Therapy of Colon Cancer with Oncolytic Adenovirus Is Enhanced by the Addition of Herpes Simplex Virus-thymidine kinase

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ABSTRACT

A major obstacle to the successful application of suicide gene therapy strategies that rely on in situ transduction of tumor cells is the poor distribution of the vector throughout the tumor mass. To address this problem, we evaluated the use of Ad.TKRC, an E1b M55,000 deleted replicating adenoviral vector engineered to express the herpes simplex virus type 1 thymidine kinase gene (HSV-tk) in combination with ganciclovir (GCV) as a treatment for human colon cancer xenografts in nude mice. We compared the efficacy of this system with that of a standard replication-deficient adenovirus expressing HSV-tk (Ad.TK) in mice bearing LS180 tumors. In this system, Ad.TKRC alone was as effective as a traditional Ad.TK vector in combination with GCV. The addition of GCV significantly enhanced the antitumor effect of Ad.TKRC. Furthermore, we demonstrated that the survival of HT-29 human colon cancer xenografted mice treated with Ad.TKRC and GCV was prolonged compared with Ad.TKRC alone or with administration of a single cycle of topotecan. These data demonstrate that the addition of direct viral oncolysis to the HSV-tk/GCV suicide gene system resulted in a striking improvement in treatment efficacy and that it may offer advantages over the use of chemotherapeutic agents for treatment of localized disease.

INTRODUCTION

With over 135,000 new cases and 57,000 deaths expected in 1998, colorectal carcinoma is the third most common cause of cancer death in the United States (1). Despite new insights into the molecular pathogenesis of colon cancer and improvements in surgery, radiotherapy, and chemotherapy, the median survival of patients with metastatic disease has remained unchanged for the last 40 years at ~15 months (2).

Suicide/prodrug gene therapy holds great promise to become an important addition to the therapy of some forms of cancer. This strategy involves the transfer and expression of nonmammalian genes encoding enzymes that convert nontoxic prodrugs into cellular toxins. The most widely used suicide gene is HSV-tk (3), which confers sensitivity to GCV (3).

Initially, replication-incompetent retroviral vectors have been used for viral-mediated HSV-tk gene delivery (4). Subsequently, replication-deficient adenoviral vectors have been used as suicide gene transfer vectors (5, 6) because they offer, in contrast to retroviral vectors, higher transduction efficiency and transgene expression in a wide variety of cell types independently of their cell cycle status. Because these vectors are replication incompetent, their use for cancer gene therapy is limited by their inability to spread and infect additional cells subsequent to the initial infection event. The fundamental physics of diffusion of vector particles in tissue spaces and the high density of viral receptors in tissue largely restrict distribution of these vectors only to the close proximity of the site of injection. The presumed need to genetically modify every tumor cell may be partially obviated by the HSV-tk/GCV system because it is often associated with a “bystander effect” (3), i.e., the extension of toxic effects to untransduced neighboring cells by diffusion of phosphorylated GCV through cellular gap junctions (7). Although the bystander effect increases tumor cell killing, it is nonetheless necessary to transduce a significant proportion of the tumor. In an effort to increase transduction efficiency, intratumoral replication of viral vectors has been attempted by cotransfection of replication-defective viral constructs expressing HSV-tk with wild-type virus (8, 9) or with viral genes critical for replication (10). Also, direct inoculation of xenogenic murine retroviral producer cells into brain tumors (11) has been performed in an effort to achieve continuous local vector production.

Exploiting the oncolytic effects of viral replication, Bischoff et al. (12) demonstrated that an E1b M55,000 deleted adenovirus preferentially replicates in p53 dysfunctional tumor cells (12). This virus, which does not express a therapeutic transgene, has entered clinical trials in combination with cisplatin in patients with advanced squamous cell carcinoma of the head and neck (13). To increase its efficacy and to be able to control the replication and spread of the virus, we and others have developed E1b M55,000 deleted adenoviral vectors expressing HSV-tk (14) or a cytosine deaminase/HSV-tk fusion gene (15).

Colorectal carcinoma has been evaluated as a potential target for HSV-tk/GCV suicide gene therapy (16–18) and appears to be an excellent tumor model for evaluation of the utility of an E1b attenuated replication competent vector because of the high prevalence of p53 abnormalities, especially in metastatic lesions (19, 20). In this report, we describe the use of an E1b M55,000-deleted, replication-competent adenovirus expressing HSV-tk in combination with GCV for the treatment of colorectal carcinoma. We compared its efficacy with those of a standard replication-deficient adenovirus expressing HSV-tk and the topoisomerase I inhibitor topotecan (reviewed in Ref. 21), which shows significant antineoplastic activity against a variety of solid tumors including colorectal carcinoma.

