Curcumin Inhibits Lung Cancer Cell Invasion and Metastasis through the Tumor Suppressor HLJ1

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Abstract

Curcumin (diferuloylmethane) is an active component of the spice turmeric and has a diversity of antitumor activities. In this study, we found that curcumin can inhibit cancer cell invasion and metastasis through activation of the tumor suppressor DnaJ-like heat shock protein 40 (HLJ1). Human lung adenocarcinoma cells (CL1-5) treated with curcumin (1–20 μmol/L) showed a concentration-dependent reduction in cell migration, invasion, and metastatic ability, and this was associated with increased HLJ1 expression. Knockdown of HLJ1 expression by siRNA was able to reverse the curcumin-induced anti-invasive and antimetastasis effects in vitro and in vivo. The HLJ1 promoter and enhancer in a luciferase reporter assay revealed that curcumin transcriptionally up-regulates HLJ1 expression through an activator protein (AP-1) site within the HLJ1 enhancer. JunD, one of the AP-1 components, was significantly up-regulated by curcumin (1–20 μmol/L) in a concentration- and time-dependent manner. Knockdown of JunD expression could partially reduce the curcumin-induced HLJ1 activation and diminish the anti-invasive effect of curcumin, indicating that JunD would seem to be involved in curcumin-induced HLJ1 expression. Curcumin was able to induce c-Jun NH2-kinase (JNK) phosphorylation, whereas the JNK inhibitor (SP-600125) could attenuate curcumin-induced JunD and HLJ1 expression. Activation of HLJ1 by curcumin further leads to up-regulation of E-cadherin and a suppression of cancer cell invasion. Our results show that curcumin induces HLJ1, through activation of the JNK/JunD pathway, and inhibits lung cancer cell invasion and metastasis by modulating E-cadherin expression. This is a novel mechanism and supports the application of curcumin in anticancer metastasis therapy. [Cancer Res 2008;68(18):7428–38]

Introduction

Curcumin (diferuloylmethane), an active component of the spice turmeric, is able to suppress cancer cell proliferation, invasion, angiogenesis, and metastasis (1) through a diversity of signaling pathways involving nuclear factor-κB, IκB kinase, Akt, activator protein (AP-1), mitogen-activated protein kinase, cyclooxygenase-2 (COX-2), lipoxygenase, inducible nitric-oxide synthase, urinary plasminogen activator, tumor necrosis factor, chemokines, cell surface adhesion molecules, cyclin D1, and others (2–5). Human clinical trials have indicated no dose-limiting toxicity when curcumin is administered at doses up to 10 g/day and when the serum concentration of curcumin was 1.77 ± 1.87 μmol/L at the dose of 8 g/day (6). These studies suggest that curcumin may have enormous potential in cancer therapy. However, a better understanding of the mechanisms of the drug would enhance the therapeutic potential of curcumin either alone or in combination with other anticancer therapy. This might be possible by synergistically modulating several of the properties of cancer such as proliferation, invasion, and metastasis. To provide a more global perspective, we examined the anticancer mechanisms of curcumin using cDNA microarray mRNA analysis of lung cancer cells and identified a panel of candidate curcumin target genes (7). Some of these target genes are highly associated with the cancer invasion and metastasis (8).

Interestingly, one of the target genes is a heat shock protein family, HLJ1, also known as DNAJB4, which was recently cloned and classified as a member of the HSP40 family (HSP40/DnaJ; ref. 9). HSPs are thought to act as molecular chaperones. They can be powerfully induced by heat shock and other chemical and physical stresses in a wide range of species (10, 11). Recently, it has been found that some HSPs are associated with progression in a variety of human cancers and may modulate cancer cell proliferation, differentiation, invasion, metastasis, and apoptosis (12, 13). Some HSPs have been suggested as able to serve as diagnostic or prognostic biomarkers and may also be useful as new targets for anticancer drug development (14).

