P21Cip1 Is a Critical Mediator of the Cytotoxic Action of Thymidylate Synthase Inhibitors in Colorectal Carcinoma Cells

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ABSTRACT

We have demonstrated previously that interferon (IFN-γ) sensitizes human colon carcinoma cell lines to the cytotoxic effects of 5-fluorouracil combined with leucovorin and to the thymidylate synthase inhibitor, ZD9331, dependent on thymineless stress-induced DNA damage, independent of p53. Here we demonstrate that the cyclin-dependent kinase (CDK) inhibitor p21Cip1 regulates thymineless stress-induced cytotoxicity in these cells. HCT116 wild-type (wt) and p53−/− cells underwent apoptosis and loss in clonogenic survival when exposed to ZD9331, whereas p21Cip1−/− cells were resistant. In contrast, IFN-γ induced marked cytotoxicity in p21Cip1−/− cells only. ZD9331 induced p21Cip1 up-regulation in all of the cell lines examined, as did thymidine deprivation in thymidylate synthase-deficient (thymidine synthase−/−) cells. Furthermore, selective induction of p21Cip1 in RKO was sufficient to induce apoptosis. P21Cip1, cdk1, cdk2, and cyclin E mRNA expression increased coincident with S-phase accumulation in HT29 cells treated with ZD9331 or 5-fluorouracil/leucovorin, as demonstrated by cDNA microarray analyses. Cell cycle analyses revealed that HCT116 wt and p21Cip1−/− cells accumulated in S phase within 24 h of ZD9331 exposure; however, wt cells exited S-phase more rapidly, where apoptosis occurred before mitosis, either in late S or G2. Finally, the CDK inhibitor roscovitine potentiated the cytotoxic activity of ZD9331 in both wt and p21Cip1−/− cells, strongly suggesting a role for p21Cip1-dependent CDK inhibition in cytotoxicity induced by thymidylate synthase inhibition. In summary, p21Cip1 positively regulates the cytotoxic action of thymidylate synthase inhibitors, negatively regulates the cytotoxic action of IFN-γ, and enhances S-phase exit after thymineless stress, possibly via interaction with CDK-cyclin complexes.

INTRODUCTION

Colorectal cancer is a leading cause of cancer-related morbidity and mortality each year in the United States. Thymidylate synthase inhibitors, such as 5-fluorouracil, continue to maintain their role as first-line chemotherapy for the treatment of colorectal cancer. Efforts to maximize efficacy of thymidylate synthase inhibitor-based regimens have addressed ways to selectively modulate the cellular response to thymidylate synthase inhibition (1), as well as to develop more specific thymidylate synthase inhibitors including ZD9331 and ZD1694 (tomudex and raltitrexed). These folate-based inhibitors of thymidylate synthase hold promise for greater therapeutic selectivity compared with 5-fluorouracil/leucovorin (2). At this time, however, our understanding of key effectors downstream of DNA damage required for thymidylate synthase inhibitor-induced cytotoxicity remains limited.

Previous work in our laboratory has demonstrated synergism between 5-fluorouracil/leucovorin combined with interferon (IFN-γ) in tumor cell killing in human colorectal cancer, with promising preclinical (3) and clinical (4) activity. The interaction mechanism requires 5-fluorouracil/leucovorin-induced DNA damage and does not affect cells that demonstrate 5-fluorouracil-induced RNA-mediated cytotoxicity that occurs in normal gastrointestinal tissues (1, 5, 6) or certain human colorectal cancer cell lines (e.g., HCT8 and HCT116) (ref. 3). Thymineless stress-induced apoptosis in colorectal cancer cells devoid of thymidylate synthase activity (thymidylate synthase−/− cells; refs. 1, 7), 5-fluorouracil/leucovorin-induced cytotoxicity (3), and the synergistic interaction between 5-fluorouracil/leucovorin and IFN-γ (3, 4) all involve the Fas death receptor and its signaling pathway. More recently, we have demonstrated that IFN-γ is also synergistic in potentiating the cytotoxic effect of ZD9331 (2). In contrast to 5-fluorouracil/leucovorin + IFN-γ, ZD9331 +/− IFN-γ displays a broader range of cytotoxicity that occurs via DNA damage-induced cell death. In expanded studies with ZD9331, we demonstrated additional “apoptosis-related” sites of interaction between thymidylate synthase inhibitors and IFN-γ, independent of Fas, and downstream of the mitochondria, involving up-regulation and activation of specific caspases (2). All of the studies confirm that thymidylate synthase inhibition, either via conventional agents (5-fluorouracil/leucovorin) or via the newer thymidylate synthase inhibitors (ZD9331), induce apoptosis and cytotoxicity in human colorectal cancer cells independent of the cell-cycle/DNA-damage response protein p53. However, the role of the multifunctional cell cycle-related protein p21Cip1 in these models is less well characterized.