MATERIALS AND METHODS

Generation of Adenoviral Vectors. The construction and generation of the adenoviral vectors were described previously (14). Briefly, Ad.TKRC harbors in the E1 region an HSV-tk/Ad5 E1a E1b M55,000 expression cassette under the transcriptional control of the human CMV-IE promoter in combination with the adenovirus tripartite leader (22). This cassette was flanked upstream by the Ad5 packaging sequence and downstream by Ad5 pIX. The E1 genes were transcriptionally linked to the HSV-tk gene via an encephalomyocarditis virus internal ribosome entry site (23) to reduce the likelihood of generating replication-competent deletion mutants lacking HSV-tk. In contrast to d1l520 (12, 24), Ad.TKRC expresses a suicide gene and the CMV-IE promoter as opposed to the endogenous E1 promoter, which drives the E1 genes. Furthermore, the E3 region of Ad.TKRC is removed, and the E1b deletion is larger than in d1l520 (1257 versus 827 bp).

For comparison, we generated the replication-deficient adenoviruses Ad.GFP and Ad.TK, which carry the GFP and the HSV-tk, respectively, driven by the CMV-IE promoter.
Tissue Culture. The colon cancer cell lines SW-620, LS180, HCT 81, WiDr, HT-29, and DLD-1 were purchased from the American Type Culture Collection (Manassas, VA) and propagated in DMEM (Life Technologies, Inc., Grand Island, NY) supplemented with 10% heat-inactivated fetal bovine serum (Life Technologies, Inc.) at 37°C and 5% CO2.

Flow Cytometric Analysis of GFP Expression. To determine the in vitro transduction efficiency of human colon carcinoma cell lines with human adeno-adenovirus type 5, subconfluent cell monolayers were exposed to Ad.GFP at a MOI of 10. Sixty h later, single-cell suspensions were analyzed by flow cytometry (FACSCalibur; Becton Dickinson, San Jose, CA).

HSV-tk Bystander Assay. We evaluated the in vitro bystander effect of different colon cancer cell lines by coculturing mixtures of different ratios of Ad.TK transduced and untransduced cells as described previously (25).

The median fraction of affected cells (F_a) was calculated according to the formula:

\[ F_a = \frac{I_{Max} - I_{100}}{I_{WT} - I_{100}} \]

Variables I_{WT}, I_{100}, and I_{Max} are the median [3H]thymidine incorporation of the unmodified wild-type cells, cells exposed to Ad.TK at an MOI of 25 for 12 h, and mixtures of the two, respectively. At this MOI, nearly all of the cells were susceptible to transduction in vitro with Ad.GFP (data not shown).

Colony-forming Assay. To evaluate the sensitivity of HT-29 to topotecan, 1 million subconfluent HT-29 cells were incubated with increasing concentrations of topotecan for 24 h. After drug exposure, the cells were washed, trypsinized, and counted. To assess the colony-forming ability of the cells after the treatment, 300 cells were dispersed in 100-mm tissue culture plates and incubated at 37°C for 10–14 days. Surviving colonies were fixed and stained with 1% crystal violet/formaldehyde, and colonies larger than 1 mm were counted. The fraction of unaffected cells (F_u) was calculated by dividing the number of colonies present in drug-treated dishes by the number present in untreated control dishes. The concentration effect curve is represented as the fraction of affected cells (F_a = 1 - F_u) versus topotecan concentration. Each experiment was performed in triplicate and was repeated twice.

Animal Experiments. Female athymic nude mice (obtained from Harlan Sprague Dawley, Inc., Indianapolis, IN) were s.c. injected with 10 million LS180 or HT-29 human colon cancer cells into the right dorsal lumbar region in 100 μl of serum-free DMEM with 10% Matrigel (Collaborative Products, Bedford, MA). Blinded, bidimensional tumor measurements were performed twice a week with calipers, and tumor volume was determined using the simplified formula of a rotational ellipsoid (l × w² × 0.5; Ref. 26). Once tumors reached a volume of 100–150 mm³, animals were treated by intratumoral injection of 10⁹ plaque-forming units of virus directly into the LS 180 xenografts. GCV treatment was started 1 and 3 days after Ad.TK and Ad.TKRC injection, respectively. The tumors were resected in toto and weighed 3 days after completion of the GCV course in animals receiving Ad.TK and 1 day after completion in those receiving Ad.TKRC.

Analysis of the tumor masses revealed that the combination of either Ad.TK or Ad.TKRC with GCV increased the treatment efficacy (P < 0.002; Fig. 1). The antitumor effect of Ad.TKRC alone was equal to that of Ad.TK followed by GCV. This demonstrates that Ad.TKRC has direct oncolytic activity in LS 180 cells and confirms our previous observations in a survival study using melanoma and cervical cancer xenografts (14). The antitumor effect in mice treated with Ad.TKRC plus GCV was significantly superior to that of Ad.TK followed by GCV (P < 0.001).

Treatment Efficacy of Ad.TKRC/GCV versus Topotecan. By clonogenic inhibition assay, we found that the EDso of topotecan in HT-29 cells was 0.02 μM (Fig. 2). Accordingly, topotecan administration to HT-29 xenografted mice resulted in a modest but significantly improved survival when compared with untreated tumor-bearing animals or GCV-treated mice with HSV-tk-negative tumors (P < 0.005; Fig. 3). However, mice treated with Ad.TKRC alone or in combination with GCV or topotecan survived longer than those treated only with topotecan (P ≤ 0.0005). There was no statistically significant difference in survival whether Ad.TKRC was administered...
alone or in combination with topotecan. By contrast, when Ad.TK<sup>RC</sup> was followed by GCV, the survival of the animals was again significantly improved ($P \leq 0.02$; Table 1).

