Cathepsin B Mediates Caspase-Independent Cell Death Induced by Microtubule Stabilizing Agents in Non-Small Cell Lung Cancer Cells

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Abstract

We have previously reported that the microtubule stabilizing agents (MSAs) paclitaxel, epothilone B and discodermolide induce caspase-independent cell death in non-small cell lung cancer (NSCLC) cells. Here we present two lines of evidence indicating a central role for the lysosomal protease cathepsin B in mediating cell death. First, inhibition of cathepsin B, and not of caspases or other proteases, such as cathepsin D or calpains, results in a strong protection against drug-induced cell death in several NSCLC cells. Second, MSAs trigger disruption of lysosomes and release and activation of cathepsin B. Interestingly, inhibition of cathepsin B prevents the appearance of multinucleated cells, an early characteristic of MSA-induced cell death, pointing to a central, proximal role for cathepsin B in this novel cell death pathway.

Introduction

Successful treatment of cancer with chemotherapy is largely dependent on its ability to trigger cell death in tumor cells. Activation of apoptosis has long been held responsible for the cell killing potential of anticancer drugs, but recent evidence suggests that there are forms of chemotherapy-induced cell death that cannot readily be classified as apoptosis or necrosis (1). Therefore, the contribution of the apoptotic cascade to the cytotoxic effects of chemotherapeutic agents and cytokines is currently being debated, and alternative forms of programmed cell death (PCD), such as “apoptosis-like PCD” and “necrosis-like PCD” have been described (2–4). Although caspases are well established as the main players in apoptosis, other proteases such as calpains, cathepsins, and serine proteases may account for alternative types of PCD (3). Cathepsins, in particular cathepsin B and D, act as the main executors of caspase-independent cell death induced by tumor necrosis factor α (TNF-α) in several systems (5, 6).

We have recently found that paclitaxel and two promising novel microtubule stabilizing drugs, epothilone B and discodermolide, exert their cytotoxic activity mainly via caspase-independent routes in non-small cell lung cancer (NSCLC) cell lines. This phenomenon was strongest for discodermolide and epothilone B, whereas paclitaxel-induced cell death was partially dependent on caspases and involved early triggering of the death receptor pathway, followed by activation of the mitochondria (7, 8). In the present study, we have investigated the potential role of other proteases in cell death induced by microtubule stabilizing agents (MSAs). Using a panel of specific protease inhibitors, we found that inhibition of the lysosomal protease cathepsin B, but not of caspases or other proteases such as calpains or cathepsin D, strongly protected against cell death, and prevented the development of multinucleated cells, an early feature of MSA-induced cell death. We further show that paclitaxel, epothilone B, and discodermolide trigger disruption of the lysosomes, followed by release and activation of cathepsin B, providing evidence for a novel, cathepsin B-dependent cell death pathway induced by MSAs in NSCLC cells.

Materials and Methods

Cell Culture and Reagents. The human NSCLC cell lines NCI-H460 (H460), SW1573, and A549 were cultured in RPMI or DMEM (BioWhittaker, Verviers, Belgium), supplemented with 10% (v/v) FCS (Life Technologies, Inc., Breda, the Netherlands), 100 units/ml penicillin, and 100 µg/ml streptomycin (Life Technologies, Inc., Grand Island, NY). Epothilone B and discodermolide were kindly provided by Dr. M. Wartmann (Novartis Pharma AG, Basel, Switzerland) and dissolved in DMSO. Paclitaxel (Bristol-Myers Squibb, Woerden, the Netherlands) was dissosed in ethanol. For each experiment, we freshly diluted stock solutions of the drugs in culture medium to their final IC\textsubscript{50} concentration (7, 8): for paclitaxel, 50 nM (H460), 15 nM (SW1573 and A549); for epothilone B, 2.0 nM (H460 and A549), 2.8 nM (SW1573); for discodermolide, 12 nM (H460), 19 nM (SW1573), and 26 nM (A549). zVAD-fmk (Enzyme Systems Products, Livermore, CA), calpeptin (Calbiochem, Darmstadt, Germany), CA-074 Me (Peptides International, Osaka, Japan), zFA-fmk (Enzyme Systems Products), and pepstatin A (Sigma Chemicals, St. Louis, MO) were dissolved in DMSO and added to the cells 1 h before the addition of the drugs at final concentrations of 50 µM (zVAD-fmk), 100 µM (calpeptin, CA-074 Me, pepstatin A), and 200 µM (zFA-fmk).

