A Novel Antisense Oligonucleotide Targeting Survivin Expression Induces Apoptosis and Sensitizes Lung Cancer Cells to Chemotherapy

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Abstract

Survivin, an inhibitor of apoptosis protein, deserves attention as a selective target for cancer therapy because it lacks expression in differentiated adult tissues but is expressed in a variety of human tumors. We designed 20-mer phosphorothioate antisense oligonucleotides targeting different regions of survivin mRNA and investigated their ability to down-regulate survivin mRNA and induce apoptosis in the lung adenocarcinoma cell line A549. Oligonucleotide 4003, which targets nucleotides 232–251 of survivin mRNA, was identified as the most potent compound. As measured by real-time PCR, 4003 down-regulated survivin mRNA in a dose-dependent manner with an IC50 of 200 nM. Its maximum effect was achieved at a concentration of 400 nM, at which mRNA was down-regulated by 70%. As revealed by increased caspase-3-like protease activity, nuclear condensation and fragmentation, and trypan blue uptake, treatment with 4003 induced apoptosis and sensitized tumor cells to the chemotherapeutic agent etoposide. Oligonucleotide 4003 did not reduce the viability of normal blood leukocytes with marginal levels of survivin mRNA.

Introduction

Diminished apoptosis plays a critical role in tumor initiation, progression, and drug resistance. Several proteins that inhibit apoptosis have been identified, including bcl-2 family members bcl-2 and bcl-xL and the IAPs. Certain members of the latter family directly inhibit terminal effector caspses engaged in the execution of cell death (1). The gene encoding the IAP survivin was cloned recently, and the protein was characterized (2). Survivin is expressed during embryonal development but lacks expression in terminally differentiated adult tissues (2, 3). Interestingly, it becomes reexpressed in transformed cell lines and in a variety of human tumors (1, 2). Survivin is expressed in the G2-M phase in a cell cycle-regulated manner, and its interaction with the mitotic spindle apparatus is essential for antiapoptotic function (4). This could imply that the IAP survivin counteracts a default induction of apoptosis in the G2-M phase of the cell cycle. Overexpression of survivin has oncogenic potential because it may overcome the G2-M-phase checkpoint to enforce progression of cells through mitosis. Because survivin inhibits processing of downstream effector caspase-3 and -7 in cells receiving an apoptotic stimulus (1), its overexpression in tumors is implicated in the resistance to a variety of apoptotic stimuli, including chemotherapy.

Lung and breast cancer are leading causes of cancer death, and their incidence continues to rise. The main reasons for the unfavorable prognosis of these tumors is their propensity to metastasize early and develop resistance to a wide range of functionally unrelated anticancer agents. Interestingly, lung and breast cancer cells express the highest levels of survivin found in human tumors (1, 2), and in agreement with its biological function, survivin expression is correlated with shorter survival in patients with non-small cell lung cancer (5). Although survivin has been widely recognized as an attractive target for cancer therapy (2–10), the use of antisense cDNA and oligonucleotides to inhibit its expression has only recently been described (11–13). Whereas these studies were designed to unravel the biological function of survivin, the promise of survivin antisense to facilitate apoptosis of tumor cells and overcome chemoresistance in cancer therapy remains to be determined.

In the present study, we developed a series of 20-mer phosphorothioate oligonucleotides targeting various regions of survivin mRNA. Using real-time PCR and the survivin-overexpressing lung adenocarcinoma cell line A549, one antisense oligonucleotide was identified that most efficiently down-regulated survivin mRNA levels and directly induced apoptosis. Moreover, in a combination experiment with the chemotherapeutic agent etoposide, evidence is provided that antisense-mediated down-regulation of survivin has the potential to sensitize tumor cells to chemotherapy.

Materials and Methods

Antisense Oligonucleotides. The secondary structure of the 1619-bp survivin mRNA (GenBank accession number NM001168) was predicted by the RNAfold program (14) to identify target sites putatively presenting in single-stranded conformation and thus likely to be accessible to antisense oligonucleotides (15, 16). Based on the identified sites, a series of six 20-mer antisense sequences targeting different regions of survivin mRNA were designed. Antisense oligonucleotide sequences are shown in Table 1. A 3-base mismatch control to the most potent antisense oligonucleotide was also used. Oligonucleotides were provided by Genset SA (Paris, France) in the form of phosphorothioate oligonucleotides. A BLASTN search of a database containing all sequences of GenBank, European Molecular Biology Laboratory, DDBJ: DNA Data Base of Japan, and Protein Data Base was performed to exclude homology of the antisense and control oligonucleotides to other human genes.

