Gene Expression Preferentially Regulated by Tamoxifen in Breast Cancer Cells and Correlations with Clinical Outcome

Jonna Frasor,1 Edmund C. Chang,1 Barry Komm,3 Chin-Yo Lin,4 Vinsensius B. Vega,4 Edison T. Liu,4 Lance D. Miller,1 Johanna Smeds,5 Jonas Bergh,2 and Benita S. Katzenellenbogen1

1Department of Molecular and Integrative Physiology and 2Cell and Developmental Biology, University of Illinois and College of Medicine, Urbana, Illinois; 3Women’s Health Research Institute, Wyeth Research, Collegeville, Pennsylvania; 4Genome Institute of Singapore, Singapore, Singapore; and 5Department of Oncology and Pathology, Radiumhemmet, Karolinska Institute and Hospital, Stockholm, Sweden

Abstract
The beneficial effect of the selective estrogen receptor (ER) modulator tamoxifen in the treatment and prevention of breast cancer is assumed to be through its ability to antagonize the stimulatory actions of estrogen, although tamoxifen can also have some estrogen-like agonist effects. Here, we report that, in addition to these mixed agonist/antagonist actions, tamoxifen can also selectively regulate a unique set of >60 genes, which are minimally regulated by estradiol (E2) or raloxifene in ERα-positive MCF-7 human breast cancer cells. This gene regulation by tamoxifen is mediated by ERα and reversed by E2 or ICI 182,780. Introduction of ERβ into MCF-7 cells reverses tamoxifen action on ~75% of these genes. To examine whether these genes might serve as markers of tamoxifen sensitivity and/or the development of resistance, their expression level was examined in breast cancers of women who had received adjuvant therapy with tamoxifen. High expression of two of the tamoxifen-stimulated genes, YWHAZ/14-3-3z and LOC441453, was found to correlate significantly with disease recurrence following tamoxifen treatment in women with ER-positive cancers and hence seem to be markers of a poor prognosis. Our data indicate a new dimension in tamoxifen action, involving gene expression regulation that is tamoxifen preferential, and identify genes that might serve as markers of tumor responsiveness or resistance to tamoxifen therapy. This may have a potential effect on the choice of tamoxifen versus aromatase inhibitors as adjuvant endocrine therapy. (Cancer Res 2006; 66(14): 7334-40)

Introduction
Tamoxifen, a selective estrogen receptor modulator (SERM), is effective in the prevention and treatment of breast cancer (1–3) and has been the most widely used endocrine therapy for breast cancer. Originally, it was termed an antiestrogen because it was assumed that the positive effects of tamoxifen arise from its ability to antagonize the stimulatory actions of estrogen acting through its receptors. However, tamoxifen is now more accurately called a SERM, reflecting its ability to display estrogen-like agonist effects, as well as antiestrogen-like antagonist effects, in a tissue or cell type-selective manner. These mixed agonist/antagonist actions of tamoxifen are associated not only with its effectiveness in the prevention and treatment of breast cancer and positive effects on the maintenance of bone mineral density but also with some deleterious side effects, such as an increased risk of endometrial cancer and thromboembolic events (4).

One of the primary functions of the ERs (ERα and ERβ) is their ability to regulate gene transcription in a ligand-dependent manner. Recently, we and several other groups have used gene expression profiling in an attempt to identify genes regulated through the ER in breast cancer cells (reviewed in refs. 5–7 and references therein). We have found that estrogen both up-regulates and down-regulates a wide range of genes, many of which would be consistent with stimulation by estrogen of breast cancer cell proliferation and survival (6). In studies using other ER ligands, such as SERMs, we and others have found that tamoxifen acts primarily to antagonize actions of estrogen but can also have an estrogen-like agonist effect on several genes known to be regulated by estradiol (E2) in breast cancer cells (8, 9). The ability of tamoxifen to have these mixed agonist/antagonist effects is consistent with the mounting evidence that different ligand-receptor complexes are capable of regulating gene expression through differential coregulator recruitment in a gene-specific manner (10, 11). In addition, because ERβ is present in ~70% of ER-positive breast cancers (12), it is important to understand the activity of tamoxifen in the presence of ERα plus ERβ.

