Positron Emission Tomography of Herpes Simplex Virus 1 Oncolyis

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Abstract

Viral oncolysis, the destruction of cancer cells by replicating viruses, is under clinical investigation for cancer therapy. Lytic viral replication in cancer cells both destroys the cells and liberates progeny virion to infect adjacent cancer cells. The safety and efficacy of this approach are dependent on selective and robust viral replication in cancer cells rather than in normal cells. Methods to detect and quantify viral replication in tissues have relied on organ sampling for molecular analyses. Preclinical and clinical studies of viral oncolysis will benefit significantly from development of a noninvasive method to repetitively measure viral replication. We have shown that positron emission tomography (PET) allows for in vivo detection of herpes simplex virus (HSV-1) replication in tumor cells using 9-(4-[18F]-fluoro-3-[hydroxymethyl]butyl)guanine ([18F]FHBG) as the substrate for HSV thymidine kinase (HSV-TK). As expected, phosphorylated [18F]FHBG is initially trapped within HSV-1–infected tumor cells and is detectable as early as 2 h following virus administration. MicroPET images reveal that [18F]FHBG accumulation in HSV-1–infected tumors peaks at 6 h. However, despite progressive accumulation of HSV-1 titers and HSV-TK protein in the tumor as viral oncolysis proceeds, tumor cell degradation resulting from viral oncolysis increases over time, which limits intracellular retention of [18F]FHBG. These observations have important consequences with regard to strategies to use [18F]FHBG PET for monitoring sites of HSV-TK expression during viral oncolysis. [Cancer Res 2007;67(7):3295–300]

Introduction

Early clinical trials in cancer gene therapy relied on viruses that are genetically engineered to prevent their replication in humans (1). This initial focus on replication-defective viruses resulted from concerns that replication of therapeutic viral vectors in humans would lead to unwanted toxicity. However, infection of tumors with replication-conditional viruses effectively reduce tumor burden as a result of cytopathic effects induced by lytic viral replication (2, 3). Following initial viral infection of tumor cells, progeny virions liberated from lysed tumor cells infect and destroy adjacent tumor cells, thereby resulting in an iterative process referred to as viral oncolysis. The safety and efficacy of this approach are dependent on selective and robust viral replication in cancer cells rather than in normal cells (4). Some replication-conditional viruses display innate tropism for cancer cells (5, 6), whereas others harbor genetically engineered alterations to increase their specificity for replication in tumor cells relative to normal cells (7, 8).

Many viruses have been examined for their potential as oncolytic viruses (9–13), including herpes simplex virus 1 (HSV-1). Several properties of this virus render it well suited to applications of viral oncolysis, including its high transgene capacity, lack of integration into the cellular genome, prevalence in the population, sensitivity to specific antiviral compounds (e.g., acyclovir), and rarity with which it produces severe medical illness. Oncolytic HSV-1 mutants have been engineered that replicate effectively and preferentially in neoplastic cells in vivo (7, 14, 15), reduce tumor burden in animal models (13, 16, 17), and carry therapeutically oriented transgenes that add to antineoplastic efficacy (18, 19). The safety and efficacy of HSV-1 mutants have been examined in clinical trials for patients with brain tumors (20), colon and rectal carcinoma liver metastases (21), and melanoma (22). Information learned from these clinical trials has been limited by the absence of methods to noninvasively determine sites of viral replication.

At present, detection of viral replication requires biopsy of tissue followed by analysis using molecular techniques such as immunohistochemical staining and PCR amplification. This strategy is suited to animal models but not suited to clinical trials. The ability to noninvasively and repetitively measure the magnitude and anatomic sites of viral infection in vivo would dramatically improve the quality of information learned from preclinical and clinical studies of viral oncolysis.

We and others have evaluated positron emission tomography (PET) for its usefulness in tracking sites of viral replication. The prototype enzyme-based PET reporter system relies on HSV thymidine kinase (HSV-TK). This viral enzyme is more promiscuous than its mammalian homologue in its spectrum of substrates and can phosphorylate a variety of substrates that are not phosphorylated by mammalian thymidine kinase. These include acycloguanosines [acyclovir, ganciclovir; 9-(4-fluoro-3-[hydroxymethyl]butyl)guanine (FHBG); and 9-[(3-fluoro-1-hydroxy-2-propoxy)methyl]guanine] and thymidine analogues [5-iodo-2′-fluoro-1′β-D-arabinofuranosyl-uracil (FIAU); ref. 23]. These compounds remain sequestered within cells once they are phosphorylated. Thus, when these compounds are radiolabeled with positron emitting isotopes, PET imaging identifies sites of HSV-TK activity.

