Estrogenic and Antiestrogenic Properties of Resveratrol in Mammary Tumor Models

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ABSTRACT

Trans-3,4',5-trihydroxystilbene (resveratrol), a phytoalexin present in grapes and grape products such as wine, has been identified as a chemopreventive agent. Recent studies performed with MCF-7 human breast cancer cells have demonstrated superestrogenic effects with resveratrol. In contrast, studies performed using estrogen receptor-transfected cell lines have shown that resveratrol acts as a mixed agonist/antagonist. The major objective of this study was to characterize the estrogen-modulatory effects of resveratrol in a variety of in vitro and in vivo mammary models. Thus, the effect of resveratrol alone and in combination with 17β-estradiol (E2) was assessed with MCF-7, T47D, LY2, and S30 mammary cancer cell lines. With cells transfected with reporter gene systems, the activation of estrogen response element-luciferase was studied, and using Western blot analysis, the expression of E2-responsive progesterone receptor (PR) and presenlin 2 protein was monitored. Furthermore, the effect of resveratrol on formation of preneoplastic lesions (induced by 7,12-dimethylbenz(a)anthracene) and PR expression (with or without E2) was evaluated with mammary glands of BALB/c mice placed in organ culture. Finally, the effect of p.o. administered resveratrol on N-methyl-N-nitrosourea-induced mammary tumors was studied in female Sprague Dawley rats. As a result, in transient transfection studies with MCF-7 cells, resveratrol showed a weak estrogenic response, but when resveratrol was combined with E2 (1 nM), a clear dose-dependent antagonism was observed. Similar mixed estrogenic/antiestrogenic effects were noted with S30 cells, whereas resveratrol functioned as a pure estrogen antagonist with T47D and LY2 cells. Furthermore, in MCF-7 cells, resveratrol induced PR protein expression, but when resveratrol was combined with E2, expression of PR was suppressed. With T47D cells, resveratrol significantly down-regulated steady-state and E2-induced protein levels of PR. With LY2 and S30 cells, resveratrol down-regulated presenlin 2 protein expression. Using the mouse mammary organ culture model, resveratrol induced PR when administered alone, but expression was suppressed in the presence of E2 (1 nM). Furthermore, resveratrol inhibited the formation of estrogen-dependent preneoplastic ductal lesions induced by 7,12-dimethylbenz(a)anthracene in these mammary glands (IC50 = 3.2 μM) and reduced N-methyl-N-nitrosourea-induced mammary tumorigenesis when administered to female Sprague Dawley rats by gavage. Therefore, in the absence of E2, resveratrol exerts mixed estrogen agonist/antagonist activities in some mammary cancer cell lines, but in the presence of E2, resveratrol functions as an antiestrogen. In rodent models, carcinogen-induced preneoplastic lesions and mammary tumors are inhibited. These data suggest that resveratrol may have beneficial effects if used as a chemopreventive agent for breast cancer.

INTRODUCTION

Despite years of intensive research, breast cancer remains a major cause of death among women in the United States (1). SERMs1 such as tamoxifen, which are used clinically for the treatment of breast cancer, act as estrogen agonists in certain tissues but exhibit antiestrogenic effects in others (2). Data from adjuvant breast cancer trials have shown that tamoxifen not only suppresses the recurrence of breast cancer but also prevents the contralateral occurrence of second primary breast tumors (3). Additional beneficial effects include maintenance of bone density and decreased mortality from coronary heart disease (4, 5). Moreover, a double-blind clinical chemoprevention study with tamoxifen revealed a 49% reduction in the incidence of breast cancer among women with increased risk of developing this disease (6). However, tamoxifen may also promote the development of endometrial cancer.Raloxifene, a related SERM, exhibits antiestrogenic effects in the breast and uterus but estrogenic effects in the bone (7). As a result, a randomized double-blinded trial comparing the effectiveness of tamoxifen with that of raloxifene (STAR) is currently being conducted with postmenopausal women at increased risk for developing breast cancer (8).

To date, no plant-derived SERMs have been evaluated in clinical trials, although a number of herbal remedies such as extracts of black cohosh and red clover are touted for the relief of postmenopausal symptoms (9, 10). The precise constituents of these extracts that might be responsible for such pharmacological effects are not known; however, flavonoids and isoflavonoids such as quercetin and genistein have been reported to display both estrogenic and antiestrogenic effects (11). Genistein is about 1000-fold less potent than E2 in its ability to bind to the ER, but its circulating concentrations in the presence of tamoxifen have been shown to be responsible for such pharmacological effects (12). Interestingly, Asian women who consume a soy-rich diet have about a 6-fold lower risk of developing breast cancer than their Western counterparts (13, 14), and population-based studies have suggested that consumption of a phytoestrogen-rich diet is protective against prostate and bowel cancer, as well as cardiovascular disease (15). Hence, it is possible that these phytoestrogens function as chemopreventive agents, and because human consumption of phytoestrogens is common, plant-derived agents are promising candidates for clinical trials.