Using gene expression analysis, we have now identified a third aspect of tamoxifen action: its ability to uniquely regulate a set of genes, which are minimally affected by E2 or other SERMs. This gene regulation by tamoxifen seems to be mediated by ERα in MCF-7 cells, and the introduction of ERβ into these ERα-containing cells represses tamoxifen action on a large proportion of these genes. The expression of these tamoxifen preferentially regulated genes was examined in a cohort of women with breast cancer treated with tamoxifen who showed either prolonged benefit or a higher frequency of breast cancer relapse. In fact, the elevated expression of two of these tamoxifen-stimulated genes in breast cancers was found to correlate with disease recurrence following tamoxifen treatment. This preferential regulation of genes by tamoxifen represents a novel dimension in SERM action in breast cancer with potential therapeutic and diagnostic implications.

Materials and Methods
Cell culture, RNA extraction, and real-time PCR. Methods for the culture and treatment of MCF-7 cells, extraction of RNA, and real-time PCR have been described previously (6, 8).

Adenoviral delivery of ERβ into MCF-7 cells. Recombinant adenoviruses were constructed and prepared as described (13). MCF-7 cells were
infected with either recombinant adenovirus containing full-length (530 amino acid) ERβ or control adenovirus containing no insert for 48 hours before ligand treatment. A multiplicity of infection of 10 was used, which results in an ~1:1 ERα to ERβ protein ratio as determined from Western blots of whole-cell extracts compared with standard curves generated with known amounts of recombinant ERα and ERβ protein from PanVera (Madison, WI).

GeneChip microarray analysis. Affymetrix human GeneChip U133A microarrays (Affymetrix, Santa Clara, CA) were used for expression profiling following infection of MCF-7 cells with adenovirus (empty virus) or AdERβ and treatment with 10⁻⁸ mol/L E₂ or 10⁻⁸ mol/L trans-hydroxytamoxifen (Tam) for 4 or 24 hours. cRNA preparation and hybridization have been described previously (6). The array data were preprocessed using the GeneChip Robust Multichip Average package in R/Bioconductor (14). A false discovery rate of ≤0.05, as determined by Significance Analysis of Microarray software (15), was used to identify genes significantly regulated by ligand treatment. Expression data were then loaded into GeneSpring software and normalized to the control (adenovirus-infected, vehicle-treated) samples. Genes preferentially regulated by Tam treatment were identified using a fold change cutoff of ≥2.0 for Tam and ≤1.3 for E₂ treatment, and for genes preferentially down-regulated by Tam a change in gene expression from the control level of 1.0 to a level of ≤0.5-fold for Tam and ≥0.77 for E₂, based on our prior findings (8). The modulation of Tam action by ERβ was determined by a 1.5-fold repression or enhancement of Tam action when compared with Tam action in the presence of no ER (empty adenovirus) because this gave robust and significant differences that were reproducibly validated by quantitative reverse transcription-PCR. Microarray data are based on triplicate arrays and three independent samples for each treatment. The entire data set will be available through National Center for Biotechnology Information (NCBI) Gene Expression Omnibus (GEO)7 and accessible through GEO Series accession number GSE4025.

Whole-cell extract preparation and Western blotting. Whole-cell extracts were prepared using radiomununoprecipitation assay buffer (1× PBS, 1% NP40, 0.5% sodium deoxycholate, 0.1% SDS, 10⁻⁶ mol/L sodium orthovanadate, 10 μg/mL phenylmethylsulfonyl fluoride, 30 μL/mL aprotinin). Protein (25-100 μg) was separated on 10% SDS-PAGE gels and transferred to nitrocellulose membranes. Western blotting was done using antibodies against IEX-1 (Santa Cruz Biotechnology, Santa Cruz, CA), β-actin (Sigma, St. Louis, MO), ERα (HC20, Santa Cruz Biotechnology), or ERβ (16). Quantitation of ERα and ERβ protein levels in MCF-7 cells before and after adenoaviral ERβ gene delivery was determined from standard curves we prepared using different known amounts of recombinant ERα or ERβ protein from PanVera and ImageQuant TL quantitation software (Amer sham Biosciences, Piscataway, NJ).

