**In Vitro Characterization of Radiolabeled Monoclonal Antibodies Specific for the Extracellular Domain of Prostate-specific Membrane Antigen**

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**ABSTRACT**

Prostate-specific membrane antigen (PSMA) is a well-characterized cell surface antigen expressed by virtually all prostate cancers (PCAs). PSMA has been successfully targeted in *in vivo* with *111*In-labeled 7E11 monoclonal antibody (mAb; ProstaScint; Cytogen, Princeton, NJ), which binds to an intracellular epitope of PSMA. This work reports the *in vitro* characterization of three recently developed mAbs that bind the extracellular domain of PSMA (PSMAext). Murine mAbs J415, J533, and J591, and 7E11 were radiolabeled with *111*In and evaluated in competitive and saturation binding studies with substrates derived from LNCaP cells. J415 and J591 were conjugated to 1,4,7,10-tetraazacyclododecane-N,N’,N”,N’’-tetraacetic acid labeled with *111*In. The uptake and cellular processing of these antibodies were evaluated in viable LNCaP cells. All four mAbs could be labeled with *111*In up to a specific activity of 350 MBq/mg with no or little apparent loss of immunoreactivity. Competition assays revealed that J415 and J591 compete for binding to PSMAext antigen, J533 bound to a region close to the J591 binding epitope, but J533 did not interfere with J415 binding to PSMA. mAb 7E11 did not inhibit the binding of J415, J533, or J591 (or vice versa), consistent with earlier work that these latter mAbs bind PSMAext whereas 7E11 binds the intracellular domain of PSMA. Saturation binding studies demonstrated that J415 and J591 bound with a similar affinity (Kd 1.76 and 1.83 nM), whereas J533 had a lower affinity (Kd 18 nM). In parallel studies, all four mAbs bound to a similar number of PSMA sites expressed by permeabilized cells (1,000,000–1,300,000 sites/cell). In parallel studies performed with viable LNCaP cells, J415, J533, and J591 bound to a similar number of PSMA sites (i.e., 600,000–800,000 sites/cell), whereas 7E11 bound only to a subpopulation of the available PSMA sites (95,000 sites/cell). This apparent binding of 7E11 to viable cells can be accounted for by a 5–7% subpopulation of permeabilized cells produced when the cells were trypsinized and suspended. Up to five DOTA chelates could be bound to either J415 or J591 without compromising immunoreactivity. A comparison of the cellular uptake and metabolic processing of the *111*In- and *131*I-labeled antibodies showed a rapid elimination of *111*In from the cell and a high retention of *131*I. All four mAbs recognized and bound to similar numbers of PSMA sites expressed by ruptured LNCaP cells (i.e., the exposed intracellular and extracellular domains of PSMA). By comparison to J415 and J591, J533 had a lower binding affinity. Both J415 and J591 recognized and bound to the same high number of PSMA sites expressed by intact LNCaP cells. By contrast, 7E11 bound to fewer sites expressed by intact LNCaP cells (i.e., the exposed extracellular domain of PSMA). Both J415 and J591 are promising mAbs for the targeting of viable PSMA-expressing tissue with diagnostic and therapeutic metallic radionuclides.

**INTRODUCTION**

PCA is the most frequently diagnosed cancer and the second most common cause of cancer mortality in United States males (1). Many groups have studied mAbs for *in vivo* diagnosis and therapy of PCA (3–8), but the only successful application to date has been the targeting of PSMA for *in vivo* imaging. PSMA is a type II membrane protein that is expressed by virtually all PCAs (9, 10). Unlike other prostate-related antigens, such as prostate-specific antigen, prostatic acid phosphatase, and prostate secretory protein, PSMA is an integral membrane protein, and therefore, it is not appreciably released into the circulation. PSMA expression has been shown to be up-regulated in both poorly differentiated (11), advanced PCAs (9) and after androgen-deprivation therapy (12). Interestingly, PSMA is expressed on the tumor vascular endothelium of other carcinomas and sarcomas (13, 14) but not on normal vascular endothelium, making it also potentially useful as an antibody-mediated diagnostic and therapeutic target across the full spectrum of solid tumors.

