A Polyepitope DNA Vaccine Targeted to Her-2/ErbB-2 Elicits a Broad Range of Human and Murine CTL Effectors to Protect against Tumor Challenge

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Abstract

A cDNA vaccine (pVax1/pet-neu) was designed to encode 12 different Her-2/ErbB-2–derived, HLA-A*0201–restricted dominant and high-affinity heteroclitic cryptic epitopes. Vaccination with pVax1/pet-neu triggered multiple and ErbB-2–specific CTL responses in HLA-A*0201 transgenic HHD mice and in HLA-A*0201 healthy donors in vitro. Human and murine CTL specific for each one of the 12 ErbB-2 peptides recognized in vitro both human and murine tumor cells overexpressing endogenous ErbB-2. Furthermore, vaccination of HHD mice with pVax1/pet-neu significantly delayed the in vivo growth of challenged ErbB-2–expressing tumor (EL4/HHD/neu murine thymoma) more actively when compared with vaccination with the empty vector (pVax1) or vehicle alone. These data indicate that the pVax1/pet-neu cDNA vaccine coding for a poly-ErbB-2 epitope is able to generate simultaneous ErbB-2–specific antitumor responses against dominant and cryptic multiple epitopes. [Cancer Res 2007;67(14):7028–36]

Introduction

The human erbB-2 proto-oncogene codifies for a transmembrane protein (Her-2/ErbB-2) member of the epidermal growth factor receptor family with tyrosine kinase activity whose ligand is still unknown. It is commonly overexpressed in several human tumors, including breast, ovarian, and lung carcinomas (1–3). Overexpression of ErbB-2 is associated with aggressive disease and poor prognosis (4, 5). ErbB-2–specific antibodies and tumor-infiltrating CTL occur naturally in breast, renal, and ovarian cancer patients, confirming the in vivo immunogenicity and ability of ErbB-2 to break self-tolerance (6–10). To date, multiple ErbB-2 epitopes presented by HLA-A*0201 or HLA-A*0301 have been identified and used as targets of specific CTL responses (9–18).

Antitumor vaccination strategies have been taking many forms, including free peptides, dendritic cells loaded with peptides or tumor lysates, and DNA-based vaccines. Although the strategies of vaccination with peptides are very attractive over other forms for the feasibility, many studies using native unmodified peptides resulted in low-level responses showing a high variability from one patient to another (9–23). Active immunotherapy strategies aimed to generate specific T-cell responses are currently being investigated in animal models or in clinical trials (24, 25). To date, increasingly powerful methods to stimulate antitumor immune responses have been developed into rat Her-2 Neu transgenic mice, including vaccination with electroporated DNA coding for the same transgene product (rat-Her-2/neu) or with “foreign” antigens through plasmid DNA xeno-immunization expressing both extracellular and transmembrane domains of the human erbB-2 oncogene product (26, 27). Yet, the stringent role of humoral or cell-mediated immune response in vivo tumor inhibition was not completely clarified: however, depletion of CD8+ T cells in ErbB-2 transgenic mice before vaccination accelerated tumor outgrowth (28, 29). The major limits of such models remain the presence of the erbB-2 transgene during ontogenesis for immune tolerance shaping and the fact that DNA vaccines coding whole proteins from extracellular and transmembrane domains of the receptor might produce unpredictable autoimmune side effects in humans (30).

To overcome epitope variability among peptide presentation of tumors and to reduce the risk of tolerance toward the ErbB-2 high-affinity epitope peptides, we explored the possibility to mobilize in vivo a large ErbB-2–derived T-cell epitope repertoire in HLA-A*0201 transgenic mice and in an in vitro human model, wherein human peripheral blood mononuclear cells (PBMC) were used to trigger a multiple antitumor CTL response aimed to target simultaneously the HLA-A*0201–restricted dominant and cryptic epitopes of the human ErbB-2 protein.

Here, we present a cDNA vaccine (pVax1/pet-neu) encoding 12 ErbB-2–derived, HLA-A*0201–restricted dominant and cryptic epitopes. The pVax1/pet-neu vaccine was able to trigger a polyspecific CTL response in HLA-A*0201 transgenic HHD mice and induce an ErbB-2–specific antitumor immunity in vivo. Moreover, pVax1/pet-neu stimulated a marked polyspecific CTL response when used to sensitize human PBMC in vitro.