Tumor Cell Line and PBMCs. The lung adenocarcinoma cell line A549, which is reported to express high levels of survivin (1), was obtained from American Type Culture Collection and maintained in RPMI 1640 supplemented with 2 mm L-glutamine and 10% FCS at 37°C in a humidified atmosphere containing 5% CO2, PBMCs, which are reported to lack survivin expression (2), were isolated from the heparin-treated blood of three healthy donors and used in transfection experiments directly upon isolation using a Ficoll-Hypaque solution (Biochrom KG, Berlin, Germany).

Treatment of Cells with Antisense and Etoposide. One day before transfection, A549 cells were plated in 6-, 24-, or 96-well tissue culture plates. Oligonucleotides were delivered in the form of complexes with Lipofectin.
Table 1  Sequences of antisense oligonucleotides and their target sites on the survivin mRNA

<table>
<thead>
<tr>
<th>Oligonucleotide</th>
<th>Sequence</th>
<th>Target site on survivin mRNA (nucleotide no.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>4018</td>
<td>GGCACCCATAGCGGCGGCCGC</td>
<td>39–58</td>
</tr>
<tr>
<td>4002</td>
<td>TGAAATGAGATGCGGTTG</td>
<td>97–116</td>
</tr>
<tr>
<td>4003</td>
<td>CCGAGGCTCGAGCTCTTG</td>
<td>232–251</td>
</tr>
<tr>
<td>4004</td>
<td>TCACACAGGTTAATCCTTC</td>
<td>329–348</td>
</tr>
<tr>
<td>4007</td>
<td>CTGGTGAACGCTCAAAACA</td>
<td>821–840</td>
</tr>
<tr>
<td>4007</td>
<td>GAGGGGATCAATACCTCAC</td>
<td>1461–1480</td>
</tr>
</tbody>
</table>

* The 1619-bp mRNA (GenBank accession number NM001168) was used.

(Life Technologies, Glasgow, United Kingdom) as described previously (16). Upon a 20-h transfection, A549 cells were harvested or the transfection medium was replaced by medium without transfection reagent, and cells were harvested 40 or 64 h after the start of transfection. In combination treatment experiments, etoposide (Sigma, St. Louis, MO) was added after the medium change at t = 20 h at a final concentration of 1.5 μg/ml. PBMCs were plated immediately before transfection in 24-well tissue culture plates. In contrast to the treatment of A549 cells, the transfection medium for PBMCs was not replaced before harvest of the cells 64 h after the start of transfection.

Real-Time PCR. Total RNA was isolated from cells by using the RNasey Mini Kit (QIAGEN Inc., Basel, Switzerland). For cDNA synthesis, 0.5 μg extracted RNA/sample and Taqman Reverse Transcription Reagents (including random hexamers) were used, as described in the user’s manual of the ABI Prism 7700 Sequence Detection System (Perkin-Elmer Applied Biosystems, Foster City, CA). This system was used for real-time monitoring of PCR amplification of the cDNA following the Taqman Universal PCR Master Mix protocol (Perkin-Elmer; Refs. 17–19). The amplification of survivin cDNA was performed using two sets of primers and Taqman probes referred to as S2 and S3. S2 consisted of 5′-AGTGAAGAGGAAGAAGGCA-3′ as a forward primer, 5′-ATTACAATGGGAAGGCTCTGC-3′ as a reverse primer, and 5′-CCGCAACAAAAGCTGTCAGCTCTAGCA-3′ as the Taqman probe. S3 consisted of 5′-ATGGTGCCCCGACGT-3′ as a forward primer, 5′-AATTAGTAGAGATCGGTGGTCTTT-3′ as a reverse primer, and 5′-CCCGCTGTGCGACGCCCCTTTC-3′ as the Taqman probe. The region in which S2 interacts with the survivin cDNA is distinct from the sites where the antisense oligonucleotides hybridize on the mRNA; therefore, this set of primers and probe was used to screen for the most potent antisense oligonucleotide. Because the sequence of the survivin gene amplified by S2 is identical to part of the EPR-1 gene, in principle S2 may also amplify the EPR-1 cDNA. Therefore, S3, which amplifies a region of the survivin gene not shared with the EPR-1 gene, was used in addition to confirm the data obtained with S2 for the most potent antisense oligonucleotide.

The survivin probes were labeled at the 5′ end with the reporter molecule 6-carboxy-fluorescein and labeled at the 3′ end with the quencher molecule 6-carboxy-tetramethylrhodamine. Relative quantification of gene expression was performed as described in the above-mentioned user’s manual, using rRNA as internal standard. Briefly, ribosomal cDNA was amplified separately as indicated in the Taqman RNA Control Reagents protocol (Perkin-Elmer). The comparative Ct (cycle threshold) method was used for relative quantification of survivin mRNA using S2, after confirming that survivin cDNA and ribosomal cDNA were amplified with the same efficiency. Using S3, survivin cDNA was not amplified with the same efficiency as ribosomal cDNA; therefore, the calibration curve method was used for relative quantification of survivin mRNA.

