Spermine Oxidation Induced by Helicobacter pylori Results in Apoptosis and DNA Damage: Implications for Gastric Carcinogenesis

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Abstract

Oxidative stress is linked to carcinogenesis due to its ability to damage DNA. The human gastric pathogen Helicobacter pylori exerts much of its pathogenicity by inducing apoptosis and DNA damage in host gastric epithelial cells. Polyamines are abundant in epithelial cells, and when oxidized by the inducible spermine oxidase SMO(PAOh1) H2O2 is generated. Here, we report that H. pylori up-regulates mRNA expression, promoter activity, and enzyme activity of SMO(PAOh1) in human gastric epithelial cells, resulting in DNA damage and apoptosis. H. pylori-induced H2O2 generation and apoptosis in these cells was equally attenuated by an inhibitor of SMO(PAOh1), by catalase, and by transient transfection with small interfering RNA targeting SMO(PAOh1). Conversely, SMO(PAOh1) overexpression induced apoptosis to the same levels as caused by H. pylori. Importantly, in H. pylori-infected tissues, there was increased expression of SMO(PAOh1) in both human and mouse gastritis. Laser capture microdissection of human gastric epithelial cells demonstrated expression of SMO(PAOh1) that was significantly attenuated by H. pylori eradication. These results identify a pathway for oxidative stress-induced epithelial cell apoptosis and DNA damage due to SMO(PAOh1) activation by H. pylori that may contribute to the pathogenesis of the infection and development of gastric cancer.

Introduction

Helicobacter pylori is a Gram-negative microaerophilic bacterium that selectively colonizes the human stomach and causes chronic gastritis, peptic ulcers, and gastric cancer. Despite inciting substantial acute and chronic immune and inflammatory responses, H. pylori infection generally persists for the life of the host, in part due to its ability to evade the antimicrobial effects of the immune response (1). We have recently reported that one potential cause of the ineffective immune response is the induction of apoptosis in macrophages caused by oxidation of polyamines resulting in generation of H2O2 (2). Apoptosis of gastric epithelial cells in H. pylori infection has been an area of focus in both in vivo (3) and in vitro studies (4). It may be an important contributing factor in the increased epithelial permeability and mucosal damage, and it has been associated with compensatory cell proliferation (5), all of which contribute to both the inflammation and risk for carcinogenesis. Additionally, DNA damage has been reported in H. pylori-infected gastric epithelial cells in vitro (6) and in vivo (7). We now report that apoptosis and DNA damage in gastric epithelial cells infected with H. pylori are mediated by spermine oxidase [SMO(PAOh1); refs. 8 and 9]. SMO(PAOh1) expression and activity are induced by H. pylori; the resulting H2O2 generation, apoptosis, and DNA damage are blocked by inhibition of polyamine oxidation; and silencing of SMO(PAOh1) expression prevents apoptosis and DNA damage. Our data are the first to demonstrate the induction of polyamine oxidation by a microbial pathogen and to link oxidative stress by this pathway to apoptosis, DNA damage, and potentially carcinogenesis.

Materials and Methods

Bacteria and Cells. H. pylori strains 60190 and SS1 were grown under microaerobic conditions as described previously (10). The human gastric epithelial cell line AGS was grown in F12 medium (11). Experiments were performed in antibiotic-free medium with 10% fetal bovine serum.

Reverse Transcription-Polymerase Chain Reaction and Real-Time Polymerase Chain Reaction. Total RNA was isolated from AGS cells and cDNA synthesized, and primer sequences and polymerase chain reaction (PCR) product sizes for β-actin and PAO1 [mouse homologue of SMO(PAOh1)] and PCR conditions for the multiplex reactions were as reported (2). Primer sequences for human SMO(PAOh1) were: sense, 5-GAC-CACAATCACGACACTGG-3, and antisense, 5-ATGACACCATG-GACACG-3, yielding a 160-bp product. Real-time PCR was performed using SYBR Green (2). For AGS experiments, relative expression of SMO(PAOh1) was determined using β-actin as the internal control (2). In tissue studies, SMO(PAOh1) expression was normalized to 18S rRNA.