Materials and Methods

Mice. The HLA-A*0201 transgenic HHD mice were previously described (31). All in vivo experiments were done in accordance with the Institut Gustave Roussy ethical guidelines.
Cell culture. Breast carcinoma cell line MCF-7-T103 (HLA-A2.1+/ErbB-2+) was purchased from the American Type Culture Collection. C1R is a human lymphoblastic cell line which is defective in the synthesis of class I HLA molecules. C1R-A2, conversely, is a derivative subline stably transfected with the HLA-A*0201 gene, which expresses large amounts of empty HLA-A*0201 on the cell membranes and does not express other class I HLA molecules (32). These cells were provided by Dr. Jeffrey Schlom (Laboratory of Tumor Immunology and Biology, Center for Cancer Research, National Cancer Institute, NIH, Bethesda, MD). RMAS/HHD and EL4/HHD cells (mouse thymoma 120-2 for murine MHCs and transduced with HHD construction) were previously described (33). ELA/HHD/ErbB-2 cells were obtained by transfection of ELA/HHD cells with a long terminal repeat-erbB-2-gpt expression retroviral vector encoding for the human ErbB-2. The ErbB-2 expression was verified by immunoblot and fluorescence-activated cell sorting analysis using the ErbB-2-specific C-1 monoclonal antibody (mAb; Calbiochem). EL4/HHD/Tel-AML transfectants [Tel/Amil is a t(12;21) translocation critical for B-cell leukaemogenesis; ref. 34] were a kind gift of Dr. P. Langlade-Demoyen (Institut Pasteur, Paris, France). Tumor cells were maintained in RPMI 1640 supplemented with 10% heat-inactivated FCS, 2 mmol/L glutamine, 100 units/mL penicillin, and 100 μg/mL streptomycin (all purchased from Life Technologies, Inc.).

Peptides. Peptides were synthesized by Syntem (Nimes, France). They were dissolved in PBS, 3% DMSO and stored at −20°C. Sequences of the human ErbB-2 peptides are illustrated in Fig. 1A.

Measurement of peptide/HLA-A*0201 relative affinity and stability. A protocol used to measure relative affinity has previously been described (35). Briefly, T2 cells were incubated with various concentrations of peptides (0.1–100 μmol/L) for 16 h and then stained with the BB7.2 mAb to quantify the expression of HLA-A*0201. For each peptide concentration, the HLA-A*0201–specific staining was calculated as the percentage of the staining obtained with 100 μmol/L of the reference peptide HIVp0991 (IVGAEFTFYV). Relative affinity was determined as concentration of each peptide / concentration of the reference peptide that was able to induce 20% of HLA-A*0201 expression. For peptide/HLA-A*0201 complex stability measure, T2 cells were incubated overnight with 100 μmol/L of each peptide at 37°C. Cells were then treated with brefeldin A for 1 h; washed; incubated at 37°C for 0, 2, 4, and 6 h; and then stained with the BB7.2 mAb. DC50 was defined as the time required for the loss of 50% of the HLA-A*0201, as previously described (35).

Design and construction of the pVax1/pet-neu polyepitope. A polyepitope cDNA (pet-neu) was designed using optimized codons from the human erbB-2 sequence and incorporating the following elements from the 5′ end: a BamHI restriction site, a Kozak consensus start sequence, 12 consecutive epitopes, a tag epitope recognized by the SV5-pk (pk) mAb, an XhoI restriction site to facilitate future modification, a translational stop codon, and an EcoRI restriction site at the 3′ end. To construct pet-neu (404 bp), six oligonucleotides were synthesized (Imperial Cancer Research Fund Oligonucleotide Synthesis Service, Leeds, United Kingdom) that overlapped by 20 bp and together covered the polyepitope sequences. Polyepitope cDNAs were assembled by PCR. Adjacent primers were dimerized (1 μg of each) in separate PCR reactions containing 1 unit of Deep Vent DNA polymerase, 1× thermopol buffer, and 200 μmol/L deoxynucleotide triphosphates (New England Biolabs, Inc.) in a final volume of 100 μL overaid with 40 μL of mineral oil (Sigma). A hot start PCR reaction was carried out at 94°C, during which time primers and DVP were added. The cycling conditions were 94°C for 20 s, 42°C for 30 s, 72°C for 30 s, five cycles. Reactions were held at 72°C and 20 μL of adjacent primer dimer reactions were combined before a further five cycles of amplification. The third primer dimer reaction (20 μL) was then added to the second primer dimer combination for the final five cycles. PCR products from the oligonucleotide splicing by overlap extension steps were used as templates for a final PCR reaction using terminal primers complementary to the polyepitope. The final PCR reaction product was purified by electrophoresis, cloned into pVAX1 (Invitrogen), and recombinant plasmids examined for mutation by DNA sequencing.