Currently, an *111*In-labeled form of the 7E11 murine mAb is approved by the Food and Drug Administration (Prostascint) for the clinical detection of recurrent and metastatic prostate cancer in soft tissue (15). 7E11 mAb binds to the intracellular portion (NH2 terminus) of the PSMA antigen and, as such, does not bind viable cells (13, 14). It is believed that successful imaging with Prostascint results from mAb binding to antigen expressed in dead or dying cells within some tumor sites (16, 17). Early clinical trials using *99*Y-labeled 7E11 resulted in no objective or biochemical (prostate-specific antigen) remissions (7).

Recently, a series of mAbs to PSMAext has been characterized and reported (13, 14, 18). In this current study, we report on the *in vitro* evaluation of radiolabeled forms of these antibodies against PSMAext and the selection of interesting candidates for *in vivo* evaluation of their diagnostic and therapeutic potential.

**MATERIALS AND METHODS**

Murine mAbs J415, J533, and J591 were produced as described earlier (13). Purified 7E11 was generously provided by Dr. Gerald P. Murphy (Pacific Northwest Research Foundation, Seattle, WA). *131*I and *111*In were purchased from Norton International (Kanata, Ontario, Canada). *99*Y was purchased from New England Nuclear (Boston, MA). DOTA was purchased from Macrocyclies, Inc. (Richardson, TX).

LNCaP cells (American Type Culture Collection, Rockville, MD) were grown in RPMI 1640, supplemented with 10% FCS, at a temperature of 37°C in an environment containing 5% CO2. Prior to use, the cells were trypsinized, counted, and suspended in serum-free medium. LNCaP cells were permeabilized by adding methanol at −80°C to the cells. The cells were maintained at −20°C for 20 min before the methanol was removed, and the cells were rehydrated by washing four times with PBS (with 5 mM CaCl2 and 5 mM MgCl2) over 20 min.

Cell membranes were prepared by lysing the cells with a Polytron in a...
hypotonic buffer [1 mm Na₂CO₃ (pH 7.4) with 1 mm EDTA and 1 mm phenylmethylsulfonyl fluoride]. Large fragments were removed by centrifuging at 2000 × g. The supernatant was centrifuged at 150,000 × g for 2 h, and the pelleted membranes were resuspended in PBS, aliquoted, and frozen at −70°C until required.

Radioiodination

The four murine mAbs were radioiodinated with 131I using the Iodogen (1,3,4,6-tetracloro-3a,6a-diphenylglycoluril) method (19). Briefly, 10-ml glass tubes were coated with 50 μg of Iodogen by adding 100 μl of a 0.5 mg/ml solution of Iodogen (Pierce, Rockford, IL) in chloroform. The chloroform was removed by blowing a gentle stream of sterile nitrogen into the tube for 30 min before the tubes were sealed and stored in the dark. The iodination reaction was initiated by adding between 4 and 40 MBq of 131I (0.01 M NaOH) to 0.08 mg of mAb in 0.1 ml of ice-cold PBS. This reaction mixture was allowed to react for 5 min on ice before being loaded onto a 10-ml Biogel-P6 column (Bio-Rad Laboratories, Hercules, CA) equilibrated with 1% BSA in PBS. Once the reaction mixture was loaded onto the column, it was washed with 2 ml of 1% BSA PBS before the main 131I-labeled mAb fraction was eluted with 2 ml of 1% BSA PBS. The amount of free iodine in the 131I-labeled mAb preparations was evaluated using instant TLC with a silica gel impregnated glass fiber support and a mobile phase of isotonic saline. Briefly, a portion of the 131I-labeled mAb was spotted on a 10-cm ITLC-SG strip (Gelman Sciences, Ann Arbor, MI) and developed in isotonic saline. Once the solvent front had reached the end of the strip, it was removed from the solvent and cut at an Rf of 0.5. The two portions were assayed for radioactivity, and the radiochemical purity determined using the following equation: radiochemical purity = activity between Rf 0 and 0.5/total activity in strip.