SMO(PAOh1) Promoter Activity. A genomic DNA plasmid library was constructed from human A549 adenocarcinoma cell DNA in the pBluescript SK(--) plasmid and screened with a cDNA probe homologous to exon 1 of SMO(PAOh1). From this library, a clone containing ~4479 bp to the transcriptional start site was identified. Deletion constructs were generated by restriction enzyme digestion and subcloned into pGL-2 basic. AGS cells were transiently transfected (2) with 200 ng of the above constructs and luciferase activity performed according to the manufacturer’s instructions (Promega, Madison, WI).

SMO(PAOh1) Activity. Lysates of AGS cells were analyzed by a chemiluminescence assay as described previously (2, 12), and the activity was expressed as nanomoles of H2O2 per minute per milligram protein.

Measurement of H2O2. AGS cells were incubated with 10 μM/L CM-H2DCFDA and intracellular H2O2 detected by flow cytometry as described previously (2). For measurement of H2O2 in supernatants, 5 × 105 cells were plated in 24-well plates. After stimulation, cells were washed and incubated with 50 μM/L Amplex Red reagent (Molecular Probes, Eugene OR) and 0.1 unit/mL horseradish peroxidase for 30 minutes at 37°C. Plates were read using a microplate reader at 560 nm, and a standard curve with varying dilutions of H2O2 was used (2).

Assessment of Apoptosis. Apoptosis was assayed using an annexin V-fluorescein isothiocyanate apoptosis detection kit (Oncogene Research Products, San Diego, CA) according to the manufacturer’s instructions. Cells reported in H. pylori-infected gastric epithelial cells in vitro (6) and in vivo (7). We now report that apoptosis and DNA damage in gastric epithelial cells infected with H. pylori are mediated by spermine oxidase [SMO(PAOh1); refs. 8 and 9]. SMO(PAOh1) expression and activity are induced by H. pylori; the resulting H2O2 generation, apoptosis, and DNA damage are blocked by inhibition of polyamine oxidation; and silencing of SMO(PAOh1) expression prevents apoptosis and DNA damage. Our data are the first to demonstrate the induction of polyamine oxidation by a microbial pathogen and to link oxidative stress by this pathway to apoptosis, DNA damage, and potentially carcinogenesis.
As shown in Fig. 1C, the time course of the induction of apoptosis closely paralleled that of SMO(PAOh1) activity.

**H. pylori-Induced Spermine Oxidation Results in H<sub>2</sub>O<sub>2</sub>-Mediated Apoptosis.** To confirm that the SMO(PAOh1)-produced H<sub>2</sub>O<sub>2</sub> was causally linked to the observed apoptosis, the effects of an oxidase inhibitor (MDL 72527), H<sub>2</sub>O<sub>2</sub> detoxifying agent (catalase), and an SMO(PAOh1)-specific small interfering RNA were examined. *H. pylori* induced a significant increase in intracellular H<sub>2</sub>O<sub>2</sub> (Fig. 1D) that was prevented by the inhibition of SMO(PAOh1) by MDL 72527 or by the addition of catalase. We also used the Amplex Red assay, specific for H<sub>2</sub>O<sub>2</sub> in the medium, to demonstrate that *H. pylori* induced a significant, 2.7 ± 0.1-fold increase in extracellular H<sub>2</sub>O<sub>2</sub> that was inhibited by 79.6 ± 7.1% with MDL 72527 and 100.4 ± 7.8% with catalase (*P < 0.01 for H. pylori versus control and for H. pylori + inhibitors versus H. pylori alone; data not shown*). Consistent with the H<sub>2</sub>O<sub>2</sub> data, apoptosis induced by *H. pylori* was significantly attenuated by MDL 72527 and catalase, as shown in Fig. 1E and F. We also confirmed these findings by analysis of apoptosis by DNA fragmentation enzyme-linked immunosorbent assay (data not shown).

Because MDL 72527 inhibits both PAO and SMO(PAOh1) (8, 9, 16), to determine whether the spermine oxidation-mediated apoptosis was specifically due to SMO(PAOh1), we transiently transfected AGS cells with a duplex small interfering RNA specific for SMO(PAOh1). This treatment significantly inhibited *H. pylori*-stimulated SMO(PAOh1) mRNA expression (Fig. 2A) and produced a 79.1 ± 8.1% inhibition of SMO(PAOh1) enzyme activity (Fig. 2B). This knockdown of SMO(PAOh1) was associated with a 71.6 ± 5.8% inhibition of apoptosis (Fig. 2C).