We have characterized microPET measurements of HSV-1 oncolysis of murine flank tumors using [18F]FHBG. Importantly, unlike any studies done to date, this study includes a sequential analysis of the evolution of viral oncolysis over time. MicroPET measurements combined with direct measurements of reporter proteins and viral titers reveal an important discordance between levels of viral replication, HSV-TK expression, and microPET measurements. We also conducted microPET imaging studies of metabolic activity of the tumors and tumor cell replication during progression of the viral oncolysis using [18F]fluorodeoxyglucose and [18F]fluorothymidine ([18F]FLT), respectively.

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Materials and Methods

Cells and viruses. MC26 murine colon carcinoma cells and African green monkey kidney cells (Vero) were maintained in DMEM supplemented with 10% bovine growth serum (HyClone, Logan, UT), 100 units/mL penicillin (Life Technologies, Inc., Carlsbad, CA), and 100 μg/mL streptomycin (Life Technologies).

HSV-TK-transformed MC26 clones were created by subcloning sr39tk cDNA (kindly provided by Dr. S. Gambhir, Stanford University, Palo Alto, CA) into an expression vector driven by an SV40 promoter. MC26 cells were transfected using Lipofectamine (Invitrogen, Carlsbad, CA), and stable G418-resistant clones were isolated and screened for thymidine kinase expression using hypoxanthine-aminopterin-thymidine medium and by assessing their sensitivity to ganciclovir. An sr39tk-expressing MC26 stable transformant was selected that shows 15-fold greater accumulation of [18F]FHBG compared with nontransformed MC26 cells. Methods used to prepare and store virus and methods used to determine viral titers in vitro and in vivo have previously been described (24).

Animal studies. Animal studies were done in accordance with the policies of the Massachusetts General Hospital Subcommittee on Research Care of Laboratory Animals and the guidelines for BALB/c mice (Charles River Laboratories, Wilmington, MA), a single-cell suspension of 1 × 10⁶ MC26 cells in 100-μL PBS without Ca²⁺ or Mg²⁺ was inoculated into the left and right upper flank tumors reached 5 mm in size, 1 × 10⁶ plaque-forming units (pfu) of hrR3 in 100-μL PBS were injected into the center of the tumor. A total of 16 microPET imaging studies were conducted in 10 tumor mice using a viral dose of 1 × 10⁷ pfu to investigate HSV-1 replication. MC26 murine colon carcinoma cells stably transformed to express HSV-TK (MC26sr39tk) implanted into the left flanks of BALB/c mice were used to confirm [18F]FHBG expression in vivo.

MicroPET imaging of HSV-1 replication in murine tumors. Once flank tumors reached 5 mm in size, 1 × 10⁶ pfu of hrR3 were injected directly into the center of the tumor. Mice were anesthetized via inhalation of isoflurane (1–1.5%) with an oxygen flow rate of 2 L/min. Depth of anesthesia was monitored by respiratory rate and eye and footpad reflex. Catheterization of tail vein was done for administration of radioligand (needle size, 30 gauge; volume of injected activity, 20–60 μL). Heparin sodium (6 units/mL) was infused into the left and upper flanks. Seven days later, 1 × 10⁶ plaque-forming units (pfu) of hrR3 in 100-μL PBS were injected into the center of the tumor. This was followed by injecting 10% glucose in 0.9% saline into the tail veins of mice on the right (treated) side, tumor injected with the virus (on the right side), and the heart cavity to obtain blood input function for quantification of the radiotracer. Data were acquired for 60 min. Regions of interest including [18F]FHBG (100–150 Ci) into the tail vein catheter to acquire dynamic volumetric data for 120 min. Volumetric images were reconstructed with filtered back projection after the data were corrected for uniformity, scatter, attenuation, decay, and injected activity using the software AsiPro4.1 provided by the manufacturer. Time-activity curves were generated from the selected regions of interest including control tumor (on the left side), tumor injected with the virus (on the right side), and the heart cavity to obtain blood input function for quantification of the data. Imaging studies with [18F]FHBG were conducted at 2, 6, 24, 48, and 72 h after administration of the virus.

