Combined DNA Methyltransferase and Histone Deacetylase Inhibition in the Treatment of Myeloid Neoplasms

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Abstract

Optimal reexpression of most genes silenced through promoter methylation requires the sequential application of DNA methyltransferase inhibitors followed by histone deacetylase inhibitors in tumor cell cultures. Patients with myelodysplastic syndrome or acute myeloid leukemia (AML) were treated with the methyltransferase inhibitor 5-azacitidine (aza-CR) followed by the histone deacetylase inhibitor sodium phenylbutyrate. Major responses associated with cytogenetic complete response developed in patients receiving prolonged dosing schedules of aza-CR. Bisulfite sequencing of the p15 promoter in marrow DNA during the first cycle of treatment showed heterogeneous allelic demethylation in three responding patients, suggesting ongoing demethylation within the tumor clone, but no demethylation in two nonresponders. Six of six responding patients with pretreatment methylation of p15 or CDH-1 promoters reversed methylation during the first cycle of therapy (methylation-specific PCR), whereas none of six nonresponders showed any demethylation. Gene demethylation correlated with the area under the aza-CR plasma concentration-time curve. Administration of both drugs was associated with the induction of acetylation of histones H3 and H4. This study provides the first demonstration that molecular mechanisms responsible for responses to DNA methyltransferase/histone deacetylase inhibitor combinations may include reversal of aberrant epigenetic gene silencing. The promising percentage of major hematologic responses justifies the testing of such combinations in prospective randomized trials. (Cancer Res 2006; 66(12): 6361-9)

Introduction

The malignant phenotype results from a combination of genetic abnormalities and epigenetic modifications, leading to dysregulation of critical genes controlling cell proliferation, differentiation, and death. Considerable data highlight the important role that methylation of cytosine residues in promoter regions plays in the transcriptional silencing of genes (1, 2). Methylated CpG dinucleotides bind specific proteins, such as MeCP2, which recruit transcriptional corepressors (3). These transcriptional inhibitory complexes include histone deacetylases (HDAC). DNA methyltransferases (DNMT), which catalyze DNA methylation, also bind to HDACs and have the potential to target these enzymes to regions of gene silencing (1, 4–7). Removal of acetyl groups from histone lysine tails is but one of several modifications made to these proteins now known to associate with transcriptional silencing (8, 9). In fact, aberrantly hypermethylated and silenced genes in cancer are now known to have key histone modifications associated with their promoter regions (10–13). However, in the context of all of these layered silencing chromatin modifications that affect such genes, DNA methylation seems to be dominant in specifying tight heritable transcriptional repression. Thus, in cultures of cancer cells, the administration of HDAC inhibitors will not result in reexpression of densely hypermethylated genes until a DNMT inhibitor is first given—the two inhibitors are then synergistic for reexpression of the genes (14, 15).

Given the above dynamics underlying the silencing of genes with hypermethylated promoters, the sequential application of HDAC inhibitors following DNMT inhibitors could potentially increase the response rate, response duration, or percentage of complete responses (CR) in myeloid leukemias through the reexpression of silenced important cell regulatory genes. Multiple such genes are known to exist in hematopoietic neoplasms, including the key cell cycle regulating gene p15

The marked clinical activity of the DNMT inhibitors 5-azacitidine (aza-CR) and 2'-deoxy-5-azacytidine (DAC) in patients with myelodysplastic syndromes (MDS) suggest that epigenetic silencing of cell regulatory genes may play important roles in the pathophysiology of this group of disorders. In addition to hematologic response rates ranging from 30% to 60% (20–23), both drugs have been shown to retard the progression of MDS to acute myeloid leukemia (AML; refs. 23, 24).