Tumor specimens and survival analysis. The original patient material consisted of freshly frozen primary breast tumors from a population-based cohort of 315 women representing 65% of all breast cancers resected in Uppsva County from January 1, 1987 to December 31, 1989 (17, 18). After exclusions based on RNA integrity and array quality control, expression profiles of 254 tumors were deemed suitable for further analysis. Clinicopathologic characteristics were derived from the patient records and from routine clinical measurements at the time of diagnosis as described elsewhere (17). Microarray analysis of patient samples was carried out on Affymetrix U133 A and B arrays, and processing of the expression data was described previously (19). We analyzed the 69 ER-positive tumors from patients who subsequently underwent endocrine therapy with tamoxifen only to assess the role of tamoxifen-regulated genes in breast tumor biology. All ER-negative tumors (n=37) and associated expression and survival data were included as controls.

Association of tamoxifen-regulated genes with patient survival and disease recurrence was determined in several ways. First, patients were sorted by disease recurrence within 10 years following tamoxifen treatment and then the expression profiles of the tamoxifen-regulated genes were assessed for significant differential expression between the recurring and nonrecurring patients by the Wilcoxon rank sum test. Second, genes that showed significant differences (P<0.05) between the two outcome groups were used to group patients using average linkage hierarchical clustering. Kaplan-Meier estimates were used to compute survival curves, and the significance of the hazard ratios between the major patient clusters was determined by the P of the likelihood ratio tests as was described previously (19). We also did Monte Carlo simulations similar to what was done previously (19) to assess the statistical validity of the two tamoxifen up-regulated genes that were found to correlate with poor outcome on tamoxifen.

Results

Tamoxifen preferential regulation of gene expression. In a previous gene expression profiling study, we compared the agonistic and antagonistic activities of several SERMs on estrogen-regulated genes (8). In the current study, we have focused on the identification and characterization of genes that are preferentially regulated by Tam but not by E₂ or other SERMs, such as ICI 182,780 (ICI) or raloxifene. In MCF-7 cells treated with 10 nmol/L E₂ or Tam for 4 or 24 hours, we have identified a total of 50 genes up-regulated by Tam and 14 genes down-regulated by Tam, with these genes being affected little if at all by treatment with E₂ (Fig. 1; Supplementary Table S1). GeneChip array expression data for this set of genes after 24 hours of treatment with E₂ or Tam are shown in Fig. 1.

Six of these genes found to be robustly up-regulated by Tam based on microarray analyses were chosen for examination in more detail. The Tam-selective stimulation of expression of these genes (FARP1, IEX-1, KRT13, PTPRG, Rab30, and SOCS1) was verified using real-time PCR. As shown in Fig. 2A, when cells were treated with E₂ or the SERMs ICI, raloxifene, or Tam, only Tam elicited strong stimulation. Next, to examine whether Tam regulation of these genes is mediated by the SERMs, we treated with 10⁻⁸ mol/L Tam in combination with 10⁻⁶ mol/L (100-fold excess) E₂ or the antiestrogen ICI (also known as fulvestrant). For all six genes, both E₂ and ICI blocked the stimulatory effect of Tam, indicating that Tam action on these genes is mediated by the ER (Fig. 2B).

To determine whether the effects of Tam represent primary responses, cells were treated with the protein synthesis inhibitor cycloheximide alone or in combination with Tam for 4 hours. All six of the genes examined seemed to be primary responses to Tam because cycloheximide had either no effect or enhanced the effect of Tam on their up-regulation (Fig. 3).