The novel candidate target of curcumin, HLJ1, belongs to HSP40 family, which occurs throughout the cell and shows a high diversity in eukaryotic genomes (15). They are at least 44 genes present in the human genome (16). We have previously reported that HLJ1 as able to inhibit lung cancer cell proliferation, anchorage-independent growth, tumorigenesis, cell motility, invasion, and cell cycle progression. High expression of HLJ1 in tumor specimens is associated with reduced cancer recurrence and longer overall survival of non–small cell lung cancer (NSCLC) patients (17). The endogenous control of HLJ1 transcription in cancer cells is regulated by transcriptional factor YY1 and AP-1 (18). This new tumor suppressor and invasion suppressor HLJ1 may be a potential treatment target for anticancer therapy (19).
Several lines of evidence have shown that curcumin is able to inhibit cancer cell invasion and metastasis (3, 7, 20, 21), and our previous microarray data suggested that not only the metalloproteinase (MMP) could be affected by curcumin, but also the tumor suppressor and invasion suppressor HLJ1 may be regulated by curcumin (7). In this study, we identified that the anti-invasive and antimetastasis effect of curcumin is mediated by up-regulation of HLJ1 and curcumin is able to transcriptionally regulated HLJ1 expression through the JNK/JunD pathway and inhibit lung cancer cell invasion and metastasis by modulating E-cadherin expression.

Materials and Methods

Cell Lines and Culture Conditions

The human lung adenocarcinoma cell lines with less invasive (CL1-0) and highly invasive capacities (CL1-5) were established previously (22). The human lung carcinoma cell line, A549 [American Type Culture Collection (ATCC), CCL-185], was obtained from ATCC. The cells were grown in DMEM (Life Technologies) supplemented with 10% fetal bovine serum (FBS; Life Technologies) and each of penicillin and streptomycin (100 mg/mL each) at 37°C in a humidified atmosphere of 5% CO₂ (7).

Migration Assay

Cancer cell migration was assayed by wound-healing assay and transwell migration assay as described previously with some modification (7, 17).

The wound-healing assay. The cancer cells were cultured in 24-well plates and grown in medium containing 10% FBS to nearly confluent cell monolayer, then carefully scratched using a plastic pipette tip to draw a linear “wound” in the cell monolayer of each well. The monolayer was washed twice with PBS to remove debris or the detached cells from the monolayer, and then curcumin was added by different concentrations (1, 5, 10, and 20 μmol/L); the control well was added with 0.1% of DMSO as the solvent control. The cultures were incubated at 37°C and photographed immediately and monitored by time lapse (photographed per 20 min for 12 h) in the Carl Zeiss Axiovert 200M/COOLSNAPfx with Bio-station system (Carl Zeiss). Under the microscope, the number of cells that migrated into the cell-free zone, base on the zero line of the linear “wound,” was evaluated. The experiments were performed thrice in triplicate and were counted double blind by at least two investigators.

The transwell migration assay. The cancer cell transwell migration assay was performed according to our previous study and partially modified (7). Briefly, Transwell membrane (8-μm pore size, 6.5-mm diameter; Corning Costar Corporation) was used. The CL1-5 cells were trypsinized, washed, and kept suspended in medium without FBS. To the lower wells of the transwells contained the same medium with 10% FBS. After 18 h of incubation, the cells on the upper well and the membranes coated with Matrigel were swabbed with a Q-tip, fixed with methanol, and stained with 20% Giemsa solution (Sigma Chemical). The cells that were attached to the lower surface of the polycarbonate filter were counted under a light microscope (magnification, ×200). The experiments were performed thrice in triplicate.

In vivo Animal Studies

We perform the in vivo metastasis assay in mice according to the protocols approved by the National Yang-Ming University Animal Care and Use Committee. The CL1-5 cells were pretreated with scramble or HLJ1 siRNA for 24 h and replate the cell survival and cell number by trypan blue; finally, 5 × 10⁵ live CL1-5 cells in 100 μL PBS were injected into the tail vein of 8-wk-old severe combined immunodeficient (SCID) mice (supplied by the animal center in the College of Medicine, National Taiwan University). To examine the effects of curcumin on the cancer colonization in lung, the mice with scramble or HLJ1 siRNA–treated CL1-5 cells were randomly grouped into curcumin-treated or untreated groups. Curcumin was suspended in the Propylene Glycol (J.T.Baker) for p.o. administration and dosed p.o. once daily at 1 g/kg for 5 wk of treatment. After 5 wk, the mice were sacrificed and analyzed for the lung colonization. The lungs (or other organs) were rinsed with PBS and then placed in a beaker containing Bouin’s solution. After 24 h, the lungs were rinsed in water to remove excess Bouin’s solution and the tumor colonies were counted under a dissecting microscope and double confirmed under a microscope in histology by H&E staining and human vimentin immunostaining.