The presently described clinical trial was designed to determine whether the sequential administration of a DNMT inhibitor followed by an HDAC inhibitor could be tolerated by patients with myeloid malignancies and to explore the possibility that such combinations would increase the response rate or the quality of responses following aza-CR administration. We combined this drug with the first HDAC inhibitor available for clinical use previously studied by our group (25, 26), sodium phenylbutyrate. This short-chain fatty acid has been successfully used to induce the production of fetal hemoglobin in patients with sickle cell anemia and β-thalassemia (27, 28). In phase I studies in patients with AML
and MDS, steady-state plasma concentrations at the maximum tolerated dose ranged between 0.3 and 0.5 mmol/L (25, 26), well within the range that induces histone acetylation in vitro (29).

In this first exploration of the feasibility of combined DNMT and HDAC inhibition for the treatment of myeloid malignancies, we sequenced these drugs in a dose-finding study. Modeling the study after in vitro models, patients were treated with the DNMT inhibitor aza-CR followed by the HDAC inhibitor phenylbutyrate. In addition to clinical toxicity and response end points, this study aimed to determine to what extent aza-CR administration was associated with reversal of promoter methylation and whether clinical administration of phenylbutyrate was associated with induction of histone acetylation.

Materials and Methods

Patient Selection

The French-American-British categorization was used to classify patients with MDS and AML; when appropriate, the WHO category of refractory cytopenias with multilineage dysplasia (RCMD) was applied. AML arising from MDS is called either AML with trilineage dysplasia (AML-TLD) or AML-MDS. Patients with MDS age ≥18 years were eligible if they had refractory anemia (RA) with excess blasts (RAEB) or chronic myelomonocytic leukemia (CMML). Patients with RA or RA with ringed sideroblasts were eligible if they met one of several criteria for significant hematologic compromise: absolute neutrophil count <1,000/μL, transfused hemoglobin <8 g/dL, platelet count <20,000/μL, and anemia or thrombocytopenia requiring transfusion or if they had high-risk cytogenetic abnormalities. Patients with relapsed or refractory AML were eligible if WBC was stably <30,000/μL and again following the last dose of phenylbutyrate and before cycle 2. Peripheral blood was obtained during cycle 1 before study; before phenylbutyrate, 4, 8, and 24 hours following phenylbutyrate; and at least once more during the infusion of phenylbutyrate as well as before cycle 2. Peripheral blood and bone marrow mononuclear cells were isolated using Ficoll-Hypaque (SG ≤1.077). Mononuclear cells were divided into aliquots for histone extraction before freezing (see below) and RNA and DNA preparation using standard techniques before freezing.

Bisulfite sequencing. Genomic bisulfite sequencing for the p15 promoter was done in selected patient samples as described to validate changes in promoter methylation (31). Primers used were 5'-TGAAGGAATTAGAATAATTTTTTTGTTTT-3' and 5'-AAAGAAGCTTTAAGACCCCTTAACCCCTA- ACTACCTAA-3' for the initial amplification, followed by a nested PCR using the internal primers 5'-GGGGATTTAGGTTGTGAAC-3' and 5'-ACCCTTAAACCCCCCTTACC-3'. The large size of this product precluded amplification from bisulfite-treated DNA in some cases. For these samples and time points, a smaller nested PCR was done initially using the primer pair 5'-TGTGGTTTTTGGTTGTTGAAA-3' and 5'-CTCTTCTCTAAAACCTTACTCCAC-3', followed by internal primers 5'-TTTTTTTTTTTTTTTTTTT-3' and 5'-AAAAATTTTTTTTTTTTTT-3'.

Methylation-specific PCR. Methylation-specific PCR (MSP) was used to detect the presence of methylated alleles for specific genes. MSP was done as previously described (32, 33). All pretreatment samples were screened for promoter methylation of p15

Histone acetylation. Histones were extracted (34) and proteins were quantified (Bio-Rad, Hercules, CA). Fifty micrograms of protein were separated by 15% SDS-PAGE gels and transferred onto nitrocellulose membranes. Primary rabbit polyclonal antibodies to human acetylated histone H3 and H4 and nonacetylated H2 (Upstate Biotechnology, Lake Placid, NY) were used for Western blot analysis, followed by horseradish peroxidase–conjugated secondary antibodies and visualization using enhanced chemiluminescence (Kodak, Scientific Imaging systems, Rochester, NY). The non-acetylated H2b band was easily resolved from the Ac-H3 and Ac-H4 band and served as a control for histone extraction and protein loading. The gels were scanned and the band density was quantified (UN-Scan-It, Silk Scientific, Orem, UT). Beginning with dose cohort 3 results for Ac-H3 and Ac-H4 were quantified as percentage of nonacetylated H2A (“acetylation index”); routine blotting for H2A was not done in all patients before this dose cohort.

