreventative agents to inhibit the development of carcinogen-induced mammary tumors in laboratory animals (1, 7, 8). When genistein was administered s.c. to neonatal or prepubertal Sprague Dawley rats followed by the initiation of mammary tumors with DMBA, a reduction in tumor incidence was observed when compared with that of control rats (9). Lamartiniere et al. (9) concluded that, like other estrogen agonists, genistein may exert its chemopreventative action by enhancing mammary cell maturation, thus reducing cell proliferation in the mammary gland and the subsequent initiation by DMBA. This presents a paradox, because it is also well documented that estrogens stimulate the growth of estrogen-dependent tumors (10). The resolution of this paradox may lie in the timing of the estrogen administration. If an estrogen is administered before mammary gland maturation and initiation with a mammary carcinogen, then the number of tumors will likely be reduced, due to the effect of estrogen in causing mammary gland maturation. However, if the estrogen is administered to an animal after the development of an estrogen-dependent tumor, the growth of this tumor will be stimulated. The present study was undertaken to test the hypothesis that dietary genistein will act as an estrogen agonist to stimulate tumor growth in athymic nude mice. Thus, we examined the effects of dietary genistein on: (a) blood genistein levels; (b) stimulation of mammary gland growth; and (c) the growth of MCF-7 cells implanted into ovariectomized athymic nude mice.

MATERIALS AND METHODS

In Vitro Studies

Cell Proliferation Study. MCF-7 cells were maintained in IMEM (Bioflu-ids, Inc., Rockville, MD) containing 5% FBS, penicillin (100 units/ml), and streptomycin (100 µg/ml). Cells were incubated at 37°C in a humidified atmosphere of 5% CO2 in air. Cells were harvested and then plated at 1.5 × 104 cells/well in 24-well tissue culture polystyrene plates in media containing 5% FBS-IMEM for 4 h. Media were removed, and the cells were washed with 150 mM PBS and then incubated with media containing 5% charcoal dextran-treated FBS (Hyclone, Inc., Logan, UT) in phenol red-free media for an additional 4 h. The media were replaced with the same media containing genistein, which was synthesized as described by Chang et al. (11). Concentrations of genistein used were 0.01, 0.1, 1.0, 10, 25, 50, and 100 µM. Proliferation was assessed by fluorometric analysis of DNA content daily for 96 h. Cells were lysed in situ with 10 min EDTA (pH 12.3) at 37°C for 30 min and neutralized with KH2PO4, and then fluorescent dye (Hoechst reagent 33258) was used to determine the DNA content. Fluorescence was measured by excitation at 350 nm and emission at 455 nm to determine the DNA content when compared with a standard of salmon sperm DNA (12, 13).

Cell Culture and Treatment for Expression of pS2 mRNA. MCF-7 cells were cultured in 100 × 20-mm polystyrene tissue culture plates with the
growth conditions and growth media described above. One week before
the initiation of experimental treatments, cells were switched to medium supple-
mented with 5% charcoal-dextran-striped FBS. Due to its potential estrogenic
effect, phenol red-free media were used 24 h before treatments, and the cells
were switched to medium supplemented with phenol red-free MEM contain-
ing penicillin (100 units/ml), streptomycin (100 µg/ml), and 5% charcoal-
dextran-striped FBS. Twenty-four h after initial plating of the MCF-7 cells,
estradiol or genistein was added. The final concentrations of estradiol in the
culture media were 200 pm and 1 nm, and the final concentrations of genistein
were 1.0 and 10 nm and 1 µM. Both estradiol and genistein were dissolved in
DMSO; the final concentration of DMSO in the medium was 0.1%. The
growth medium was changed every 36 h after initial plating, and fresh estradiol
and genistein were added with each media change.

RNA Isolation. RNA was isolated 72 h after the addition of estradiol or
genistein using the method of Chomczynski et al. (14) as modified by Xie and
Rothblum (15). Ten µg of RNA were used for Northern blot analysis.