Flow Cytometric Analysis. The extent of apoptosis was determined by flow cytometry, using propidium iodide (Sigma) staining of hypodiploid DNA-content, as previously described (9). We performed lysosomal integrity assays with the lysosomotropic probe Lysotracker Red (Molecular Probes, Eugene, OR), according to the literature (10).

Plasmid Construction and Transfection. We created an expression plasmid encoding yellow fluorescent protein-tagged cathepsin B by cloning full-length human cathepsin B cDNA (generated by reverse transcription-PCR using primers 5’-ATC TGT TCT CGA GAT ATG CTC TGG GCC TCC-3’ and 5’-ATG GTA GGA TCC TTA GAT CTT TTC CCA GTA CTG-3’) into the pEYFPc1 vector (Clontech Laboratories, Inc., Palo Alto, CA). Cells were seeded onto glass coverslips in a 6-well tray and were transfected with 1.5 g of plasmid DNA using the FuGene6 transfection reagent (Roche Molecular Biochemicals, Mannheim, Germany), according to the manufacturer’s protocol.

Fluorescence Microscopy. We performed fluorescence stainings as described previously (7), using Hoechst 33342 (Sigma) or Lysotracker Red (Molecular Probes), according to the manufacturer’s protocol. Slides were mounted in Vectashield (Vector Laboratories, Burlingame, CA) and were analyzed with a Leica DM IRBE fluorescence microscope.

Protein extraction and Western Blotting. Preparation of total cell extracts as well as Western blot analysis was performed as described previously (9). For immunodetection, anti-cathepsin B monoclonal antibody (Oncogene Research Products, Boston, MA; 1:400 dilution) and horseradish peroxidase-conjugated secondary antibody (Amersham, Braunschweig, Germany; 1:2000 dilution) were used. Enhanced chemiluminescence (Amersham) was used for detection.

Statistics. We analyzed quantitative experiments by the Student’s t test. All P\textsubscript{s} results from the use of two-sided tests and were considered significant when <0.05.
Results

Identification of Cathepsin B as a Mediator of Caspase-Independent Cell Death Triggered by MSAs. To elucidate the caspase-independent pathways mediating cell death induced by MSAs, we evaluated the contribution of different cellular proteases, calpains, and cathepsin B and D, to the cytotoxic potential of paclitaxel, epothilone B and discodermolide. For this purpose, we screened several protease inhibitors for their ability to interfere with MSA-induced cell death in H460 cells. As shown in Table 1, the selective calpain inhibitor calpeptin did not reduce the cytotoxic effects of IC50 concentrations of either drug, as determined by propidium iodide-staining based fluorescence-activated cell sorter-analysis of hypodiploid cells, a method that we previously reported to correlate well with induction of cell death (7, 8). The lack of effect of calpain-inhibition was independent of the duration of drug exposure (24–72 h) or concentration of the inhibitor (20–200 μM; data not shown).

Interestingly, inhibition of cathepsin B by the selective chemical inhibitors CA-074 Me and zFA-fmk strongly protected H460 cells from induction of a hypodiploid cell population by paclitaxel, epothilone B, and discodermolide, thereby reducing their cell killing potential by 42 up to 70% (Ps < 0.05). In a dose-response experiment, low concentrations of the inhibitors that also protect against TNF-α-induced cell death (25 μM of CA-074 Me and 100 μM of zFA-fmk (5)) already reduced the cytotoxic effects of MSAs significantly (data not shown), whereas optimal results were obtained when higher concentrations were used (100 and 200 μM respectively; Table 1). Additional studies indicated that cathepsin B inhibition provided protection from 16 up to 96 h posttreatment (data not shown). Protection was not observed when cells were pretreated with pepstatin A, a selective inhibitor of cathepsin D. As a control in these experiments, we used the pancaspase inhibitor zVAD-fmk. In line with our previous reports (7, 8), zVAD-fmk did not protect significantly against the cytotoxic effects of epothilone B or discodermolide, and it only reduced paclitaxel-induced cell death from 32.6 to 26.5%. We next analyzed the effects of epothilone B or discodermolide, and it only reduced paclitaxel-induced cell death from 32.6 to 26.5%.