Measurement of Cell Growth and Viability. Growth inhibition of A549 cells was determined by use of the colorimetric MTT cell viability/proliferation assay as described previously (20, 21). Cells were transfected for 20 h at 37°C and incubated for another 44 h at 37°C. Subsequently, 10 μl of MIT reagent (Sigma; 10 mg/ml) was added and allowed to react for 1.5 h at 37°C before the addition of solubilization reagent [100 μl of 20% SDS in 50% dimethyl formamide and 50% H2O (pH 4.7); pH adjusting solution, 80% acetate and 20% 1 M HCl]. Substrate cleavage was monitored at 570 nm by use of a SPECTRAMax 340 microplate reader and analyzed using SOFTmax PRO software (Molecular Devices). To confirm that substrate cleavage was due to caspase activity, extracts were incubated in the presence of 10 μM of the caspase-3-specific inhibitor DEVD-CHO (Bachem, Dübendorf, Switzerland) and incubated at 37°C. Subsequently, substrate cleavage was monitored at 405 nm using a SPECTRAmax 340 microplate reader and analyzed using SOFTmax PRO software (Molecular Devices). To confirm the corresponding data, extracts were incubated in the presence of 10 μM of the caspase-3-specific inhibitor DEVD-CHO (Bachem) for 30 min at 37°C, before the addition of substrate. The value (in arbitrary absorbance units) of the absorbance signal of the inhibited sample was subtracted from that of the noninhibited sample.

Nuclear Staining. Detached cells harvested from the culture medium by centrifugation at 1000 rpm (170 × g) for 5 min were pooled with attached cells collected in a similar way after trypsinization. Cells were pelletted, resus-
The coding strand of the survivin gene shows extensive complementarity to the EPR-1 gene (11), and S2 amplifies a region that is identical in both genes. Therefore S2 is also likely to amplify EPR-1 cDNA, which may lead to a false positive survivin signal in cases where only EPR-1 is expressed or to an underestimation of survivin down-regulation on antisense treatment if both genes are coexpressed. The expression of both genes, however, is reported to be mutually exclusive (2, 3), and the use of antisense oligonucleotide 4003 strongly reduced the survivin mRNA signal measured by real-time PCR. This suggests that the S2-amplified transcript corresponds to the survivin mRNA and not the EPR-1 mRNA. To confirm the data obtained with S2, the effect of the most potent antisense oligonucleotide, 4003, at a dose of 600 nM was also analyzed using the S3 primers and probe set, which amplifies a region unique to survivin and not shared with EPR-1. As shown in the inset in Fig. 1B, the data obtained with S3 confirm the ability of 4003 to specifically down-regulate survivin mRNA expression in A549 cells.

**Inhibition of Cell Growth by Oligonucleotide 4003.** To analyze the biological effect associated with the down-regulation of survivin expression, the growth of A549 cells on treatment with oligonucleotide 4003 was investigated by use of the MTT assay. As shown in Fig. 2A, 64 h after the start of transfection, oligonucleotide 4003 had reduced A549 cell growth dose dependently, with an IC50 of 300 nM. The unspecific growth-inhibitory effect of the mismatch control oligonucleotide was comparatively low. Oligonucleotide 4003 induced death in A549 cells, as revealed by detachment from the culture surface (Fig. 2, B–D).

**Induction of Apoptosis by Oligonucleotide 4003.** Having demonstrated that down-regulation of survivin expression reduced the viability of A549 cells, we analyzed whether cell death was due to the...
induction of apoptosis. As shown in Fig. 3, lysates from oligonucleotide 4003-treated cells revealed increased caspase-3-like protease activity compared with untreated cells. Moreover, on treatment with oligonucleotide 4003, cells contained nuclei with condensed and fragmented chromatin, providing further evidence for the induction of apoptosis as a consequence of survivin antisense treatment (inset in Fig. 3). In cultures treated with the mismatch control oligonucleotide, very few cells showed signs of apoptosis.

To verify that the induction of apoptosis in A549 cells was due to the down-regulation of survivin mRNA, the effect of oligonucleotide 4003 was also examined on PBMCs, which represent a normal cell population reported to lack survivin expression, as analyzed by Northern blotting (2). In agreement with this finding, we found PBMCs to express marginal levels of survivin mRNA as analyzed by real-time PCR using the survivin-specific S3 primers and probe set (inset in Fig. 4A). Trypan blue exclusion assays revealed that PBMCs, in contrast to A549 cells, were not susceptible to death induction by oligonucleotide 4003 (Fig. 4A).