Antibody Conjugation

J415 and J591 antibodies were modified with DOTA by an analogous method to that used by Lewis et al. (20). This method uses the direct coupling of one of the four carboxylic acid groups of DOTA to the primary amines present in the protein structure (Fig. 1). Twenty-five mg of antibody were concentrated in a Am 30,000 Microsep centrifugal concentrator (Pall Filtron, Northborough, MA) and washed with 5 × 4 ml of 1% DTPA (pH 5.0) over a period of 24 h. The antibody buffer was then changed to 0.1 m phosphate (pH 7.0) using the same centrifugal technique. An active ester of DOTA was created by dissolving 146 mg of DOTA (0.361 mmol) and 36 mg of N-hydroxysuccinimide (0.313 mmol) in 2 ml of water and adjusting the pH to 7.3 with NaOH, prior to the addition of 10 mg of 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide. This reaction mixture was cooled on ice for 1 h before being added to the J591 solution. The resultant DOTA-antibody conjugate was separated from the excess DOTA and other reactants by repeated washing with 0.3 m NH₂OAc (20 × 4 ml) and centrifugal concentration.

1) Preparation of active ester

2) Conjugation

Fig. 1. Two-step conjugation of DOTA to free amines displayed by either J415 or J591. The first step used N-hydroxysuccinimide and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide to create an active ester with DOTA. In the second step, the unpurified active ester is allowed to react with the monoclonal antibody.

Assay of Binding Site Number

DOTA-J591 conjugate concentration was assayed by determining the UV absorption at 280 nm. Two 50-μl aliquots of DOTA-J591 were mixed with either 20 or 40 μl of a 1:30 m/solution of InCl₃ (0.01 M HCl) spiked with a tracer amount of 111In. The mixture was incubated at 37°C for 16 h and then analyzed by ITLC, using a silica gel-impregnated glass fiber 10-cm strip (ITLC-SG, pH 6.0). Antibody-bound activity remains at the origin, and free In³⁺ moves with the solvent front as an [In-DTPA]⁻ complex. The relative amounts of In³⁺ and In-DOTA-J591 were determined by cutting the ITLC strip at Rf 0.5 and counting the two halves with a Na(Tl)I detector. The number of binding sites was calculated by considering the molar reaction ratio between In and DOTA-mu-J591, and the observed ratio of 111In and 111In-DOTA-mu-J591 was detected.

111In and 90Y Labeling of DOTA Conjugate

Radio-labeling of DOTA-J591 with 111In was achieved by adding the radiouclide (in dilute HCl) to the ammonium acetate-buffered DOTA-J591. Briefly, a mixture composed of 20 μl of 111InCl₃ (300 MBq), 0.01 M HCl and 400 μl of DOTA-J591 (4 mg/ml; 0.3 m NH₂OAc, pH 7) was allowed to react at 37°C for 20 min. The reaction mixture was then separated on a 20-ml Biogel-P6 column equilibrated with 4 × 10 ml of sterile 1% HSA in PBS. After the reaction mixture was loaded onto the column, it was washed with an additional 5 ml of 1% HSA PBS before the main 111In-DOTA-J591 fraction was eluted with 3 ml of 1% HSA PBS. A similar procedure was used for radio-labeling with 90Y, but an incubation time of 5 min was used, and the labeling mixture included 50 mm ascorbic acid.

Free 111In in the radiolabeled DOTA-J591 preparations was determined using the ITLC method with a silica gel-impregnated glass fiber support and a mobile phase of 1% DTPA (pH 5.5). A portion of the radiolabeled DOTA-J591 was spotted on a 10-cm ITLC-SG strip and developed in 1% DTPA (pH 5.5). Once the solvent front had reached the end of the strip, it was removed from the solvent and cut at a Rf of 0.5. The two portions were assayed for radioactivity, and the radiochemical purity was determined using the equation described earlier.