Resveratrol, a stilbene found in the diet of humans, was identified as a potential chemopreventive agent (16). Subsequently, the compound was considered a phytoestrogen due to potent estrogenic and even superestrogenic (when combined with E2) properties in MCF-7 mammary cancer cells (17), but additional studies conducted with these cells demonstrated antagonist activity in the presence of E2 (18, 19). With ER-transfected cell lines, resveratrol has been reported to function as a mixed agonist/antagonist (20–22). For example, while...
acting as an agonist with mammary tissues containing ER-α, estrogen antagonist activity was observed with ER-β (22). Using in vivo models, resveratrol has been shown to be an estrogen agonist in stroke-prone spontaneously hypertensive rats, in which it attenuated an increase in systolic blood pressure, enhanced endothelin-dependent vascular relaxation in response to acetylcholine, and prevented ovariectomy-induced decreases in femoral bone strength in a manner similar to E2 (23). In contrast, however, using rat uterotrophic assays, it was shown that high doses of resveratrol antagonized the serum cholesterol-lowering activity of E2 (24), mildly decreased uterine weight, and suppressed the expression of ER-α protein, similar to pure antiestrogens (25). Using a human endometrial cell line, we have recently demonstrated estrogen antagonistic responses with resveratrol (26).

Thus, the estrogen-modulatory potential of resveratrol in mammary tissues is not well delineated. In this study, a variety of cell lines were used to assess the potential of resveratrol to serve as a SERM. In addition, estrogen-dependent MMOC studies were performed, as well as full-term tumorigenesis studies in rats. The results obtained from both in vitro and in vivo studies suggest that resveratrol may serve as a cancer chemopreventive agent in the breast.

MATERIALS AND METHODS

Cell Culture. MCF-7 (ER-α/PR/pS2+) and T47D (ER-α/PR/pS2+) cells were obtained from American Type Culture Collection (Manassas, VA). LY2 cells (ER-α/pS2+) were provided by Dr. Marc E. Lippman (Georgetown University, Washington, D.C.), and S30 cells (ER-α/pS2+) were obtained from Dr. V. C. Jordan (Northwestern University, Chicago, IL). MCF-7, T47D, and LY2 cells were routinely maintained in MEME supplemented with 5% FBS, 100 μg/ml streptomycin, 100 units/ml penicillin, 10 μg/ml insulin, 2 mM L-glutamine, 1 mM sodium pyruvate, and nonessential amino acids at 37°C in a 5% CO2 incubator. S30 cells were maintained in MEME supplemented with 5% DCC-FBS, 100 μg/ml streptomycin, 100 units/ml penicillin, 6 mg/ml insulin, 2 mM glutamateMAX-1, 1 mM sodium pyruvate, and 500 μg/ml G418. All media components and reagents were purchased from Life Technologies, Inc. (Grand Island, NY), with the exception of FBS, which was purchased from Atlanta Biologicals (Atlanta, GA). One week before plating the cells (MCF-7, T47D, and LY2), the medium was changed to a phenol red-free formulation of MEME containing DCC-FBS.

Transfection and Luciferase Assays. The plasmid pUC 18 (provided by Dr. Peter J. Kushner; University of California at San Francisco) is a construct consisting of two copies of Xenopus vitellogenin A2 ERE (GGTCACAGTGACCT) inserted upstream (~331 to ~289) of a minimal thymidine kinase promoter (~109 to +45) linked to the luciferase gene. For transfection assays, cells were grown in estrogen-free media for 7 days and then plated in 12-well plates at a density of 10 × 104 cells/well. Experimental media containing various concentrations of resveratrol and E2 were added 24 h later. Two h after changing to experimental media, the cells were transfected with 2 μg/well lipid-plasmid complex using FUGene 6 reagent (Roche Biochemicals, Indianapolis, IN). Luciferase activity was detected using a Luciferase Assay System (Promega, Madison, WI) approximately 24 h after transfection using a TD-20/20 luminometer (Turner Designs, Sunnyvale, CA). Luciferase induction responses for each treatment group were normalized to β-galactosidase activity and expressed as the means ± SD of three replicate experiments for each treatment group.

