Mitogen-activated Protein Kinase Kinase 4 (MKK4) Acts as a Metastasis Suppressor Gene in Human Ovarian Carcinoma

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ABSTRACT

Despite improvements in chemotherapy and the recognition that aggressive surgical cytoreduction is beneficial, the majority of patients diagnosed with ovarian cancer will die as a result of metastatic disease. The molecular changes associated with acquisition of metastatic ability in ovarian cancer are poorly understood. We hypothesize that metastasis suppressor gene inactivation or down-regulation plays a role in ovarian cancer progression.

Mitogen-activated protein kinase kinase 4 (MKK4), a member of the stress-activated protein kinase signaling cascade, has been identified recently as a metastasis-suppressor gene. An immunohistochemical approach was taken to test the possibility that MKK4 dysregulation occurs during the development of clinical ovarian cancer metastases. MKK4 expression was evaluated in normal and metastatic ovarian tissues. Normal ovarian epithelial cells showed high intensity staining for MKK4, whereas metastatic tissues showed a statistically significant decrease in expression. These results support a role for MKK4 dysregulation in the development of clinical disease.

A functional approach was taken to test the ability of MKK4 to suppress metastatic colonization, the process whereby disseminated cancer cells lodge and grow at a secondary site in vivo. The SKOV3ip.1 human ovarian cancer cell line was chosen for these studies because it lacks endogenous MKK4 expression but retains both upstream and downstream components of the signaling cascade of MKK4. Ectopic expression of MKK4 in these cells, when injected into female SCID mice, suppressed the number of overt metastatic implants by nearly 90%. Furthermore, MKK4 expression increased the life span of the animals by 70%. Taken together, these data support a role for MKK4 in the suppression of metastatic colonization in ovarian cancer.

INTRODUCTION

Ovarian cancer causes more deaths than any other gynecologic malignancy despite improved therapeutic regimens, such as aggressive surgical cytoreduction and the combination of paclitaxel and platinum-based chemotherapy. In the year 2002, it is estimated there will be 23,300 new cases of ovarian cancer and 13,900 deaths (1). The significant mortality associated with ovarian cancer is the result of advanced stage disease at the time of diagnosis because of lack of early, recognizable symptoms, and the development of peritoneal and extraperitoneal metastases that become chemotherapy-resistant.

The acquisition of metastatic ability requires the coordinated expression of metastasis-promoting genes and the down-regulation of metastasis-suppressing genes. In recent years, a number of metastasis-suppressor genes have been identified in various organ systems (2–8). Metastasis-suppressor genes have been shown to suppress the growth of metastases without affecting the growth of the primary tumor (9). Thus, these genes are distinct from tumor suppressor genes, which suppress primary tumor growth. Particularly important in this process is the concept of metastatic colonization whereby disseminated cancer cells lodge and grow at a secondary site. Metastatic colonization requires disseminated cells to initiate context-dependent signaling cascades that allow them to survive, enter the cell cycle, and divide to become microscopic metastases, and subsequently, overt metastases (10). Growth control mechanisms at these secondary sites of disease govern whether cells will form clinically significant metastases or not (10, 11). Although a plethora of data exists in other organ systems, little specific information is available on metastasis suppressor gene expression and function in ovarian cancer (12–16). Such information would be of considerable value in identifying the molecular alterations, which regulate the dissemination of ovarian cancer cells from the primary tumor and subsequent colonization of extra-ovarian sites.

We identified recently the MKK44/INKK1/SEK1 (hereafter referred to as MKK4), which is located on chromosome 17p11.2, as a metastasis suppressor gene in prostate cancer. MKK4 is a member of the SAPK signaling pathway that ultimately controls transcription factors such as c-jun (17). In this way, MKK4 plays a role in such processes as cellular proliferation, differentiation, and apoptosis. In highly metastatic AT6.1 prostate cancer cells, ectopic expression of MKK4 reduces overt lung metastases by ~80% without affecting primary tumor growth (18). An association between decreased MKK4 expression and increasing Gleason grade, a predictor of metastatic potential, supports a role for MKK4 down-regulation in clinical prostate disease (19). Although the metastatic patterns of spread in prostate and ovarian cancer are clinically different, we speculated that fundamental mechanisms governing metastatic colonization would be conserved. Of interest is the observation that loss of 17p is a frequent event in ovarian cancers. Therefore, we hypothesized that MKK4, or members of its signaling pathway, would be down-regulated in clinical metastases and that ectopic expression of MKK4 would suppress metastatic colonization in a model of ovarian cancer.