MicroPET imaging of tumor cell replication. Once the flank tumors reached 5 mm, the animals were prepared for imaging as described above. Tumor cell replication prior to and at sequential time points after hrR3 injection was determined by microPET using [18F]FLT (100–150 μCi) as a radiotracer. Data were acquired for 60 min. Regions of interest including tumors on the right (treated) and left (control) sides as well as the liver were drawn and the corresponding time-activity curves were generated. Accumulation of the [18F]FLT was integrated in the regions of interest at a time interval 20 to 40 min and the standard unit values of the accumulation were calculated.

Western blot analysis of HSV-TK expression. Tumor cell lysates containing equal amounts of protein were resolved by PAGE, and proteins were transferred onto polyvinylidine difluoride membrane (Millipore, Billerica, MA). Membranes were incubated overnight at 4°C with rabbit sera raised against HSV-TK (kindly provided by Margaret Black, Washington State University, Pullman, WA) diluted 1:10000. Specific proteins were detected with peroxidase-linked antirabbit immunoglobulin (NA 931, Amersham Life Sciences, Inc., Buckinghamshire, United Kingdom) diluted 1:2000 and an enhanced chemiluminescence system according to the manufacturer's instructions (Amersham Life Sciences). Anti-β-actin antibody (Abcam, Cambridge, MA) was used as the internal control.

Measurement of viral titers in tumors. Excised flank tumors were frozen on dry ice and stored at –80°C. Tumor was weighed, minced, and digested with 1 mg/mL collagenase (Life Technologies) in HBSS (Cambrex, Rockland, ME) for 1 h at 37°C. The samples were subjected to three freeze-thaw cycles before centrifuging at 3,000 rpm for 30 min on a benchtop centrifuge to pellet cell debris. The supernatant was collected into freshly labeled tubes. These supernatants were diluted and assayed for infectious virions on Vero cell monolayers as previously described (27).

Staining of frozen and paraffin-embedded tumor sections. Five-micrometer-thick sections of frozen tumors mounted on superfrrost-coated microscope slides (Fisher Scientific, Pittsburgh, PA) were fixed in 3% glutaraldehyde. β-Galactosidase activity was measured by incubating the slides in 5-bromo-4-chloro-3-indolyl–β-D-galactopyranoside (X-gal) solution at 37°C overnight. X-gal solution was prepared as follows: X-gal at 20 mg/mL in dimethyl formamide, 0.1 mol/L potassium ferricyanide, 0.1 mol/L potassium ferrocyanide, and 0.1 mol/L MgCl₂ (all from Sigma, St. Louis, MO). Sections were coveredslipped before examination under a light microscope.

Formalin-fixed tumors were embedded in paraffin and sequential 5-μm sections were mounted for staining [H&E, HSV-TK, apoptosis, and proliferating cell nuclear antigen (PCNA)]. For immunohistochemical detection of HSV-TK, one serial section was deparaffinized in 2-5 min exposures to xylene; rehydrated in decreasing ethanol concentrations of 100%, 70%, and 50%; and subsequently washed in water and PBS. Antigen was retrieved with proteinase K (Zymed, San Francisco, CA) for 20 min and washed with PBS. Endogenous peroxidase and nonspecific protein binding were sequentially blocked with 3% H₂O₂ for 30 min and 10% bovine serum albumin for 20 min. Avidin and biotin were blocked for 15 min. HSV-TK antisera was applied overnight at 1:1,000. After washing in PBS, biotinylated goat anti-rabbit secondary antibody was applied for 1.5 h, followed by horseradish peroxidase for 30 min (R&D Systems, Minneapolis, MN). Each step was preceded by three 5-min PBS washes. Freshly prepared peroxidase substrate 3,3′-diaminobenzidine was applied onto the sections for 6 min. The slides were rinsed, dehydrated, and coverslipped using Permount. When viewed under a light microscope, HSV-TK–positive cells appeared brown. An adjacent section was deparaffinized in xylene, rehydrated in decreasing ethanol concentrations, and stained with H&E (Sigma).