CTL generation in vaccinated HHD mice. For peptide in vivo priming, HHD mice were injected s.c. with 100 μg of peptide emulsified in incomplete Freund’s adjuvant in the presence of 140 μg of the IAβ−restricted HBVcore252–268 T-helper (Th) epitope. After 11 days, spleen cells (5 × 107 in 10 mL) were then stimulated in vitro with cognate peptide (10 μmol/L). On day 6 of culture, the bulk responder populations were tested for specific cytotoxicity. For pVax1/pet-neu in vivo priming, HHD mice were injected i.m. with 10 μmol/L cardiotoxin (Latoxan) in 50-μL PBS. Twenty-five days later, mice were injected i.m. with 100 μg of pVax1/pet-neu. Spleen cells were collected 1 week after the second vaccination and stimulated with lipopolysaccharide (LPS) blasts loaded with the cognate peptide (splenocytes/LPS blasts ratio, 3:1) in the presence of 10% T-cell growth factor–enriched supernatant. Effector CTL were tested after four cycles of in vitro stimulation. Positive responses were considered when peptide-specific lysis was at least 10% above the background level (RMAS/HHD targets pulsed with the irrelevant HVIgag22 peptide).

ELISPOT assay. Peptide-specific T cells from vaccinated mice were counted by IFN-γ ELISpot (36). IFN-γ–secreting cells were counted using the automated image analysis system ELISpot Reader (AID). The Wilcoxon two-tail rank test was done to determine whether there was a statistically significant difference between the numbers of IFN-γ–secreting cells in the wells stimulated with the ErbB2– or the HVIgag22–derived peptides.

Generation of human CTL. Blood samples were obtained from normal healthy donors. Dendritic cells were obtained by stimulation of adherent cells from PBMC with 50 ng/mL granulocyte macrophage colony-stimulating factor and 0.5 ng/mL IL-4 (both purchased from R&D Systems) for 7 days. The PBMC for CTL primary cultures were washed thrice in Dulbecco’s PBS and then resuspended in AIM-V medium (Life Technologies), supplemented with 5% pooled human AB serum (Valley Biomedical), 2 mmol/L glutamine, 100 units/mL penicillin, and 100 μg/mL streptomycin (Life Technologies). Cells (2 × 106) in a 100-μL volume of complete medium were added to each well of a 96-well flat-bottomed assay plate (Corning, Costar Corp.). One hundred thousand dendritic cells were grown in six-well microplates at 37°C and transfectioned with 1 μg of pVax1/pet-neu using the Effectene transfection reagent (Qiagen) according to the manufacturer’s instructions. Transfected dendritic cells were irradiated and added to the lymphocyte cultures at a final dendritic cell/PBMC ratio of 1:5. The cocultures were then incubated for 5 days at 37°C in a humidified atmosphere containing 5% CO2. The cultures were fed with 20 IU/mL human interleukin-2 (IL-2; Cetus Corp.) for 10 days and IL-2–containing medium was replenished every 3 days. Incubation for 5 days with pVax1/pet-neu–pulsed dendritic cells plus 10 days with IL-2 constitutes one in vitro stimulation cycle. Cytotoxicity was tested after four in vitro stimulation cycles.

Cytotoxicity assay. Targets were labeled with 150 μCi of 51Cr, plated in 96-well round-bottomed plates (104 per well in 100 μL of RPMI 1640 + 3% FCS), and, when necessary, loaded with the peptide (1 μmol/L) for 90 min. Effectors were added to targets at different ratios and plates were incubated at 37°C for 6 h. After incubation, supernatants were collected and radioactivity was measured in a gamma counter. Percentage of specific lysis was calculated as (experimental spontaneous / maximal spontaneous 51Cr release) × 100.