To begin to test this hypothesis, the expression of MKK4 in normal and metastatic ovarian tissues was examined. In the simplest scenario, if MKK4 inactivation played a role in the development of metastases, then the level of MKK4 expression in ovarian cancer metastases would be decreased as compared with normal ovarian epithelial cells. To test the ability of MKK4 to function as a metastasis suppressor in vivo, MKK4 was ectopically expressed in the highly metastatic...
SKOV3ip.1 human ovarian cancer cell line to determine the effect on metastatic colonization.

MATERIALS AND METHODS

Evaluation of MKK4 Staining in Clinical Samples

Normal and cancerous ovary samples were identified from the Department of Pathology database at our institution. In accordance with a protocol approved by the Institutional Review Board at the University of Chicago, 34 normal ovary specimens from patients undergoing surgery for benign conditions that specifically contained normal surface epithelium or epithelial inclusion cysts containing normal epithelial cells were identified. Metastatic tumor specimens from 34 patients undergoing primary cytoreductive surgery for ovarian cancer were similarly identified.

Specimen preparation and immunohistochemical studies were conducted as described previously (19). In brief, slides were prepared for immunostaining and incubated overnight at 4°C using rabbit anti-MKK4/MEK4 (5 μg/ml; Santa Cruz Biotechnologies, Inc.) or isotype control (5 μg/ml; whole rabbit IgG) prepared in protein blocking solution (5% horse serum +1% goat serum in PBS). Slides were subsequently incubated for 3 h at 37°C with goat anti-rabbit biotin-conjugated IgG, (3 μg/ml; Santa Cruz Biotechnologies, Inc.) followed by incubation with ABC Elite Vectastain kit (Vector Labs). Immunostaining was detected using a 3.3'-diaminobenzidine peroxidase substrate kit (Vector Laboratories), and slides were counterstained with Fast Green (Fisher Scientific). Controls for sensitivity and specificity of the immunostaining included an epitope control (peptide competition using 5-fold excess of peptide; Santa Cruz Biotechnologies, Inc.), secondary antibody alone, and avidin/biotin alone (described in detail in Ref. 19).

A semiquantitative evaluation scale was developed based on intensity of MKK4 staining. A numerical scale of 0, 1, 2, or 3 was assigned for designated for those epithelial areas that did not stain for MKK4 and 3 assigned to epithelial cells that stained with greatest intensity for MKK4. Samples were interpreted and scored by two independent reviewers, a pathologist with dedicated expertise in gynecologic pathology and a gynecologic oncologist (A. M. and S. D. Y., respectively).

Human Ovarian Cancer Cell Lines and Control Cell Lines

Caov-3 was obtained from American Type Culture Collection and grown in DMEM supplemented with 8% FCS, 1% penicillin, and 1% streptomycin. SKOV-3, SKOV3ip.1, and HEY-A8 (the generous gifts of Dr. Gordon Mills, M.D. Anderson Cancer Center, Houston TX) were grown in DMEM supplemented with 5% FCS, 1× sodium pyrovate, 1× nonessential amino acids, 1× t-glutamine, and 2× vitamin solution (Life Technologies, Inc.). ASCPC-1, a pancreatic cell line, which contains a homozygous deletion from Cn5% CO2.

Immunoblotting

Ovarian cancer cell lines were grown in monolayer to 80% confluence. Cell lysates were prepared in boiling cell lysis buffer [100 mM Tris (pH 7.5), 1% SDS, and 100 mM NaVO3]. Protein concentrations were determined using a bicinchoninic assay reduction kit (Pierce). Either 20 μg or 100 μg of total protein from each cell line was resolved by SDS-PAGE (10% acrylamide) and transferred onto nitrocellulose membrane. The polyclonal antibodies (Santa Cruz Biotechnologies, Inc.) and working dilutions used for immunoblotting were as follows: MEK1 (1:1,000), MKK6 (1:5,000), MKK7 (1:250), MKK3 (1:10,000), MKK4 (1:5,000), JNK1 (1:50,000), p38 (1:10,000), and c-Jun (1:5,000).

Membranes were incubated with a horseradish peroxidase-conjugated IgG secondary antibody as per the manufacturer’s instructions and detected using enhanced (Femto) chemiluminescence (Pierce). As a loading control, membranes were subsequently reprobed for actin (Oncogene Research Products). Protein lysates from ASCPC-1 cells (20 μg), which have a homozygous deletion in exon C of MKK4, and rat brain (3 μg), which has high endogenous levels of MKK4, served as negative and positive controls, respectively.