Protease d pS2. A plasmid containing a 559-bp cDNA fragment encoding
full-length pS2 (16) was obtained from the American Type Culture Collection.
PstI restriction endonuclease digestion of the plasmid (16) produced DNA
fragments of 4.4, 0.32, and 0.24 kb. These fragments were isolated from the
agarose gel slice using methods adapted from Moore et al. (17). Twenty-five
ng of pS2 cDNA (0.32 and 0.24 kb) and 50 µCi of [α-32P]dCTP were used to
label the DNA by the random primed labeling method using a commercial kit
and following the manufacturer’s instructions (Life Technologies, Inc., Gaith-
ersburg, MD).

Northern Blot Hybridization and Detection. For detection of pS2 expres-
sion, 10 µg of total RNA were separated on 1.2% formaldehyde agarose gels
and transferred to a Hybond-N Nylon membrane (Amersham, Arlington
Heights, IL). The RNA was cross-linked onto the membrane using UV light for
3 min by a 25-W transilluminator (Hoefer Scientific Instruments, San Fran-
cisco, CA; wavelength, 312 nm). The membrane was then hybridized with the
32P-labeled DNA probe, and hybridizing RNA molecules were detected by
performing autoradiography. Northern blots were also probed for human
glyceraldehyde-3-phosphate-dehydrogenase cDNA (Clontech Laboratories,
Inc., Palo Alto, CA) as a housekeeping gene to confirm equal loading of RNA
among treatment and control groups.

In Vivo Studies

Diet Selection. Diet selection was based on recommendations of the AIN
(18). The AIN-93G semipurified diet has been established as meeting all of the
nutritional requirements of mice. One concern for the use of a commercial diet
is the presence of phytosterogens from plant sources such as soybean meal.
Protein in the AIN-93G diet is derived from casein; thus, the potential inter-
ference of phytosterogens in soy-based protein supplements is eliminated.

Protein Pellet Preparation. Each protein pellet contained 2 mg of
17β-estradiol mixed with 18 µg of cholesterol as a carrier. The 20-mg mixture
containing estradiol and cholesterol was placed into a pellet mold and pressed
into a compact disk approximately 4.5 mm in diameter and approximately 2.5
mm in depth. These pellets then were placed s.c. in the interscapular region of
mice as described previously (19).

Analysis of Uterine Weight and Mammary Gland Growth in Ovari-
tomized Athymic Female Mice Fed Genistein. Athymic nude mice (Har-
land, Indianapolis, IN) were ovariectomized at 21 days of age. Dietary treat-
ments were initiated at 28 days of age for a period of 5 days. Mice were fed
by the AIN-93G semipurified diet containing estradiol at 1 ppm or genistein
at 750 ppm. Control animals received only the AIN-93G diet. Mice were killed,
and the uteri were removed for assessment of uterine weights and mammary
gland growth. Mammary glands were removed and fixed and stained as whole
mounts (20). Mammary gland growth was assessed by determining the si-
Ze and number of end buds and the extent of ductal development as viewed under
a dissecting scope (21).

Analysis of Tumor Growth in Athymic Nude Mice Fed Genistein.
Female 21-week-old athymic nude mice (Harland) were allowed to acclimate
for 1 week. Mice were ovariectomized at 28 days of age and then allowed to
recover for 7 days before any treatment. Estrogen-dependent human breast
 cancer (MCF-7) cells were grown to confluence and collected using trypsin-
EDTA. Cells were counted using a hemocytometer (Fisher Scientific, Pitts-
burgh, PA) and diluted in culture media to a concentration of 1 × 106 cells/ml.