Vaccination and analysis of antitumor activity in vivo. HHD mice were vaccinated twice either with pVax1/pet-neu or with the empty pVax1 at 2-week intervals or with the vehicle PBS. Ten days after the second vaccination, mice were challenged with 2 × 105 EL4/HHD/Tel-AML or EL4/HHD/ErbB-2 cells. For each immunogen, groups of nine HHD mice were vaccinated. The number of injected tumor cells was previously defined as 2-fold the minimal dose that gives 100% of tumors in HHD mice. Tumor volumes were measured by a caliper in two dimensions, and the volumes were calculated using the formula (width2 × length) / 2. Animals were monitored twice a week for the development of palpable tumors. Mice were sacrificed when they showed any symptom of very sickness, hard to breathe and move. Animals were monitored twice a week for the development of palpable tumors. Mice were sacrificed when they showed any symptom of very sickness, hard to breathe or move, or when their tumors grew larger than 1,000 mm3 to avoid unnecessary suffering. Survival was recorded until 65 days. All in vivo experiments were done in accordance with ethical guidelines of the Institut Gustave Roussy.

Statistical analysis. Statistical significance of median survival was determined by survival data by Mantel-Haenszel log-rank test. Differences
A

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B

Figure 1. A, affinity for HLA-A*0201 of ErbB-2 peptides (RA, relative affinity; DC_{50}, 50% of stably bound peptide to MHC molecules at given time). In boldface are amino acids differing from Her-2 murine sequence. B, generation of specific CTL peptide–vaccinated HHD mice. Spleen cells from peptide–primed HHD mice were in vitro stimulated as described in Materials and Methods. CTL activity was tested against RMAS/HHD cells loaded with the HIVgag_{36} (■) or the cognate ErbB-2 native (■) peptides. E/T ratio, effector/target ratio. Bars, SD. C, design of the pVax1/pet-neu polyepitope.
in average tumor volume were evaluated by a two-tailed t test, whereas statistical differences in tumor incidence at defined time points were evaluated using the Fisher exact test. The Wilcoxon two-tail rank test was done to determine whether there was a statistically significant difference between the numbers of IFN-γ-secreting cells in the wells stimulated with the ErbB-2– or the HIVgag76-derived peptides.

## Results

### Affinity for HLA-A*0201 and immunogenicity of ErbB-2 peptides

Twelve HLA-A*0201–restricted ErbB-2 epitopes were included in the polypeptidic construct (pet-neu; Fig. L4). Eight of them (p5, p48, p369, p689, p773, p879, p799, and p1023) were described to be targets of tumor-infiltrating lymphocytes in breast, ovarian, and gastric cancers and renal cell carcinoma (8–11, 15). They exhibited high affinity for HLA-A*0201 (relative affinity <5; DC50 >4 h) except pN02, which is considered an intermediate binder (relative affinity >5; DC50 >4 h). All epitopes show full homology with murine Her-2, with the exception of ErbB peptide pVax1/pet-neu pVax1

A second set of experiments, spleen cells from three pVax1/pet-neu–vaccinated mice were processed epitopes on ErbB-2–expressing tumors. In vitro restimulated with peptide and then

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<th>Table 1. Polyspecific anti-ErbB-2 response generated in HHD mice vaccinated with pVax1/pet-neu</th>
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NOTE: Results represent the mean of IFN-γ-producing cells/10^5 CD8 cells. Means in boldface are statistically significant compared with HIVgag76 (P < 0.05, Wilcoxon two-tail rank test).
to evaluate the in vitro antitumor ability of the multiple CTL response induced in pVax1/pet-neu–primed mice using the syngeneic murine tumor cell line EL4/HHD expressing human ErbB-2 (EL4/HHD/ErbB-2) or Tel-Aml (EL4/HHD/Tel-Aml) as target cells. Figure 3A shows the CTL activity from 2 representative HHD mice of the 11 vaccinated with pVax1/pet-neu; results indicate that EL4/HHD/ErbB-2 cells were killed by CTL specific for ErbB-2 peptides, whereas EL4/HHD/Tel-Aml cells were not, showing specific CTL being able to recognize the corresponding natural epitopes. To show that generation of ErbB-2–specific CTL was due to the in vivo priming with pVax1/pet-neu and not to the repetitive in vitro stimulation with peptide-loaded LPS-activated spleen cells, we vaccinated mice with either pVax1/pet-neu or the empty pVax1 and stimulated pooled spleen cells from each group with LPS-generated blasts pulsed with peptide in the same conditions. Results in Fig. 3B show that in vitro stimulated spleen cells from pVax1/pet-neu mice developed ErbB-2–specific CTL whereas in vitro stimulated spleen cells from mice vaccinated with pVax1 vector alone did not.