In Vivo Metastasis Assays

Clonal Cell Lines. Hemagglutinin (HA)-tagged MKK4 cDNA was subcloned into the retroviral vector pLNCX2 (Clontech, Palo Alto, CA). Retrovirus was produced by transient transfection of retroviral vector alone (vector only; transfectants) or HA-MKK4-pLNCX2 into amphotrophic Phoenix cells (American Type Tissue Collection, with permission of Dr. Gary Nolan, Stanford University, Palo Alto, CA) using an Effectene transfection reagent per manufacturer’s instructions (Qiagen, Santa Clarita, CA). This approach routinely yields high efficiency transfer of the target DNA into recipient cells. Transfection medium was collected 48 h later. SKOV3ip.1 cells (2×106) were plated 24 h before infection, and incubated with transfection medium and Polybrene for 8 h. Transfection medium was replaced with fresh medium, and the cells were then diluted in growth medium containing G418 (500 μg/ml; Sigma). Cells were split for selection and establishment of clonal cell lines.

HA-MKK4 expression was verified by immunoblotting using an anti-HA.11 monoclonal antibody (1:500 dilution; Covance Inc.), a horseradish peroxidase-conjugated goat antimouse secondary antibody (1:2000 dilution; Sigma), and biotin followed by detection with an enhanced chemiluminescence kit (Pierce). The positive and negative controls used were AT6.1-SR3-HA-JNK1, the AT6.1 prostate cancer cells transiently transfected with HA-MKK4 (JNK1) using the SR3 plasmid (hereafter referred to as AT6.1-SR3-HA-MKK4), and the parental SKOV3ip.1 cell line, respectively.

Metastasis Assays. Parental SKOV3ip.1 cells were injected i.p. (1×106 cells) into female SCID mice (Frederick Laboratories) to determine the time course for metastasis formation, as well as the location and number of metastases. A similar model has been used by other investigators (20, 21). On the basis of our preliminary experiments in which animals survived an average of 40 days, 7 SKOV3ip.1-HA-MKK4 clones and 5 vector control clones, SKOV3ip.1-pLNCX2, were injected i.p. (1×106 cells/mouse) into 60 different mice (5 mice/clone). The number and extent of overt metastases were quantitated 20 days after injection. This time point was chosen because it gave a reproducible number of overt metastases without significant morbidity. Metastases were defined as visible tumor implants >1 mm in diameter. Two independent studies were conducted. To determine total survival time, each clonal cell line was also injected into an additional 5 mice/clone and the mice followed until moribund.

In Vitro Characteristics of SKOV3ip.1-HA-MKK4 Clones and SKOV3ip.1-pLNCX2 Clones

Cell Proliferation. To determine whether the introduction of MKK4 into the SKOV3ip.1 cells affected inherent growth rate, each clonal cell line (15,000 cells/well) was plated in triplicate in a six-well plate and allowed to grow under standard tissue culture conditions. A Coulter counter was used to count cells at interval time points of 0, 72, 96, 120, 144, 168, and 240 h after plating. At the final time point, cells were 95–100% confluent.

DAPI Staining. To assess whether ectopic MKK4 expression induced apoptosis, DAPI staining and flow cytometry were performed on SKOV3ip.1-HA-MKK4 clones and results compared with those obtained from SKOV3ip.1-pLNCX2 clones. In brief, each clonal cell line was plated in a six-well plate and allowed to grow under standard tissue culture conditions until 70–80% confluent. Cells were fixed for 30 min at room temperature in 3.7% parformaldehyde (Sigma) followed by detection with an enhanced chemiluminescence kit (Pierce). The positive and negative controls used were AT6.1-SR3-HA-JNK1, the AT6.1 prostate cancer cells transiently transfected with HA-MKK4 (JNK1) using the SR3 plasmid (hereafter referred to as AT6.1-SR3-HA-MKK4), and the parental SKOV3ip.1 cell line, respectively.