Real-time Quantitative Reverse Transcription-PCR

The expression level of HLJ1 was detected with real-time PCR on ABI prism 7900 sequence detection system (Applied Biosystems). The HLJ1 primers are as follows: forward primer QHLJ1-F, 5’-CCACGACGACATTTTTATCATC-3’; and reverse primer QHLJ1-R, 5’-CCATCGTGTGTTGTTGATCATATT-3’. TATA-box binding protein (TBP) was used as the internal control (Genbank X54993). The primers and probe used for quantitative reverse transcription-PCR (RT-PCR) of TBP mRNA were as described previously (23, 24). The relative expression level of HLJ1 compared with that of TBP was defined as ΔCT = ΔCTHLJ1 – ΔCTX̄. The HLJ1 mRNA/TBP mRNA ratio was calculated as 2ΔCTHTBP·K, in which K is a constant. Experiments were performed thrice in triplicate.

Western Blot, Immunofluorescent Staining, and Flow Cytometry

Western blot and immunohistochemistry were used to examine the protein expression levels of HLJ1, c-Jun, JunB, JunD, Fra-1, Fra-2, and E-cadherin before and after curcumin treatment, whereas, the flow cytometry was used to confirm the E-cadherin expression in curcin-treated cells. The detailed procedures were as described previously (18). The primary antibodies for c-Jun, JunB, JunD, c-Fos, Fos-B, FRA-1, and FRA-2 were purchased from Santa Cruz Biotechnology, Inc., E-cadherin was purchased from signaling transduction (BD Biosciences), and primary antibody for HLJ1 was made in house (16). Monoclonal mouse anti–α-tubulin (1:500; Calbiochem) was used for loading control. The membranes were then washed thrice with TBST, followed by incubation with horseradish peroxidase–conjugated secondary antibody (1:1,000) in TBST/2% skim milk. Bound antibody was detected using the Enhanced Chemiluminescence System. Chemiluminescence signals were captured using the FujiFilm LAS 3000 system (FujiFilm). All experiments were performed at least thrice in duplicate.

FITC-conjugated secondary antibody (Biotech, Inc.) was used for immunofluorescent staining. Propidium iodide was used as counter staining for cell nuclei. The stained cells were then examined by a confocal laser scanning microscope (C1si; Nikon) at ×400 magnifications. The cells stained with E-cadherin antibody, and FITC-conjugated second antibody were analyzed by flow cytometry (Beckman-Coulter; EPICS-XL). Experiments were performed thrice in triplicate.

Promoter and Enhancer and Luciferase Reporter Gene Constructs

To clone the putative promoter and/or enhancer region of the *HLJ1* gene, a PCR-based method was used and specific primers were designed from the 5'-end of the known *HLJ1* promoter sequence from our previous study (25) and from the bioinformatic search in Genbank. The amplified DNA fragment of 2,302 bp was cloned into the promoterless pGL3-Basic vector (Promega) to construct pGL3-FRER as from the previous study (25).

![Figure 1](image-url)
According to our previous works, varying lengths of the 5'-flanking region of the *HLJ1* gene for luciferase assays were generated (17, 25). The pGL3-Control, a positive control plasmid, was also obtained from Promega. The putative enhancer (-2,125 to -1,039) element and its various deletion mutants were generated by PCR and subcloned into pGL3-promoter vector as our previous study described (25). This vector contains the luciferase gene driven by SV40 promoter. The same cloning strategy was used to generate the minimal enhancer construct pGL3-p-Emi. For site-directed mutagenesis assay, the pGL3-p-Emi construct served as a template for the generation of mutations in the SPI1 and AP-1 binding sites. All mutant constructs were prepared by PCR using appropriate primers containing the mutations. The composition of all of the constructs was confirmed by restriction endonuclease digestion and DNA sequencing.