Antitumor protective immunity in pVax1/pet-neu immunized mice. Next, we determined whether the administration of the pVax1/pet-neu polyepitope vaccine could prevent the
establishment of ErbB-2–expressing tumor transplants. Ten days after the second vaccination with the pVax1/pet-neu or pVax1 plasmid, HHD mice were challenged with $2 \times 10^4$ EL4/HHD/ErbB-2 cells. To test the specificity of the antitumor immune response, a group of pVax1/pet-neu–vaccinated mice challenged with $2 \times 10^4$ EL4/HHD/Tel-Aml cells was added. Tumor growth was monitored twice a week until day 32 when all the unvaccinated mice died or were sacrificed. Mice of the remaining groups were followed for time survival. Results of tumor growth are shown in Fig. 4A. By day 17, tumor overexpressing ErbB-2 had developed in all mice vaccinated with the control vector or vehicle alone. By contrast, none of the mice immunized with the pVax1/pet-neu vaccine exhibited tumor growth. In this group of vaccinated mice, tumor protection continued through day 23 in all animals ($P = 0.00004$). Conversely, tumor growth of EL-4/HHD/Tel-Aml cells was not affected by the pVax1/pet-neu vaccination. These results indicated the appearance of an anti–ErbB-2–specific immune interference with tumor growth elicited by the pVax1/pet-neu immunization. At day 28, tumors developed also in seven of the nine pVax1/pet-neu–vaccinated mice. However, the tumors developed in these animals (116 ± 66) were smaller than those observed in mice immunized with vehicle only (772 ± 268; $P < 0.001$) or with vector alone (404 ± 121; $P < 0.001$). It is worth noting that at this stage of tumor development, vaccination with pVax1 vector alone affected tumor growth with respect to that with vehicle alone ($P = 0.0027$). The in vivo antitumor protective immunity elicited by the pVax1/pet-neu immunization was specific for ErbB-2 because vaccinated mice were not protected against tumor cells lacking ErbB-2 (EL4/HHD/Tel-Aml tumor cell line). All these mice developed tumors by day 17 and the average of tumor volume measured at day 28 was $578 \pm 231$ ($P < 0.001$). Furthermore, pVax1/pet-neu vaccination increased the survival of tumor-bearing mice. Results are presented in Fig. 4B. Whereas unvaccinated mice died at day 32, the mean survival for the pVax1-vaccinated mice was $373 \pm 4.4$ days ($P = 0.04$, compared with unvaccinated mice). The time of survival was significantly improved in the group of pVax1/pet-neu–vaccinated mice; five of nine mice were still alive at day 65 ($P = 0.02$ and $P = 0.0007$, compared with pVax1 and PBS immunized mice, respectively). Overall, these results showed that interference with tumor growth of cells overexpressing ErbB-2 was obtained by active immunization with pVax1/pet-neu.

**Human ErbB-2–specific CTL response to pVax1/pet-neu.** The pVax1/pet-neu vaccine capacity to mobilize a human T-lymphocyte repertoire specific for the epitopes encoded in the construction was investigated by CTL priming with pVax1/pet-neu–transduced dendritic cells. CTL lines were established

