MicroRNA Expression Profiling in Prostate Cancer

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Abstract

MicroRNAs (miRNA) are small, endogenously expressed non-coding RNAs that negatively regulate expression of protein-coding genes at the translational level. Accumulating evidence, such as aberrant expression of miRNAs, suggests that they are involved in the development of cancer. They have been identified in various tumor types, showing that different sets of miRNAs are usually deregulated in different cancers. To identify the miRNA signature specific for prostate cancer, microRNA expression profiling of 6 prostate cancer cell lines, 9 prostate cancer xenograft samples, 4 benign prostatic hyperplasia (BPH), and 9 prostate carcinoma samples was carried out by using an oligonucleotide array hybridization method. Differential expression of 51 individual miRNAs between benign tumors and carcinoma tumors was detected, 37 of them showing down-regulation and 14 up-regulation in carcinoma samples, thus identifying those miRNAs that could be significant in prostate cancer development and/or growth. There was a significant trend (P = 0.029) between the expression of miRNAs and miRNA locus copy number determined by array comparative genomic hybridization, indicating that genetic aberrations may target miRNAs. Hierarchical clustering of the tumor samples by their microRNA expression accurately separated the carcinomas from the BPH samples and also further classified the carcinoma tumors according to their androgen dependence (hormone naive versus hormone refractory), indicating the potential of miRNAs as a novel diagnostic and prognostic tool for prostate cancer. [Cancer Res 2007;67(13):6130–5]

Introduction

MicroRNAs (miRNA) are a class of small, noncoding RNAs that are endogenously expressed in animal and plant cells. They regulate the expression of protein-coding genes at the translational level, either by triggering degradation or preventing translation of the target mRNAs (1). Primary transcripts of miRNAs (pri-miRNA) are generated by RNA polymerase II (2), after which they are sequentially processed by RNase III class enzymes, Drosha and Dicer, to first produce ~70-nt-long intermediate hairpin structures (pre-miRNAs) and finally the 17-24-nt-long, mature miRNAs (3). The single-stranded, mature miRNAs bind to their target sequences that most often are located in the 3′-prime untranslated regions of the mRNA, usually showing only partial bp complementarity with the miRNA sequence. According to the current view, imperfect complementarity leads to repression of translation of the target mRNA and is the main mechanism of miRNA regulation in animals, whereas perfect complementarity induces degradation of the target mRNA and is mainly detected in plants (3).

At their discovery, miRNAs were shown to control fundamental cellular processes, such as differentiation of cells and timing of development of the organism (4, 5), suggesting that aberrations of miRNAs could be involved in various human diseases, including cancer. In the year 2002, the first piece of evidence connecting miRNAs and cancer was obtained when Calin et al. (6) showed that miR-15 and miR-16 are the target genes of the 13q14 deletion that is common in chronic lymphatic leukemia. After this initial finding, more than half of the known miRNAs have been reported to be located in cancer-associated genomic regions and to show copy number alterations in cancer (7, 8), and deregulation of several miRNAs, such as miR-143, miR-145, miR-21, BIC/miR-155, and the let-7 family, has been detected in various cancers (9–16). Functional studies of individual miRNAs have shown that they can act as oncogenes (the so-called onco-miRs) or tumor suppressor genes (11, 17–20). Although targets of most miRNAs have not been identified, some have been shown to negatively regulate well-known oncogenes: miR-15 and miR-16 have been reported to repress the antiapoptotic factor BCL2 (21) and the let-7 miRNA family to target the RAS oncogene (22).

Recently, microRNA expression profiling studies have been done to identify cancer-specific miRNA signatures (10, 12, 16, 23–31). Although there are some miRNAs, such as miR-143 and miR-145, that have been reported to be aberrantly expressed in several different cancer types (9, 11, 12), the miRNA signatures of cancers of different cellular origin seem to be unique. In this study, expression profiling of 319 miRNAs in 6 prostate cancer cell lines, 9 prostate cancer xenograft samples, and 13 clinical prostate tissue samples was carried out, revealing the miRNA signature of prostate cancer. Furthermore, hierarchical clustering of the clinical tumor samples by their microRNA expression was done, showing the high accuracy of miRNA expression profiling in classifying prostate tumors.