P21Cip1 functions at multiple levels (8), and although knowledge regarding the multifunctionality of this protein continues to increase, many of its functions remain to be determined. P21Cip1 is well known as a p53 response gene (9) capable of inhibiting multiple cyclin-dependent kinases (CDKs), resulting in the induction of G1 or G2 cell cycle arrest, assuring the normal progression of cells through the cell cycle (10). CDKs are protein kinases that, when complexed to the appropriate cyclin, enhance cell cycle progression (reviewed in ref. 10). P21Cip1 null cells, paradoxically, appear more susceptible to accumulation in an arrested phase of the cell cycle, including S phase (11). Additionally, deletion of p21Cip1 has been shown to induce uncoupling of S phase and mitosis in cells treated with anticancer agents (12), and p21Cip1-induced G1 arrest itself can lead to subsequent depletion of mitosis-control proteins and abnormal mitosis with endoreduplication in recovering cells (13). These cell cycle phenomena are presumed to be important in determining whether or not a cell will recover from a given form of DNA damage.

As an important mediator of such cell cycle phenomena, which are now known to play a critical role in apoptotic responses (14), it is not surprising that p21Cip1 has been reported to have both pro- and antiapoptotic roles. Overexpression of p21Cip1 in human cancer cell lines has been reported to induce apoptosis (15), implying a proapoptotic role, whereas overexpression of p21Cip1, leading to G1 arrest, can protect against p53-mediated apoptosis in other human cancer models (16). P21Cip1 may also be involved in the regulation of apoptosis induced by tumor necrosis factor α (17), tumor necrosis factor-related apoptosis-inducing ligand (18, 19), and Fas (20), as well as cytotoxic agents (11, 19, 21).

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In the current study we have demonstrated, using HCT116 isogenic cell lines, that p21CIP1−/− cells are highly resistant to the cytotoxic effects of thymidylate synthase inhibition mediated by ZD9331 and are highly sensitive to IFN-γ. In contrast, wild-type (wt) and p53 −/− cells are sensitive to ZD9331 and resistant to IFN-γ treatment alone, whereas the cytotoxic action of ZD9331 is potentiated by IFN-γ (2). All of the HCT116 isogenic cell lines were equally sensitive to 5-fluorouracil/leucovorin, mediated by RNA- dependent cytotoxicity and also to etoposide and doxorubicin. Taken together, the data suggest that p21CIP1 plays a critical role in the positive regulation of apoptosis and in the cytotoxic response to thymidylate synthase inhibitors, specific to thymineless stress-induced DNA damage, independent of p53 (2), whereas p21CIP1 is an important negative regulator of cytotoxicity induced by IFN-γ in human colorectal cancer cells. Additional investigations into cell cycle-related events are presented.

MATERIALS AND METHODS

Cell Lines. HT29 was obtained from American Tissue Type Culture Collection (Manassas, VA). The thymidylate synthase− cell line (GC1/thymidylate synthase c363) is a doubly cloned mutant isolated from wt GC/c (7). HCT 116 wt, p53−/−, and p21CIP1−/− cells were provided by Dr. Bert Vogelstein (Johns Hopkins University, Baltimore, MD), and RKO was obtained from Dr. Michael Kastan (St Jude Children’s Research Hospital, Memphis, TN). RKO cells with the edcsynone-ducible expression of p21CIP1 or p27KIP1, referred to as RKO p21 IND or RKO p27 IND, respectively, were obtained from Drs. Zhen Fan and Mathias Schmidt (The University of Texas M.D. Anderson Cancer Center, Houston, TX). Cultures were maintained in low folate-containing, thymidine (dThd)-free media composed of RPMI 1640 containing 10% dialyzed fetal bovine serum, 80 μM [6RS]-5-methyltetrahydrofolate, and 712 μM Ca2+ (2, 3). 

Clonogenic Survival Assays. Cell lines were plated at a density of 1,500 cells/well in six-well plates (Costar). After overnight attachment, cells were treated, in triplicate, with the various treatment conditions for 72 hours. Subsequently the media were aspirated and replaced with media supplemented with dThd (20 μM). Cells were harvested 5–7 days after treatment and clonogenic survival was evaluated as described previously (2, 3, 7, 22). ZD9331 was a generous gift from AstraZeneca.

Apoptosis Assays. Cell lines were plated at a density of 100,000–200,000 cells/well in six-well plates. After overnight attachment, HCT116 isogenic cell lines were each treated, in duplicate, with ZD9331 (50 μM) for 72 hours. RKO, RKO p21 IND, and RKO p27 IND cells were treated with ZD9331 (50 μM), Muristerone A (3 μM, Invitrogen, Carlsbad, CA), alone or in combination, for total exposures of 48 hours or 72 hours. All of the cells, both floating and attached, were combined after trypsinization. After fixation with 70% ethanol, samples were stored at −20°C. Before analysis, samples were prepared in 1 ml of Na2HPO4 buffer (pH 7.8) containing 50 μg/ml RNase and filtered. Apoptotic cells were detected as a sub-G1 fraction after propidium iodide staining and analyzed using a Becton Dickinson FACScan (2, 22).