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**Figure 4.** A, effects of immunization with pVax1/pet-neu on the growth of EL4/HHD cells transduced with the cDNA encoding for ErbB-2. *, number of mice with tumors/total number of mice; †, average tumor volume (mm$^3$) ± SD; ‡, average tumor volume of pVax1/pet-neu–vaccinated mice versus vehicle (Student’s t test); ††, average tumor volume of pVax1/pet-neu–vaccinated mice versus vehicle (Student’s t test); †††, average tumor volume of pVax1/pet-neu–vaccinated mice challenged with EL4/HHD/ErbB-2 versus pVax1/pet-neu–vaccinated mice challenged with EL4/HHD/ErbB-2 (Student’s t test). B, effects of immunization with pVax1/pet-neu on time survival of mice challenged with EL4/HHD cells transduced with the cDNA encoding for ErbB-2. Nine HHD mice in each group were vaccinated with vehicle (PBS), empty pVax1 vector, or pVax1/pet-neu encoding the polyepitope ErbB-2 vaccine. Ten days after the second vaccination, mice immunized with pVax1/pet-neu were challenged with $2 \times 10^4$ EL4/HHD/ErbB-2 or EL4/HHD/Tel-Aml tumor cells, whereas mice immunized with vehicle alone or pVax1 vector were challenged with $2 \times 10^4$ EL4/HHD/ErbB-2 cells. The time of survival determined until day 65 was significantly improved in the pVax1/pet-neu–vaccinated mice group compared with the other groups of immunized mice ($P = 0.02$ and $P = 0.0007$, pVax1/pet-neu versus pVax1 or PBS, respectively).
by in vitro priming and cyclic restimulations with autologous dendritic cells pulsed with pVax1/pet-neu in the presence of IL-2. After the fourth stimulation, lymphocyte cultures were tested for ErbB-2–specific cytolytic activity against CIITAC2 cells loaded with ErbB-2 peptides (A) or ErbB-2 CMYC-103 in the presence of the HLA-A*0201 specific A2.69 and the irrelevant UPC-10 mAbs (B). Bars, SD.

Figure 5. In vitro stimulation of ErbB-2 polyspecific human CTL by pVax1/pet-neu. PBMC from HLA-A*0201 healthy donors were stimulated with autologous dendritic cells transduced with pVax1/pet-neu. CTL were tested after the fourth in vitro stimulation against CIITAC2 cells loaded with ErbB-2 peptides (A) or ErbB-2 CMYC-103 in the presence of the HLA-A*0201 specific A2.69 and the irrelevant UPC-10 mAbs (B). Bars, SD.

Discussion

We evaluated the in vivo efficacy of a polyepitope vaccine (pVax1/pet-neu) composed of 12 ErbB-2–derived, HLA-A*0201–restricted epitopes. We showed that HLA-A*0201 transgenic HHD mice vaccinated with pVax1/pet-neu developed simultaneously CTL against the majority of these epitopes. These CTL recognize the endogenous ErbB-2 epitopes and provide partial protection of HHD mice from the growth of the marine tumor EL4, engineered to express human ErbB-2. Moreover, pVax1/pet-neu stimulates in vitro from HLA-A*0201 human PBMC a multispecific CTL response capable to lyse tumor cells.