Materials and Methods

Cell lines, xenografts, and tumor samples. The prostate cancer cell lines PC-3, DU-145, LNCaP, and 22Rv1 were obtained from American Type Culture Collection, whereas LAPC4 and VCaP were kindly provided by Dr. Charles Sawyers (University of California at Los Angeles, Los Angeles, CA) and Dr. Jack Schalken (Radboud University Nijmegen Medical Center, Nijmegen, the Netherlands), respectively. All cell lines were cultured under the recommended conditions. Freshly frozen samples of prostate cancer xenografts (LuCaPs 41, 49, 69, 70, 73, 86, 2, 92, 1, 93, and 115) were made available for the analyses by one of the coauthors (R.L. Vessella). The characteristics of the xenografts have been reported previously (32, 33).
Freshly frozen clinical tissue samples from 4 benign prostatic hyperplasias (BPH), 5 untreated prostate carcinomas, and 4 hormone-refractory prostate cancers were obtained from Tampere University Hospital (Tampere, Finland). The carcinoma samples were histologically examined for the presence of tumor tissue (>60%) using H&E-stained sections. The BPH samples were obtained from prostatectomy or cystoprostatectomy specimens from patients treated for BPH or bladder cancer. The specimens were histologically verified not to contain any prostate cancer cells. The clinicopathologic characteristics of the tissue specimens are given in Supplementary Table S1.

miRNA array design and preparation. Antisense oligonucleotide probes for all 319 different human miRNA sequences found from the miRNA database of the Sanger Institute5 (34, 35) in October 2005 were designed and purchased from Invitrogen. The probes were 34-48 nt long, unmodified DNA oligonucleotides, in which the antisense sequence of the target miRNA was repeated twice. Antisense oligonucleotide probes for tRNA-Met and tRNA-Ala were used as positive controls. To confirm the specificity of the array hybridization signals, antisense oligonucleotide probes for miR-16, miR-21, and miR-28 containing one, two, or three mismatching bases (G to C) per target sequence were used. The sequences and lengths of probes included in the array are summarized in Supplementary Table S2. All the probes were spotted in duplicate at 50 μmol/L concentration onto nylon membranes (GeneScreen Plus, Perkin-Elmer) by using a 96-pin VJP 409 replicator tool (V&P Scientific, Inc.) followed by immobilization of the probes on the membranes as described previously (36).

RNA extraction and miRNA array hybridization. RNA extraction, labeling, and hybridization were done essentially as described previously (36). Briefly, total RNA from the cell lines, xenografts, and clinical tissue samples was extracted by using the Trizol reagent (Invitrogen). Low-molecular weight (LMW) RNA (<300 nt) from 2 μg total RNA was purified by using the Microcon YM-100 columns (Millipore), dephosphorylated with the calf intestine alkaline phosphatase (Fermentas International, Inc.), and 5’-end labeled with [γ-32P]ATP (3,000 Ci/mmol; Amersham Biosciences/GE Healthcare) using T4 polynucleotide kinase (Fermentas). Unincorporated nucleotides were removed from the labeling reaction with the Microspin G-25 column (Amersham Biosciences/GE Healthcare) followed by immobilization of the labeled RNA on nylon membranes (GeneScreen Plus, GE Healthcare) using T4 polynucleotide kinase (Fermentas). Unincorporated nucleotides were removed from the labeling reaction with the Microspin G-25 column (Amersham Biosciences/GE Healthcare). miRNA array membranes were prehybridized in MicroHyb hybridization solution (Invitrogen) for 1 h at +37°C, after which the labeled LMW RNA was added to the solution. Hybridization was carried out at +37°C overnight followed by washes. After exposing the membranes to the Storage Phosphor screen (Amersham) and immobilization at +80°C for 15 min, the membranes were cross-linked by UV (IBI Ultralinkr, Kodak). DNA oligonucleotide probes for miR-99a (5’-CACAAGATCGGATCTACGGGTT-3’), miR-141 (5’-CCATCTTTACCAGA-CGTTTA-3’), miR-198 (5’-CCATCTCTCCCTCTGGAC-3’), and miR-205 (5’-CAGACTCCGGTGGAATGAGGA-3’) were end labeled with [γ-32P]ATP using T4 polynucleotide kinase. Unincorporated nucleotides were removed, and after the prehybridization, the dot blot membranes were hybridized with the labeled probe at +37°C overnight followed by washes. After exposing the membranes to the Storage Phosphor screen, signals were detected by using the Typhoon Trio scanner. To normalize the miRNA hybridization signals, the membranes were stripped and rehybridized with a probe for the 5S rRNA (5’-CAGGCCCGACGCCGCTAGCTTCCGAGATCACGAGGAT-3’). The dot blot hybridization signals were normalized by calculating the miRNA/5S rRNA expression ratios for each miRNA.