Oligonucleotide 4003 Sensitizes A549 Cells to Etoposide. To investigate whether down-regulation of survivin expression has the potential to sensitize A549 cells to chemotherapy, a combination treatment with antisense oligonucleotide 4003 and etoposide was performed. Fig. 4B shows that in cultures treated with a combination of 600 nm antisense oligonucleotide and 1.5 μg/ml etoposide, approximately 85% of the cells showed signs of cell death 64 h after the start of transfection. Compared to treatments with either oligonucleotide or etoposide alone, this represents about twice or even six times the number of dead cells, respectively. Similarly, nuclear fragmentation and condensation were also increased in combination-treated cells compared with single-agent treated cells (data not shown).

Discussion

The IAP survivin deserves attention as a target for cancer therapy due to its differential expression in tumors versus normal tissues. It is expressed during embryonal development, lacks expression in terminally differentiated adult tissues, and becomes reexpressed in transformed cell lines and a variety of human tumors, with highest levels being found in breast and lung cancer (1). The expression of survivin in tumors is correlated with drug resistance and/or shorter survival of patients with non-small cell lung cancer (5), colorectal cancer (7), and neuroblastoma (9). Despite recognition that survivin represents an attractive target for cancer therapy (2–10), mainly survivin antisense cDNA fragments (11, 12) and very recently also antisense oligonucleotides (13) have been used to elucidate the role of survivin during cell division and apoptosis. In the present study, we describe an antisense oligonucleotide approach to down-regulate survivin expression in the lung adenocarcinoma cell line A549 and demonstrate its ability to induce apoptosis and sensitize tumor cells to chemotherapy.

Most antisense approaches rely on empirical targeting of oligonucleotides against the translation initiation site of mRNAs, where the ATG start codon lies. The rationale for choosing this site is that it likely presents in single-stranded conformation and thus should be accessible to antisense oligonucleotides. In a previous bcl-2 antisense study, we have shown that oligonucleotides hybridizing to the coding region of the mRNA may offer additional targeting options that are equally or even more effective (20). To predict putative binding sites likely to present in single-stranded confirmation, the RNAView program (14) was used as described previously (15). One of the phosphorothioate antisense oligonucleotides developed in the present study targets the ATG start codon site (4018), whereas the others were designed to hybridize to sequences located in the coding region. From these oligonucleotides, 4003 most efficiently down-regulated the survivin mRNA level in A549 cells, achieving its maximum effect at a
concentration of 400 nm, at which mRNA was down-regulated by 70%.

Cells harboring multiple genetic alterations are normally eliminated by apoptosis. For survival, they depend on the overexpression of antiapoptotic molecules such as bcl-2, bcl-xL, or survivin, and down-regulation of these proteins is likely to reduce the apoptotic potential in cells. Oligonucleotide 4003 indeed induced a strong growth-inhibitory effect and apoptosis in A549 cells in the absence of any further cytotoxic stimulus. Similar results were obtained with various other tumor cell lines (data not shown). This observation is in agreement with the findings of others (4) describing the necessity of interaction between survivin and microtubules of the mitotic spindle apparatus to prevent a default induction of apoptosis at the G2-M phase of the cell cycle. This mode of action may constitute a safeguard mechanism against the proliferation of cells with unwanted properties. Whether down-regulation of survivin expression sensitizes cells to apoptosis only when passing the G2-M checkpoint remains to be investigated. The specificity of our survivin antisense approach was further corroborated by the finding that oligonucleotide 4003 did not induce death in normal PBMCs. In these cells, survivin mRNA was hardly detectable using real-time PCR, in agreement with the reported lack of survivin expression in PBMCs (2).

Two major apoptosis signaling pathways are known: (a) the mitochondrion pathway; and (b) the death receptor pathway. The antiapoptotic proteins bcl-2 and bcl-xL block the apoptotic event of mitochondrial cytochrome c release into the cytosol and have been shown to inhibit mainly the first of these two pathways. We have previously shown that in lung cancer cells, antisense oligonucleotides down-regulating bcl-2 or bcl-xL expression induce apoptosis and synergistically interact with chemotherapy (20, 21, 23). Because survivin directly blocks the processing and activation of effector caspase-3 and -7, which act at a common downstream part of the two major apoptosis pathways, the survivin antisense approach has the potential to facilitate apoptosis through both pathways. Using oligonucleotide 4003 in combination with etoposide, which triggers cell death via mitochondrial pathway, the death receptor pathway (and (b) the antiapoptosis gene, survivin, expressed in cancer and lymphoma. Nat. Med., 3: 917–921. 1997.


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