**DISCUSSION**

To enhance tumor transduction by in situ amplification and spread of the initial viral inoculum within a tumor mass, we developed the recombinant adenovirus Ad.TK<sup>RC</sup> that combines viral replication and oncolysis with the HSV-<i>tk</i>/GCV suicide gene system to kill cancer cells. We compared its antitumor effect with that of a standard replication-deficient Ad.TK in the rapidly growing LS180 colon cancer xenograft model. In addition, we studied the antitumor response and survival of mice bearing xenografts of HT-29 colon adenocarcinoma cells, a tumor known to be resistant to most chemotherapeutic agents (27). Because of its slower growth (48-h doubling time), this tumor model more closely reflects clinical colon cancer, where only a small fraction of tumor cells are cycling at any given time. The HT-29 xenografted animals were treated with Ad.TK<sup>RC</sup> alone or in combination with GCV or topotecan.

Our studies in the LS180 xenograft model revealed that there was no significant difference in the antitumor effect whether animals received the replication-deficient Ad.TK in combination with GCV or Ad.TK<sup>RC</sup> alone, confirming that replicating human adenovirus has intrinsic oncolytic activity (12, 28). In addition, in both the LS180 and HT-29 colon cancer models, treatment with Ad.TK<sup>RC</sup> in combination with GCV starting 3 days after virus injection showed the best antineoplastic effects.

<table>
<thead>
<tr>
<th>Group</th>
<th>Group size (n)</th>
<th>Long-term survivors (n)</th>
<th>Median survival (days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Untreated</td>
<td>5</td>
<td>0</td>
<td>22</td>
</tr>
<tr>
<td>GCV alone</td>
<td>5</td>
<td>0</td>
<td>22</td>
</tr>
<tr>
<td>TPT alone</td>
<td>10</td>
<td>0</td>
<td>33</td>
</tr>
<tr>
<td>Ad. TK&lt;sup&gt;RC&lt;/sup&gt;</td>
<td>10</td>
<td>1</td>
<td>65</td>
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<tr>
<td>Ad. TK&lt;sup&gt;RC&lt;/sup&gt; GCV</td>
<td>10</td>
<td>4&lt;sup&gt;a&lt;/sup&gt;</td>
<td>91</td>
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<tr>
<td>Ad. TK&lt;sup&gt;RC&lt;/sup&gt; TPT&lt;sup&gt;b&lt;/sup&gt;</td>
<td>10</td>
<td>2</td>
<td>65</td>
</tr>
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<sup>a</sup> Two of the four animals were tumor free at day 114; all other long-term survivors had tumors ($V \leq 350 \text{ mm}^3$).

<sup>b</sup> TPT, topotecan.

Mice bearing HT-29 tumors treated with Ad.TK<sup>RC</sup> alone or in combination with GCV survived longer than animals treated with the topoisomerase I inhibitor topotecan. Topotecan is active against many colon carcinoma cell lines that are resistant to other frequently used antineoplastic agents. Preclinical drug screening models using athymic nude mice have shown that topotecan is the most active agent in HT-29 tumors (27).

The results of the experiments described here confirm our previous results obtained in human melanoma and cervical cancer models, demonstrating that the combination of direct viral oncolysis together with an increase in suicide gene distribution within the tumor results in a significant improvement of treatment efficacy (14). Furthermore, the addition of HSV-<i>tk</i> to the adenovirus also provides a safety mechanism that allows the viral infection to be aborted by GCV treatment should undesired spread of the infection occur during therapy.

Previous reports indicate that the binding of adenovirus E1a proteins to the cellular pRB and p300 proteins can induce quiescent cells to enter S-phase (29) and thereby conceivably increases susceptibility to S-phase-specific antineoplastic agents, like HSV-<i>tk</i>/GCV or topotecan (30). Furthermore, adenovirus E1a increases topoisomerase levels (31, 32) and thus potentially enhances the cytotoxicity of topoisomerase I inhibitors (33). However, we did not observe an increased survival of animals treated with Ad.TK<sup>RC</sup> followed by topotecan administration relative to those receiving Ad.TK<sup>RC</sup> alone, suggesting that the inhibition of viral replication and oncolysis by topotecan (34–36) might have counterbalanced the potential increased cytotoxicity of topotecan. In contrast, despite abolition of viral replication by GCV, we demonstrated that the combination of direct viral oncolysis together with HSV-<i>tk</i>/GCV mediated cell killing resulted in a striking improvement of treatment efficacy. Indeed, this combination proved to be more efficacious than a standard replication-deficient adenovirus expressing HSV-<i>tk</i>, as well as a single treatment cycle with one of few antineoplastic agents active against HT-29 colon cancer, topotecan. It is tempting to speculate that the efficacy of cancer treatment could be enhanced by the combined use of standard chemotherapy and gene therapy. Studies are presently in progress to verify this hypothesis.

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**REFERENCES**


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