Chelate Stability Studies

111In-labeled DOTA-J591 and DTPA-7E11 were mixed with an equal volume of 50 mM DTPA and maintained at 37°C. Periodically, samples were removed and spotted on a 10-cm ITLC-SG strip and developed in 0.9% NaCl. Once the solvent front had reached the end of the strip, it was removed from the solvent and cut at a Rf of 0.5. The two portions were assayed for radioactivity, and the amount of intact chelate was determined using the equation described earlier.

Binding Studies

Immunoreactivity. The immunoreactivity of the 111In- and 131I-labeled mAb preparations was assessed by the method of Lindmo et al. (21), which extrapolates the binding of the radiolabeled antibody at an infinite excess antigen. Briefly, six test solutions were prepared (in duplicate) and contained 20,000 cpm of the radioiodinated antibody, and increasing amounts of membranes were prepared from LNCaP cells in a total test volume of 250 μl of PBS (0.2% BSA, pH 7.4). The solutions were incubated at 37°C for 45 min prior to being filtered through a glass membrane filter and washed with ice-cold 10 mM Tris-0.9% NaCl buffer. Filters were counted in a gamma counter with standards representing the total radioactivity added. Data were then plotted as the reciprocal of the substrate concentration (X axis) against the reciprocal of the fraction bound (Y axis). The data were then fitted according to a least squares linear regression method using Origin software (Microcal Software, Inc., Northampton, MA). The Y intercept gave the reciprocal of the immunoreactive fraction. A similar method using intact or permeated LNCaP cells and centrifugal isolation of the cells gave the same results.

Competitive Binding Studies. Competitive binding studies were performed with each of the radioiodinated antibodies and the four unlabeled antibodies using either LNCaP tumor sections or membranes derived from LNCaP tumors. Acetone fixed and frozen 10-μm tumor sections were soaked in Tris buffer [170 mM (pH 7.4), with 2 mM CaCl₂ and 5 mM KCl] for 15 min and then washed with Tris buffer (170 mM, pH 7.4). The sections were then
incubated with the radiiodinated antibodies in the presence of 100 nm concentrations of each of the unmodified mAbs for 1 h at 4°C. Sections were washed three times with PBS (0.2% BSA) and once with Tris buffer (170 mM, pH 7.4) prior to being fixed with acetone and exposed with BioMax film (Kodak). The assay using the membranes typically used 50 µg of membranes, 10 fmol of iodinated antibody, and amounts of competing antibody from 0.25 fmol to 25 pmol in a 250-µl volume of PBS (0.2% BSA). Membranes were isolated as described above, and data were analyzed by a least squares regression method and Origin software (Microcal Software, Inc.) was used to determine the IC50.

Saturation Binding Studies. Saturation binding studies were performed with each of the radiolabeled antibodies using substrates of intact and permeated LNCaP cells. Briefly, 10 test solutions were prepared (in duplicate) and they contained increasing amounts of the radiiodinated antibodies, 500,000 LNCaP cells in a total volume of 250 µl of PBS (0.2% BSA, pH 7.4). The solutions were incubated at 4°C for 1 h and centrifuged and washed twice with ice-cold PBS (0.2% BSA). For each concentration of radiolabeled antibody, nonspecific binding was determined in the presence of 100 nm of the unmodified antibody. The data were analyzed with a least squares regression method (Origin; Microcal Software, Inc.) to determine the Kd and Bmax values, and a Scatchard transformation was performed.