To stain for PCNA, another adjacent section was deparaffinized in xylene and rehydrated in decreasing ethanol concentrations before antigen retrieval in citrate buffer at 95°C for 5 min. A coplin jar containing citrate buffer was placed in a beaker containing water and microwaved for 55 s to reach the required temperature. The slides were placed in the coplin jar once the temperature had stabilized. Endogenous peroxidase, unspecific protein binding, avidin, and biotin were sequentially blocked as described above. PCNA antibody raised in mice (Chemicon, Temecula, CA) was applied overnight at 1:1,000 and detected with biotinylated antiamine antibody as described above. PCNA-positive cells were counted in five random areas (~1,000 cells) and a PCNA-positive cell index was expressed as the mean percentage of PCNA-positive cells relative to the total cell count.

The ApoTag Fluorescein in situ apoptosis detection kit (Chemicon) was used according to the manufacturer’s instruction to detect apoptosis in situ. Sections were deparaffinized in two 5-min exposures to xylene and rehydrated in decreasing ethanol concentrations of 100%, 70%, and 50% before treatment with proteinase K for 15 min. Positive control sections were treated with DNase for 10 min. Cells were quenched with 3% (v/v) H₂O₂ for 5 min followed by washing with PBS. Terminal deoxynucleotidyl transferase enzyme was added to the pre-equilibrated cells and incubated for 1 h at 37°C. Stop buffer was added to the slide and agitation for 15 s followed by 10-min incubation at room temperature.
After washing thrice with PBS for 1 min each, antidigoxigenin-peroxidase conjugate was added to the slides and incubated for 30 min. The slides were washed twice with PBS and then counterstained with propidium iodide for 15 min. The sections were dehydrated in xylene for 2 min and then mounted with glass coverslip. ApopTag-positive cells were determined under a light microscope with appropriate filters for fluorescence.

**Statistical analysis.** Data, expressed as the mean ± SD, were compared using the unpaired two-tailed $t$ test.

**Results**

Detection of anatomic sites of [$^{18}$F]FHBG sequestration with microPET. MC26 murine colon carcinoma cells transformed to

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**Figure 1.** Determining the expression of HSV1-TK as a PET reporter probe for imaging hrR3 expression in tumors. A, microPET image of HSV-TK–transformed mouse colon cancer tumor (left) and nontransformed mouse colon cancer MC26 tumor (right) on the flanks radiolabeled with [$^{18}$F]FHBG. B, time-activity curves of [$^{18}$F]FHBG accumulation in left tumor (blue curve), right tumor (green curve), heart (red curve), and liver (yellow curve). C, microPET imaging following hrR3 injection correlated with tumor pathology. Row 1, transverse microPET scans of the upper body to include the left and right tumor regions of the mouse at different time points after virus injection. Mice were imaged at 2, 6, 24, 48, and 72 h after virus (hrR3) injection. Row 2, H&E-stained sections of the hrR3-injected (right) tumors. Row 3, X-gal staining for ß-galactosidase expression by hrR3. Row 4, immunohistochemical staining for HSV1-TK expression in right tumors. Row 5, terminal deoxyribonucleotidyl transferase–mediated dUTP nick end labeling (TUNEL) staining for apoptosis in the hrR3-treated (right) tumors. Row 6, immunohistochemical staining for PCNA in right tumors.
express HSV-TK (MC26sr39tk) were implanted into the left flanks of BALB/c mice and untransformed control tumors (MC26) were implanted into the right flanks. MicroPET imaging with $[^{18}F]$FHBG was done once tumors reached 5 mm. As expected, gradual accumulation of $[^{18}F]$FHBG was observed in the HSV-TK–transformed (left flank) tumor over time (Fig. 1A) as a result of intracellular entrapment of HSV-TK–phosphorylated $[^{18}F]$FHBG. Time-dependent washout of $[^{18}F]$FHBG was observed in the right-flank tumor that was comparable to that of heart and liver (Fig. 1B), which relates to blood flow.

Dynamic $[^{18}F]$FHBG PET study following intratumoral injection of an oncolytic HSV-1 mutant. The progression of HSV-1 oncolysis and HSV-TK activity was studied over time with microPET. Bilateral MC26 tumors were established in BALB/c mice flanks, and once the tumors reached 5 mm in size, $1 \times 10^9$ pfu of hrR3 were injected into the right-flank tumor. PET scans were obtained with $[^{18}F]$FHBG freshly prepared for each time point: 2, 6, 24, 48, and 72 h after hrR3 administration. As a result of disruption of the gene encoding the large subunit of the viral ribonucleotide reductase, hrR3 replicated preferentially in mitotically active cancer cells compared with quiescent normal cells (14).