**Figure 2.** Curcumin (*cur*) induces *HLJ1* expression and inhibits cancer cell migration and invasion. A, real-time quantitative RT-PCR reveals that *HLJ1* mRNA expression was induced by curcumin in a concentration-dependent manner (1–20 μmol/L) after 24 h incubation. B, concentration-dependent and time course–dependent curcumin-induced *HLJ1* expression at the protein level were confirmed by Western blotting analysis. The results indicated that the protein level of *HLJ1* was significantly increased after curcumin (10 μmol/L) treatment for 12 to 24 h. The expression levels were qualified by densitometer and calculated relative to the expression level of α-tubulin (as the internal control) and are shown as the mean expression levels from three independent experiments; columns, mean (n = 3); bars, SD. *, P < 0.05, statistically significantly compared with vehicle-treated control. C, Western blotting showed the *HLJ1* expression levels of *HLJ1* siRNA or scrambled siRNA–transfected (control) CL1-5 cells with or without curcumin treatment. Data are representative of at least three independent experiments with α-tubulin used as the internal control. *HLJ1* protein expression was reduced by *HLJ1* siRNA both in the base level and curcumin-treated cells (lane 3 and 4), whereas there was no effect within the scrambled siRNA control group (lane 5 and 6). Bottom, the *HLJ1* knockdown specifically reduced the anti-invasive activity of curcumin. Cells transfected with control or *HLJ1* siRNA were treated with or without curcumin and then tested in invasion assays across a Transwell membrane coated with Matrigel. Columns, mean derived from three separate experiments done in quadruplicate; bars, SD. **, P < 0.05, statistically significant compared with vehicle-treated control; *, P < 0.05, statistically significant compared with the scramble control.
Transfection and Luciferase Reporter Assays

All transfections were performed in triplicate in 6-well plates, and the detailed protocol was described in previous studies (18, 25). In brief, CL1-5 cells (2 × 10^5 cells per well) were seeded for 24 h prior to transfection. Plasmids were transfected into cells using Lipofectamine 2000 reagent according to the manufacturer’s instructions (Invitrogen). The HLJ1 promoter and enhancer luciferase reporter constructs (25), along with the control plasmid, was cotransfected with a β-galactosidase construct, pSV-β-gal (Promega). The ratio of the DNA amounts for luciferase reporter constructs versus β-galactosidase construct was 3:1. The cells were incubated in transfection mixture for 4 h and then harvested after 44 h in culture. An aliquot of cell lysate (10–25 μL) was used to assay luciferase activity using a luciferase assay kit (Tropix, Inc.). Another aliquot (10–25 μL) was used to measure β-galactosidase activity using the Galacto-Light chemiluminescent assay kit (Tropix, Inc.). Luminescence was measured with a Victor2 1420 Multilabel Counter (Wallac). Transfection efficiency was normalized with β-gal activity. Each experiment was repeated at least thrice.

Actin Staining

Cells were washed twice with PBS and fixed in a 3.7% formaldehyde-PBS solution for 10 min at room temperature. After two additional washes with PBS, cells were permeabilized with a solution of 0.1% Triton X-100 in PBS for 3 to 5 min and washed again with PBS. Phallolidin-tetramethylrhodamine isothiocyanate (0.2 μg/mL) and Alexa Fluor 488 DNsA1 conjugate (9 μg/mL) were used to localize filamentous actin (F-actin) and G-actin, respectively, as described by Cramer and colleagues (26). Fluorescent dyes were diluted on blocking solution (1% bovine serum albumin and 0.025% saponin in PBS) and added to coverslips for 40 min at room temperature. After washing with PBS, coverslips were mounted on a microscopy slide with mounting media (Vector Laboratories). Time of image acquisition and image intensity gain were optimally adjusted and kept constant for all experiments. F-actin to G-actin fluorescence ratio was quantified using fields containing >30 cells imaged with a inverted fluorescence microscope (Eclipse TE2000; Nikon) and a 12-bit resolution cooled charge-coupled device camera (Orca; Hamamatsu Photonics) at ×10 magnification as previously report (27). F-actin cytoskeleton imaging was performed with a confocal laser scanning microscope (C1si; Nikon) at ×400 magnifications.

Statistical Analysis

All experiments were performed in triplicate and analyzed by ANOVA (Excel; Microsoft) for significant differences. *P* values of <0.05 were considered statistically significant. Where appropriate, the data are presented as the mean ± SD.