Internalization and Cellular Processing of J415 and J591

LNCaP cells were plated in 8-cm² Petri dishes and allowed to grow until confluent. One µCi of either the 131I- or 111In-labeled forms of J415, J591, or 7E11 (0.1–0.2 µg) were added to cells and allowed to incubate for 1 h. The medium was then removed, and the cells were washed once with fresh media. One ml of fresh medium was added, and the cells were incubated for up to 2 days at 37°C. Triplicate samples were periodically removed, and the medium was isolated. Surface bound activity was stripped and collected with an ice-cold acid wash (100 mM acetic acid, 100 mM glycine, pH 3.0). The cells were then treated with 1 ml of a 1% solution of Triton X-100 (containing 5 µg/ml each of antipain, pepstatin, and leupeptin as well as 1 mM phenylmethyisulfonyl fluoride) and kept at on ice for 20 min. The resultant suspension was then centrifuged, and the three samples were counted with a gamma counter. The medium and supernatants were also analyzed by ITLC and size exclusion HPLC to determine the amounts of free iodide produced or the size of the radioactive species created.

RESULTS

Radiolabeling and Quality Control. The radiiodination yield for the four mAbs was typically 70–80%, and the amounts of free iodine in the purified mAbs was <0.3%. Specific activities of 350 MBq/mg were routinely achieved. The immunoreactivities of the 131I-labeled mAbs were determined by extrapolating the binding of a fixed amount of 131I-labeled mAb to an infinite amount of PSMA (Lindmo method; Fig. 2). This method gave immunoreactivities of >75% for all mAbs tested. When labeling conditions were increased to produce specific activities >350 MBq/mg, the immunoreactivity was compromised.

An average of five DOTA molecules could be randomly conjugated to J591 and J415, with little apparent loss of immunoreactivity. Conjugation of an average of eight DOTA molecules to J591 resulted in a 20% reduction in immunoreactivity. A 90% incorporation of 111In could be achieved within 15 min. A 90% incorporation of 90Y could be achieved within 5 min. Using the DOTA-J591 conjugate with an average of five DOTA molecules attached, specific activities of 280 MBq 111In/mg DOTA-J591 and 360 MBq 90Y/mg DOTA-J591 were achieved.

111In-Chelate Stability Studies. A direct comparison of the chelate stability of 111In-DTPA-7E11 and 111In-DOTA-J591 showed that 111In was lost from DTPA-7E11 with an apparent half-life of 11 h, whereas the DOTA chelate had an apparent half-life exceeding 1000 h (Fig. 3).

Competitive Binding Studies: Membranes. Radiolabeled J415 could be displaced from binding to LNCaP cell membranes by both J415 and J591 but not J533 (Fig. 4A). The J415 mAb had an mean IC50 of 1.5 nm (±0.9; n = 6), and J591 had a mean IC50 of 6.6 nm (±4.5; n = 6). Similarly, 131I-labeled J353 could be displaced by J533 and J591 but not by J415 (Fig. 4B). In these studies, J533 had a mean IC50 of 2.3 nm (±1.5; n = 3), and J591 had a mean IC50 of 1.7 nm (±1.3; n = 3). Finally, 131I-labeled J591 could be displaced by J415, J533, and J591 (Fig. 4C). The observed IC50s were 1.3 nm (±0.9; n = 6) for J415, 7.6 nm (±5.5; n = 6) for J533, and 3.1 nm (±1.5; n = 6) for J591. The 7E11 mAb did not inhibit the binding of J415, J533, or J591 (or vice versa). These data are consistent with earlier data (11) that J415, J533, or J591 bind to the extracellular domain of PSMA, whereas 7E11 binds to the intracellular domain of PSMA (13).