Fas Expression. HCT116 isogenic cell lines were plated at a density of 200,000 cells/well in 12-well plates. After overnight attachment, cells were either untreated or treated with ZD9331 (100 ng/ml), in duplicate, for 24 hours. Cell surface Fas expression was determined by fluorescence-activated cell sorter analysis using standard procedures, as described previously (2, 22).

Cell Cycle Analysis. HCT116 wt and p21CIP1−/− cells were plated at a density of 200,000–400,000 cells/well in six-well plates. After overnight attachment, cells were either untreated or treated with ZD9331 (50 μM) for 24, 48, or 72 hours, in duplicate. At collection, the media (and floating cells) were discarded, and the remaining attached cells were washed with Hank’s solution, trypsinized, suspended in PBS solution and centrifuged at 1400 rpm × 10 min at 4°C. The liquid was aspirated and replaced with 1 ml of cell cycle buffer [10 mM Tris, 1.4-piperazineethanesulfonic acid (pH 6.8), 0.1 mol/L NaCl, 2 mMol/L MgCl₂, and 0.1% Triton X-100] mixed with RNa (100 ng/ml). The cells were suspended by repetitive pipetting and then incubated at 37°C for 30 min. Propidium iodide (20 μg/mL; 1 mg/ml) was added and cell cycle distribution determined by fluorescence-activated cell sorter analysis. G1−, S−, and G2/M-phase cells were quantitated using the Modfit software program (Verity Software House, Topsham, ME). M-phase analysis was conducted using immunofluorescent detection of phosphorylated histone H3 (23). HCT116 wt and p21CIP1−/− cells were either untreated, or treated with ZD9331 (50 μM) for 24, 48, 72, and 96 hours. Harvested cells (attached and floating) were washed in PBS, fixed in 70% ethanol, and stored at −20°C for up to 72 hours. After fixation, the cells were washed twice with PBS, resuspended in 1 ml of PBS containing 0.25% Triton X-100, incubated at room temperature for 15 min, and then resuspended in 100 μl of primary antibody solution [0.075 μg rabbit polyclonal antiphosphohistone 3 antibody (Upstate Biotechnology, Lake Placid, NY) in 100 μl of antibody buffer (sterile PBS containing 0.5% Tween and 5% fetal bovine serum)]. After 3-hour incubation at room temperature, the cells were rinsed with 1 ml of antibody buffer and resuspended in 100 μl of secondary fluorescein isothiocyanate-conjugated goat antirabbit antibody (Sigma-Aldrich, St. Louis, MO) diluted 1:30 in antibody buffer. After 1 hour at room temperature, cells were rinsed with antibody buffer and resuspended in 1 ml of 0.035 mg propidium iodide/1 ml PBS solution. After 1 hour of incubation at room temperature in the dark, the cells were analyzed using a Becton Dickinson FACScan.

cDNA Microarrays. HT29 cells, cultured in a 75 flask, were treated when 50–70% confluent with ZD9331 (50 μM) or 5-fluorouracil/leucovorin (3 μM/1 μM) for 24 hours. RNA was collected using standard techniques using RNAzolB (TEL-TEST Inc., Friendswood, TX), and subjected to quality control as described previously (2). The human cDNA microarray, constructed in the St Jude Children’s Research Hospital Hartwell Center for Bioinformatics, contains ~9,400 PCR products corresponding to human expressed sequence tags (the Human Unigene 1 collection) obtained from Incyte Genomics (Palo Alto, CA) designated as the H1 chip. Data files generated from the chip experiments were imported to Spotfire Decisionsite for Functional Genomics (described by Somerville, MA) for additional analysis. Outliers were identified using Cy5/Cy3 ratios after background subtraction, filtering of inappropriate spots (improper morphology, small size, and intensity/background ratio <1.4), and global normalization. Dye flight replicates were performed for every experiment. Average fold-changes and SDs were generated from analysis of four experiments for each treatment condition.

RNome Protection Assay. HT29 cells were either untreated or treated with various agents [IFN-γ (100 units/ml), 5-fluorouracil/leucovorin (3 μM/1 μM), or ZD9331 (50 μM)] for 24 hours, and RNA was extracted as described above. The RibovoQuant Multi-Probe RNome protection assay was performed according to the manufacturer’s specifications (PharMingen). The hCC-2 multiprobe template panel was used (p130, Rb, p107, p53, p57, p27KIP1, p21CIP1, p19, p18, p16, p14/15, L32, and GAPDH). Thymidylate synthase cells were grown in dThd-replete or -depleted media for 72 hours, and RNA was extracted as described above. The RNome protection assay was performed using the hStress-1 multiprobe template panel (p53, GADD45, c-Fos, p21, Bax, Mcl-1, L32, and GAPDH).