Results

Curcumin inhibits cancer cell migration and invasion. CL1-5 is a highly invasive cancer cell line both *in vitro* and *in vivo* (22). Due to the fact that curcumin could induce cell apoptosis and reduce cell viability as the concentrations of curcumin over 20 μmol/L (the IC50 is 33.2 ± 3.7 μmol/L in CL1-5 cells; ref. 7), the effects of curcumin on migration and invasion abilities of CL1-5 cells were performed at the concentration range below 20 μmol/L and determined by the wound-healing assay, the Transwell migration model, and the Matrigel-based Transwell invasion assay. Concentration dependently, curcumin (5–20 μmol/L) was able to significantly inhibit CL1-5 cancer cell migration, and this effect was consistent across both the wound-healing and transwell migration models (Fig. 1A and B). In addition, the Matrigel-based transwell assay indicated that curcumin was able to inhibit cell invasion (Fig. 1C). Similar results whereby curcumin inhibited cell migration and invasion were confirmed for another invasive lung cancer cell line, A549 (data not shown), indicating that this observation would seem to be neither cell specific nor a coincidence.

Suppression of cancer cell invasion and metastasis by curcumin is associated with up-regulation of HLJ1. To elucidate the anti-invasion and antimetastasis mechanisms of curcumin, curcumin-regulated differentially expressed genes in CL1-5 cells were profiled in our previous study (7), and some of these genes are highly correlated with the invasive ability of cancer cells (8). Herein, one curcumin-regulated tumor suppressor and invasion suppressor gene *HLJ1* (17) is examined. As shown in Fig. 2, we confirmed that curcumin was able to increase both the mRNA and protein expression levels of *HLJ1* in CL1-5 cells in a concentration- and time-dependent manner (Fig. 2A–B).

To evaluate the importance of *HLJ1* in the anti-invasive effect of curcumin in CL1-5 cells, a specific siRNA for *HLJ1* was used to knockdown *HLJ1* expression in curcumin-treated cells. We found that *HLJ1* expression decreased after 24 hours of 20 nmol/L *HLJ1* siRNA treatment; furthermore, the effect of curcumin-induced *HLJ1* expression was also reduced by 67% ± 7.2% compared with curcumin-treated group by *HLJ1* siRNA (*P < 0.05; Fig. 2C). In the same experiment, the anti-invasive effect of curcumin was reversed by pretreated with *HLJ1* siRNA (Fig. 2C). These results showed that 51.4% ± 8.7% of cancer cell invasion could be inhibited by curcumin and that this effect was reduced to 20.1% ± 12.6% if cells were preincubated with *HLJ1* siRNA before curcumin treatment. On the other hand, the scrambled siRNA control showed no difference compared with the control group.

To examine the role of *HLJ1* in the antimetastatic effects of curcumin *in vivo*, the i.v. experimental lung metastasis model in SCID mice was used. CL1-5 cells transfected with scrambled siRNA (mock) or *HLJ1* siRNA were counted (5 × 10^5 cells) and injected into tail vein of the SCID mice. These mice were then treated with or without curcumin (p.o., 1 g/kg/day). After 5 weeks, the numbers of colonized pulmonary tumor nodules were counted. Most of the pulmonary tumor colonies in the mice were found near or on the surface of the lungs (Fig. 2D). The data showed that injection with mock cancer cells could result 21.80 ± 13.84 pulmonary colonized tumor nodule formation per mice. The curcumin-treated group showed a significantly reduced number of the colonized tumor nodules in the lungs compared with the mock control group (3.89 ± 2.28 nodules per mice; *P < 0.05). The knockdown *HLJ1* was able to attenuate the antimetastatic effects of curcumin *in vivo* as shown by the fact that the *HLJ1* siRNA–transfected plus curcumin-treated group showed many nodules (11.78 nodules per mice) compared with the scrambled siRNA plus curcumin-treated group (3.89 nodules per mice). These results indicated that *HLJ1* would seem to play a role in the antimetastasis effects of curcumin.