An aliquot of cell suspension (100 µl) was injected into four sites in the flank
of 35-day-old female athymic mice. MCF-7 cells will not produce tumors in
ovariectomized athymic nude mice unless the mice are supplemented with
estrogen. Estradiol was administered in pellets containing 2 mg of estradiol
(19, 22) implanted s.c. in the interscapular region of each mouse. Tumors were
allowed to develop for 5–6 weeks and were measured weekly, and the
cross-sectional areas were determined using the formula described previously
(22). When tumors reached an average cross-sectional area of 35–40 mm2,
animals were divided into three treatment groups (four to six animals/group),
with each group normalized for tumor number, tumor size, and animal number.
Estradiol pellets were removed from all animals. Positive controls were reim-
planted with a fresh estradiol pellet, negative control mice received the
AIN-93G diet, and dietary treatments with 750 ppm genistein were initiated.
Tumor areas were measured weekly for the duration of the study. At the
completion of the study, mice were killed, and the uteri were removed for the
assessment of wet weights. Blood was collected from the animals by cardiac
puncture for the analysis of genistein content.

Plasma Genistein Concentration Analysis in Athymic Nude Mice Fed
Genistein. Blood from cardiac puncture was placed into EDTA-containing
vials and centrifuged at 500 × g for 10 min. Plasma was separated and stored
at −20°C until analysis. A known amount of daidzein, which differs from
genistein in chemical structure by one hydroxyl group, was added to each
sample to serve as an internal standard. Daidzein and genistein have similar
chromatographic properties. Therefore, the isolation and quantification of
daidzein provided a recovery standard that could be used to analyze the
effectiveness of the recovery of genistein. Each sample was split into two
 aliquots. One aliquot was incubated with β-glucuronidase/sulfatase for 24 h at
37°C to liberate free genistein from its conjugated form. The other aliquot was
handled in a similar manner without β-glucuronidase/sulfatase. Samples were
applied to a C18 Sep-Pack columns washed with 10 ml of H2O (HPLC grade)
and eluted with 10 ml of 85% methanol. The samples were dried in a vacuum
spin dryer (Savant, Farmingdale, NY) and reconstituted in 100 µl of 85% methanol.
Twenty µl were used for HPLC analysis using a reverse phase C18 column.
The mobile phase consisted of 50% methanol, 50% water, and 0.1% acetic acid,
with a flow rate of 1 ml/min (23). The absorbance in samples was monitored
using a spectrophotometer at 260 nm.

Statistical Analysis. Tumor area data were analyzed statistically using a
repeated-measures protocol in a split plot design (24) and the general liner
models procedure in the SAS program (25). Independent variables were
 treatments (estrogen, genistein, or control), tumor nested within treatments,
week, and the treatment by week interaction. Treatment means for each week
were compared using the least significant difference method.

RESULTS

In Vitro Studies

Effect of Genistein on MCF-7 Cell Proliferation. To determine the
minimum concentration of genistein that will stimulate or inhibit
MCF-7 cell growth, we conducted a cell proliferation dose-response
study. MCF-7 cells were monitored in response to estradiol (1 nM) and
various concentrations of genistein ranging from 0.01–100 µM (Fig. 1).
Data are expressed as a percentage of the control cell cultures.
These levels were chosen because the genistein blood levels reported
in animals and humans consuming diets high in genistein (such as
soy-containing diets) have blood concentrations ranging from 0.1–6
µM (26). Estradiol (1 nM) stimulated cell proliferation 2.4-fold over that
of the control MCF-7 cells. Genistein increased cell growth in a dose-
dependent manner in the range of 0.01–1 µM. Maximal growth
stimulation (approximately 3-fold over that of the control) was ob-
served at 1 µM and was sustained at this level of stimulation at a dose
up to 10 µM. In contrast, higher concentrations (25–100 µM) of
genistein produced a dose-dependent decrease in cell growth when compared
with that of untreated controls.

Effect of Genistein on Estrogen-dependent pS2 mRNA Expression
in MCF-7 Cells. To evaluate the potential of genistein to en-
hance the expression of an estrogen-responsive gene, we conducted

3834

Downloaded from cancerres.aacrjournals.org on October 22, 2017. © 1998 American Association for Cancer Research.
estradiol and genistein (Fig. 3). The average end bud number was with the end buds as the major growth points (26). Increases in the ppm) in the AIN-93G diet or from controls fed the AIN-93G diet only.