Table 1

<table>
<thead>
<tr>
<th>Inhibitor (target protease)</th>
<th>Percentage of cells with a hypodiploid DNA-content</th>
<th>Paclitaxel</th>
<th>Discodermolide</th>
<th>Epothilone B</th>
</tr>
</thead>
<tbody>
<tr>
<td>None (control)</td>
<td></td>
<td>32.6 ± 2.1%</td>
<td>32.5 ± 1.7%</td>
<td>33.2 ± 2.7%</td>
</tr>
<tr>
<td>zVAD-fmk (caspases)</td>
<td></td>
<td>26.5 ± 1.7%</td>
<td>30.8 ± 1.2%</td>
<td>31.2 ± 1.7%</td>
</tr>
<tr>
<td>Calpeptin (calpains)</td>
<td></td>
<td>35.8 ± 3.6%</td>
<td>32.0 ± 3.7%</td>
<td>31.9 ± 3.8%</td>
</tr>
<tr>
<td>CA-074 Me (cathepsin B)</td>
<td></td>
<td>10.8 ± 1.3%</td>
<td>14.2 ± 2.09%</td>
<td>14.9 ± 2.39%</td>
</tr>
<tr>
<td>zFA-fmk (cathepsin B)</td>
<td></td>
<td>14.9 ± 0.6%</td>
<td>18.9 ± 1.99%</td>
<td>14.5 ± 2.69%</td>
</tr>
<tr>
<td>Pepstatin A (cathepsin D)</td>
<td></td>
<td>32.0 ± 3.9%</td>
<td>30.5 ± 1.0%</td>
<td>34.1 ± 3.6%</td>
</tr>
</tbody>
</table>

*P < 0.05.

For epothilone B and discodermolide, the protection rates ranged from 2.1% to 2.7% and the active 25,000 or 31,000 forms (12). As shown in Fig. 1, the cathepsin B inhibitors protected against paclitaxel-induced cell death in all of the investigated cell lines (Ps < 0.05). It was observed that the reduction of cell death in SW1573 cells, in which CA-074 Me reduced paclitaxel-induced cell death from 33 to 2.5%. For epothilone B and discodermolide, the protection rates ranged from 45% in A549 cells to 83% in SW1573 cells and from 42% in H460 cells to 89% in SW1573 cells (Ps < 0.05; data not shown). Taken together, these results provide evidence for a central role of cathepsin B as a mediator of caspase-independent cell death induced by MSAs in NSCLC cell lines.

Cathepsin B Is Released from the Lysosomes and Activated on Treatment with Paclitaxel, Epothilone B, and Discodermolide. The lysosomal protease cathepsin B can be released from its cellular compartment on triggering with apoptotic stimuli such as TNF-α, resulting in tumor cell apoptosis (5). To assess whether treatment with MSAs results in a similar lysosomal membrane permeabilization, we performed lysosomal integrity assays using the lysosomotropic fluorescence probe Lysotracker Red. In H460 cells treated with paclitaxel, increased numbers of cells with a weak fluorescence (so-called “pale cells”) were detected (Fig. 2A), a phenomenon reported to be due to leakage of the probe into the cytoplasm, indicating lysosomal rupture (10). This effect was also observed in SW1573 cells (data not shown), or when cells were treated with epothilone B or discodermolide, and was already present from 8 h after treatment (Fig. 2B). Next, we studied whether the disruption of the lysosomal integrity would facilitate the release of cathepsin B. We, therefore, engineered a yellow fluorescent protein-tagged cathepsin B fusion protein into the expression vector pEYFP-C1, to allow easy detection by fluorescence microscopy. Co-localization studies with Lysotracker Red confirmed that cathepsin B-yellow fluorescent protein was present mainly in the lysosomes (data not shown). As shown in Fig. 2C, cathepsin B was released from the lysosomes of SW1573 cells on incubation with either drug, as indicated by the diffuse staining pattern of the fusion protein throughout the cells after treatment. As expected based on the integrity studies, this release was evident after 16 h of incubation, and was also observed in H460 cells (data not shown). Inhibition of cathepsin B only slightly prevented disruption of the lysosomes (data not shown), suggesting that cathepsin B can stimulate its own release, as reported previously (11), and confirming that cathepsin B release occurs downstream of the disturbance of the lysosomal membrane integrity. To examine the activation of cathepsin B after its release into the cytoplasm, we performed Western Blot analysis on cellular extracts derived from SW1573 cells, treated for 16 up to 72 h with paclitaxel or epothilone B. Cathepsin B is synthesized as an inactive pro-enzyme (M, 43,000) which is processed into active 25,000 or 31,000 forms (12). As shown in Fig. 2D, a clear increase in the cellular amount of the cleaved, active M, 25,000 product of cathepsin B was observed from 48 h after treatment with either agent, whereas levels of the inactive M, 43,000 pro-enzyme and the active M, 31,000 product remained unchanged. Incubation of H460 cells with paclitaxel or epothilone B produced similar results (data not shown). Taken together, these results indicate that lysosomal disruption and release and subsequent activation of cathepsin B accompany the induction of cell death in NSCLC cells by MSAs.