Flow Cytometry. The fractions of apoptotic and proliferating cells were also assessed by flow cytometry. Each clonal cell line was plated in a six-well plate and allowed to grow under standard tissue culture conditions until 70–80% confluent. Cells were trypsinized and added to supernatant, spun at 1200 rpm, and washed twice in FACS buffer (0.2% BSA/0.1% NaN3 in 1× PBS). Cells were resuspended in 250 μl of FACS buffer. Ten μl of 100 μg/ml propidium iodide (Molecular Probes) was added to each tube and immediately analyzed via flow cytometry.
MKK4 Suppresses Metastasis in Ovarian Cancer

Statistical Analyses

Immunohistochemistry. For the expression of MKK4 in the clinical specimens, the proportion of normal ovary specimens that demonstrated MKK4 staining intensity ≥2 was compared with the proportion of metastatic samples demonstrating MKK4 staining intensity ≥2. A power calculation indicated that a sample size of 28 normal specimens and 28 cancer specimens would provide >90% power to detect a true reduction from 95% of the normal ovaries staining at the designated intensity level to 55% of the cancer specimens staining at the designated intensity level. A continuity corrected χ² analysis was used to compare the observed proportions in the two groups. A Wilcoxon rank sum test was also used to compare the full set of ordinal responses (intensity score 0, 1, 2, or 3) in the different groups. P < 0.05 was considered statistically significant.

Metastasis Assays. The mean number of metastases at 20 days after injection in the 5 SKOV3ip.1-pLNCX2 clones was compared with the mean number of metastases in the 7 SKOV3ip.1-HA-MKK4 clones using a mixed model ANOVA treating each clone as a random effect because observations within each clone were correlated (23). To stabilize the variance, the data were transformed to the square root scale before analysis although results are reported as mean number of metastases. For the mice injected with the SKOV3ip.1-pLNCX2 clones or the SKOV3ip.1-HA-MKK4 clones and followed until moribund, an estimated survival distribution was calculated by the method of Kaplan and Meier (24). To allow for the within-clone correlation, survival times were averaged across mice within the same clone and the averages compared between the two groups using a Wilcoxon rank-sum test. (All but 5 of the mice had complete observations. Five mice in the MKK4 group were censored at 53 days when the study was terminated.)

Doubling Times and Apoptosis Assays. Mean doubling time for the SKOV3ip.1-pLNCX2 clones was compared with the SKOV3ip.1-HA-MKK4 clones using a mixed two-step procedure. A separate linear growth curve was first fit to each clone over the period of logarithmic growth (72–144 h), and the doubling time was calculated as log₂/slope. The mean doubling time was compared between the two groups using a two-sample t test. The DAPI experiments and flow cytometry data were analyzed using a two-factor (treatment group by experiment) ANOVA. A P < 0.05 was considered statistically significant.

RESULTS

MKK4 Is Down-Regulated in Clinical Ovarian Cancer Specimens. To test the potential role of MKK4 down-regulation in metastasis, we compared intensity of MKK4 expression in the epithelial component of normal ovary samples to that in metastatic ovarian cancer specimens. Of the 34 patients with metastatic ovarian cancer, 3 had stage II disease, 19 had stage III disease, and 12 had stage IV disease. A scoring system was devised to assign a staining intensity score for MKK4 expression ranging from 0 (no expression) to 3 (highest intensity staining) in the epithelial components. Fig. 1, A and B, show MKK4 staining in the normal epithelium corresponding to level 3 staining (Fig. 1A) and its corresponding negative control (Fig. 1B). In contrast, Fig. 1, C and D show level 1 staining in the metastatic epithelial component and no staining in the corresponding IgG control. Table 1 shows the distribution of MKK4 staining intensity scores among the normal ovary and metastatic cancer specimens. The majority of normal ovary specimens (31 of 34, 91%) stained for MKK4 at an intensity level ≥2 with 20 (58%) samples staining at the highest intensity level, 3. The staining was uniform and epithelial-specific, with relatively little stromal staining noted. In contrast, only 11 of 34 (32%) metastatic samples stained at an intensity level ≥2, and none of these samples stained at the highest intensity level, 3 (P < 0.0001). The Wilcoxon test also yielded highly significant results (P < 0.0001). These data support a role for MKK4 down-regulation in ovarian cancer cells that have acquired the ability to metastasize.