In Vivo Studies

In mature mice, mammary gland growth occurs via ductal elongation with the end buds as the major growth points (26). Increases in the number and size of the end buds were observed in animals consuming estradiol and genistein (Fig. 3). The average end bud number was 2.9 ± 0.85 in the ovariectomized control group, 9.1 ± 0.67 in the estradiol group, and 6.4 ± 0.89 in the genistein-treated groups (Fig. 4). Mammary gland growth was significantly greater (P < 0.05) in mice fed genistein or estradiol when compared with that of ovariectomized controls. Genistein- or estradiol-treated animals were not significantly different from each other (P > 0.05). The uterine weight of the genistein treatment group was also increased in comparison with that of the control group (data not shown). These data suggest that dietary genistein has the potential to stimulate estrogenic responses in vivo.

Effect of Genistein on Tumor Growth in Athymic Nude Mice. Because genistein stimulated both mammary gland and uterine growth, it was of interest to determine the effects of an estradiol (2 mg) pellet or dietary genistein (750 ppm) on estrogen-dependent tumor growth. Ovariectomized athymic mice implanted with MCF-7 cell tumors were divided into three treatment groups after tumors reached an average cross-sectional area of approximately 55 mm². The three treatment groups were: (a) negative controls (without estriadiol pellets or genistein treatments); (b) positive controls (estradiol pellets); and (c) dietary genistein (750 ppm) treatment groups. Tumor growth was monitored weekly. Tumors in the positive control group grew rapidly, and after an additional 2 weeks on estrogen treatment, the average tumor cross-sectional area had reached 120 mm² (Fig. 5). After 2 weeks of estradiol treatment, the average tumor weight was 750 mg, and the total tumor weight/mouse was approximately 3 g (approximately 10% of the body weight). Therefore, mice were killed after 2 additional weeks of estrogen treatment due to the size of the tumors. After the estrogen pellets were removed in the negative control group, tumor growth stopped, and tumor size was maintained at an average cross-sectional area of approximately 58 mm² for an additional 12 weeks study (Fig. 5). In the dietary genistein treatment group, after the estrogen pellets were removed and the mice were switched to 750-ppm genistein diets, tumor growth increased. The average tumor cross-sectional area in the dietary genistein treatment group increased from 55 to 120 mm² after 12 additional weeks on genistein treatment. These MCF-7 cell tumors were similar in size to that of tumors after 2 weeks on estradiol treatment. These results indicated that dietary genistein acts as an estrogen agonist in vivo to stimulate the growth of estrogen-dependent MCF-7 tumor cells implanted into athymic mice.

Plasma Genistein Concentration Analysis in Athymic Nude Mice Fed Genistein. Plasma genistein levels were measured by HPLC. The concentration of free genistein in plasma was 0.24 ± 0.08 μM. The concentration of total genistein including free and conjugated forms in plasma was 2.1 ± 0.14 μM. These concentrations are within

Northern blot analysis using RNA isolated from genistein-treated MCF-7 cells and probing with a human pS2 cDNA probe (16). The cell proliferation studies (Fig. 1) suggested that genistein acts via the estrogen receptor to enhance cell proliferation at low concentrations. Estradiol at concentrations of 0.2 nm and genistein at concentrations of 1–10 μM were observed to induce pS2 gene expression (Fig. 2). Additionally, we have observed an increase in pS2 mRNA expression by genistein at concentrations up to 50 μM (data not shown). These data indicate that genistein can act as a weak estrogen agonist in vitro as measured by estrogen-dependent (pS2) gene expression.

In Vivo Studies

Effect of Genistein on Uterine Weight and Mammary Gland and End Bud Development in Ovariectomized Athymic Female Mice. Mammary gland whole mounts were obtained from ovariectomized mice consuming either estradiol (1 ppm) or genistein (750 ppm) in the AIN-93G diet or from controls fed the AIN-93G diet only.