The AP-1 site within the *HLJ1* enhancer and the JNK/JunD signaling pathway are responsible for HLJ1 up-regulation by curcumin. We have previously identified that the YY1 and AP-1 binding sites in the *HLJ1* promoter and enhancer regions play a major role in *HLJ1* transcriptional regulation (18, 25). To study whether curcumin could regulate *HLJ1* expression through the promoter and/or enhanced regions of *HLJ1*, several constructs with deletions or mutations were used and have been previously described in detail (25). The full length of the *HLJ1* promoter with enhancer construct (PGL3-FRER) was used to show that curcumin could concentration-dependently induce luciferase activity (Fig. 3A), whereas curcumin has less effect on the promoter only construct (PGL3-F2RER). Interestingly, curcumin was able to significantly increase the luciferase activity of the minimum enhancer region (Emi-F/R; data not shown), which contains the AP-1 and SP-1 binding sites. Site-directed mutagenesis indicated that the AP-1 site (−1,457 to −1,451) and not the SP-1 site was essential for the curcumin-regulated *HLJ1* enhancer region activity (Fig. 3B).
are several protein components that make up the AP-1 complex, including c-Jun, c-Fos, JunD, JunB, Fra-1, and Fra-2. On studying the effects of curcumin on the AP-1 subunit family, we found that only JunD was able to be significantly induced by curcumin in CL1-5 cells (Fig. 3C). In contrast to c-jun, which was shown to be slightly down-regulated, JunB shows an ascending trend, whereas Fra-1, Fra-2, and c-Fos are not affected significantly by curcumin over the concentration range used (1–10 μmol/L; Fig. 3C). Furthermore, time course studies showed that the dynamic effects of curcumin during the induction of JunD expression occurred after 2 hours of incubation at the mRNA level and after 4 hours at protein level. This is just a little earlier than the induction of HLJ1 expression in curcumin-treated cells (Fig. 3D).

To confirm the role of JunD in curcumin-induced HLJ1 expression, JunD siRNA was applied to curcumin-treated CL1-5 cells to knock down JunD expression. The Fig. 4A showed that...
pretreatment with JunD siRNA was able to reduce curcumin-induced HLJ1 mRNA expression (Fig. 4A). This effect was confirmed at the protein levels for JunD and HLJ1 by Western blot analysis as shown in Fig. 4B. In the same preparation, the anti-invasive effect of curcumin was reduced by pretreatment with JunD siRNA (Fig. 4B).

To further investigate the regulation of curcumin on the JunD upstream modulator, c-Jun-NH2-kinase (JNK) and phosphorylation of JNK were examined (Fig. 4C). The data showed that two isoforms of JNK, JNK1, and JNK2 were both expressed in CL1-5 cells and were not affected by curcumin treatment. The phospho-form of JNK1 and JNK2, p-JNK1 and p-JNK2, were significantly induced by curcumin with dynamic changes (Fig. 4C). This contrasts with the effect of the JNK inhibitor (SP600125), which was able to reduce the effects of curcumin-induced JunD and HLJ1 expression in CL1-5 cells (Fig. 4D). These results showed that JNK phosphorylation and JunD would seem to be involved in the curcumin-induced HLJ1 signaling pathway in CL1-5 cells.

Curcumin up-regulates HLJ1 and E-cadherin expression, suppresses filopodia formation, and inhibits cancer cell invasion. The question remains as to how curcumin-induced HLJ1 expression is able to affect cancer cell invasion. Our previous study suggested that HLJ1 was able to inhibit cell invasion through...
the induction of E-cadherin expression (18), and other evidence has suggested that these DnaJ-like proteins might interact with actin, tubulin, or various intermediate filaments to restrict cellular activity (28). Immunostaining of curcumin-treated CL1-5 cells showed that there was up-regulation and localization of E-cadherin into the cell-cell junctions together with a mesenchymal-epithelial phenotype transition (Fig. 5A). Western blot and flow cytometry analysis confirmed that the E-cadherin protein levels are up-regulated by curcumin in a concentration-dependent manner (5–20 μmol/L; Fig. 5B and C). HLJ1 siRNA was able to specifically knockdown HLJ1 protein levels and this significantly diminished curcumin-induced E-cadherin expression (P < 0.05; Fig. 5D).