Evidence of HSV-TK expression was observed as early as 2 h after viral injection (Fig. 1C). MicroPET scans revealed an area of intense radiolabel uptake in the treated tumor compared with the contralateral untreated tumor. The radiolabel uptake in the right tumor peaked by 6 h after virus injection to gradually decrease its intensity by 72 h. The accumulation of $[^{18}F]$FHBG in the virus-injected right tumor compared with the untreated left tumor was considerable (Fig. 2A).

The observation that $[^{18}F]$FHBG peaked as early as 6 h following injection of virus into the tumor was unexpected considering the kinetics of viral replication and the process of iterative infection of tumor cells by progeny virion liberated from the first wave of virus-infected cells. We therefore measured hrR3 titers in infected tumors and observed that the viral titer peaked 24 h after hrR3 administration. Peak hrR3 at this time point was 4-fold greater than the titer recovered immediately after intratumoral injection (Fig. 2B). Thus, the discordance between the time to peak $[^{18}F]$FHBG accumulation and time to peak hrR3 titers in treated tumors may be a result of tumor cell lysis taking place following viral oncolysis.

Pathologic evaluation of hrR3-treated tumors. To further investigate the kinetics of intratumoral $[^{18}F]$FHBG accumulation and viral titers, HSV-TK expression following hrR3 injection was assessed by immunohistochemical staining of sections from harvested tumors. These studies showed gradually increasing expression of HSV-TK over 72 h (Fig. 1C). Similar results were observed in an analysis of HSV-TK in tumor by Western blot (Fig. 2C). Similarly, expression of the lacZ transgene seemed to increase over 72 h following hrR3 administration. These data suggest that the decline in $[^{18}F]$FHBG accumulation in tumors starting after 24 h post intratumoral injection of hrR3 is not caused by a decline in HSV-TK expression.

Histologic examination of hrR3-treated flank tumors revealed small regions of tumor destruction at the earliest time point of viral infection (2 h), which continued to increase in size over time.
Tumor destruction following viral oncolysis was characterized by cellular degradation, nuclear polymorphism, and loss of cellular outline together with nuclear fragmentation leading to necrosis. Apoptosis continued to increase over the 72-h duration of the study. Of note, these destroyed cells were unable to retain phosphorylated \( [^{18}F] \)FHBG intracellularly.

**Measurement of tumor cell proliferation during viral oncolysis with hrR3 by microPET imaging with \( [^{18}F] \)FLT.** The effect of viral oncolysis on tumor cell proliferation was studied by microPET imaging of proliferating cells using \( [^{18}F] \)FLT. FLT data obtained in untreated tumors showed considerable radiolabel uptake by both left and right tumors indicative of robust tumor cell proliferation at a rate of 0.012 \( \pm \) 0.004% per hour, corresponding to a tumor doubling time of 58 \( \pm \) 16 h. After virus injection into the right tumor, FLT uptake in the right tumor decreased whereas the FLT uptake continued to increase in the untreated left tumor (Figs. 2D and 3). These measurements were in accordance with results of immunohistochemical staining for PCNA in virus-treated tumors, which revealed a decrease in the percentage of proliferating cells after viral oncolysis. PCNA is a nuclear polypeptide whose synthesis correlates with G1 and S phases of the cell cycle in proliferating cells. The percentage of PCNA-positive cells decreased from 60% at 2 h after virus infection to 37% at 72 h.

**Discussion**

Based largely on promising results of viral oncolysis observed in preclinical models, several replication-conditional viruses are currently under examination in clinical trials (20, 22, 28–30). Notably, the elements of success observed in preclinical models are both efficacy in reduction of tumor burden and preferential infection of tumor cells rather than normal cells. This latter observation in animal models has generally been studied by molecular analyses of potential sites of viral infection; harvested organs from sacrificed animals are analyzed by PCR, immunohistochemistry, or fluorescent microscopy for evidence of transgene expression or viral gene expression using immunohistochemical staining, PCR, and fluorescent microscopy (16, 19, 31–34). An assessment of sites and magnitude of viral replication in vivo is absolutely necessary to correlate vector design as well as dose schedule with specificity of infection, toxicity, and antineoplastic efficacy.