As shown in Fig. 3, IEX-1 up-regulation is superinduced by cycloheximide treatment, which is consistent with the findings that IEX-1 is an immediate early response gene to many stimuli (20). In addition, because the up-regulation of IEX-1 in breast cancer cells is associated with an inhibition of breast cancer cell proliferation (21), we decided to further characterize the regulation of this gene. Dose-response and time course experiments were carried out, and the findings are shown in Fig. 4. An increase in IEX-1 mRNA could be detected within 30 minutes, and an increase in IEX-1 protein was detected over 2 to 8 hours of Tam treatment (Fig. 4A and B). Dose-response studies indicate that the maximum response of IEX-1 was observed at 10 nmol/L to 1 μmol/L Tam (Fig. 4C), whereas raloxifene had no effect across the broad range of doses. We also observed that the RNA synthesis inhibitor actinomycin D caused a rapid down-regulation of IEX-1 mRNA levels (Fig. 4D). The time course for this down-regulation was identical in the presence or absence of Tam.

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indicating that Tam is not acting to regulate IEX-1 mRNA stability and further supporting the suggestion that IEX-1 is a primary transcriptional response to Tam.

**Effect of ERβ on tamoxifen regulation of gene expression.**

Several lines of evidence indicate that Tam can have different effects through the two ERs, ERα and ERβ (22–24). To examine whether ERβ can influence tamoxifen activity on these preferentially regulated genes in MCF-7 cells, which only express ERα, adenoviral delivery of ERβ was used and additional microarrays were carried out. In cells expressing both ERα and ERβ, we find that the major effect of ERβ was to repress the response of MCF-7 cells to Tam. Specifically, the regulation of 38 of the 50 genes up-regulated by Tam and 7 of the 14 genes down-regulated by Tam was reduced >1.5-fold in MCF-7 cells coexpressing ERβ (Fig. 5A). This inhibitory effect of ERβ on Tam activity was examined and verified for several representative genes (FARP1, IEX-1, Rab30, and SOCS1) by real-time PCR (Fig. 5B). Of note, the

![Figure 1. Identification of genes preferentially regulated by tamoxifen. Microarrays were carried out using RNA from MCF-7 cells treated with 10 nmol/L E2 or Tam for 24 hours. Genes preferentially regulated by Tam were identified (as described in Materials and Methods) and clustered based on their expression profile in response to the two ligands.](image1)

![Figure 2. Verification by real-time PCR of the preferential up-regulation of genes by tamoxifen (A) and that this gene regulation by tamoxifen is mediated by the ER (B). A, gene regulation was verified using RNA obtained from MCF-7 cells treated for 24 hours with 0.1% ethanol control vehicle (C), 10 −6 mol/L E2, 10 −6 mol/L ICI, 10 −6 mol/L raloxifene (Ral), or 10 −6 mol/L Tam. The gene abbreviations used, full gene names, and Genbank accession numbers are as follows: FARP1 (FERM, RhoGEF ARHGEF and pleckstrin domain protein 1, AI701049), IEX-1 (immediate early response 3, S81914), KRT13 (keratin 13, X14640), PTPRG (protein tyrosine phosphatase, receptor type G, U46116), Rab30 (Ras oncogene family member, U57092), and SOCS1 (suppressor of cytokine signaling 1, AB00734). The primers used for real-time PCR are provided in Supplementary Table S2. Columns, mean of three determinations; bars, SE. B, real-time PCR was carried out on RNA obtained from MCF-7 cells treated for 24 hours with 0.1% ethanol control vehicle, 10 −6 mol/L Tam, 10 −6 mol/L E2, or 10 −6 mol/L ICI alone or with 10 −6 mol/L Tam in combination with control vehicle or 10 −6 mol/L E2 or ICI. Columns, mean of three determinations; bars, SE.)
significant association (64 genes were ranked (Wilcoxon rank sum) based on their association of the 64 genes with response to Tam as measured by tamoxifen following surgery. As a group, there was no significant expression profiles of the 64 Tam-regulated genes were analyzed had been treated with tamoxifen and followed over time. The responses to Tam.

stimulations are not blocked by cycloheximide and hence seem to be primary synthesis inhibitor, for 4 hours. Real-time PCR was then carried out. These gene stimulations are not blocked by cycloheximide and hence seem to be primary responses to Tam.

presence of ERβ in vehicle-treated cells (+ERβ, vehicle) elicited changes in expression of some genes similar to that of Tam but often more weakly, consistent with the ligand-independent activity of this receptor form (25). This was seen both by microarray analysis and quantitative PCR (Fig. 5A and B). For KRT13 and PTORG, the basal level of mRNA expression was enhanced by ERβ in the absence of ligand treatment, whereas the stimulation by Tam was not affected (Fig. 5B).