Because HLJ1, such as other DnaJ family members, might modulate cell invasion through regulating the kinetics of actin, tubulin, or various other intermediate filaments (28), we examined the effects of curcumin on actin polymerization. As shown in Fig. 6A, curcumin did not significantly affect the total amount of actin. However, the F-actin (polymer)/G-actin (monomer) ratio was significantly decreased by curcumin treatment in a concentration-dependent manner (Fig. 6B). Figure 6C shows that curcumin also decreased cell membrane filopodia protrusions and induced an abnormal aggregation of actins in cytosol of the curcumin-treated cells (Fig. 6C).

Discussion
Curcumin has long been known to act as a therapeutic or preventive agent for several major human cancers by regulating a variety of antitumor signaling pathways (29, 30). In this study, we have confirmed that curcumin can inhibit the invasion and metastasis of lung cancer cells in vitro and in vivo. We found that curcumin can suppress cancer cell invasion and metastasis through the up-regulation of the tumor suppressor HLJ1. Curcumin is able to transcriptionally activate HLJ1 expression through the JNK/JunD pathway. Activation of HLJ1 by curcumin further leads to up-regulation of E-cadherin, and this inhibits cancer cell invasion and metastasis. Our results strengthen the potential of curcumin as a multitarget drug in anticancer therapy (1, 31, 32).

The emerging concept of integrating gene expression profiles after small molecule treatment of specific diseases has been suggested to be an effective strategy for the identification the new drug development targets (33). We have previously generated two cDNA microarray data sets: namely the cancer invasion/metastasis associated gene data set obtained from cell lines with different invasive abilities (CL1-0 to CL1-5; ref. 8) and the curcumin-regulated gene expression signature data set (7). By combining and sorting the curcumin-regulated genes and the invasion/metastasis-associated genes from these two data sets, the tumor suppressor HLJ1 was one of the selected candidate targets for control of metastasis and invasion. This was further confirmed in this study, which showed that HLJ1 is a potential new treatment target for the control cancer cell invasion by curcumin. These results are consistent with our previous study and support the hypothesis that the tumor suppressor HLJ1 is an important target for anti-invasion, antimetastasis, and antiangiogenesis in cancer therapy (17, 19). Our findings help to shed some light on the development of new drugs such as curcumin analogues that may inhibit cancer invasion and metastasis by targeting HLJ1.

A previous study showed HLJ1 to be regulated by AP-1 through the enhancer region of HLJ1 (18, 25). The AP-1 transcription factor complex, composed mainly of Jun-Jun and Jun-Fos protein dimer,
is involved in various biological processes, including normal cell growth and cancer development (34). Curcumin has been suggested to inhibit tumor growth through an inhibition of AP-1 activation (35). However, in this study, we found that curcumin significantly and concentration-dependently induced JunD expression and it could positively enhance the promoter activity of \textit{HLJ1} via AP-1 binding site. JunD is a versatile AP-1 component and is diverse from other AP-1 components in transcription regulation and cellular function (36). Previous evidence has shown that AP-1 complexes enriched with c-Jun and JunB may result in morphologic alterations and anchorage-independent cell growth, whereas complexes enriched with JunD had an antiproliferative effect (37). The induction of JunD by curcumin has also been reported in a previous study, which showed that curcumin exposure could up-regulate JunD expression and may lead to an increase in the enzymes involved in glutathione synthesis in HBE1 cells (38), the other JunD