Real-time quantitative PCR. Total RNA from the clinical tissue samples was extracted by using the Trizol reagent. The quantitative real-time PCR (qRT-PCR) expression analysis of miR-145 was done by using the miRNA-specific mirVana qRT-PCR Primer Set (Ambion Ltd.). First, 20 ng total RNA was reverse transcribed by using the mirVana RT Primer (Ambion) and the SuperScript III reverse transcriptase (Invitrogen) at +37°C for 30 min, after which the enzyme was inactivated at +95°C for 10 min. Subsequently, the reverse-transcribed miRNAs were used as templates in the qRT-PCR analysis using the LightCycler instrument (Roche Diagnostics) with the mirVana PCR primers (Ambion) and the LightCycler FastStart DNA Master SYBR Green I kit (Roche Diagnostics). After the activation of the polymerase enzyme at +95°C for 10 min, 45 cycles of +95°C for 1 s, +60°C for 10 s, and +72°C for 10 s were done. Both melting curve analysis and agarose gel run were used to confirm the specificity of the amplification reactions. For normalization, expression of the TATA box binding protein miRNA was measured from the each RNA sample as described previously (38).

Results

miRNA expression in prostate cancer cell lines and xenografts. An oligonucleotide array hybridization method was set up to study expression of 319 human miRNAs in prostate cancer. First, the method was set up and validated by hybridizing the arrays with radiolabeled LMW RNA from 6 prostate cancer cell lines and 9 prostate cancer xenografts. About 40% (128 of 319) of the miRNA probes gave detectable signal in at least one of the samples. To validate the array hybridization results, expression of four miRNAs (miR-99a, miR-141, miR-198, and miR-205) that showed expression differences between the cell lines was studied in five cell lines (PC-3, DU-145, LNCaP, 22Rv1, and LAPC4) using the dot blot hybridization. The dot blot hybridization results were in concordance with the array hybridization results (Supplementary Fig. S1).

After the validation of the array analysis, the samples were clustered by their miRNA expression profiles using the hierarchical clustering algorithm of the Cluster software (37). One major characteristic by which the cell line and xenograft samples differ from each other is their androgen receptor (AR) status: four of the samples studied (the cell lines PC-3 and DU-145 as well as the xenografts LuCaPs 49 and 93) are AR negative, whereas the others express AR at variable levels. In cluster analysis, the AR-positive cell lines (LNCaP, 22Rv1, LAPC4, and VCaP) were all clustered in the same branch together with one of the xenograft samples (LuCaP 92.1). On the other hand, the two AR-negative cell lines (PC-3 and DU-145) were clustered next to each other, distantly from the other cell lines. Similarly, the two AR-negative xenograft samples (LuCaPs 49 and 93) formed a separate node of their own (Fig. 1; Supplementary Fig. S2).
Association of gene copy number and expression of miRNAs.

To study association between miRNA locus copy number and expression of miRNAs, previously published array comparative genomic hybridization (aCGH) data (32) of the cell lines (PC-3, DU-145, LNCaP, LACC4, and 22Rv1) and seven of the xenografts (LuCaP49, 69, 70, 73, 86, 93, and 115) were used. For the analysis, the expression values of miRNAs were divided into three groups according to whether the samples showed deletions, amplifications, or no alterations of the corresponding miRNA loci according to the aCGH data (32). There was a moderate association ($P = 0.0290$, Kruskal-Wallis test) between the expression of the miRNAs and the copy number alterations of the loci. On average, the miRNAs located in the chromosomal regions of deletion showed the lowest level of expression, whereas the miRNAs located in the regions of amplifications showed the highest expression (Fig. 2).

**miRNA expression in clinical tissue samples.** After establishing the miRNA array hybridization system with the cell line and xenograft samples, the method was applied for miRNA expression profiling of clinical prostate tissue samples from 4 BPHs, 5 untreated prostate carcinomas, and 4 hormone-refractory prostate carcinomas. As shown in Fig. 3, the miRNA expression patterns in BPH, untreated prostate carcinoma, and hormone-refractory prostate carcinoma were clearly distinguishable from each other.

To validate the array hybridization results, expression of miR-145 that showed clear expression differences between the tissue samples was studied by using the qRT-PCR, the result of which up and validated. The confirmation of the array data was done by dot blot hybridization as well as qRT-PCR. Expression of 128 of 319 (40%) miRNAs was detectable in, at least, one of the samples by array hybridization, indicating that the majority of the miRNAs were either not expressed in these samples or that the expression was too low to be detected by this method. Because the expression of miRNAs is highly tissue specific (39, 40), it was actually not expected that the expression of all the miRNAs would be detected in prostate cancer cells.