Western Blot Analysis for Protein Expression. Cells (50 × 104 cells/dish) were plated in 100-mm2 Petri dishes, and compounds were added 24 h later. After 48 h of incubation, the cells were lysed using a 2× electrophoresis sample buffer [250 mM Tris-HCl (pH 6.8), 4% SDS, 10% glycerol, 0.006% bromophenol blue, and 2% β-mercaptoethanol]. Twenty μg of protein lysate were fractionated using SDS-PAGE on 7.5% polyacrylamide gels. Anti-PR mouse monoclonal antibody was purchased from Lab Vision Corp. (Fremont, CA). Anti-pS2 goat monoclonal antibody was obtained from Santa Cruz Biotechnology (Santa Cruz, CA). Anti-β-actin mouse polyclonal antibody was purchased from Sigma Chemical Co. (St. Louis, MO). The proteins were transferred onto nitrocellulose membranes by electroblotting and incubated with primary antibody (anti-PR, anti-pS2, and anti-β-actin) for 2 h. Secondary antibodies (antimouse or antirabbit) conjugated to horseradish peroxidase and streptavidin-horseradish peroxidase conjugate (Amersham Pharmacia Biotech, Piscataway, NJ) were diluted 1:2000 and incubated with the membrane for 30 min. Proteins were detected using an enhanced chemiluminescence plus detection system according to the manufacturer’s instructions (Amersham Pharmacia Biotech) and quantified using Scion imaging software (Scion Corp., Frederick, MD).

Induction of Atypical Ductal Lesions in MMOC. BALB/c female mice (Charles River, Wilmington, MA), 3–4 weeks of age, were housed 5 mice/polycarbonate cage and maintained in accord with institutional guidelines. The animals were pretreated daily for 7 days with 1 μg of E2 and 1 mg of progesterone by s.c. injections. The organ culture procedure has been described in detail elsewhere (27). Briefly, mice were killed by cervical dislocation, and the thoracic pair of mammary glands were dissected out on silk rafts and incubated for 10 days in Waymouth MB752/1 medium (5 glands/5 ml/dish; Life Technologies, Inc.) at 37°C in a 95% O2 and 5% CO2 environment. The medium was supplemented with 350 μg/ml glutamine, antibiotics (100 units/ml penicillin and 100 μg/ml streptomycin), and growth-promoting hormones (5 μg of insulin, 5 μg of prolactin, 1 ng of E2, and 1 μg of progesterone per milliliter of medium; Sigma Chemical Co.). DMBA (2 μg/ml) was added to the medium between days 3 and 4, and control dishes contained DMSO as vehicle. On day 4, DMBA was removed from the medium by rinsing the glands with fresh medium, and the glands were transferred to new dishes containing carcinogen-free medium. Various concentrations of resveratrol (1–10 μM in DMSO) were included in the incubated media for the first 10 days of culture (10 glands/treatment). After 10 days of incubation, the glands were maintained for an additional 14 days in medium containing only insulin (5 μg/ml). At the end of the incubation period, glands were embedded in paraffin, and 5-μm longitudinal sections were cut. The sections were processed and stained with H&E, and ductal lesions were identified as hyperproliferative ductal epithelium with a ≥3-layer thickness of epithelial cells. Atypical ductal lesions were defined as ducts that were completely occluded with ductal epithelial cells that demonstrated nuclear pleomorphism and variability in size, form, staining intensity, chromatin composition, and nuclear organization. The histological sections of the gland were divided into three regions, and each region was evaluated for the number of normal (single-layer thickness of epithelial cells), hyperplastic (≥3-layers thick), or atypical (highly occluded, with no or very little lumen) areas. The percentage of multiplicity of lesions in control and treated glands was expressed as the mean ± SD, and the IC50 of resveratrol was calculated.