F-cell quantification. Red cells containing hemoglobin F (F cells) were quantified using a standard flow cytometric technique (28).
**Table 1. Patient characteristics**

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<tr>
<th>UPN</th>
<th>Age</th>
<th>Sex</th>
<th>Diagnosis</th>
<th>Dose cohort</th>
<th>Cytogenetics</th>
<th>International Prognostic Score</th>
<th>Previously treated</th>
<th>Comments</th>
<th>Response</th>
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<td>HI-Nmaj</td>
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<td>NE</td>
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<td>NE</td>
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<td>RAEB</td>
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<td>NE</td>
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<td>Hi-Pmaj, Hi-Nmaj</td>
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<td>Hi-Pmaj, Hi-Nmaj</td>
<td>CR</td>
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<td>CR</td>
</tr>
<tr>
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<td>M</td>
<td>RAEB</td>
<td>5</td>
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<tr>
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<td>NR</td>
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<td>NR</td>
<td>NR</td>
<td>NR</td>
</tr>
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</table>

NOTE: WNL indicates normal cytogenetic analysis. Patients who consented to study but were screen failures or withdrew consent before treatment are not displayed. Four screen failures were not included.

Abbreviations: NR, no response; HI-Nmaj, hematologic improvement neutrophils, major; HI-Pmaj, hematologic improvement, platelet, major; HI-Emin, hematologic improvement, erythroid, major.
### Results

#### Study Population

Thirty-six patients signed consent; four patients were screen failures. Characteristics of treated patients are listed in Table 1. Median age was 66 years (41-85 years). Thirteen patients had MDS (including two treatment-induced patients), 1 CMMoL, 15 AML-TLD (MDS-AML), and 3 relapsed AML. Ten of fourteen (71%, 95% confidence interval, 52-94) patients with MDS or CMMoL and 14 of 18 (78%, 95% confidence interval, 52-94) patients with AML had clonal cytogenetic abnormalities. International Prognostic Score categories among MDS/CMMoL patients were as follows: low, 1; Int-1, 7; Int-2, 4; high, 2 (40).

#### Toxicity

No unusual toxicities were discovered in this trial. Three patients withdrew consent before completing a single cycle: one because of needle phobia, one because of poor performance status, and one for undisclosed personal reasons. Mild nausea was frequent in patients receiving aza-CR and occasional in patients receiving phenylbutyrate; nausea was alleviated in all cases with either 10 mg prochlorperazine or 100 mg dolasetron. Injection site reactions were common in response to aza-CR. One patient in dose cohort 1 developed severe asthenia requiring a dose reduction of aza-CR to 50 mg/m²/d.

#### Aza-CR pharmacokinetics

Samples were obtained before treatment and on days 1, 2, 5, 7, 14, and 21 after aza-CR administration. Samples were processed and frozen within 30 minutes by centrifugation at 3,000 × g for 5 minutes in a refrigerated centrifuge. Plasma was divided into multiple aliquots to avoid freeze/thaw cycling and was stored at −80°C until analysis. Initially, samples were collected without tetrahydrouridine. Following observation of samples in which aza-CR could not be detected, presumably due to presumed breakdown by cytidine deaminase, tetrahydrouridine was added to the plasma supernatant at a final concentration of 100 μmol/L to increase the stability of drug in plasma during storage in the freezer. Tetrahydrouridine increased freezer stability from 7 to 21 days when frozen at −80°C (35, 36). Aza-CR was quantified in plasma using a previously described validated liquid chromatography/tandem mass spectrometry method over the range of 5 to 500 ng/mL (20.4 to 2,041.6 nmol/L; ref. 36).