Clinical correlations: expression of tamoxifen preferentially regulated genes in breast cancer patients and association with probability of disease recurrence on tamoxifen. To evaluate the possible prognostic value of these tamoxifen-regulated genes, we examined the expression of these genes in a cohort of women with ER-positive breast tumors (n = 69) who had been treated with tamoxifen and followed over time. The expression profiles of the 64 Tam-regulated genes were analyzed in tumor samples from the 69 ER-positive patients treated with tamoxifen following surgery. As a group, there was no significant association of the 64 genes with response to Tam as measured by disease recurrence (i.e., disease-free survival). However, when the 64 genes were ranked (Wilcoxon rank sum) based on their individual association with disease recurrence, 2 genes showed significant association (P < 0.05). These 2 genes were YWHAZ/14-3-3ε (P = 0.0199) and LOC441453 (P = 0.0327). When the patient samples were clustered using this two-gene cassette, patients clustered into good and poor outcome groups (P = 0.000621), with the higher expression of the two genes associated with a higher probability of recurrence (Fig. 6). Three other tamoxifen selectively regulated genes (RAP1A0, MYLIP, and GPR56) showed some borderline significance (P > 0.05 < 0.07) with outcome to tamoxifen. By Monte Carlo simulations, we estimated the statistical likelihood of finding two random genes whose hierarchical clustering could produce a Kaplan-Meier curve likelihood ratio P ≤ 0.000621. The Monte Carlo P obtained was 0.015 (based on ~100,000 iterations of Monte Carlo simulation), indicating that it is unlikely that the two tamoxifen-responsive, survival-associated genes (YWHAZ and LOC441453) were selected by chance.

Consistent with the hypothesis of ER regulation, YWHAZ and LOC441453 were associated with disease recurrence only in ER-positive patients on tamoxifen; no correlation was observed in the 37 women with ER-negative breast cancers. For the ER-negative patients, Ps of the “high” and “low” expression groups for disease-specific survival and disease-free survival were at P = 0.213 and 0.328, respectively. A list of all Tam-regulated genes and the Wilcoxon rank sum Ps for their association with disease recurrence are presented in Supplementary Table S3.

Discussion

Tamoxifen has been and is still for many patients a cornerstone in the management of hormone-responsive breast cancer. In the adjuvant setting, the recurrence rate is reduced by ~50% (26) and the mechanism of its effectiveness is primarily thought to be due to its ability to antagonize estrogen action through interaction with the ER (1, 4, 8, 11). However, tamoxifen often acts as a partial agonist/antagonist or a full agonist, depending on the gene as well as the cellular background (8–12). Eventually, treatment with tamoxifen can lead to resistance where breast cancer cells no longer see tamoxifen as an antagonist and it may actually stimulate breast cancer cell proliferation (1, 27, 28). Our studies show that, in addition to the antagonistic, partial agonist/antagonistic, or even full agonist activities of Tam on genes regulated by E2, Tam is also capable of regulating the expression of some genes that are not regulated by either E2 or other SERMs. This regulation, like that of the other categories of tamoxifen-modulated gene expression, is mediated via the ER. Although these genes must be viewed within the context of all the genes, on which Tam exerts agonist or antagonist actions, we investigated whether some of these genes preferentially regulated by Tam might serve as markers of Tam responsiveness or the development of resistance in breast cancer.

Figure 3. Cycloheximide does not prevent the up-regulation of gene expression by Tam. MCF-7 cells were treated with 0.1% ethanol control vehicle or 10−8 mol/L Tam with or without 10 μg/mL cycloheximide (CHX), a protein synthesis inhibitor, for 4 hours. Real-time PCR was then carried out. These gene stimulations are not blocked by cycloheximide and hence seem to be primary responses to Tam.