Evaluation of PR Induction in MMOC. As described above, mouse mammary glands were incubated with growth-promoting hormones (insulin and prolactin) either alone or in the presence of resveratrol for 6 days. Glands were not exposed to DMBA, and the hormone withdrawal stage was omitted. The glands were fixed in buffered formalin, and 5-μm histological sections were prepared as described above. The sections were mounted on adhesive-coated slides (Superfrost; Fisher Scientific, Pittsburgh, PA), dried at 60°C overnight, deparaffinized in xylene, dehydrated in alcohol, and washed with PBS. To block nonspecific antibody reactions, sections were treated with 5% dried skim milk for 10 min and then incubated with a primary antibody solution against PR (2 μg/ml; Neomarkers, Fremont, CA) overnight at 0°C to 4°C. The tissues were rinsed in PBS and incubated with biotinylated antirabbit antiamouse-linked antibody (Dako Corp., Carpinteria, CA) for 10 min, followed by a 10-min incubation with peroxidase-conjugated streptavidin (Dako Corp.) and AEC chromogen (BioGenex, San Ramon, CA) in H2O2 substrate for 5 min. The tissues were counterstained with Gill’s hematoxylin (Fisher Scientific, Fair Lawn, NJ), dehydrated through a graded series of alcohol and xylene, and mounted in Permount (Fisher Scientific). Slides were evaluated semiquantitatively for the expression of PR according to the proportion of positively stained cells (28). The cells were scored as follows: −, no staining; +, observable staining; ++, marked staining; and +++, intense staining. The proportion of positively stained cells was calculated using the following formula: percentage of PR+ cells = 100(nn+n−+nn+n−+n+n−+n+n−y/n+n−+n+n−+n+n−+n+n−+n+n−+n+n−+n+n−+n+n−+n+n−+n+n−), where n is number of cells in each category of staining. The percentage of induction of PR in the control and treated glands were represented as the mean ± SD, and the IC50 of resveratrol was calculated.

Induction of Mammary Tumors in Rats. Virgin female Sprague Dawley rats were received from Harlan Sprague Dawley (Indianapolis, IN) at 35 days old.
Fig. 1. Effects of resveratrol on ERE-luciferase activity with transiently transfected breast cancer cells. Panels represent data obtained with MCF-7, S30, T47D, and LY2 cells. Cells were treated with the indicated concentrations of resveratrol alone or in combination with 1 nM E2 as described in “Materials and Methods.” Two h later, cells were transfected with ERE-luciferase plasmid, and luciferase activity was measured after 24 h. Induction values are averaged from triplicates and presented with standard error (SE). Columns, average of triplicate samples; bars, SE. Values with an asterisk are significantly different from the respective controls. *P < 0.05; **P < 0.005 (Student’s t test).

RESULTS

Modulation of ERE-Luciferase Activity in Mammary Tumor Cell Lines. In the first series of studies, we compared the effect of resveratrol on ERE-luciferase activity with four different cell lines: MCF-7, T47D, LY2, and S30. In these models, ligand-activated ER binds to the ERE, and its functional activation is monitored using a luciferase reporter gene linked to its enhancer (ERE) driven by the minimal thymidine kinase promoter. Relative to DMSO controls, E2 (1 nM) caused a 3–10-fold increase in luciferase activity (Fig. 1). With MCF-7 and S30 cells, resveratrol (10 μM) caused a significant increase in luciferase activity when administered alone (P < 0.05), but when combined with E2 (1 nM), resveratrol antagonized luciferase induction with each of the cell lines tested (Fig. 1). No statistically significant induction was observed at a concentration of 1 μM, and, unlike previous reports (17), no additive or superagonist activities were seen. In T47D and LY2 cells, resveratrol behaved as a pure antiestrogen, with no statistically significant induction of luciferase observed at the three concentrations tested (Fig. 1). When resveratrol was coadministered with E2, potent antagonism was observed in a dose-dependent manner, with approximately 50% inhibition at 10 μM (P < 0.005) in T47D cells. In tamoxifen-resistant LY2 cells, resveratrol also responded as an antagonist in the presence of E2 at a concentration of 15 μM (P < 0.005).

Modulation of Estrogen-inducible Protein Expression in Mammary Cancer Cell Lines. We compared the expression of PR and pS2 in the four different mammary cancer cell lines. MCF-7 cells expressed both PR and pS2, T47D cells expressed PR but not pS2, and S30 and LY2 cells expressed pS2 but not PR (Fig. 2). Cells were treated with resveratrol in the presence or absence of E2 for 48 h and analyzed for protein expression. Treatment of MCF-7 cells with E2 caused a significant increase in PR expression. Resveratrol caused similar induction of PR protein expression at concentrations of 10 and 15 μM, but no effect was seen at 1 μM. When combined with E2, resveratrol suppressed PR expression in a dose-dependent manner. At a concentration of 15 μM, resveratrol blocked the induction of PR by about 90%.

With T47D cells, even after incubation with phenol red-free media containing DCC-FBS for 1 week, basal levels of PR expression were not minimized, and the expression was not increased by the addition of E2. However, resveratrol significantly suppressed basal levels of PR expression and down-regulated PR levels in the presence of E2 in a dose-dependent fashion (Fig. 2).