For these same reasons, determining the sites and magnitude of viral replication during viral oncolysis of tumors is important in clinical trials. Clearly, organ sampling for molecular analyses is much less feasible in clinical studies compared with animal models, as these procedures are invasive, cannot be done repetitively in multiple organs, and are subject to sampling errors. It is therefore critically important to develop techniques for noninvasively imaging viral replication.

Several imaging modalities have been tested for imaging cancer gene therapy, including PET, optical imaging, single-photon emission computed tomography, and magnetic resonance imaging (26, 35). PET has many advantages, including high spatial resolution and methods for imaging molecular processes that are quantitative, repeatable, and three-dimensional. As a single imaging modality, PET allows for accurate tumor localization as well as determination of response to treatment. Thus, successful development of PET for imaging of viral replication would theoretically enable PET to simultaneously acquire information on tumor location, sites and magnitude of viral replication, and response to therapy. Importantly, concepts developed in preclinical studies with microPET are easily translated to clinical PET studies. Previously published studies have reported on nuclear imaging of HSV-1 viral oncolysis. For example, HSV-1 has been labeled passively with \( [^{111} \)In-oxine, and viral biodistribution in animals was then assessed with gamma camera imaging (36). One drawback to this approach is that the process of passive labeling with \( [^{111} \)In-oxine labels both infectious and noninfectious viral particles. Moreover, this technique measures only the initial biodistribution of viral particles and is not capable of imaging successive waves of viral infection and replication. In another study, \( [^{124} \)I]FIAU was used for PET imaging of HSV-TK expression following HSV-1 infection of tumors (37). The investigators showed that different HSV-1 mutant viruses produced different levels of \( [^{124} \)I]FIAU accumulation in vivo following infection of cells. \( [^{124} \)I]FIAU label accumulation in vivo correlated with the amount of virus inoculated directly into a tumor, but, importantly, these investigators did not study HSV-1 replication within the same tumor over time (e.g., repetitive imaging for quantification in the same animal). In a separate study, HSV-TK reporter was imaged with PET using \( [^{124} \)I]FIAU following direct inoculation of hrR3 into rat 9L gliosarcoma tumors (38). The rate of FIAU accumulation in hrR3-infected rat gliosarcoma cells was correlated with the levels of HSV-TK gene expression. These studies involved selection of a single time point to carry out PET imaging and did not attempt to study viral replication in a single tumor over time.

Our imaging studies show the ability to detect HSV-TK phosphorylation of \( [^{18}F] \)FHBG as early as 2 h after administration of \( 1 \times 10^9 \) pfu into a tumor. Unlike previous studies, we have imaged the treated tumors sequentially over time. Radiolabel accumulation is seen to peak by 6 h after virus injection. As viral replication gathers momentum and viral oncolysis proceeds, HSV-TK increases in the tumor. However, with increasing cell lysis resulting from viral oncolysis, \( [^{18}F] \)FHBG does not remain sequestered within the cells and PET signal within the tumor declines. While it is possible that the nature of the tumor microenvironment at later time points of viral oncolysis impedes...
[18F]FHBG access to the remaining viable tumor cells, PET studies in HSV-1-treated tumors with [18F]fluorodeoxyglucose argue against this hypothesis (data not shown). On the other hand, [18F]FLT data suggest a decrease in viable tumor cells following virus treatment over time, which is indicative of viral oncolysis. We initiated this study with the prospect of imaging sites and magnitude of viral replication during viral oncolysis of tumors over time. Our data suggest that viral replication in tumors can be imaged by microPET during the early stages of viral oncolysis. However, as viral oncolysis creates tumor destruction, the imaging system fails to correlate with the magnitude of viral replication. An intact cell is required for intracellular sequestration of phosphorylated [18F]FHBG, but cell lysis, which is the hallmark of viral oncolysis, makes imaging the latter stages of viral oncolysis in tumors unachievable. The important interaction between HSV-1 oncolysis and PET imaging of phosphorylated [18F]FHBG that we have described herein does not eliminate PET as a useful modality for imaging viral oncolysis. PET retains its usefulness for identification of sites of initial HSV-1 infection, and a reduction in [18F]FHBG accumulation following an early peak correlates with effective oncolysis. Additional experiments are necessary to determine whether PET imaging of HSV-1 infection is applicable following intravascular administration (rather than intratumoral administration). Clinical trials of viral oncolysis would benefit significantly from noninvasive measurements of these variables.

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