To confirm that SMO(PAOh1) has a causal role in gastric epithelial cell apoptosis, we transiently transfected AGS cells with a full-length cDNA for SMO(PAOh1). There was a significant increase in apoptosis with SMO(PAOh1) transfection that was similar to the level of increase with *H. pylori* stimulation in mock-transfected cells (Fig. 2D).

**H. pylori-Induced Deoxyribonucleic Acid Damage in Gastric Epithelial Cells Is Mediated by SMO(PAOh1).** The comet assay was used to directly visualize DNA damage morphologically. There was a marked increase in the size and intensity of the tail of the DNA fluorescence of the cells in the *H. pylori*-treated (Fig. 3B) versus untreated cells (Fig. 3A). Treatment of AGS cells with MDL 72527 (Fig. 3C) or catalase (Fig. 3D) resulted in reduction of damage. We quantitated the tail moment in >270 cells for each condition and found that there was a 4.4 ± 0.1-fold increase with *H. pylori* (*P < 0.01 versus control*) that was inhibited by 90.3 ± 4.1% with MDL 72527 and 87.6 ± 4.2% with catalase (*P < 0.01 for both inhibitors versus *H. pylori* alone). When we assessed 8-oxoguanosine binding by flow cytometry as an indicator of oxidatively damaged DNA, we found that stimulation with *H. pylori* resulted in increased fluorescence that was significantly attenuated with MDL 72527 or catalase (Fig. 3E) or transfection with SMO(PAOh1) small interfering RNA (Fig. 3F).

**SMO(PAOh1) Is Up-Regulated in H. pylori Gastritis Tissues and Down-Regulated with H. pylori Eradication.** To determine whether the *in vitro* observations were relevant in an *in vivo* setting, mouse and human tissues from *H. pylori*-induced gastritis were examined. There was a significant increase in mRNA expression of mouse PAO1 (Fig. 4A) and human SMO(PAOh1) (Fig. 4B) in *H. pylori* gastritis tissues. Levels of SMO(PAOh1) in human gastritis tissues from *H. pylori*-negative patients (Fig. 4B) were only modestly increased, whereas tissues from *H. pylori*-infected patients exhibited consistently higher levels of expression. Real-time PCR analysis in
the mouse tissues revealed a 3.8 ± 0.4-fold increase in *H. pylori* gastritis versus uninfected tissues (*P* < 0.01), and in human samples, there was a 2.5 ± 0.4-fold increase in *H. pylori*-negative gastritis and a 4.9 ± 1.5-fold increase in *H. pylori*-positive gastritis (*P* < 0.05 versus normal for *H. pylori* positive only). To confirm that SMO(PAOh1) was expressed in vivo in the gastric epithelium, we used RNA extracted from epithelial cells harvested by laser capture microdissection from *H. pylori*-infected gastric tissues (Fig. 4C). After eradication therapy, SMO(PAOh1) mRNA levels determined by real-time PCR decreased in each patient, with an 85.4 ± 7.5% inhibition compared with levels before treatment (*P* < 0.001).

**Discussion**

The involvement of oxidative stress in carcinogenesis is well established, both generally and in gastrointestinal cancers (17). However, the origins of the reactive oxygen species leading to DNA damage and cancer, in many cases, have not been identified. The results presented here describe the pathway that establishes the source of oxidative stress in human gastric epithelial cells as *H₂O₂* that is specifically produced from *H. pylori*-induced spermine oxidase activity and results in both apoptosis and DNA damage. These results suggest that one link between *H. pylori* infection and gastric cancer may be SMO(PAOh1)-produced *H₂O₂*.