Western Analysis. HT29, RKO, RKO p21 IND, RKO p27 IND, and HCT116wt were either untreated or treated with ZD9331 (100 ng/ml) or muristerone (3 μM) for either 48 hours only or for 2, 8, 24, 48, and 96 hours, followed by determination of the level of expression of p21CIP1 (Oncogene/Calbiochem), p27KIP1 (BD PharMingen), or actin (Sigma) by Western analysis, as described previously (2, 24).

RESULTS

p21CIP1 Regulates the Cytotoxic Response to Thymidylate Synthase Inhibitors and IFN-γ in Colorectal Cancer. In clonogenic survival assays, ZD9331 demonstrated cytotoxicity in the nM range (IC50, approximately 30–50 nM) in HCT116wt and HCT116p53−/− cell lines (Fig. 1A; 2). In each case ZD9331-induced cytotoxicity was reversible by dThd, consistent with thymineless stress (and subsequent DNA damage) induced by thymidylate synthase inhibition as the predominant mechanism of cell death. IFN-γ potentiated the cytotoxic activity of ZD9331 in a synergistic manner in these cell lines, an effect also reversed by dThd, demonstrating the requirement for ZD9331-induced DNA damage in the mechanism of sensitization by IFN-γ (Fig. 1A; 2). In contrast, p21CIP1−/− cells were highly resistant to the effects of thymidylate synthase inhibition by ZD9331.
P21<sup>CIP1</sup> is required for ZD9331-induced loss in clonogenic survival and such requirement is specific to thymineless stress-induced cell death, whereas p21<sup>CIP1</sup> mediates resistance to IFN-γ. HCT116 isogenic cell lines were treated with various concentrations of ZD9331 (0–50 nm) either in the absence or presence of IFN-γ (100 units/ml) or dThd (20 μM) for 72 hours, or IFN-γ (0–1000 units/ml) in the absence or presence of ZD9331 (50 nm) for 72 hours (A). HCT116 isogenic cell lines (wt, p53<sup>−/−</sup>, p21<sup>CIP1</sup><sup>−/−</sup>) were treated with various concentrations of 5-fluorouracil/leucovorin (0–10 μM 5-fluorouracil/1 μM leucovorin), doxorubicin (0–50 nm), or etoposide (0–1000 nm) for 72 hours (B). Clonogenic survival is reported as percentage of control (untreated). Data (% of control) represent the mean of three determinations at each concentration of ZD9331; bars, ±SD.

**Fig. 1.** P21<sup>CIP1</sup> is required for ZD9331-induced loss in clonogenic survival and such requirement is specific to thymineless stress-induced cell death, whereas p21<sup>CIP1</sup> mediates resistance to IFN-γ. HCT116 isogenic cell lines were treated with various concentrations of ZD9331 (0–50 nm) either in the absence or presence of IFN-γ (100 units/ml) or dThd (20 μM) for 72 hours, or IFN-γ (0–1000 units/ml) in the absence or presence of ZD9331 (50 nm) for 72 hours (A). HCT116 isogenic cell lines (wt, p53<sup>−/−</sup>, p21<sup>CIP1</sup><sup>−/−</sup>) were treated with various concentrations of 5-fluorouracil/leucovorin (0–10 μM 5-fluorouracil/1 μM leucovorin), doxorubicin (0–50 nm), or etoposide (0–1000 nm) for 72 hours (B). Clonogenic survival is reported as percentage of control (untreated). Data (% of control) represent the mean of three determinations at each concentration of ZD9331; bars, ±SD.

(Fig. 1A; IC<sub>50</sub> approximately 1 μM; data not shown) but were exquisitely sensitive to IFN-γ (Fig. 1, A and B). Treatment of HCT116wt and p21<sup>CIP1</sup><sup>−/−</sup> cells with another pure thymidylate synthase inhibitor, Tomudex, yielded similar results (data not shown). Of note, whereas HCT116wt cells were completely resistant to IFN-γ alone (Fig. 1A), pharmacologically relevant IFN-γ concentrations (10 units/ml; 4) were sufficient to sensitize HCT116wt to ZD9331 (28% versus 16% survival) as well as to sensitize HT29 to 5-fluorouracil/leucovorin treatment (data not shown). Extremely low concentrations of IFN-γ alone were sufficient to induce a cytotoxic response in HCT116 p21<sup>−/−</sup> cells (25% loss in clonogenic survival at 1 units/ml; IC<sub>50</sub> < 3 units/ml; Fig. 1A).

p21<sup>CIP1</sup> Does Not Regulate the Cytotoxic Response to Other Investigated Agents. Agents that induce uridine-reversible RNA-mediated damage in HCT116 cells (5-fluorouracil/leucovorin), alkylation or topoisomerase I inhibition (doxorubicin), or topoisomerase II inhibition (etoposide) did not demonstrate preferential cytotoxicity in HCT116wt, p53<sup>−/−</sup>, or p21<sup>CIP1</sup><sup>−/−</sup> isogenic cell lines, with the exception that p53<sup>−/−</sup> cells maintained a slight increased resistance to 5-fluorouracil/leucovorin in survival assays when compared with wt or p21<sup>−/−</sup> cells (Fig. 1B). Data demonstrate that p21<sup>CIP1</sup> is required for thymineless-stress-induced cell death in HCT116, and the requirement for p21Cip1 is specific for cytotoxicity induced by this mode of cellular stress.