**Discussion**

Expression profiling of 319 miRNAs in 6 prostate cancer cell lines, 9 prostate cancer xenograft samples, and 13 clinical prostate tissue samples (4 BPH, 5 untreated prostate carcinomas, and 4 hormone-refractory prostate carcinomas) was carried out to identify the prostate cancer miRNA signature and to use the miRNA expression profiling in classifying prostate tumors. The antisense oligonucleotide array hybridization method was first set up and validated. The confirmation of the array data was done by dot blot hybridization as well as qRT-PCR. Expression of 128 of 319 (40%) miRNAs was detectable in, at least, one of the samples by array hybridization, indicating that the majority of the miRNAs were either not expressed in these samples or that the expression was too low to be detected by this method. Because the expression of miRNAs is highly tissue specific (39, 40), it was actually not expected that the expression of all the miRNAs would be detected in prostate cancer cells.
Surprisingly, clustering analysis of the cell lines and xenografts did not simply separate the cell lines and xenograft samples in their own subclusters, but instead they were mixed. They seemed to form subnodes according to their AR status: all AR-positive cell lines (LNCaP, 22Rv1, LAPC4, and VCaP) were located close to each other in the same subcluster, whereas the two AR-negative cell lines (PC-3 and DU-145) formed a separate subnode in the other subcluster. Similarly, the two AR-negative xenograft samples (LuCaPs 49 and 93), representing small-cell carcinomas of prostate, were clustered in their own subnode, separately from the other xenografts. The results suggest that androgens might regulate the expression of some miRNAs. Very little is known about the regulation of the expression of miRNAs themselves, and no studies on their androgen regulation have been published thus far. Alternatively, the differential expression of miRNAs in AR-positive and AR-negative cells may reflect general fundamental differences between AR-positive and AR-negative prostate tumors that are not necessarily directly under the regulation of androgens.

When the expression of individual miRNAs between the clinical BPH and carcinoma samples was compared, 51 miRNAs were differently expressed.

Table 1. miRNAs showing differential expression in the prostate carcinoma samples compared with the BPH samples

<table>
<thead>
<tr>
<th>miRNA</th>
<th>In all carcinomas</th>
<th>Only in hormone-refractory carcinomas</th>
<th>In all carcinomas</th>
<th>Only in hormone-refractory carcinomas</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(n = 22)</td>
<td>(n = 15)</td>
<td>(n = 8)</td>
<td>(n = 6)</td>
</tr>
<tr>
<td>let-7a</td>
<td></td>
<td>let-7f</td>
<td>miR-202</td>
<td></td>
</tr>
<tr>
<td>let-7b</td>
<td></td>
<td>let-7f</td>
<td>miR-210</td>
<td></td>
</tr>
<tr>
<td>let-7c</td>
<td></td>
<td>miR-19b</td>
<td>miR-296</td>
<td></td>
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<tr>
<td>let-7d</td>
<td></td>
<td>miR-22</td>
<td>miR-320</td>
<td></td>
</tr>
<tr>
<td>let-7g</td>
<td></td>
<td>miR-26b</td>
<td>miR-370</td>
<td></td>
</tr>
<tr>
<td>miR-16</td>
<td></td>
<td>miR-27a</td>
<td>miR-373*</td>
<td></td>
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<tr>
<td>miR-23a</td>
<td></td>
<td>miR-27b</td>
<td>miR-498</td>
<td></td>
</tr>
<tr>
<td>miR-23b</td>
<td></td>
<td>miR-29a</td>
<td>miR-503</td>
<td></td>
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<tr>
<td>miR-26a</td>
<td></td>
<td>miR-29b</td>
<td></td>
<td></td>
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<tr>
<td>miR-92</td>
<td></td>
<td>miR-30a_5p</td>
<td></td>
<td></td>
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<tr>
<td>miR-99a</td>
<td></td>
<td>miR-30b</td>
<td></td>
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<tr>
<td>miR-103</td>
<td></td>
<td>miR-30c</td>
<td></td>
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<tr>
<td>miR-125a</td>
<td></td>
<td>miR-100</td>
<td></td>
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<tr>
<td>miR-125b</td>
<td></td>
<td>miR-141</td>
<td>miR-205</td>
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<tr>
<td>miR-143</td>
<td></td>
<td>miR-148a</td>
<td></td>
<td></td>
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<tr>
<td>miR-145</td>
<td></td>
<td>miR-221</td>
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<tr>
<td>miR-195</td>
<td></td>
<td>miR-222</td>
<td></td>
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<tr>
<td>miR-199a</td>
<td></td>
<td>miR-497</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

1 P value of 0.0028 (Mann-Whitney U test) for all miRNAs in this category.
2 P value of 0.0286 (Mann-Whitney U test) for all miRNAs in this category.
3 P value 0.0040 (Mann-Whitney U test) because of one missing value in the category of carcinomas.
The fundamental methodologic differences used in the studies...