We then analyzed the expression of pS2, another estrogen-inducible protein, in S30 cells. Based on the analysis of pS2 protein expression by Western blots, high basal levels were present, and E2 did not cause any increase. However, resveratrol down-regulated pS2 expression at all concentrations tested as a single agent, and in the presence of E2, suppression was observed in a dose-dependent manner (Fig. 2).
LY2 cells responded to E₂ treatment by a slight increase of pS2 expression relative to controls. However, production of pS2 protein was down-regulated by resveratrol in the presence or absence of E₂ in a dose-dependent manner (Fig. 2).

**Atypical Ductal Lesions in MMOC.** Experiments were carried out to evaluate the effect of resveratrol on estrogen-dependent DMBA-induced ductal lesions in MMOC. DMSO-treated control glands exhibited normal morphology with branching ducts and alveolar structures, whereas alveoli in glands treated with DMBA in the presence of E₂ and progesterone were sparse (data not shown). On histological examination, it was found that DMSO-treated glands showed a single layer of epithelium in the lumen (Fig. 3A). When treated with DMBA, however, 4–5 layers of epithelial cells resulting in thickened ducts exhibiting a hyperplastic nature (Fig. 3B) were observed. In some ductal structures, the intraductal outgrowths were aggressive and resulted in a complete occlusion of the ducts (Fig. 3C). The percentage of incidence of hyperplastic and aggressive ductal lesions induced by DMBA (63.0 ± 8.1%; P < 0.005) was reduced by resveratrol in a dose-dependent manner (Fig. 3D). The percentage of areas with abnormal histology (hyperplastic and atypical ductal lesions) in control and resveratrol groups is shown in Table 1. Based on these data, the IC₅₀ of resveratrol was approximately 3 μM.

**Modulation of PR Expression in MMOC.** To evaluate a functional aspect of estrogen action in MMOC, induction of PR was examined. Mammary glands were treated for 6 days with estrogen in insulin- and prolactin-containing medium, sections were prepared, and expression of PR was assessed and quantified by immunohistochemistry. With control glands (in the absence of E₂), basal levels of PR were expressed with a proportion of positively stained cells equal to 22.8 ± 4.7% (Fig. 4A). In the presence of insulin, prolactin, and E₂, both the intensity (data not shown) and the proportion of positively stained cells for PR expression (85 ± 14.8%; control versus estrogen-induced PR expression, P < 0.005) were increased (Fig. 4B). When glands were treated in a similar manner without E₂ in the presence of resveratrol, expression of PR was induced to a proportion of 58.3 ± 10.5% (control versus resveratrol-induced PR expression, P < 0.005) at 10 μM (Fig. 4C). Lower concentrations of resveratrol did not cause statistically significant induction of PR. Nonetheless, when E₂ (1 nM) was combined with resveratrol, estrogen-induced PR expression was suppressed to 23.9 ± 7% (estrogen-induced PR expression versus resveratrol + estrogen-induced PR expression, P < 0.005; Fig. 4D) at 10 μM, and the effect was dose-dependent (Fig. 4E). Resveratrol also decreased the intensity of PR staining in estrogen-induced tissues (data not shown). When tested at a concentration of 20 μM, diffuse cellular structures indicative of toxicity were observed in the glands. Based on these data, the IC₅₀ of resveratrol was approximately 4.0 μM.

**NMU-induced Mammary Carcinogenesis.** To determine the effect of resveratrol on mammary tumorigenesis, the NMU model with rats was used. Resveratrol was administered i.g. (5 days/week) for the entire period of study, starting 7 days before NMU administration. Doses of 10 and 100 mg/kg body weight were selected on the basis of previous studies. As shown in Fig. 5A, under these experimental conditions, resveratrol had no effect on body weight. Tumors were palpable in the control group by day 21 after NMU treatment (5.3%), and the percentage of incidence reached 100% on day 111 (Fig. 5B). The high dose of resveratrol delayed tumorigenesis. Notably, on day 40, 0% incidence was observed in the resveratrol-treated group.
resveratrol, as described above. The histological sections of the glands were divided into normal, hyperplastic, or aggressive (highly occluded duct with atypical epithelial cells), hyperplastic (3 layers of thickness), or aggressive (highly occluded duct with atypical epithelial cells). Ductal lesions were induced by addition of DMBA (2 µg/ml) on day 3 for 24 h. Glands were then fixed in formalin, processed, and stained with H&E for histopathological evaluation. A, control glands receiving only DMSO (without DMBA) for the first 10 days show a single layer of epithelial cells lining the ducts. Glands treated with DMBA exhibit (B) hyperplastic lesions or (C) completely occluded duct with atypical epithelial cells. D, glands treated with 10 µM resveratrol for the first 10 days of culture demonstrate normal mammary ductal structures comparable to control. E, dose-response curve for the inhibition of DMBA-induced atypical ductal (hyperplastic and aggressive) lesions by resveratrol. Mouse mammary glands were treated with the indicated concentrations of resveratrol, as described above. The histological sections of the glands were divided into three regions, and each region was evaluated for the number of normal (single-layer thickness of epithelial cells), hyperplastic (≥3 layers of thickness), or aggressive (highly occluded, with no or very little lumen) areas. Data points indicate the mean percentage of incidence of atypical lesions in resveratrol-treated glands. Original magnification for A–D, ×40.