**p21<sup>CIP1</sup> Is Required for Thymineless Stress-Induced Apoptosis, Independent of Induced Elevation of Fas Cell-Surface Expression.** HCT116wt and p53<sup>−/−</sup> cells readily underwent an equivalent degree of apoptosis when exposed to ZD9331 (50 nm, 72 hours), whereas p21<sup>CIP1</sup><sup>−/−</sup> cells demonstrated minimal apoptosis above baseline after such exposure (Fig. 2A). Because the Fas signaling pathway is important in thymineless stress-induced apoptosis (3, 4, 7), we explored the possible role of deregulated Fas expression in resistance of p21Cip1<sup>−/−</sup> cells to the consequences of thymidylate synthase inhibition. Cell surface Fas expression was determined in HCT116 isogenic cell lines both before and after treatment with ZD9331 (50 nm, 24 hours). Data demonstrated that Fas expression was reduced in p53<sup>−/−</sup> cells but increased in p21<sup>CIP1</sup><sup>−/−</sup> cells, compared with wt controls, and all of the cell lines demonstrated an ~2.5-fold increase in Fas expression after 24-hour exposure to ZD9331 (Fig. 2B).

**Fig. 2.** ZD9331 induces apoptosis in HCT116 wt and p53<sup>−/−</sup> cells, dependent on p21<sup>CIP1</sup>, independent of Fas cell-surface expression. A, HCT116 isogenic cell lines were treated with ZD9331 (50 nm) for 72 hours and the sub-G<sub>1</sub> (apoptotic) fraction quantified via fluorescence-activated cell sorter analysis. B, fluorescence-activated cell sorter analysis of cell surface Fas expression in HCT116 isogenic cell lines after 24-hour exposure to ZD9331 (100 nm).
P21<sup>Cip1</sup> Promotes Exit from S Phase and Accumulation of Cells in Sub-G<sub>1</sub> after Thymidylate Synthase Inhibition. After treatment with ZD9331 (50 nM), HCT116 wt and p21<sup>Cip1</sup>-/- cells each accumulated in S phase by 24 hours (Fig. 3, A and B). By 48 hours, wt cells were exiting S phase and progressing into G<sub>2</sub>M, with evidence of a sub-G<sub>1</sub> apoptotic fraction (data not shown), in contrast to p21<sup>Cip1</sup>-/- cells, which remained in S phase. At 72 hours, the S-phase fraction in wt cells approximated control levels, and the sub-G<sub>1</sub> fraction was >35% (Fig. 2A). In contrast, S-phase arrest in p21<sup>Cip1</sup>-/- cells persisted, and no significant sub-G<sub>1</sub> fraction was detected. Data support a functional role for p21<sup>Cip1</sup> in facilitating S-phase exit in HCT116 wt cells after thymidylate synthase inhibition.

HCT116 wt Cells Undergo Apoptosis before M-Phase Entry after Thymidylate Synthase Inhibition. HCT116 wt and p21<sup>Cip1</sup>-/- cells were either untreated or treated with ZD9331 (50 nM) for up to 96 hours, and their M-phase fraction was analyzed. Representative graphic analyses at 72 hours are shown for each condition (Fig. 3C), and the average of duplicate independent experiments for all of the conditions and time points is tabulated (Fig. 3D). At 24 hours, the M-phase fraction was greatly reduced in wt and p21<sup>Cip1</sup>-/- cells, consistent with S-phase accumulation in both cell lines after treatment with ZD9331. As wt cells progressed through S and into G<sub>2</sub>M phases, the low M-phase fraction persisted, in contrast to p21<sup>Cip1</sup>-/- cells, which demonstrated an increased M-phase fraction at 48 hours. By 72 hours, dramatic differences were observed in control versus ZD9331-treated wt cells. Despite a decrease in M-phase in control cells due to increasing confluence (observed by confocal microscopy), treated cells demonstrated an almost nonexistent M-phase fraction at this time. At 96 hours, the M-phase fraction had returned to control levels in p21<sup>Cip1</sup>-/-treated cells. The sub-G<sub>1</sub> fraction, detected in ZD9331-treated wt cells, did not label positive for phosphohistone-H3, indicating that cells were actively undergoing apoptosis before entry into M-phase (Fig. 3C). Therefore, data are consistent with entry of wt cells into the sub-G<sub>1</sub> compartment after thymidylate synthase inhibition either late in S phase or in the G<sub>2</sub> phase of the cell cycle.