The concentration of total genistein including free and conjugated forms in plasma was 2.1 ± 0.14 μM. These concentrations are within
GENISTEIN STIMULATES TUMOR CELL GROWTH IN VIVO

The range of concentrations that stimulated pS2 mRNA expression (Fig. 2) and the growth of MCF-7 cells (Fig. 1).

DISCUSSION

Genistein elicits a concentration-dependent dual threshold effect with regard to growth stimulation or inhibition on cultured estrogen-dependent human breast cancer (MCF-7) cells in vitro. At low concentrations of genistein in dextran-charcoal-stripped FBS, we observed a dose-dependent increase in MCF-7 cell proliferation from 0.01 to 1 μM (Fig. 1). Maximal growth stimulation was observed at 1 μM, and this level of growth was sustained up to 10 μM. At higher concentrations, we observed a dose-dependent inhibition in cell growth when concentrations were 25–100 μM. These results are consistent with those reported by Martin et al. (4), in which growth stimulation in MCF-7 cells was observed at a concentration of 200 nM, and Wang et al. (5), in which growth stimulation in MCF-7 cells was observed in a concentration-dependent manner between 10 nm and 1 μM. Wang et al. (5) also observed growth inhibition at concentrations higher than 10 μM. These results suggest that there are at least two dose-dependent mechanisms by which genistein can alter cell proliferation. One mechanism stimulates proliferation at low concentrations of genistein (10 nm to 1 μM) and is likely to be mediated via the estrogen receptor. The other mechanism, which is antiproliferative, is active at high concentrations of genistein (25–100 μM) and is likely to be mediated via anti-tyrosine phosphorylation and inhibition of cell cycle progression (6). The threshold for stimulation of proliferation...
was in the concentration range required to stimulate expression of pS2 mRNA (Fig. 2). Previous studies have determined that tamoxifen inhibits the induction of pS2 gene expression (5). Additionally, in our studies, pS2 gene expression was enhanced at concentrations up to 50 μM. However, cell proliferation was inhibited at concentrations above 25 μM. These observations suggest that the estrogen receptor-mediated mechanism is active at the higher concentrations; however, the growth-inhibitory mechanism associated with genistein overrides the growth-stimulatory effects from genistein.

Genistein binds to the estrogen receptor with an affinity approximately 100-fold less than that of estradiol (5, 23). To further confirm whether genistein is acting via an estrogen receptor mechanism, we evaluated the expression of the pS2 gene in cultured MCF-7 cells. The human pS2 gene was initially characterized as a gene whose expression is specifically controlled by estrogen in the breast cancer cell line MCF-7 (27). The increase in pS2 mRNA after the addition of estradiol to the culture medium is a primary transcriptional event, suggesting that control of pS2 gene promoter activity by the estrogen receptor is mediated by a cis-acting estrogen-responsive element that could be located in the 5'-flanking region of the pS2 gene (16). The results on pS2 stimulation by genistein (Fig. 2) are consistent with the results observed on cell proliferation in that 1 μM (Fig. 1) was effective at increasing proliferation and pS2 gene expression. Additionally, Wang et al. (5) reported that tamoxifen inhibits genistein-stimulated pS2 gene expression in MCF-7 cells. These data further support that genistein can act via an estrogen receptor-mediated mechanism.