Figure 4. Time- and dose-dependent up-regulation of IEX-1 mRNA and protein levels by Tam. MCF-7 cells were treated with 10−8 mol/L Tam for different times (A and B), with different doses of Tam or raloxifene for 4 hours (C), or with 10−8 mol/L Tam for up to 120 minutes in the presence or absence of the RNA synthesis inhibitor actinomycin D (ActD, D). RNA extraction and PCR analysis (A and C–D) or Western blot (B) was then carried out for IEX-1. β-Actin was also examined by Western blot as a loading control.
Our data show that Tam action, in preferentially regulating this set of genes, is mediated by the ER because both E2 and ICI block Tam action. Several lines of evidence, including X-ray crystallography, ER mutagenesis, and peptide mapping, have all indicated that different ER ligands are capable of inducing different ER structures (11, 29–31). In addition, Tam is known to require different regions of the ER for enhanced transcriptional activity than does E2 (32) and to differentially recruit coregulatory proteins to estrogen-responsive genes (33). That these genes are stimulated very preferentially by Tam suggests that they might be emblematic of pharmacologic differences between Tam and the other SERMs, raloxifene and ICI.

Figure 5. ERβ modulates tamoxifen activity on genes preferentially regulated by tamoxifen. Microarrays were carried out using RNA from MCF-7 cells infected with a control empty adenovirus (no ERβ) or with adenovirus expressing ERβ (+ERβ) for 48 hours and then treated with 10 nmol/L Tam for 24 hours. A, genes preferentially regulated by Tam were clustered based on their expression profile in response to tamoxifen in ERβ-negative or ERβ-positive (+ERβ) MCF-7 cells. B, verification of ERβ modulation of tamoxifen activity on gene expression. PCR analysis was carried out for six genes preferentially regulated by Tam. The presence of ERβ inhibited responsiveness to Tam for some genes (FARP1, IEX-1, Rab30, and SOCS1) but not for other genes (KRT13 and PTPRG). For KRT13 and PTPRG, ERβ altered basal gene expression in the absence of Tam.
The fact that ERβ could reverse the regulation of ~75% of these genes by Tam, including reversal of the Tam stimulation of YWHAZ/14-3-3z and LOC441453, is of note because human breast tumors often express ERβ along with ERα, and these women are commonly treated with Tam (12). However, the mechanism by which ERβ modulates Tam gene activity is not clear. In response to E2 or other ER ligands, ERα and ERβ can have both common and distinctly different gene regulatory activity (24, 34, 35). In addition, heterodimers of ERα/ERβ may have distinct activity from either ERα or ERβ homodimers (24, 36). For these Tam-regulated genes, it is possible that the activation function-1 (AF-1) domain of ERα, which is essential for Tam agonist activity, may be repressed by the strong AF-1 activity of ERβ (36). Other evidence suggests that different surfaces of ERα and ERβ may be exposed on Tam binding, which may lead to the recruitment of different coactivator complexes to each receptor in the presence of Tam compared with E2 (37). The relationship between ERβ and response to endocrine therapy still remains unclear and under active investigation (38).

The genes preferentially regulated by Tam encode proteins that have diverse roles in multiple functional pathways and gene networks. For example, PKIA, an inhibitor of cyclic AMP–dependent protein kinase A activity, PTPRG, a receptor-type protein tyrosine phosphatase, and SOCS1, an inhibitor of Janus-activated kinase (JAK)/signal transducers and activators of transcription (STAT) signaling, all could potentially alter different cellular signaling pathways and, thus, responsiveness of breast cancer cells to other hormones, growth factors, or cytokines. In addition, IEX-1 has been shown to have growth-inhibitory effects on MCF-7 cells (20, 21), suggestive of a beneficial effect of Tam. In addition, several other Tam preferentially regulated genes might have potential tumor suppressor roles. SOCS1, a negative regulator of cytokine action through the JAK/STAT pathway, has been proposed to suppress growth of hepatocellular carcinomas (39). Similarly, PTPRG was identified as a potential tumor suppressor because it is located in a region rearranged in some cancers and deleted in some breast cancers (40).