ZD9331-Induced S-Phase Arrest Leads to Increased Expression of Specific Cell Cycle-Related Genes. Treatment with ZD9331 (data not shown) or 5-fluorouracil/leucovorin (22) led to S-phase accumulation in various colorectal carcinoma cell lines, including HT29, by 24 hours. HT29 cells expressing mp53 were exposed to either ZD9331 (50 nM) or 5-fluorouracil/leucovorin (3 μM/1 μM) for 24 hours, and both treated and untreated cells were harvested for RNA isolation and genomic comparison via cDNA microarray analysis. Genes involved in cell cycle regulation were specifically assessed for fold changes after S-phase arrest. (The microarray data were mined via Spotfire software using the search phrases “cyclin” or “cell cycle.”) Genes that were either up-regulated or down-regulated by >50% are shown in Fig. 4A. Of the ~70 genes examined, only 6 genes demonstrated significant up-regulation by either ZD9331 or 5-fluorouracil/leucovorin (p21<sup>Cip1</sup>, CDK1, CDK2, CDC45, CDC6, and cyclin E1), and 4 (p27, CDK1, CDC45, and CDC6) were up-regulated by both drugs. (Complete analyses of the array data are available at the www tjudegersearch.org.) P21<sup>Cip1</sup> mRNA increased to the greatest degree (>3-fold) followed by CDK1 (>2-fold). CDCA20 was the only cell cycle-related gene assessed in our search to be down-regulated by either treatment. Verification of these findings was provided by RNase protection assay that focused on CDK inhibitors (p16, p18, p19, p21<sup>Cip1</sup>, p27<sup>Kip1</sup>, and p57), including p21<sup>Cip1</sup>. Only p21<sup>Cip1</sup> was up-regulated by either ZD9331 or 5-fluorouracil/leucovorin treatment in HT29 cells (Fig. 4B). Optical densitometry determinations demonstrated a selective 4–5-fold increase in p21<sup>Cip1</sup> mRNA expression in HT29 after thymidylate synthase inhibition (data not shown), closely correlating with the >3-fold up-regulation of this gene from cDNA microarray analyses. Furthermore, both assays were concordant with isolating p21<sup>Cip1</sup> as the only CDK inhibitor induced by thymineless stress in HT29 cells. In G<sub>c</sub>/c1 thymidylate synthase- cells, also expressing mp53, p21<sup>Cip1</sup> mRNA was up-regulated by 72 hours after dThd deprivation (Fig. 4B), at the time the cells were undergoing apoptosis and 50% loss in clonogenic survival (1, 7). Thymineless stress induced up-regulation of p21<sup>Cip1</sup> was confirmed at the protein level in all of the cell lines tested (HT29, HCT116 wt, and RKO; Fig. 4C and Fig. 5A).

Selective Overexpression of p21<sup>Cip1</sup> Is Toxic to RKO But Does Not Enhance ZD9331-Induced Cytotoxicity. Induction of p21<sup>Cip1</sup> or p27<sup>Kip1</sup> in isogenic edcsyone-inducible RKO cell lines demonstrated that ZD9331 induced expression of p21<sup>Cip1</sup> protein but not p27<sup>Kip1</sup> in all of the RKO isogenic cell lines (Fig. 5A). Furthermore, muristerone induced p21<sup>Cip1</sup> in RKO p21 IND by 8 hours and p27<sup>Kip1</sup> in RKO p27 IND by 2 hours (Fig. 5B). Significant G<sub>i</sub> arrest was evident by either induction by 24 hours (data not shown). A time-dependent increase in apoptosis was observed when p21<sup>Cip1</sup> was selectively induced, without any other intervention (Fig. 5C). P27<sup>Kip1</sup>
induction was also capable of inducing apoptosis at later time points (72 hours) in these cells but to a much lesser extent compared with p21Cip1 induction and serves as a vector-transfected control (Fig. 5C). Selective induction and overexpression of p21Cip1 or p27Kip1 simultaneously with ZD9331 treatment led to attenuated cytotoxicity compared with ZD9331 alone. Delayed induction of p21Cip1 or p27Kip1, 24 hours after ZD9331 exposure, did not attenuate or enhance ZD9331-induced cytotoxicity (Fig. 5, D [SCAP] and E). These data suggest that whereas induction of p21Cip1 is independently sufficient to induce apoptosis in this colorectal carcinoma cell line, such abnormally elevated p21Cip1 (or p27Kip1) expression causes the cells to undergo a G1 arrest that mitigates the effects of S phase-specific drugs. Similarly, pharmacological induction of p21Cip1 in various cell lines (HCT116 wt and HT29) with sodium butyrate demonstrated similar sequence-dependent attenuation of the effects of thymidylate synthase inhibition (data not shown).