*H. pylori* infection has been reported to cause production of *H₂O₂* in AGS human gastric epithelial cells, and it has been suggested that this may contribute to the carcinogenic process in *H. pylori*-induced gastric cancer (18). However, the source of the *H₂O₂* was not determined. Recently, it has been demonstrated that in glutathione peroxidase 1 and 2 (*Gpx1* and *Gpx2*) knockout mice, there is a high incidence of ileocolitis and microflora-associated cancers when mice are raised in conventional housing that includes infection with *Helicobacter* species (19), but when these mice are raised under germ-free conditions, they do not develop tumors. Because Gpx1 and Gpx2 are major detoxifying enzymes for *H₂O₂*, these results indicate that the...
reactive oxygen species responsible for the inflammation and carcinogenesis is bacterial infection-induced \( \text{H}_2\text{O}_2 \) production. Here, we demonstrate that \( \text{H. pylori} \) exposure leads to increased \( \text{H}_2\text{O}_2 \) production in AGS cells that can be inhibited by MDL72527, indicating that a polyamine oxidase activity is responsible for the \( \text{H}_2\text{O}_2 \) production. It should also be noted that our results are not restricted to a single cell line, because we have found similar effects of MDL 72527 in MKN-28 gastric epithelial stimulated with \( \text{H. pylori} \) (data not shown). The SMO(PAOh1) activity is sufficient to produce DNA-damaging amounts of \( \text{H}_2\text{O}_2 \), as evidenced by both 8-oxoguanosine production and comet assay. Although generation of reactive oxygen species in response to \( \text{H. pylori} \) has been previously linked to DNA damage (6, 7), our studies provide a new mechanism for generation of reactive oxygen species in epithelial cells and directly demonstrate that the polyamine oxidation causes both apoptosis and DNA damage by this mechanism. The DNA damage and apoptotic cell death produced by \( \text{H}_2\text{O}_2 \) was inhibited by MDL 72527, catalase, and, most importantly, SMO(PAOh1)-specific small interfering RNA, thus demonstrating that the oxidase in question is, in fact, the spermine oxidase SMO(PAOh1) and not the classical \( \text{N}^1\text{-acetyl-polyamine oxidase}, \text{PAO} \) (16). These data combined with those from the transient transfection studies demonstrating that SMO(PAOh1) produces the same effects as \( \text{H. pylori} \) exposure confirm that the oxidation of spermine by SMO(PAOh1) is the source of \( \text{H}_2\text{O}_2 \) in \( \text{H. pylori} \)-induced DNA damage in gastric epithelial cells is ameliorated by MDL 72527, catalase, or transfection with SMO(PAOh1) small interfering RNA. A, reverse transcription-PCR analysis of cells transfected with either scrambled small interfering RNA (−) or SMO(PAOh1) small interfering RNA (+) in the absence and presence of \( \text{H. pylori} \) for 6 hours. Results from two separate experiments are shown. B, SMO(PAOh1) enzyme activity measured after 16 hours. C, summary data for total apoptosis (annexin V−) at 24 hours. In B and C, **, \( P < 0.01 \) versus unstimulated cells transfected with scrambled small interfering RNA control; §, \( P < 0.05 \); §§, \( P < 0.01 \) versus \( \text{H. pylori} \)-stimulated cells transfected with scrambled small interfering RNA. D, AGS cells were transfected with human SMO(PAOh1) in the pcDNA3.1 vector. Summary data of apoptosis determined by flow cytometry with total annexin V− cells shown. **, \( P < 0.01 \) versus unstimulated cells transfected with empty vector (Mock). n = 3 separate experiments in duplicate.
pylori-exposed AGS cells and is directly responsible for the observed downstream effects.

It is important to note the potential that some reactive oxygen species-induced mutations may not be lethal and may lead to growth and/or survival advantages. 8-Hydroxydeoxyguanosine, a common adduct downstream from H$_2$O$_2$ production, is known to produce G$\rightarrow$T and A$\rightarrow$C substitutions.

In summary, the results presented here demonstrate that *H. pylori* infection leads to the increased expression of an important polyamine catabolic enzyme, the spermine oxidase, SMO(PAOh1). This enzyme oxidizes spermine producing the DNA-damaging reactive oxygen species, H$_2$O$_2$. Because reactive oxygen species have been directly linked to the etiology of multiple cancers including *H. pylori*-induced gastric cancer, these data are entirely consistent with the hypothesis that *H. pylori*-induced SMO(PAOh1) activity is responsible for the genotoxic insult that results in tumorigenic transformation of affected gastric epithelial cells.

References

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