The two genes whose expression we found to correlate with decreased disease-free survival, YWHAZ and LOC441453, have yet to be implicated in breast cancer progression. Both genes were preferentially up-regulated by tamoxifen in MCF-7 cells as seen in Figs. 1 and 5 and Supplementary Table S1. YWHAZ, also known as 14-3-3c, belongs to the highly conserved 14-3-3 family of proteins, which regulate the cell cycle and prevent apoptosis and mediate signal transduction by binding to phosphoserine-containing proteins. YWHAZ is a scaffolding protein that is phosphorylated on specific serine residues by protein kinase C and AKT. YWHAZ is thought to play a central role in insulin receptor and epidermal growth factor receptor signaling and in cell cycle regulation (41–44). The YWHAZ protein binds and stabilizes key signaling proteins, such as IRS1, protein kinase B/AKT1, β-catenin, and BIN1, and activates c-raf, BAD, and cbl (41, 42, 44). High expression of a protein central in cell signaling, such as YWHAZ, under the influence of tamoxifen might contribute to the promotion of tumor recurrence. Little is known about LOC441453, which is also known as similar to olfactory receptor, family 7, subfamily A, member 17. The protein contains a conserved seven transmembrane receptor motif (NCBI RefSeq, gene ID 441453), but its role in cells is virtually unexplored currently. Further study of the biological roles of both proteins would be of interest.

It is known that some ER-positive breast cancers, even if small and from lymph node-negative patients, can do poorly on tamoxifen. High expression of YWHAZ/14-3-3z and LOC441453 might indicate a more aggressive cancer. YWHAZ/14-3-3z is a critical player in numerous cell signaling pathways, so elevated expression is likely to be associated with a more aggressive phenotype. In fact, Oncomine database analysis8 of two published clinical studies (45, 46) confirms that high YWHAZ/14-3-3z expression is associated with poor outcome on endocrine therapy; there is no information on LOC441453 available. Therefore, on tamoxifen treatment, which would further increase expression of these two genes, one might predict a poor outcome on tamoxifen. It is possible that women with such breast cancers might do better with aromatase inhibitor therapy to avoid tamoxifen stimulation of such genes. Further work to study the predictive value of these genes would require a separate neoadjuvant therapy study, where tumor biopsies would be obtained before and shortly after the initiation of tamoxifen treatment to measure the effect of tamoxifen on expression of these genes.

In conclusion, we have identified several genes preferentially regulated by Tam in ER-positive breast cancer cells, suggesting a novel dimension in the action of this SERM. The correlation of high expression of some of these genes with disease recurrence on tamoxifen indicates that these genes are likely to be useful in predicting the effectiveness of tamoxifen in treating ER-positive breast cancer and assessing the development of tamoxifen resistance. Such approaches are already proving useful in the prediction of risk of recurrence following tamoxifen treatment (47, 48) and in the molecular classification of tamoxifen-resistant breast cancers (49). However, it is clear that additional genes, such as those observed in this report to be preferentially regulated by tamoxifen and to correlate with a shorter disease-free survival and time to recurrence, might serve as valuable prognostic markers to improve the accuracy of tamoxifen resistance and recurrence prediction that might potentially affect the type of endocrine therapy selected. Aromatase inhibitors are increasingly being used in the adjuvant setting with good results (50). As with tamoxifen, however, there are also side effects of concern with

Figure 6. Correlation of tamoxifen-regulated genes with patient outcome. A, hierarchical clustering of 69 ER-positive tumor samples from patients treated with tamoxifen into two groups of high (black dendrogram; cluster 1) and low (red dendrogram; cluster 2) YWHAZ and LOC441453 expression clusters. B, Kaplan-Meier plots of disease recurrence for clusters 1 (black) and 2 (red) patients with the associated P from likelihood ratio analysis.

aromatase inhibitors (i.e., a higher incidence of bone fractures and joint pain; ref. 50). These issues and the findings presented in this article should stimulate researchers to use endocrine therapies in a more tailored way based on a more detailed understanding of the mechanisms of action for tamoxifen, which therefore may still be the drug of choice for many breast cancer patients. Potentially, they might also assist in the development of new SERMs with an optimized complement of positive effects in breast cancer.

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