Fig. 3. Effect of resveratrol on DMBA-induced atypical ductal lesions in MMOC. Mouse mammary glands in culture were incubated with insulin, prolactin, E2, and progesterone, with or without resveratrol, for the first 10 days in culture, followed by withdrawal of all hormones (and resveratrol) except insulin and incubation for an additional 14 days. Ductal lesions were induced by addition of DMBA (2 µg/ml) on day 3 for 24 h. Glands were then fixed in formalin, processed, and stained with H&E for histopathological evaluation. A, control glands receiving only DMSO (without DMBA) for the first 10 days show a single layer of epithelial cells lining the ducts. Glands treated with DMBA exhibit (B) hyperplastic lesions or (C) completely occluded duct with atypical epithelial cells. D, glands treated with 10 µM resveratrol for the first 10 days of culture demonstrate normal mammary ductal structures comparable to control. E, dose-response curve for the inhibition of DMBA-induced atypical ductal (hyperplastic and aggressive) lesions by resveratrol. Mouse mammary glands were treated with the indicated concentrations of resveratrol, as described above. The histological sections of the glands were divided into three regions, and each region was evaluated for the number of normal (single-layer thickness of epithelial cells), hyperplastic (≥3 layers of thickness), or aggressive (highly occluded, with no or very little lumen) areas. Data points indicate the mean percentage of incidence of atypical lesions in resveratrol-treated glands. Original magnification for A–D, ×40.

Table 1. Effect of resveratrol on percentage of incidence of atypical ductal lesions in mouse mammary organ culture

<table>
<thead>
<tr>
<th>Group</th>
<th>Normal regions</th>
<th>Hyperplastic lesions</th>
<th>Atypical lesions</th>
</tr>
</thead>
<tbody>
<tr>
<td>DMBA</td>
<td>37.0 ± 8.1</td>
<td>23.4 ± 6.8</td>
<td>39.6 ± 11.1</td>
</tr>
<tr>
<td>DMBA + resveratrol (10 µM)</td>
<td>79.2 ± 15.4 (&lt;0.001)</td>
<td>15.3 ± 1.5 (&lt;0.001)</td>
<td>5.5 ± 0.2 (&lt;0.001)</td>
</tr>
<tr>
<td>DMBA + resveratrol (5 µM)</td>
<td>79.4 ± 23.6 (&lt;0.001)</td>
<td>16.2 ± 5.1 (&lt;0.001)</td>
<td>4.9 ± 1.0 (&lt;0.001)</td>
</tr>
<tr>
<td>DMBA + resveratrol (2.5 µM)</td>
<td>43.5 ± 12.3b</td>
<td>18.4 ± 2.5b</td>
<td>38.1 ± 7.8b</td>
</tr>
<tr>
<td>DMBA + resveratrol (1 µM)</td>
<td>31.5 ± 12.1b</td>
<td>26.8 ± 4.5b</td>
<td>41.7 ± 12.7b</td>
</tr>
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</table>

*Values represent the mean percentage of each region in total glands that were classified as normal, hyperplastic, and aggressive, as defined in "Materials and Methods." Numbers shown in parentheses are P values of a two-tailed t test for independent samples.