...up-regulation or down-regulation. This discrepancy may be due to the fundamental methodologic differences used in the studies.

Volinia et al. (26) actually measured the expression of both the primary miRNA transcripts (pri-miRNAs) and the active, mature miRNAs, simultaneously. Considering that the pri-miRNAs are transient products in the miRNA biogenesis and presumably are rapidly further processed after their synthesis (46), the expression status of the pri-miRNA and the mature miRNA is not necessarily the same. Therefore, the profiles of combined expression of the primary miRNAs and the mature miRNAs and the profiles of expression of only the mature miRNAs probably should not be compared with each other.

Because of the tissue specificity of the miRNAs (39, 40), different sets of miRNAs are likely to be up-regulated or down-regulated in cancers of different cellular origins, although it has been reported that the miRNA signatures of different cancer types could share some individual miRNAs (26). Of the miRNAs that were detected here to be up-regulated or down-regulated in prostate carcinoma samples, some have been reported to be similarly deregulated in other cancers: down-regulation of miR-125a and miR-125b in breast cancer (12), the let-7 miRNAs in lung cancer (15, 16), and miR-143 and miR-145 in several different cancer types (9, 11, 12). However, most of the differentially expressed miRNAs detected in this study have not been commonly reported to be deregulated in other cancers, suggesting that prostate cancer has a miRNA signature that is specific for this cancer type.

The differential expression of miRNAs in carcinoma samples suggests that miRNAs could be mechanistically involved in the development of cancer. To elucidate the role of the individual miRNAs in prostate cancer, genes and pathways that are regulated by the differentially expressed miRNAs should be studied. Unfortunately, only a few targets of miRNAs have been identified thus far. Of the differentially expressed miRNAs detected in this study, the let-7 family has been shown to target the RAS oncogene (22), miR-16 to repress the antiapoptotic factor BCL2 (21), and miR-125a and miR-125b to suppress the oncogenes ERBB2 and ERBB3 (47). Experimental evidence for targets of the other differentially expressed miRNAs is not available. Because of the imperfect complementarity of miRNAs and their target sequences, target prediction, even with the aid of special prediction programs, is very challenging and inaccurate. Therefore, targets of most of the miRNAs, and thereby also their functions, still remain to be studied.

Hierarchical clustering of the BPH, untreated, and hormone-refractory prostate carcinoma samples successfully separated the BPH samples from the carcinoma samples. Moreover, the carcinoma samples were quite accurately further classified according to their androgen dependence, with four of the five untreated tumors forming one subcluster and the four hormone-refractory, more aggressive tumors plus the remaining untreated tumor forming the other subcluster. In addition, the untreated tumor in the hormone-refractory subcluster was separated into its own branch. Therefore, it seems that the miRNA expression profiles of the hormone-refractory tumors were more similar to each other than those of the untreated ones. The surprisingly high accuracy of clustering suggests that, in spite of the relatively low number of miRNAs known by now, classifying tumors by their miRNA expression can be very informative. In fact, it has already been evidenced that miRNA profiling is more accurate in classifying poorly differentiated tumors than miRNA profiling (29). Therefore, miRNA expression profiling could be used, for example, in developing new diagnostic tools for cancers that currently are lacking good molecular markers, such as prostate cancer.
Individual miRNAs and miRNA signatures have already been shown to have prognostic value in some cancers, such as chronic lymphatic leukemia, diffuse large B-cell lymphoma, and lung carcinomas (14–16, 24). To identify miRNAs that could be used as prognostic or predictive markers in prostate cancer, more miRNA expression profiling studies with larger number of tumor samples are warranted.

In summary, expression profiling revealed several miRNAs that are underexpressed or overexpressed in prostate cancer, thereby identifying miRNAs that could be significant in prostate cancer development. The data also suggest that one mechanism for differential expression could be miRNA gene copy number.

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References


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