Several variables could influence the vaccine efficacy of a polyepitope, such as the presence of a Th epitope, the optimal cleavage of the different epitopes by the proteasome which depends on the epitope rearrangement, the addition of spacers between the epitopes, the creation of new junctional epitopes, and, finally, the role of affinity of the peptide epitopes included in the construction for immunodominance and skewing of the multiple specific CTL response in vitro. However, data in the literature do not allow for the establishment of a consensus on the role of each one of these variables on polyepitope efficacy. For instance, results of Mateo et al. (37) suggest that there is a correlation between affinity and immunogenicity of peptides included in a melanoma polyepitope. Furthermore, whereas Ishioka et al. (38) showed that, using an HIV polyepitope, the affinity of the epitopes does not interfere with their capacity to induce a CTL response. Palmowski et al. (39) showed the role of affinity diversity among peptide epitopes included within a polyepitope construction as crucial for skewing of immune response and CTL immunodominance in HLA-A*0201 transgenic mice. Concerning the need for a Th epitope, Ishioka et al. (38) have also shown the usefulness of including the PADRE Th epitope for the improvement of the vaccination efficacy, whereas Velders et al. (40) did not find any difference in antitumor immunity between normal and CD4+/- alleles in mice vaccinated with a polypeptide containing the TT Th epitope. A consensus, however, seems to be established for the role of the polyepitope organization (epitope rearrangement, addition of spacers) that should allow for the appropriate cleavage of all the epitopes and should prevent the creation of new junctional epitopes with high affinity for the HLA molecule (38, 39). Therefore, to design the pVax1/pet-neu polyepitope, we took care that the arrangement of the 12 epitopes (a) allows the processing of each one at its COOH-terminal position; (b) does not create new junctional peptides with high affinity for HLA-A*0201; and (c) includes native or high-affinity heteroclitic PIY variants of peptide epitopes showing similar strong binding affinity in vitro for the MHC-I molecule and high immunogenicity in vivo into mouse models. The processing at the COOH-terminal position was evaluated by two predictive models of proteasome cleavage (41, 42). Among the 10 virtual arrangements screened, we have chosen the pet-neu construct that fit better with the two criteria, p773, p369, p1023, p5, p466Y, p402Y, p391Y, and p650Y were predicted to be processed and only five new junctional peptides with predicted high affinity for HLA-A*0201 could be generated (p679, p949, p179, p9110, and p6310) but only one (p9110) was predicted to be processed. Although we have not quantified the processing of the ErbB-2 and the junctional specific A2.69 mAb abrogated the MCF-7-T103 lysis whereas the irrelevant UPC-10 did not, thus showing the HLA-A*0201 restriction of the MCF-7-T103 recognition.
epitopes, our results showing that HMD mice vaccinated with pVax1/pet-neu develop CTL against all the ErbB-2 epitopes show that all these epitopes are processed.

The major point addressed in our work is the possibility to include heteroclitic cryptic tumor epitopes in polyepitopic vaccines. The *ex vivo* ELISPOT assay done after mice vaccination with pVax1/pet-neu did not show any immunodominance among native or heteroclitic peptide epitopes included within the construction, suggesting the possibility to increase *in vivo* a wide response directed simultaneously against multiple epitopes independently of their nature. Furthermore, results on CTL specific for ErbB-2 showed variability of response among different mice. Interpretation of such results might also involve sequence differences between some ErbB-2–derived epitopes (p5, p466Y, p391Y, and p650Y) and the murine Her-2 sequence, but the possibility to increase a multiple and diverse CTL response having antitumor activity was strongly indicated. Furthermore, results on human PBMC showed a CTL response variability among two individuals showing the possibility to mobilize a human T-cell repertoire specific for multiple ErbB-2 epitopes including cryptic epitopes. The multiple CTL response was capable to kill MCF-7 tumor cells and no correlation with the ErbB-2 specificity of CTL response evoked into mice was observed. Thus, the major evidence of the study is that pVax1/pet-neu generates a multiple CTL response and that response was addressed concomitantly to dominant and cryptic epitopes with antitumor effects *in vitro* and *in vivo*.

The interest of cryptic tumor epitopes in tumor immunotherapy is suggested in our previous work (17, 35) and is mainly based on the fact that the specific CTL repertoire is not submitted to the thymic or peripheral negative selection and must, therefore, be large and avid. The mechanisms of tolerance to self are particularly important for the design of new cDNA vaccines for the prevention of ErbB-2–expressing malignancies. Previous data from vaccines tested into mice models transgenic for rat-p185 cannot be directly translated to humans because the mechanisms of tolerance to human self-p185neu could be different from those of rat p185neu expressed in transgenic mice and the escape of human prneu to T-cell receptor.


References


Correction: Polyepitope DNA Vaccine and Tumor Challenge

In the article on polyepitope DNA vaccine and tumor challenge in the July 15, 2007 issue of *Cancer Research* (1), Poulam M. Patel should have been included as the 13th author, and the correct name of the 15th author is Andrew Mark Jackson. The correct affiliation for Dr. Patel and Dr. Jackson is Cancer Research UK Clinical Cancer Centre, Leeds, United Kingdom and Academic Division of Oncology, University of Nottingham, City Hospital, Nottingham, United Kingdom. Also, the correct affiliation for Steven G. Smith is Cancer Research UK Clinical Cancer Centre, Leeds, United Kingdom and London School of Tropical Hygiene and Infectious Disease, University of London, London, United Kingdom.

A Polyepitope DNA Vaccine Targeted to Her-2/ErbB-2 Elicits a Broad Range of Human and Murine CTL Effectors to Protect against Tumor Challenge

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