CDK Inhibition by Roscovitine Potentiates ZD9331-Induced Cytotoxicity in HCT116 wt and p21Cip1−/− Cells. Exposure of HCT116 wt and p21Cip1−/− cell lines to the CDK inhibitor roscovitine (72 hours) demonstrated a dose-dependent reduction in clonogenic survival in both cell lines, with p21Cip1−/− cells demonstrating slightly increased sensitivity to this agent (IC50: wt, 13 μM; p21Cip1−/−, 7 μM; data not shown). Co-exposure with roscovitine (3 μM) and ZD9331 (50 nm and 100 nm) for 72 hours demonstrated potentiation of the cytotoxic activity of ZD9331 in wt and p21Cip1−/− cells. Roscovitine (3 μM) initiated ~10% loss in clonogenic survival in both cell lines, ZD9331 (100 nm) alone yielded ~90% loss in clonogenic survival in wt cells and ~20% in p21Cip1−/− cells, which are considerably more resistant to ZD9331. However, roscovitine (3 μM) + ZD9331 (100 nm) yielded 65% loss in clonogenic survival in p21Cip1−/− cells, more than double the anticipated additive effect. Similar potentiation was evident in wild type cells (Fig. 6) and p53−/− cells (data not shown). Data suggest that inhibition of CDK function, normally performed endogenously by p21Cip1, may be important in the mechanism of thymineless stress-induced cytotoxicity.

DISCUSSION

The role of p21Cip1 in influencing chemosensitivity in colorectal cancer remains to be clearly defined. The isogenic HCT116 wt and p21Cip1−/− cell lines have been used by several investigators to explore the role of p21Cip1 in chemotherapeutic response in vitro. Studies have demonstrated increased sensitivity of p21Cip1−/− cells to anticancer agents compared with wt cells, supporting the contention that p21Cip1 functions as an antiapoptotic protein (11, 25, 26). It has been reported that p21Cip1−/− cells are more sensitive to the apoptosis-inducing effects of doxorubicin (27); however, we have demonstrated that neither doxorubicin nor etoposide induce preferential cytotoxicity in p21Cip1−/− cells, determined in clonogenic survival assays, in contrast to apoptosis assays (27). More importantly, we have demonstrated that p21Cip1 is required for maximal cytotoxicity induced by thymineless stress in colorectal carcinoma, suggesting a proapoptotic role for p21Cip1 in this setting.

The endogenous CDK inhibitor, p21Cip1, has been reported to play a role in the regulation and promotion of apoptosis, in contrast to its antiapoptotic role, in other settings. One report demonstrated 50% reduction in sensitivity of HCT116 p21Cip1−/− cells to apoptosis induced by manumycin and paclitaxel, with concomitant decrease in caspase-3 activity (28). Induced p21Cip1 expression in human carcinoma cell lines also induced apoptosis, similar to our findings in RKO (14, 29). Li et al. (30) demonstrated that flavopiridol-induced p21Cip1 expression in Rb-deficient osteosarcoma cells, with increased cdk2 inhibition, yielded enhanced doxorubicin sensitivity, placing p21Cip1 and cdk2 interactions in the center of the apoptotic pathway in this model. The CDK inhibitor olomoucine, p21Cip1 antisense, or cdk1 antisense have each inhibited the induction of apoptosis in human Hep3 cells, confirming an important role for p21Cip1 and cdk1 in the induction of retinoic acid-induced apoptosis (31). Importantly, in addition to colorectal cancer cell lines HCT116 and RKO, p21Cip1 has been shown to enhance cytotoxicity from platinum drugs in the colorectal cancer cell line DLD-1 (32). Thus, p21Cip1 (14, 29, 30, 32) and its CDK substrates (21, 30, 31, 33–35) appear to be able to promote apoptosis, with or without concomitant chemotherapeutic...
agents, and the consistent enhancing effect of \( p21^{\text{Cip1}} \) in colorectal cancer is noteworthy given the prognostic role of \( p21^{\text{Cip1}} \) in such patient tumor samples.

This report and others (25) also demonstrate that cells lacking \( p21^{\text{Cip1}} \) are intrinsically sensitive to the cytotoxic effects of IFN-\( \gamma \) in contrast to wt cells that demonstrate resistance to IFN-\( \gamma \). Therefore, within the same isogenic cellular system (HCT116), \( p21^{\text{Cip1}} \) appears to promote cytotoxicity from thymineless stress (proapoptotic) as well as to antagonize cytotoxicity from IFN-\( \gamma \) (antiapoptotic). Several hypotheses have been proposed to explain the antiapoptotic role of \( p21^{\text{Cip1}} \). Firstly, \( p21^{\text{Cip1}} \) can induce cell cycle arrest in G1-phase thereby protecting cells from DNA damage-inducing agents. Secondly, \( p21^{\text{Cip1}} \) can bind to procaspase-3, thereby preventing cleavage to active caspase-3. Thirdly, either direct inhibition of stress-activated protein kinases by \( p21^{\text{Cip1}} \) or association between cytoplasmic \( p21^{\text{Cip1}} \) and ASK1 can suppress ASK1 activity and mitogen-activated protein kinase cascade activation (8). With regard to the ability of \( p21^{\text{Cip1}} \) to induce G1 arrest, we have demonstrated that induction and overexpression of \( p21^{\text{Cip1}} \) alone in RKO (RKO p21 IND) induces apoptosis not attributable to the inducing agent muristerone. However, this overexpression of \( p21^{\text{Cip1}} \) also promotes an initial G1 arrest in RKO (data not shown), thereby attenuating the cytotoxic effect of the S-phase-specific thymidylate synthase inhibitor, ZD9331. This is similar to a reported role for \( p21^{\text{Cip1}} \) in protection of RKO cells when exposed to cytotoxic agents (36). Consistent with this hypothesis is the finding by Rau et al. (37) who demonstrated that induction of \( p21^{\text{Cip1}} \) over basal expression levels led to radiochemotherapeutic resistance in vivo in patients with rectal carcinoma. Thus, the cell cycle modulating effect of \( p21^{\text{Cip1}} \), specifically G1 arrest, is likely to be antagonistic to the cytotoxic effects of various chemotherapeutic agents, as is overexpression of \( p21^{\text{Cip1}} \). It is likely that the endogenous level of \( p21^{\text{Cip1}} \) expression, its subcellular localization, and other factors regulating its function each play a role in determining cellular fate after drug exposure.