Saturation Binding Studies. The saturation binding curves generated were characteristic of high affinity binding of an antibody to a single class of antigen. These studies, performed with intact LNCaP cells (Fig. 5), demonstrated that J415 and J591 bound with a similar
affinity ($K_d$ 1.76 ± 0.69 and 1.83 ± 1.21 nM), whereas J533 had a lower affinity ($K_d$ 18 ± 5 nM). In parallel studies, all four mAbs bound to a similar number of PSMA sites expressed by permeabilized cells (1,000,000–1,300,000 sites/cell). In parallel studies performed with viable LNCaP cells, J415, J533, and J591 bound to a similar number of PSMA sites (i.e., 600,000–800,000 sites/cell). In contrast, 7E11 specifically bound to only 10–15% of the PSMA sites expressed by apparently intact LNCaP Cells ($K_d$, 6.69 nM); but when the cells were deliberately ruptured (Fig. 6), 7E11 bound to a similar number of antigen sites as the other three mAbs. In parallel studies, using 121-I-labeled J591, permeabilized cells expressed about twice the amount of PSMA as intact LNCaP cells, suggesting that not all available PSMA is simultaneously expressed on the cell surface.

Internalization and Cellular Processing of J415 and J591. Both 121-I-labeled J415 and J591 demonstrated a poor cellular retention of...
radioactivity (Fig. 7). For both mAbs, a biexponential curve fit of the data showed that ~10% of the radioactivity was released from the cells with an apparent half-life of 1 h, and the remaining 90% was released into the medium with an apparent half-lives of 31 and 38 h for J415 and J591, respectively. In parallel studies, 131I-labeled J415 consistently showed a faster release of radioactivity than 131I-labeled J591. Little or no activity (<1%) was associated with the Triton X-100 (or NaOH) insoluble cell pellet. Analysis of the Triton X-100 soluble fractions indicated that there were no appreciable amounts of free 131I present (<1%). HPLC and TLC analysis of the culture medium showed that a large iodinated species, which corresponded to the same size as the intact mAbs, was being released from the cells, but this never amounted to >10% of the total activity, and after 4–6 h, no further release of this radioactive species was observed. The predominant metabolite of the iodinated mAbs found in the cell medium had the same HPLC and TLC elution profile as free 131I present (<1%). HPLC and TLC elution profile of the intact mAbs, was being released from the cells, but this never amounted to >10% of the total activity, and after 4–6 h, no further release of this radioactive species was observed. For both J415 and J591, two main 111In-labeled metabolites were observed in the medium. Analysis of the cell-associated radioactivity (Fig. 8) demonstrated the rapid formation of two groups of metabolites (based on molecular size). One group of metabolites achieved a maximum concentration after 1–2 h, after which it began to steadily decline. The second group of metabolites, however, demonstrated an ever-increasing intracellular concentration. This second metabolite did not behave the same as 111In3+ (HPLC or TLC), but rather it had a similar molecular weight as an 111In-DOTA or an 111In-DOTA-amino acid fragment. The first metabolite had a molecular weight between that of the intact mAb and the second metabolite (Mw ~10,000–30,000).

The uptake rates of 111In-labeled J415, J591, and 7E11 by LNCaP cells showed a similar initial uptake rate for J415 and J591, which was 10–20 times faster than that of 7E11 (Fig. 9). However, by 4 h after the addition of the radioactivity, the cells treated with 111In-DOTA-J591 have incorporated and retained more activity than those treated with 111In-DOTA-J415.
e.g., stay in circulation for prolonged periods of time in the presence of competing ligand. This is in agreement with other studies (20, 26) and could be modified with sufficient DOTA to produce high specific activities.111 In- and 90 Y-labeled mAbs for both antigen binding domain. High specific activities are often required for noreactivity attributable to the presence of a lysine residue in the when this type of random DOTA coupling results in loss of immunoreactivity attributable to the presence of a lysine residue in the antigen binding domain. High specific activities are often required for accurate mAb characterization and particularly when large amounts of the radiotherapeutic agent are administered to a patient. J451 and J591 could be modified with sufficient DOTA to produce high specific activity111In- and 90Y-labeled mAbs for both in vitro binding studies and eventual in vivo studies. Early approaches to labeling mAbs with radiometals used DTPA, which in its dicyclic anhydride form could be conveniently coupled to mAbs (22). Unfortunately, this simple coupling chemistry produced a more labile chelate than bifunctional forms of the same unconjugated DTPA chelator (23). Macrocyclic chelators have shown even higher kinetic stability (24), but they are even more time consuming to chemically synthesize (25). DOTA can be coupled directly to mAbs using simple chemistry and commercially available materials (20).