Metastatic Human Ovarian Cancer Cell Lines Lack MKK4 Expression but Retain Other Components of the SAPK Pathway. To test the hypothesis that ectopic expression of MKK4 would suppress metastatic colonization in vivo, we needed to identify an appropriate model system. The expression of endogenous MKK4 and components of the SAPK signaling cascade (Fig. 2A) were evaluated in four human ovarian cancer cell lines. ASPC-1 cells, which have a homozygous deletion in the MKK4 gene and do not express the protein, were used as a negative control, whereas rat brain, which expresses high levels of MKK4, was used as a positive control (Fig. 2B). HEY-A8, SKOV-3ip.1, and SKOV-3, which have been shown previously to form metastases in vivo (20), had undetectable levels of MKK4 expression even when 100 µg of protein was loaded per sample (Fig. 2B). MEKK1, which is known to phosphorylate MKK4 in response to cellular stimuli, was expressed in all four of the cell lines as were the MAP kinases JNK1, JNK2, and p38, and the transcription factor, c-jun. Of the MAP kinase kinases, MKK3 was detected in all four of the cell lines. Representative data are shown in...
Table 1. Intensity of MKK4 expression in epithelium of normal ovaries compared with metastatic ovarian cancer specimens

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<thead>
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<th>MKK4 staining intensity</th>
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<tr>
<td>Normal ovary specimen</td>
<td>0 1 2 3</td>
</tr>
<tr>
<td>Ovarian cancer metastases</td>
<td>5 18 11 0</td>
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The proportion of specimens with an intensity score ≥2 in the normal ovary group is 31 of 34 (91%) as compared with 11 of 34 (32%) in the metastatic ovarian cancer group, χ² = 22.5, P < 0.0001.

Fig. 2. Immunoblotting for constituents of the SAPK signaling pathway in human ovarian cancer cell lines. A, representative members of the SAPK signaling pathway are analyzed. B, four human ovarian cancer cell lines (HEY-A8, Caov-3, SKOV3ip.1, and SKOV-3) were subjected to immunoblotting for upstream (MEKK1) and downstream (JNK and c-jun) constituents of the MKK4 signaling pathway. In addition to MKK4, MKK3, MKK6, MKK7, and p38 were also analyzed for protein expression. Rat brain and ASPC-1 cells, a pancreatic cell line that has a homozygous deletion of MKK4, serve as positive and negative controls, respectively. Three of four (HEY-A8, SKOV3ip.1, and SKOV-3) human ovarian cancer cell lines lacked detectable levels of MKK4 expression even when 100 μg protein (B, top) was resolved. MEKK1 (data not shown) and the downstream components of the MKK4 pathway (JNK and c-jun) were present, as was p38. In the Caov-3 cell line, MKK4 protein was present; however, other MAP kinase kinases, MKK6 and MKK7, were undetectable.

Fig. 3. Western analysis demonstrating expression of HA-MKK4 in SKOV3ip.1, SKOV3ip.1-pLNCX2, and AT6.1-HA-MKK4. Actin was used as a loading control and is shown below each immunoblot.

Ectopic MKK4 Expression Suppresses Metastatic Ability in Vivo. To test the ability of MKK4 protein to suppress metastatic colonization in vivo, 5 SKOV3ip.1-pLNCX2 clonal vector-only control cell lines (Fig. 3, left) and 7 SKOV3ip.1-HA-MKK4 clonal cell lines (Fig. 3, right) were established. HA-MKK4 transgene expression was confirmed by immunoblotting. AT6.1 cells that transiently expressed HA-MKK4 (e.g., AT6.1-SRα-HA-MKK4) were used as a positive control, whereas parental SKOV3ip.1 cells were used as a negative control. The effect of ectopic MKK4 expression on metastatic colonization was assessed by injection of 1 × 10⁶ cells i.p. into female SCID mice. Twenty days after injection, mice were sacrificed and the number of overt (i.e., > 1 mm diameter) surface metastases counted (Fig. 4). Animals injected with SKOV3ip.1-pLNCX2 vector-only control cell lines had overt metastatic implants on the liver, the small bowel, and near the stomach and spleen (Fig. 4, A and B). On histological section, the metastatic implants retained the papillary serous histology that is characteristic of the parental cell line and the majority of clinical human ovarian cancers (Fig. 4C). The data from two independent experiments were tabulated and are summarized in Table 2. Ectopic expression of MKK4 suppressed the number of overt metastases by 88% (Fig. 4D). The mean number of metastases in mice injected with the vector control clones was 27.2 ± 14.8 as compared with the mean number in the mice injected with the SKOV3ip.1-HA-MKK4 clones, which was 3 ± 4.6 (Table 2, left).

Ectopic MKK4 Expression Does Not Affect Proliferation or Induce Apoptosis in Vitro. To determine whether ectopic HA-MKK4 expression affected cell proliferation, thereby reducing the inherent proliferative rate of metastatic implants, the in vitro doubling times of the cell lines were determined (Table 2, right). Mean doubling time for the vector control clones of 34.7 h (95% confidence interval, 27.2–42.1) was not significantly different when compared with that of the HA-MKK4 transfected, 37.8 h (95% confidence interval, 31.3–44.2; P = 0.44).