CDK activity, controlled by cyclins, \( p21^{\text{Cip1}} \), and other CDK inhibitor-related proteins and processes, in the context of thymidylate synthase inhibition, has received limited investigation. Yin et al. (38) demonstrated in head and neck carcinoma cell lines deficient in both p53 and \( p21^{\text{Cip1}} \) that thymidylate synthase inhibition by tomudex led to increased cyclin E, cdk2, and cyclin A expression. Increased cyclinE-cdk2 activity correlated with effective thymidylate synthase inhibition, decreased E2F1, decreased DNA synthesis, and S-phase arrest. Furthermore, they showed that thymidylate synthase inhibition by tomudex in HCT-8 led to induction of \( p21^{\text{Cip1}} \) (38). Although no direct correlation of increased CDK activity to cytotoxicity was reported, these findings are consistent with our findings of thymidylate synthase inhibitor-induced expression of \( p21^{\text{Cip1}} \), cdk2, and cyclin E in HT29 arrested in S phase after ZD9331-induced thymidylate synthase inhibition. In other p21+/− cellular systems, human breast cancer cell lines forced to arrest in S phase after thymidylate synthase inhibition by tomudex demonstrated increased levels of E2F-1, cyclin A, and cdk2 levels that appeared to correlate with nonspecific cell cycle changes rather than to cytotoxicity. They also showed that induction of \( p21^{\text{Cip1}} \) reversed tomudex resistance in MDA-bcl7 cells (bcl-2 overexpressing clones), consistent with our findings of the requirement for \( p21^{\text{Cip1}} \) in the apoptosis-promoting effect of thymineless stress in colorectal carcinoma (39). Li et al. (40) also demonstrated increased sensitivity to tomudex after induction of \( p21^{\text{Cip1}} \) in SaOS-2 osteosarcoma cells, extending the connection between \( p21^{\text{Cip1}} \) and thymidylate synthase inhibition to sarcomas. Interestingly, thymidylate synthase itself has been found to down-regulate \( p21^{\text{Cip1}} \) expression in cancer models (41).

The role of \( p21^{\text{Cip1}} \) in mediating S-phase exit and the consequences of S-M phase asynchrony are particularly intriguing. Prolonged S-phase accumulation in \( p21^{\text{Cip1}} \)-/− cells compared with wt cells, supporting a role for \( p21^{\text{Cip1}} \) in mediating G0/S transition (42), has also been demonstrated. The role of \( p21^{\text{Cip1}} \) in mediating S-phase exit and the consequences of S-M phase asynchrony are particularly intriguing. Prolonged S-phase accumulation in \( p21^{\text{Cip1}} \)-/− cells compared with wt cells, supporting a role for \( p21^{\text{Cip1}} \) in mediating G0/S transition (42), has also been demonstrated.
tion of thymineless stress, as demonstrated herein, is similar to the effect reported after exposure of these cells to nitrogen mustard (11). Additionally we demonstrate that pure thymineless stress (thymidylate synthase- cells) or drug-induced thymineless stress (ZD9331) each strongly induce p21Cip1 expression, independent of p53. Reports have suggested that at low expression levels, p21Cip1 may enhance thymidylate synthase inhibition in the p21Cip1/H9253 line (3), has been shown by others to be cytotoxic in various models including colorectal carcinoma cells (45, 46). The finding that potentiating synergistic interaction in multiple colorectal cancer cell lines as presented here and elsewhere (2–4). Induction or overexpression of p21Cip1 via genetic or pharmacological means, which would promote G1 arrest is not likely to be successful in exploiting this finding, and it is likely that endogenous levels of p21Cip1, its subcellular distribution, as well as cell type- and cell cycle-specific effects are important in determining the ultimate outcome from manipulation of this protein pharmacologically. Clarification of how the key downstream effectors of cytotoxicity from these agents play their part in the induction of apoptosis requires additional investigation.

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P21\textsuperscript{Cip1} Is a Critical Mediator of the Cytotoxic Action of Thymidylate Synthase Inhibitors in Colorectal Carcinoma Cells


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