Individual concentration-time data were analyzed using noncompartmental methods using WinNonlin Professional version 3.1 as reported elsewhere (Pharsight, Mountain View, CA; ref. 35). Pharmacokinetic variables were summarized using descriptive statistics.

#### Phenylbutyrate pharmacokinetics

Samples were obtained before treatment and on days 1 and 7 of phenylbutyrate infusion. Samples were collected in heparinized tubes (5 mL) and centrifuged at 3,000 rpm for 5 minutes; plasma was aliquoted and stored at −80°C until the time of analysis. Samples were analyzed for phenylbutyrate and two metabolites, phenylacetylglutamine and phenylacetic acid, using a high-performance liquid chromatography method using UV detection (25, 26, 37–39). The linear calibration curves were generated over the range of 10 to 3,000 μmol/L.

Average steady-state concentrations (Css) were calculated for phenylbutyrate, phenylacetylglutamine, and phenylacetic acid based on graphical presentation of concentration-time profiles.

#### Statistical considerations

The primary end points of this trial were to evaluate the toxicities and responses associated with the different dosing regimens. Toxicities were recorded for each individual. The responses were graded as described above and response rates were calculated as percentages with 95% confidence intervals both for the entire cohort as well as within each treatment group. Among the responders, duration of individual responses and the median response time were calculated. In addition, median cycle times and ranges are reported for duration of outcomes of interest, such as time until response.

Changes in biological variables over time were represented graphically with additional tests for associations. For each dose group, the association between the mean percentage of F cells and time was tested using a one-way ANOVA. ANOVA was used to determine the association between aza-CR or phenylbutyrate exposure and clinical response expressed as a categorical variable [i.e., 1, a major response (CR or partial response, PR); 2, minor response (i.e., hematologic improvement/lineage response); 3, no response]. A paired t test was used to determine the association between aza-CR or phenylbutyrate exposure and methylation changes, methylation reversal–induced F cells, and acetylation changes expressed as a categorical variable (i.e., response or no response). The aza-CR exposure variables explored were maximum plasma concentrations (Cmax), area under the concentration-time curve (AUC) from time 0 to 2 hours (AUC0-2), AUC from time 0 to 4 hours (AUC0-4), AUC from time 0 to the last quantifiable time point (AUC0-tl), and AUC extrapolated to infinity (AUC∞). The phenylbutyrate exposure variables explored were C∞ for phenylbutyrate, phenylacetylglutamine, and phenylacetic acid. Statistical analysis was done using JMP Statistical Discovery Software version 3.2.6 (SAS Institute, Cary, NC). The a priori level of significance was P < 0.05.

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### Table 2. Accrual to dose-finding trial of aza-CR plus phenylbutyrate in MDS and AML

<table>
<thead>
<tr>
<th>Cohort</th>
<th>Aza-CR dose (mg/m²/d)</th>
<th>Days of aza-CR</th>
<th>Patients assigned to cohort</th>
<th>Number not evaluable</th>
<th>Number reduced to this cohort</th>
<th>Number increased to this cohort</th>
<th>No. patients evaluated at cohort*</th>
<th>Responses total/major*</th>
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<tbody>
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<td>75</td>
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</table>

*For patients who underwent dose escalation or dose reduction, responses are tallied in their final dose cohort.
†Patient originally in dose cohort 4, reduced to dose cohort 3, then reduced to cohort 1.
‡Patient reduced from dose cohort 1.
§Both patients reduced from dose cohort 4; one of these patients was subsequently further reduced to dose cohort 1.
¶Patient dose increased from cohort 5 following lack of response after four cycles.
*Only nine patients were evaluated at this dose cohort because the patient dose reduced from dose cohort 4 was further dose reduced to cohort 1 and is tabulated in that cohort. The patient dose escalated from cohort 5 was a nonresponder after an additional four cycles at cohort 3, but is not included in this primary analysis of response.
encephalopathy was mild and the 7-day infusion could be successfully completed. Three patients received dose reductions of phenylbutyrate in subsequent treatment cycles (325 mg/kg/d). Two patients tolerated the lower dose; one patient developed severe recurrent encephalopathy and was removed from the trial following his third cycle. The final patient decided not to resume treatment upon recovery from initial symptoms of encephalopathy.