**Not significantly different from the control group that received only DMBA.

DISCUSSION

Although a number of previous studies have been conducted, the potential of resveratrol to function as a SERM remains unclear. For example, whereas it was shown with MCF-7 cells in culture that resveratrol is a superagonist when combined with E2 (17), resveratrol was also shown to be antiestrogenic with the same cell line (18, 19). Similarly, the ability of resveratrol to interact with ER is not conclusive. Whereas some researchers have shown that resveratrol binds to ER from MCF-7 cells (IC50 = ~10 µM) and rat uterine cytosolic extracts (IC50 = ~100 µM; Refs. 17 and 29), other studies have shown that resveratrol does not bind to ER in cell extracts from PR1 pituitary cells or partially purified mouse uterus, even at concentrations as high as 100 µM (21, 30). More recently, it was shown that resveratrol acts as a mixed agonist/antagonist in cells transiently transfected with ER and mediates higher transcriptional activity when bound to ER-β than to ER-α (22). Moreover, resveratrol showed antagonist activity with ER-α, but not with ER-β (22). In accord with these data, we have recently observed that resveratrol mediates antiestrogenic effects in endometrial cancer (Ishikawa) cells by a novel mechanism that involves selective down-regulation of ER-α but not ER-β manifested as suppression of estrogen-dependent alkaline phos-
phatase and ERE-luciferase activities, as well as PR and α1 integrin expression (26). To further clarify the estrogen-modulatory effects of resveratrol, we examined its effect with in vitro and in vivo mammary models.

In the first set of experiments, we examined the effect of resveratrol on ERE-luciferase activity in various transiently transfected mammary cancer cell lines. Consistent with previous reports, resveratrol, when administered alone, stimulated luciferase activity at concentrations ranging from 5–15 μM in MCF-7 cells. However, luciferase activity induced by 1 nM E2 was antagonized by resveratrol in the same concentration range, and, in contrast to previous work (17), no superagonist activity was observed under any experimental conditions. Because previous studies (17) had used ethanol as the solvent, and ethanol has the potential to induce ER-α mammary cancer cells (31, 32), it may be speculated that synergistic effects of resveratrol and ethanol could explain the reported superagonist activity of resveratrol. Also, because it is known that resveratrol is UV light and pH sensitive in solution (33), we speculated that improper storage could lead to degraded products capable of mediating superagonism. However, in studies conducted with MCF-7 cells and resveratrol that was intentionally degraded, superagonist activity was not observed (34). In the current report, a profile similar to that of MCF-7 cells (estrogenic alone and antiestrogenic when combined with E2) was observed when S30 cells were treated with resveratrol, whereas with T47D and LY2 cells, resveratrol acted as a pure antagonist.

Induction of breast cancer cell growth, up-regulation of estrogen-inducible genes, and tissue-, species-, and promoter-specific suppression of these parameters in the presence of E2 are some of the classic effects of partial estrogens such as 4-hydroxytamoxifen (35–37). However, in the case of resveratrol, the differential activities observed with various cell lines may be due to the relative expression levels of ER-α and ER-β. As demonstrated by Pettersson et al. (38), heterodimerization of ER-α and ER-β has a major influence on cellular responses to agonists and antagonists; ER-β has the ability to dominantly regulate the activity of the α/β heterodimer positively (genistein) and negatively (tamoxifen), probably by modifying the activity of ER-α AF-1. Although a comprehensive study to establish the relative levels of these receptors in mammary cancer cell lines remains to be done, some reports have indicated that ER-β is virtually absent in MCF-7 cells but is expressed highly in T47D cells (39–41). S30 cells have been stably transfected with ER-α and contain low levels of ER-β (42), and data are not available with respect to the relative levels of the ER subtypes in LY2 cells. Nonetheless, based on the response profile currently reported (Fig. 1), it is reasonable to suggest that resveratrol exerts partial agonist activity in tissues that express predominantly ER-α (e.g., MCF-7 and S30) but exerts antagonist activity when ER-β is coexpressed (e.g., T47D). Interestingly, both T47D and LY2, in which resveratrol acted as a pure antagonist, are antiestrogen-resistant cell lines (43). Therefore, it would be reasonable to evaluate resveratrol in combination with an agent such as tamoxifen.