The reaction kinetics for 111In and DOTA are longer than for DTPA, but an incubation period of 15 min can give high labeling yields. The DOTA chelator was immensely superior to DTPA in its ability to tightly chelate 111In in the presence of an excess of competing ligand. This is in agreement with other studies (20, 26) and underlies the importance of using stable chelates with mAbs that can stay in circulation for prolonged periods of time in the presence of competing ligands (e.g., transferrin). The higher stability of the 111In-labeled DOTA complex relative to the 111In-labeled DTPA complex also applies for the 90Y complex (20, 26) and is an important prerequisite for radiolabeled mAbs used for either diagnosis or therapy because optimal tumor:non-tumor ratios are often achieved after 2–4 days. Because 111In-DOTA-J591 is stable to DTPA competition, it enables nonspecifically bound 111In to be removed by challenging with DTPA and a simple column separation to yield a highly pure radiopharmaceutical.

High binding affinity between the mAb and the target antigen is another prerequisite to in vivo targeting of tumor antigens. The binding studies with the iodinated mAbs showed that two of three of these mAbs against PSMA (i.e., J415 and J591) and 7E11 have similar nanomolar binding affinities. The use of intact and ruptured cells showed clearly that 131I-labeled 7E11 binds to the intracellular domain of PMSA, consistent with other results (13, 17). There was some binding of 7E11 to “intact” LNCaP cells, but that could be explained by the presence of a small population of cells ruptured during the trypsinization of the cells from the cell culture flasks and subsequent handling during resuspension. Because J591 recognized and specifically bound to twice the number of PSMA sites in permeabilized cells as opposed to intact cells, this suggests that only 50% of all cellular PSMA is exposed extracellularly. Also, 10–15% binding of 7E11 could be explained by the presence of a population of 5–7% of permeabilized cell in the “intact” cell preparation. This was confirmed in other studies that examined the binding and cellular uptake of the mAbs with plated LNCaP cells and showed a 7E11 uptake of 3–4% that of J415 and J591. This quantitative difference seen might explain why one group claims that the 7E11 binds to apparently intact LNCaP cells (27), whereas other groups report no such binding (13, 17, 18). Additionally, the Barren study (27) used cells that were scrapped from the monolayer, a procedure known to create significant cell rupture.

Effective systemic targeting of tumors has been achieved with both iodinated and metallochelated antibodies. A significant factor in selecting the two approaches is in whether the antibody is internalized. In the case of an internalizing antibody, directly iodinated antibodies are metabolized within the cell, and the main metabolites of iodide and iodotyrosine are then freely released from the cell. Conversely, the metal chelate-labeled antibody is metabolized to leave a chelate-amino acid fragment that is typically not released from the cell and is trapped for further degradation. This effect can produce vastly different residence times for two differently labeled forms of the same antibody (28). In the LNCaP cell model, PSMA is known to be an internalizing surface bound glycoprotein (18). These studies clearly support the notion that metalloabeled mAbs are superior for cells expressing internalizable PSMA and that metabolites from the DOTA-mAb conjugate are not appreciably released from LNCaP cells.

In conclusion, both J415 and J591 have similar nanomolar affinities to PSMA as 7E11. Similarly, these two mAbs are far more readily bound and were internalized by live LNCaP cells than 7E11. The 111In-labeled DOTA conjugates are able to associate more radioactivity with LNCaP cells than the comparable iodinated forms. The 111In-labeled DOTA conjugates are also more stable to loss of 111In than DTPA-7E11. These findings make DOTA-J415 and DOTA-J591 attractive candidates for further evaluation as either diagnostic or radiotherapeutic agents in patients with various cancers that express PSMA.

**REFERENCES**


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