Dose-limiting hematologic toxicity was observed in dose cohort 4 (50 mg/m²/d × 14 days). These toxicities include dose-limiting neutropenia (one patient), dose-limiting thrombocytopenia (two patients), and septic death during neutropenia in cycle 2 (one patient). One surviving patient tolerated dose reduction to 50 mg/m²/d × 10 days; the other patient had recurrent dose-limiting neutropenia at both 50 mg/m²/d × 10 days and 75 mg/m²/d × 5 days and was removed from study. No dose-limiting toxicities occurred in dose cohort 5.

**Clinical Outcomes**

Three patients received less than one complete cycle of therapy and are not considered evaluable for response. Overall, 11 of 29 evaluable patients responded, including 5 of 9 evaluable patients in dose cohort 3 (one patient reduced from dose cohort 4) and 3 of 6 evaluable patients in dose cohort 5. Five were major responders (four CR/one PR); these responders were all treated in dose cohorts 3 (four patients) and 5 (one patient). Seven of 11 responders, including three of the major responders, had clonal cytogenetic abnormalities before treatment. The three major responders had complete cytogenetic responses; none of the minor responders developed cytogenetic remission. Major responses developed in two patients with AML-TLD, one patient with RAEB, one patient with RA with steroid-refractory MDS-associated vasculitis (complete hematologic and rheumatologic response), and one patient with RCDM (complete hematologic response with evidence of ongoing dysplasia in bone marrow). Response duration among major responders was 8, 10, 15, and 19+ months. The patient considered to have "progressed" at 8 months showed cytologic evidence of recurrent MDS with recurrent neutropenia following AML-TLD; complete cytogenetic response persisted.

Minor responses (IWG hematologic improvement-major) occurred in four patients with RAEB (including a patient with therapy-related MDS) and two patients with AML-TLD including four monolineage and two bilineage responses (Table 2). Median duration of response among minor responders was 3 months; four patients were removed from protocol with ongoing response because of persistent cytopenias in other lineages or persistent increases in blasts. Among patients who achieved major hematologic improvement, the median number of treatment cycles to observe response (range) were as follows: platelets 1.5 (1-4, n = 7); reticulocytes 3 (2-3, n = 6); hemoglobin 3 (2-4, n = 6); neutrophils 1 (1-5, n = 7). No trend was observed in time to response according to theaza-CR dose cohort.

**Changes in Promoter Methylation**

DNA methylation in bone marrow cells was examined during the first course of drug administration in dose cohorts 3 to 5 to monitor changes within tumor clones rather than monitoring alterations stemming from replacement of neoplastic cells by normal cells. Genomic bisulfite sequencing of a hypermethylated p15 promoter was done in five patients (Fig. 1). Sequential sequencing of DNA from patients who had significant responses to the combined therapy, patients 13 (PR, cohort 3), 19 (CR, cohort 3), and 24 (hematologic improvement, cohort 4, reduced to dose cohort 3) showed significant decrement in p15 methylation followingaza-CR treatment. Importantly, we saw a pattern of heterogeneous loss of CpG methylation within each single allele examined, strongly supporting demethylation within the tumor clone. The bisulfite sequencing of patient 19 showed evidence of remethylation before initiation of the next drug cycle. No demethylation was observed in thep15 promoter in two clinical nonresponders (patient 15, cohort 3; patient 34, cohort 5).