We then studied the expression of two estrogen-responsive proteins, PR and pS2. MCF-7 and T47D cells were compared for expression of PR. With MCF-7 cells, consistent with the transfection experiments, PR was induced by treatment with resveratrol as a single agent and suppressed when resveratrol was given in combination with E2. With T47D cells, even after long-term replacement with DCC-FBS, basal levels of PR were not reduced. Nonetheless, PR protein levels were down-regulated on treatment with resveratrol. In LY2 and S30 cells, resveratrol suppressed pS2 expression irrespective of the presence of E2. Although these data do not correlate directly with the transient transfection studies performed with these cells, it is possible that the natural, imperfect ERE sequences for PR and pS2, which differ slightly from the consensus ERE, have the ability to dominantly regulate the activity of the α/β heterodimer positively (genistein) and negatively (tamoxifen), probably by modifying the activity of ER-α AF-1. Although a comprehensive study to establish the relative levels of these receptors in mammary cancer cell lines remains to be done, some reports have indicated that ER-β is virtually absent in MCF-7 cells but is expressed highly in T47D cells (39–41). S30 cells have been stably transfected with ER-α and contain low levels of ER-β (42), and data are not available with respect to the relative levels of the ER subtypes in LY2 cells. Nonetheless, based on the response profile currently reported (Fig. 1), it is reasonable to suggest that resveratrol exerts partial agonist activity in tissues that express predominantly ER-α (e.g., MCF-7 and S30) but exerts antagonist activity when ER-β is coexpressed (e.g., T47D). Interestingly, both T47D and LY2, in which resveratrol acted as a pure antagonist, are antiestrogen-resistant cell lines (43). Therefore, it would be reasonable to evaluate resveratrol in combination with an agent such as tamoxifen.
resveratrol functions as an antiestrogen. Furthermore, one of the initial effects of E₂ in mammary cell differentiation is the induction of estrogen-inducible genes including PR (46, 47). In the current report, we have shown that expression of PR is localized in the epithelial lining of glands incubated with insulin, prolactin, and E₂ and that we have shown that expression of PR is localized in the epithelial lining of glands incubated with insulin, prolactin, and E₂ and that treatment with resveratrol for 6 days reduced induction of PR. These data indirectly suggest a regulatory role of progesterone and PR in carcinogenesis and are consistent with a recent report in which PR knockout mice developed fewer DMBA-induced tumors as compared with the isogenic wild-type mice (48). It would be of interest to evaluate resveratrol in this model.

In the current study, we did examine the effect of resveratrol on the formation of NMU-induced mammary tumors in Sprague Dawley rats. The etiology of this model is generally considered relevant to human breast cancer, and it has been widely used in various chemoprevention studies (49, 50). Resveratrol caused a significant suppression of tumor multiplicity and increased the latency period by 27 days. Tumor incidence in resveratrol-treated animals was reduced by ~50% for 69 days, after which it gradually increased until the end of the experiment, when it reached that of control animals (95%). Histopathological examination revealed response to resveratrol treatment, as evidenced by increased alveolar and adipocyte differentiation and the occurrence of apoptotic cells within peripheral tumor areas, which are rare in these areas. It is, of course, difficult to affirm whether the chemopreventive response mediated by resveratrol is due exclusively to antiestrogenic effects. However, our observation that resveratrol delays the occurrence of mammary tumors and suppresses incidence at the early time points (until day 69) suggests that the agent may be more efficacious in early stages, when hyperplastic and premalignant lesions predominate in the mammary gland. These lesions are also small in size and composed entirely of ER⁻ cells (51, 52), and their response to ER agonists or antagonists should be more profound than the response of large mammary tumors, in which a selection of ER⁺ tumor cells may also occur and thus nullify antiestrogenic effects. Studying the effect of resveratrol in rats in which E₂ administration reactivates NMU tumor growth after an ovariectomy-induced tumor regression would be significant in this regard. Also, it may be of interest to investigate whether higher doses of resveratrol (>100 mg/kg) can cause further reduction of the incidence and multiplicity of tumors. Although relatively high doses of a chemopreventive agent may be scrutinized for physiological relevance, justification for using such doses is provided by the necessity of inducing tumors in model systems by treatment with high doses of chemical carcinogens. In addition, the effect of metabolism needs to be investigated because studies have shown that resveratrol, when administered p.o. in rats, is sulfated, and hepatic and duodenal sulfation might limit the bioavailability of this compound (53). Alternatively, metabolites themselves may mediate biological responses, as in the case of tamoxifen.

In conclusion, the data presented in this study emphasize the mixed estrogenic/antiestrogenic activity of resveratrol in mammary tissues. In the absence of E₂, resveratrol weakly induced ER-dependent transcriptional events in some mammary tumor cell lines, but down-regulation was invariably observed when the agent was coadministered with the
RESVERATROL AND MAMMARY TUMORS

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hormone. In mouse mammary glands grown in culture, resveratrol inhibited

the formation of DMBA-induced, E2-promoted, atypical ductal hyperplasia. Furthermore, resveratrol inhibited the early stages of NMU-induced mammary carcinogenesis when administered to female Sprague Dawley rats. In conjunction with previous work reported in the literature, these studies support the suggestion that resveratrol is a novel SERM that may be useful for the chemoprevention of breast cancer.


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