To test the possibility that ectopic expression of MKK4 increased the number of cells undergoing apoptosis, two independent methods, DAPI and flow cytometry, were used to quantitate the fraction of apoptotic cells. The percentage of apoptotic cells was determined in three representative SKOV3ip.1-pLNCX2 clones and four representative SKOV3ip.1-HA-MKK4 clones used in the in vitro experiments described above. Experiments were performed in duplicate and triplicate for DAPI staining and flow cytometry, respectively, and the mean number of apoptotic cells in the control clones and MKK4 containing clones compared. By DAPI staining, the majority of cells in both groups were not apoptotic: 97.4 ± 0.93% in the vector controls versus 97.5 ± 1.13% in the SKOV3ip.1-HA-MKK4 clones (P = 0.94). These results were similar when flow cytometry was used with 95.4 ± 2.4% of the vector control cells and 94.7 ± 4.0% of the MKK4-expressing clones being nonapoptotic (P = 0.30). The 95% confidence intervals for the true difference (MKK4-control) were −1.3% to 1.4% and −3.0% to 1.0%, respectively. Taken together, results of these in vitro studies suggest that ectopic expression of MKK4 does not alter the inherent growth properties of SKOV3ip.1.
cells. Future studies will be conducted to examine these parameters in vivo.

**Ectopic MKK4 Expression Prolongs Survival.** To test the effect of MKK4-mediated metastasis suppression on animal survival, additional mice (n = 5/clone) injected with SKOV3ip.1-pLNCX2 and SKOV3ip.1-HA-MKK4 clones were followed until moribund and sacrificed (Table 2, middle). Overall mean survival was significantly increased by 70%, from 37 days in the mice injected with vector control clones to 63 days in mice injected with the SKOV3ip.1-HA-MKK4 clones (Wilcoxon P = 0.0045; see also Fig. 5).

**DISCUSSION**

Ovarian cancer is, by far, the most lethal of the gynecologic malignancies. Screening techniques for early detection of disease and an adequate understanding of the molecular mechanisms contributing to development of disease are ineffective and lacking, respectively. Given that most women present with metastatic disease, the greatest impact on reducing the death rate from ovarian cancer lies particularly important to defining the critical steps that regulate metastatic colonization. These studies have shown that MKK4 expressing prostate cancer cells are capable of reaching remote sites, yet, at these sites, are growth arrested in comparison to control cells. A comprehensive understanding of MKK4-mediated growth regulation is particularly important to defining the critical steps that regulate metastatic colonization.

In the current study, we have demonstrated that the normal surface epithelium, from which 90% of epithelial ovarian cancers originate, express high levels of MKK4. In contrast, the majority of metastatic ovarian cancer specimens show significantly decreased expression suggesting that MKK4 protein levels are down-regulated in clinical signaling pathway that responds to a number of cellular and environmental stressors (17). By phosphorylating MAP kinases such as JNK, MKK4 can ultimately transmit stress signals to nuclear transcription factors that mediate various processes including proliferation, apoptosis, and differentiation. In recent years, distinct biological functions have been identified for MKK4 including a role in development, hepatogenesis, and metastasis suppression (18, 19, 25). The characterization of MKK4 as a prostate cancer metastasis suppressor gene is supported by work from both animal model and clinical correlative studies (18, 19). These studies have shown that MKK4 expressing prostate cancer cells are capable of reaching remote sites, yet, at these sites, are growth arrested in comparison to control cells. A comprehensive understanding of MKK4-mediated growth regulation is particularly important to defining the critical steps that regulate metastatic colonization.

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Additional mechanistic studies will be required before appropriate therapeutic interventions can be formulated, but, certainly, a first step will be to analyze in vivo the combined effects of MKK4 reconstitution with standard therapies such as chemotherapy. Our ultimate goal is to identify targets to improve ovarian cancer treatment. To this end, we find the effect on animal survival encouraging. As we better understand the role the MKK4 signaling pathway plays in metastatic colonization, perhaps we will be able to identify novel modulators of this pathway that may act as targets for treatment and ultimately, improved patient survival.

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Mitogen-activated Protein Kinase Kinase 4 (M KK4) Acts as a Metastasis Suppressor Gene in Human Ovarian Carcinoma


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