Twelve patients evaluable for clinical response were subsequently studied by MSP, a sensitive PCR assay for the methylation of multiple CpG sites in a CpG island (33) for p15 and/or CDH-1. The primers completely overlap the region of the p15 promoter sequenced in the above studies and a region of the CDH-1 gene, which we have used in multiple previous studies. All 12 patients were informative for pretreatment hypermethylation of either or both genes. Six patients developed reversal of methylation for either or both genes during the first cycle of treatment. The methylation status over time of the two genes changed in parallel in three of four patients informative at both loci (Fig. 2A). Particularly noteworthy is patient 19: No evidence of methylation of either gene was present followingaza-CR (day 10); however, some evidence of remethylation of both genes was present following phenylbutyrate (day 17). Most importantly, these six patients with reversed methylation of either gene all developed hematologic responses, including three CR patients, one PR, and two patients who achieved hematologic improvement. In contrast, the six patients who did not show methylation reversal were all nonresponders. The difference in response rates between the two groups was statistically significant (P = 0.002, Fisher’s exact test).

**Histone acetylation.** Changes in histone acetylation in peripheral blood and bone marrow mononuclear cells were monitored using Western analysis. A representative example is displayed in Fig. 3A. Eleven of 23 evaluable patients showed increases (minimum 50%) in global acetylation of H3 and/or H4 in response toaza-CR alone, similar to the example shown in Fig. 3A. This was observed in one of five patients in dose cohort 1, three of four in dose cohort 2, five of seven in cohort 3, and one of three each in cohorts 4 and 5 (P = not significant between cohorts). Further increases in acetylation in response to phenylbutyrate (minimum 50%) were found in 12 of 23 patients. Overall, 17 of 23 patients developed increased histone acetylation following eitheraza-CR or phenylbutyrate. Figure 3B displays the changes in histone acetylation from patients in dose cohort 3 semiquantitatively. Blots were stripped and reprobed with antibody to nonacetylated histone H2A. Gels were scanned and band intensity was quantified. The data were then reported as an acetylation index: band intensity acetylated H3 or H4/band intensity nonacetylated H2A. The greatest increment in acetylation appeared followingaza-CR administration; phenylbutyrate induced a smaller increment in histone acetylation in many patients. Acetylation remained elevated compared with baseline during phenylbutyrate infusions even in patients who showed no further increment in acetylation in response to phenylbutyrate.

**F cells.** Changes in percentage ofRBC containing hemoglobinF (F cells) were monitored as an indirect measure of induction of expression of a surrogate gene silenced through epigenetic mechanisms (globin). Baseline F-cell data were obtained on 26 patients who consented to study. The mean percentage F cells was 7.9% (median 4.9%, range 0.9-48.3%). There was no statistical difference between the mean percentage F cells of patients with the diagnoses of MDS or AML nor was there a statistical difference in the proportion of patients with each diagnosis in patients whose F cells were above or below the mean.
Sequential F-cell data were obtained on 24 patients (Fig. 4). Six patients increased percentage of F cells over time. Three patterns of F-cell induction were observed: a gradual induction throughout the first cycle of therapy (three patients, all in dose cohort 3); delayed induction, coincident with clinical response (one patient in dose cohort 3 and one in dose cohort 5); and transient increase in F-cell production in response to phenylbutyrate (one patient in dose cohort 2). Each cohort was examined for a correlation between time point and F-cell percentage by ANOVA. An association between F cell and treatment time, indicating an increase in F cells over time, was found only dose cohort 3. Three of four clinical responders in cohort 3 who were evaluated for F-cell response developed significant increases in F cells. The other patient in this cohort with an F-cell response did not respond clinically. The patients in cohorts 2 and 5 who developed significant increases in F cells were clinical nonresponders. No significant correlation between changes in F cells and clinical response was found when all evaluable patients were examined.