Figure 6. The effects of curcumin on the F-actin/G-actin ratio and filopodia formation. A, Western blotting analysis shows that total actin protein expression was not affected by curcumin (5–10 μmol/L). B, Western blotting analysis shows the fractions of F-actin and G-actin in CL1-5 cells treated with or without curcumin. The ratio of F-actin/G-actin is shown below (n = 3). Cells treated with 0.1% DMSO were used as the solvent control. Columns, mean (n = 3); bars, SD. *, P < 0.05, statistically significant compared with the vehicle-treated control. These results are representative of three independent experiments. C, CL1-5 cells treated with or without curcumin were stained with phalloidin-tetramethylrhodamine isothiocyanate and Alexa Fluor 488 DNase I conjugate to detect F-actin (red) and G-actin (green). F-actin labeling with phalloidin-tetramethylrhodamine isothiocyanate reveals that CL1-5 cells exhibit numerous filopodia, whereas the curcumin-treated cells exhibit fewer filopodia fibers. D, the hypothetical mechanism by which curcumin causes HLJ1 induction and related signaling events during the drug’s anti-invasive effects.
downstream gene, ferritin H mRNA expression, can be transcriptionally regulated by curcumin (36, 39). JunB and JunD are able to bind to the putative AP-1 site in the HLJ1 enhancer and the expression of HLJ1 is positively correlated with JunB and JunD expression in a dose-dependent manner (25). Therefore, we suggest that HLJ1 may be regulated by curcumin through JunD induction and that this curcumin-induced JunD expression may have the tumor suppressor–like effects. This hypothesis is further supported by other recent studies, which have shown that specific AP-1 proteins, such as Fra-1, c-Fos, JunB and JunD, have tumor suppressor activity and that these effects depend on the antagonistic activities of the different Jun proteins, the tumor type, the tumor stage, and the genetic background (25, 34, 40). Previous evidences showed that the regulation of JunD expression diverges from the well-characterized growth factor-inducible pattern of the c-jun early response genes and AP-1 autoregulation (36). Herein, we report that curcumin might regulate HLJ1 through modulating the JunD composition of the AP-1 transcription factor complex. Indeed, more studies are needed to clarify the AP-1 dimerization pattern and the activation of specific tumor suppressors in different cells and tissues.

E-cadherin, as the key component for adherence junctions between epithelial cells, has been investigated for its roles in tumorigenesis and as an invasion suppressor molecule in carcinoma cells (41). E-cadherin has also been suggested to be involved in epithelial-mesenchymal transition, a process associated with loss of cell-cell adhesion and epithelial polarity during cancer development and progression (42). Tumor cells with limited E-cadherin expression are more likely to detach from a tumor mass, resulting in a poor prognosis. This has been suggested to be an independent prognostic factor for NSCLC survival (41, 43). Our previous study indicated that HLJ1 might regulate cell invasion through E-cadherin (18), and herein, we have confirmed that curcumin is able to regulate E-cadherin expression in the previous finding (44); furthermore, we have also identified that this effect would seem to be at least partly mediated through HLJ1 and results in an inhibition of cancer cell invasion.

In this study, we found that the curcumin-treated cells showed a higher level of HLJ1 expression with a decreased F/G actin ratios and an aggregation of actins. Although, some HSPs have been suggested to be involved in cytokinesis arrangement and play important roles in cell growth and motility regulations. For example, the major components of the intermediate filaments of simple epithelia, keratins 8/18, have been reported to directly bind with Mrj, a DnaJ/Hsp40 family protein (28). Our previous study found that the Hsp70-like protein, HLJ1, Hsp27, and Hsp70 were significantly increased by curcumin (7). These results suggest that the HSPs may play a role and can explain some pharmacologic effects of curcumin. Although we did find the actin polymerization was affected by curcumin, whether this event was HLJ1 dependent or independent still need to be evaluated.

The key finding of this study is that it provides evidence supporting the idea that curcumin is able to up-regulate the expression of the tumor suppressor HLJ1 by transcriptionally activating its enhancer. This indicates that the anti-invasive and antitumorigenesis effects of curcumin are likely to be mediated through the induction of HLJ1, at least in part. Figure 6D summarizes our hypothetical signaling pathways for curcumin, its regulation of HLJ1 expression, and its inhibition of cancer cell invasion. Furthermore, curcumin is able to increase JunD expression, stimulate HLJ1 enhancer activity, and trigger the expression of HLJ1, which would seem to subsequently reduce filopodia formation and up-regulate E-cadherin expression. These effects ought to restrain cancer cells from invasion and metastasis.

Based on the molecular heterogeneity and complexity of the signaling pathways involved in cancer development, multiple targets strategy may be a winning strategy for anticancer treatment (19, 29). According to this study and our previous findings, curcumin could inhibit cancer cell migration and invasion, not only through the HLJ1/E-cadherin induction, but also involve the inhibition of MMP14 mRNA and protein expression, as well as the enzyme activities of MMP2 and MMP9 (7). These evidences strengthen the possibility for that curcumin may have the potential as a multitarget drug in anticancer therapy because this drug is able to target diverse mechanisms involved in the cell cycle, apoptosis, angiogenesis, invasion, and metastasis (1, 29–32). This study further suggests that curcumin may be a template for new antitumor drug developments that are able to target the tumor suppressor HLJ1.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

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