Here, we report that dietary genistein (750 ppm) fed to 28-day-old athymic ovariectomized mice will stimulate mammary gland growth (Figs. 3 and 4). This is the first report in which dietary genistein has been shown to enhance mammary gland growth in vivo. Reports (9, 28) in which high dosages of genistein were administered s.c. to rats demonstrate that genistein stimulates mammary gland differentiation, promoting lobular-alveolar development in immature Sprague Dawley rats. No studies have been reported in which the effect of genistein was evaluated for its potential to stimulate the growth of estrogen-dependent tumors in vivo. The studies presented here provide compelling evidence that MCF-7 estrogen-dependent tumors can be stimulated to grow in ovariectomized athymic host mice supplemented with dietary genistein (750 ppm). It is important to point out that both estrogen and genistein have been shown to be chemopreventive in the carcinogen-induced rat DMBA animal model (9, 28, 29). Neonatal administration of estrogen to rats has been shown to reduce incidences of DMBA-induced mammary tumors (29). In the studies with genistein, three s.c. injections of genistein were given to young rats, which reduced the number of mammary tumors initiated by DMBA. Naganawa et al. suggest that genistein acts as an estrogen agonist to stimulate mammary gland differentiation. This enhanced mammary gland differentiation may be the mechanism responsible for the chemopreventive effects of genistein in the rat DMBA mammary carcinogenesis model. Thus, there seems to be a paradox with regard to the action of genistein on tumor growth in rodent models. Our results in the ovariectomized athymic mouse suggest that genistein stimulates mouse mammary gland growth without maturation, which is different from the effect of genistein in the immature rat, in which genistein stimulates prolactin secretion (23) and ultimately results in lobular-alveolar development and mammary gland maturation (26). Genistein may act similar to estradiol, because estradiol is known to act to reduce tumor incidence by chemoprevention (30), whereas there are numerous reports (10) that indicate that estrogens act as tumor promoters. Thus, whether genistein acts as a chemopreventative agent or as an agent to stimulate tumor growth will likely depend on the timing of genistein administration.

The present study focused on the estrogenic effect of genistein in vitro and in vivo systems. Genistein binds to the estrogen receptor with an affinity approximately 100-fold lower than that of estradiol, resulting in enhanced proliferation activity. In cultured cells, genistein stimulates MCF-7 cell growth at concentrations as low as 100 nm, achieving similar effects to that of estradiol at 1 nm (Fig. 1). We observed a 240% increase in growth when treated with 100 nm genistein compared with that of the control. Expression of the estrogen-responsive gene pS2 was also induced in response to treatment with genistein at concentrations higher than 100 nm (Fig. 2). In vivo, using ovariectomized athymic nude mice implanted with estrogen-responsive MCF-7 cells, we have demonstrated that treatment with 750 ppm dietary genistein was able to stimulate the growth of these estrogen-responsive MCF-7 cell tumors in the absence of estrogen (Fig. 5). Plasma genistein was 2.1 μM, of which 0.24 μM present was in the free form. These concentrations are above the concentration required for MCF-7 cell growth stimulation observed in vitro. One can predict from the in vitro cell growth studies that the dosage of dietary genistein used in these studies will produce a blood concentration of genistein that will stimulate the growth of implanted MCF-7 cell tumors. Thus, these results are consistent with in vivo cell culture studies. Even if the conjugated form is biologically active, these levels are below the high concentrations (above 25 μM) required in vitro to produce a dose-dependent decrease in MCF-7 cell growth. It is unlikely that the concentration required to inhibit MCF-7 cell growth can be achieved in vivo.

In summary, genistein can act as an estrogen agonist resulting in the proliferation of human breast cancer cells (MCF-7) in vitro and enhanced growth of MCF-7 cell tumors implanted into ovariectomized athymic nude mice in vivo. Thus, there is the potential for dietary genistein to stimulate the growth of estrogen-dependent tumors in humans with low circulating endogenous estrogen levels, such as those found in postmenopausal women.

ACKNOWLEDGMENTS

We thank Drs. Y. C. Chang and M. G. Nair for providing genistein. Thanks are also due to Dr. Patty Dickerson-Weber for technical assistance and to Dr. Les Bourquin for statistical consultation. Sincere appreciation is extended to Drs. J. Mark Cline and John Linz for critical review of the manuscript.

REFERENCES

Estrogenic Effects of Genistein on the Growth of Estrogen Receptor-positive Human Breast Cancer (MCF-7) Cells  *in Vitro* and  *in Vivo*


*Cancer Res* 1998;58:3833-3838.

Updated version  Access the most recent version of this article at:  
http://cancerres.aacrjournals.org/content/58/17/3833

E-mail alerts  Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions  To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions  To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.