**Aza-CR pharmacokinetics.** The results of the aza-CR pharmacokinetics, combined with results from a trial in nonhematologic tumors, have been reported elsewhere (35). The current analysis includes data beginning with patient 18; tetrahydrouridine, required for stability of aza-CR, was added to the plasma starting with this patient. Aza-CR was rapidly absorbed with the mean $T_{\text{max}}$ occurring at 0.50 hour. Average maximum concentration ($C_{\text{max}}$) and area under the curve (AUC$_{0-\infty}$) values increased in a dose-proportionate manner with increasing dose from 25 to 75 mg/m$^2$/d; the mean ± SD $C_{\text{max}}$ and AUC$_{0-\infty}$ at 25 mg/m$^2$/d were 1,612 ± 1,042 nmol/L and 1,941 ± 980 h × nmol/L, respectively, and at 75 mg/m$^2$/d was 6,207 nmol/L and 5,831 h × nmol/L, respectively. Aza-CR has a short terminal half-life of 1.04 ± 0.88 hours.

A trend to correlation between aza-CR AUC$_{0-\infty}$ and reversal of methylation was detected among the small number of patients and clinical response was found when all evaluable patients were examined.
with complete methylation and pharmacokinetic data \((n = 9, P = 0.0658; \text{Fig. 2B})\). Aza-CR exposure was not correlated with clinical response, changes in histone acetylation, or induced F cells.

**Phenylbutyrate pharmacokinetics.** The median steady-state phenylbutyrate plasma concentration was 322.5 \(\mu\text{mol/L}\) (mean 562.7 \(\pm\) 818.9, range 19.5-4,371.2). For phenylacetic acid, the median steady-state plasma concentration was 723.9 \(\mu\text{mol/L}\) (mean 1,617.1 \(\pm\) 1987.6, range 250.2-6,769.2). For phenylacetylglutamine, the median steady-state plasma concentration was 440.3 \(\mu\text{mol/L}\) (mean 677.6 \(\pm\) 457.1, range 153.4-1,520.9). A total of six patients displayed an unidentified phenylbutyrate metabolite in the plasma samples, which was quantified from the phenylbutyrate standard curve. For the unknown metabolite, the median steady-state plasma concentration was 187.1 \(\mu\text{mol/L}\) (mean 192.8 \(\pm\) 71.6, range 84.6-292.9). The median steady-state plasma concentration of patients who showed an increase in acetylation following phenylbutyrate was 237 \(\mu\text{mol/L}\) (mean 317.1 \(\pm\) 100.3). The median steady-state plasma concentration of patients who did not increase acetylation during phenylbutyrate was 343.4 \(\mu\text{mol/L}\) (mean 976.4 \(\pm\) 457.5). Phenylbutyrate and metabolite exposure was not correlated with clinical response, changes in histone acetylation, induced F cells, or reversal of methylation.

**Discussion**

The DNMT inhibitors aza-CR and DAC comprise two of the most active agents for the treatment of MDS. Despite data demonstrating an \(~50\%\) hematologic response rate \((23, 41, 42)\), these agents were developed empirically without careful dose finding, and the molecular mechanism underlying their clinical activity remains unclear. Issa et al. \((22)\) studied DAC administered from 5 to 20 mg/m\(^2\)/d i.v. for 10 to 20 doses in patients with a variety of hematologic malignancies. The data suggested a potentially improved response rate \((65\%)\) in the patients treated at 15 mg/m\(^2\)/d for 10 days. Methylation of p15 and CDH-1 promoters was determined using MSP. The four representative patients were all treated in dose cohort 3 and cycle 2 in the first dose of drug for nine patients with complete set of methylation and pharmacokinetic data. Correlation approached statistical significance \((P = 0.0658)\). Dotted line, median concentration. Solid line, mean concentration.

