In Vivo Monitoring of Capecitabine Metabolism in Human Liver by 19 Fluorine Magnetic Resonance Spectroscopy at 1.5 and 3 Tesla Field Strength

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Materials and Methods

Because the liver is the primary site of capecitabine metabolism as well as the predominant site for metastasis of colorectal cancer, we performed nine 19F MRS measurements in the liver of five patients with advanced colorectal cancer (Table 1). Patients were treated with oral capecitabine during 2 weeks. All of the patients had a Karnofsky performance status ≥90% and a normal liver function. 19F MRS measurements were performed during at least 40 min, starting ~1 h after oral intake of capecitabine in the 2-week period of oral capecitabine intake. Patients gave written informed consent, and the experiments were approved by the local ethical committee.

19F MRS measurements were performed on both a clinical 1.5 T and a 3 T whole body Siemens MR system. A flexible 16-cm 19F MR coil was used to enable optimal positioning across the liver region to receive the MR signals of capecitabine and its metabolites from that region. For all of the measurements a pulse-acquire sequence was used with a repetition time of 470 ms. In patients 2, 3, and 4 (Table 1) pulse-acquire measurements were interleaved with localized measurements by 19F MRSI (8). The use of MRSI gives the opportunity to differentiate between the conversion of capecitabine in tumor and normal liver tissue. An 8 × 8 × 8 MRSI was used with a voxel size of 4 × 4 × 4 cm. Both pulse-acquire and MRSI measurements were optimized for signal to noise (9) with a temporal resolution of 4 min. No respiratory gating was used. Resonances in the 19F MR spectra were analyzed using MRUI software. 2 Percentage changes during the measurement interval in 19F MR spectral peak areas obtained by the pulse acquire sequence were determined for DFUR + DFUR and the 5FU catabolites. For DFUR + DFUR the difference in MR peak area of the last spectrum that showed the MR peak of DFUR referenced to the spectral position of the 5FU signal, urine of patient 2 collected from 3–10 h after capecitabine intake was measured at 1.5 T using the same MR protocol, after which 5DFUR, 5DFUR (Roche, Mijdrecht, the Netherlands), and 5FU (Teva Pharma, Mijdrecht, the Netherlands) were added, consecutively. The dependence of spectral peak position on pH was examined by adding HCl and NaOH to solutions of 5FU with 5DFUR and the peak area by the aforementioned MR protocol.

To confirm peak assignments of the capecitabine metabolites 5DFUR and 5DFUR referenced to the spectral position of the 5FU signal, urine of patient 2 collected from 3–10 h after capecitabine intake was measured at 1.5 T using the same MR protocol, after which 5DFUR, 5DFUR (Roche, Mijdrecht, the Netherlands), and 5FU (Teva Pharma, Mijdrecht, the Netherlands) were added, consecutively. The dependence of spectral peak position on pH was examined by adding HCl and NaOH to solutions of 5FU with 5DFUR and the peak area by the aforementioned MR protocol.

To quantify the improvement of SNR at 3 T in comparison with 1.5 T we determined the amplitude of the catabolite peaks in the 19F MR spectra of patients 2, 3, and 4 obtained at a similar time point after capecitabine intake toxicity (7). Here we report for the first time that the metabolism of capecitabine after oral intake can be monitored in vivo by 19F MRS.

Until now human 19F MRS studies have been performed at standard clinical MR systems operating at 1.5 T field strength. 3 T MR scanners have become available recently for clinical application. For 19F MRS the use of higher field strengths is expected to result in an increased SNR and an improvement of spectral resolution, which would better facilitate its use in clinical examinations of fluorinated drugs. Therefore, in this study we have included a first investigation in patients of 19F MRS at 3 T.

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from the 1.5 T and 3 T measurements. The amplitude was divided by the SD of the noise from the same spectrum. This ratio acquired for the measurement at 3 T was divided by the ratio for that at 1.5 T to obtain the factor of improvement in SNR for each patient.

**Results and Discussion**

$^{19}$F MR spectra taken from the liver showed distinct resonances for capecitabine and its metabolic products (Fig. 1). Spectra obtained at 3 T showed a factor 1.3–3 higher SNR and an improved spectral resolution in comparison with those obtained at 1.5 T (Fig. 2), as may be expected from measurements at higher field strengths.