Inhibition of Cathepsin B Potently Protects Against Drug-Induced Nuclear Changes. Next, we studied the effect of cathepsin B inhibition on nuclear changes elicited by MSAs. Paclitaxel, epothilone B, and discodermolide trigger mitotic anomalies, resulting in abnormal chromosome segregation and the appearance of so-called “multinucleated cells” with multiple nuclei of various sizes. This early event is then...
Caspase-independent cell death triggered by paclitaxel, epothilone B, and discodermolide is protected against cell death, indicating that cathepsin B is essential for the caspases or other proteases such as calpains or cathepsin D, strongly supporting the role of cathepsin B in the cell death process in this type of cell. Inhibition of cathepsin B, and not of other proteases, effectively reduced cell death in NSCLC cells (7, 8). In this study, we demonstrate that MSAs can trigger cell death primarily via caspase-independent routes in NSCLC cells (7, 8), with discodermolide (data not shown). These findings confirm the pivotal role of cathepsin B in cell death in NSCLC cells, because the apoptotic cascade is functional and accounts for the cytotoxic activity of DNA-damaging agents (9). Our study, therefore, provides further evidence for the view that the cellular death response depends on the type and dose of chemotherapeutic stress within the cellular context.

Discussion

We have previously reported that paclitaxel, epothilone B, and discodermolide trigger cell death primarily via caspase-independent routes in NSCLC cells (7, 8). In this study, we demonstrate that MSAs can trigger disruption of lysosomes leading to release and subsequent activation of the lysosomal protease cathepsin B, representing an early stage of the cell death process in this type of cell. Inhibition of cathepsin B, and not of caspases or other proteases such as calpains or cathepsin D, strongly protected against cell death, indicating that cathepsin B is essential for the caspase-independent cell death triggered by paclitaxel, epothilone B, and discodermolide. Although our observation of cathepsin-B dependency in MSA-induced cell death is novel, involvement of lysosomal proteases in cell death is not unprecedented. Cathepsin D mediates PCD induced by IFN-γ, Fas, and TNF-α in HeLa cells (14), and this protease can activate cathepsin B in hepatocytes on treatment with camptothecin and thereby lead to cell death (6). The pivotal role of cathepsin B in hepatocyte apoptosis has further been demonstrated in cathepsin B knockout mice, which were resistant to TNF-α-mediated apoptosis and liver injury (15).

Strong evidence is now accumulating for the involvement of alternative proteases, such as cathepsin B, in PCD (16), but the molecular identity of the mediators and the necessity of activation of the apoptotic pathways remains to be elucidated in most cases and may vary on the type of cells and the applied death stimulus (17). For example, cathepsin B has been reported to contribute to apoptosis via induction of mitochondrial membrane permeabilization, possibly via cleavage of BID, in some systems, thereby acting upstream of the caspase cascade (18–21). Others, however, have found that cathepsin B can act as an effector protease, downstream of caspases (5, 22). Additionally, cathepsin B is capable of executing cell death completely independent of the apoptotic machinery in WEHI-S fibrosarcoma cells (5). In our system, the inhibition of caspases did not affect the cytotoxicity of epothilone B or discodermolide, and it only slightly reduced cell death by paclitaxel, indicating that cathepsin B is the main mediator of MSA-induced cell death in NSCLC cells, without requirement of the apoptotic cascade. Loss of mitochondrial membrane potential may, however, contribute to this process as a secondary event, because we have previously reported that these drugs trigger release of cytochrome c in a late stage of the cell death process (7, 8). In this respect, our data indicate that the three drugs may not act fully identically; whereas paclitaxel may partially exploit caspases as effector proteases, epothilone B and discodermolide induce cell death completely independently from the apoptotic machinery in NSCLC cells. Caspase-independent cell death is, however, not an intrinsic characteristic of this type of cell, because the apoptotic cascade is functional and accounts for the cytotoxic activity of DNA-damaging agents (9). Our study, therefore, provides further evidence for the view that the cellular death response depends on the type and dose of chemotherapeutic stress within the cellular context.
induced by microtubule-interacting agents and to evaluate a potential role of cathepsin B as a determinant of sensitivity to MSA-based chemotherapy.

Acknowledgments

We are grateful to Dr. M. Wartmann (Novartis Pharma AG, Basel, Switzerland), who kindly provided discodermolide and epothilone B.

References

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