**Figure 2.** Changes in methylation of p15 and CDH-1 promoters during cycle 1 of treatment. A, bone marrow mononuclear cells were procured before therapy \((\dagger)\), following the last injection of aza-CR \((\dagger)\), following discontinuation of phenylbutyrate \((\dagger)\), and before cycle 2 \((\dagger)\). In one patient, a fifth time point was procured after four cycles of treatment. Methylation of p15 and CDH-1 promoters was determined using MSP. The four representative patients were all treated in dose cohort 3 \((50 \text{ mg/m}^2/\text{d aza-CR for 10 days})\). NI, normal peripheral blood lymphocytes. IVD, positive methylation control. B, reversal of promoter methylation for either p15 and or CDH-1 after one cycle of therapy is plotted against the aza-CR AUC for the first dose of drug for nine patients with complete set of methylation and pharmacokinetic data. Correlation approached statistical significance \((P = 0.0658)\). Dotted line, median concentration. Solid line, mean concentration.
densely hypermethylated genes once some demethylation is initiated by drugs, such as aza-CR, as shown in vitro (14, 15).

The pharmacokinetic/pharmacodynamic studies suggest that reversal of promoter methylation may correlate with AUC for aza-CR. Nonresponse may be due to failure of adequate exposure of the target to drug, raising hopes that adjusted dosing schedules might improve clinical outcomes further. The relationship between aza-CR AUC and methylation reversal could be due to genetic polymorphisms affecting drug metabolism; one candidate would be differences in cytidine deaminase, the primary catabolic enzyme for aza-CR. Polymorphisms and differences in expression of cytidine deaminase and other catabolizing enzymes have been associated with in vivo resistance to cytarabine (45–48). These data suggest that more easily administered formulations of aza-CR could be developed, with potential for an adaptive control mechanism to increase the percentage of patients with promoter methylation reversal.

The current study design does not allow assessment of the specific biochemical contribution of the HDAC inhibitor. Ultimately, a randomized trial of a DNMT inhibitor with and without an HDAC inhibitor will be required. Increases in global histone acetylation were observed in the majority of patients studied. The induction of histone acetylation in response to aza-CR before the HDAC inhibitor was unexpected. The mechanism by which aza-CR led to histone acetylation is currently unknown; it is not clear whether this is a DNMT-dependent phenomenon and whether similar induction of histone acetylation occurs in response to in vivo administration of DAC. Increases in acetylation could be specifically attributed to phenylbutyrate in only 12 of 23 patients. This may be due to maximal induction of acetylation in response to aza-CR alone. The median steady-state plasma concentration of phenylbutyrate was consistent with previous phase I studies and would be predicted to be adequate for the induction of acetylation (25, 26, 37–39). However, the large percentage of patients evaluable for acetylation who developed increments in this endpoint (17 of 23) precludes assessment of the contribution of HDAC inhibition to clinical response.

From a clinical perspective, larger trials will be required to assess the efficacy of combining aza-CR with HDAC inhibitors in the treatment of MDS and AML. The current trial shows the clinical feasibility of combining a DNMT and HDAC inhibitor. Our current study also suggests that aza-CR in combination with phenylbutyrate yields a potential enrichment in major responses in patients who received aza-CR at 50 mg/m²/d × 10 days followed by phenylbutyrate. In the definitive Cancer and Leukemia Group B study of aza-CR, the CR plus PR rate was 23% (23). In dose cohort 3, four of nine evaluable patients achieved CR or PR. These numbers are small and require confirmation in a phase II setting; however, they suggest the possibility that prolonged exposure to lower doses of a DNMT inhibitor and/or the addition of an HDAC inhibitor may increase the major response rate.
In summary, we have developed a well-tolerated combination schedule of aza-CR and HDAC inhibitor, which induces recommended methylation reversal, global histone acetylation, F-cell production, and clinical responses. This first study provides a recommended phase II dose schedule for aza-CR (50 mg/m²/d × 10 days s.c.) in studies involving comparisons without an HDAC inhibitor to fully evaluate the efficacy of combining such drugs. We have also shown the first solid evidence that demethylation of hypermethylated genes in tumor clones is a target effect of aza-CR in patients who respond to therapy with this drug. This provides a possible assay to make early judgments about efficacy in individual patients and may be evidence that demethylation may underlie the responses themselves.

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