We confirmed peak assignments to 5’DFUR and 5’DFCR in measurements of urine samples (pH 5.87) at 4.0 and 3.4 ppm, respectively. The spectral peak position of 5’DFUR falls within the physiological pH range and as this range corresponds with a relatively large shift in ppm values, 5’DFUR can be used as an in vivo marker of tissue pH. From our spectra the pH of liver tissue was found to be 7.39, which agrees with in vivo pH values in liver tissue measured by $^{31}$P MRS (11). In isolated tumor cells intra/extracellular 5FU ratio correlated with extracellular pH, intracellular pH, and the pH gradient across the cell membrane (12). In an animal model a decrease in local tissue pH from 7.3 to 6.9 was associated with a 2.5-times increase of the $t_{1/2}$ of 5FU, indicating a trapping of 5FU in the tumor (13). Because of the relation between trapping of 5FU in the tumor and tumor response (1), in vivo measurement of pH by the 5’DFUR signal shift can be useful in the prediction of therapy outcome.

The time course of capecitabine metabolism is shown in Fig. 1 for patient 1. In this patient the concentration of capecitabine in the liver declined to below MR detectable levels within 80 min after intake due to its conversion and clearance (Fig. 1). The conversion and clearance

![Fig. 1. Sequential $^{19}$F MR spectra of the liver of patient 1, obtained by a pulse acquire sequence at 1.5 T and starting 60 min after oral capecitabine intake. Inset, expanded spectrum of the region from $-5$ to 15 ppm, showing the average of spectra taken from 60 to 76 min after capecitabine intake.](image-url)
Fig. 2. $^{19}$F MR spectra of the liver of patient 4, obtained by a pulse acquire sequence at 1.5 T (left) and 3 T (right), showing the average of two spectra taken 68 and 76 min after oral capecitabine intake. To facilitate comparison the 3 T spectrum have been scaled to equal noise levels with the 1.5 T spectrum.

Fig. 3. Localized spectra obtained by $^{19}$F MR spectroscopic imaging at 1.5 T of patient 2, 60 min after oral capecitabine intake on day 10. A and B show the spectra from five tumor voxels and four liver voxels as indicated on the T1 weighted image (D), respectively. The tumor has a maximum diameter of $\sim 7$ cm, and its center is located at a distance of at least 8 cm from the coil. C shows the spectrum from four gallbladder voxels as indicated on the T2 weighted image (E). A, anterior; r, right; S, superior; other abbreviations as in Fig. 1.
of capecitabine metabolites was highly variable between patients, as is indicated in Table 1. In all of the patients 5’DFCR and 5’DFUR were MR detectable for a prolonged time compared with capecitabine. This is in accordance with pharmacokinetic parameters in plasma, which showed a longer t1/2 for 5’DFCR and 5’DFUR compared with capecitabine (14). In fasting patients a delay in occurrence of maximum plasma concentration and area under the plasma concentration curve has been described for capecitabine and its metabolites, with no effect on the apparent elimination half-life (15). In all of our patients capecitabine was administered in nonfasting condition, as has been the procedure in clinical trials.

5FU concentration was below MR detectable levels due to its rapid metabolic conversion. FBAL, a 5FU catabolite, was detected in both unlocalized (Fig. 1) and localized spectra (Fig. 3). Others have shown in humans that in urine FBAL was the major metabolite of capecitabine (16). The large peak in our spectra 2–2.5 ppm downfield from FBAL is usually assigned to the 5FU catabolite 5-fluoro-uridine-propanoic acid (17). At this spectral position a contribution from FBAL-bile acid conjugate has been described recently (18). Evidence from both 19 F nuclear MR spectroscopic studies (19) and high-performance liquid chromatography (20) indicate that the major biliary metabolites of 5FU are conjugates of FBAL. The amplitude of the peak in localized spectra from gallbladder voxels (Fig. 3C) supports this latter assignment. FBAL-bile acid conjugates would be able to undergo enterohepatic recirculation. PVI of 5FU may establish a higher pool of recirculating catabolite compared with bolus infusion of 5FU, resulting in higher biliary catabolite levels (18). The mimicking of conventional PVI of 5FU is one of the claimed properties of oral fluoropyrimidines. Because hepatic catabolite levels are correlated with toxicity in patients receiving PVI of 5FU (21), FBAL kinetics measured by 19 F MRS may be an early predictor of capecitabine-related toxicity.

If the correlation between the rate of 5’DFUR conversion and tumor response (5, 6) is confirmed in the clinical setting, 19 F MRS can also be used as a noninvasive method for predicting the antitumor response of capecitabine. This would be highly relevant, because advanced colorectal cancer is a frequently occurring cancer, and only a subset of patients responds to fluoropyrimidine-based therapy. We conclude that 19 F MRS of capecitabine metabolism or of other orally administered produgs (22) is a promising tool for the individualization of chemotherapy, especially when higher field strengths can be applied for better sensitivity.

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References

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