Metastamir: The Field of Metastasis-Regulatory microRNA Is Spreading

Douglas R. Hurst,1,5 Mick D. Edmonds,1,5 and Danny R. Welch1,2,3,4,5

Departments of 1Pathology, 2Cell Biology, 3Pharmacology/Toxicology, Comprehensive Cancer Center, and 4National Foundation for Cancer Research - Center for Metastasis Research, University of Alabama at Birmingham, Birmingham, Alabama

Abstract

Despite advancements in knowledge from more than a century of metastasis research, the genetic programs and molecular mechanisms required for cancer metastasis are still incompletely understood. Genes that specifically regulate the process of metastasis are useful tools to elucidate molecular mechanisms and may become markers and/or targets for antimetastatic therapy. Recently, several noncoding regulatory RNA genes, microRNA (miRNA), were identified, which play roles in various steps of metastasis, some with obvious roles in tumorigenesis. Understanding how these metastasis-associated miRNA, which we term metastamir, are involved in metastasis will help identify possible biomarkers or targets for the most lethal attribute of cancer: metastasis. [Cancer Res 2009;69(19):7495–8]

Introduction

Cancer is currently the second leading cause of death in the United States and, if current trends continue, will soon be the leading killer. The major reasons for cancer deaths are complications arising from metastasis. Therefore, improved morbidity and mortality will require effective treatments targeting metastatic disease. Despite many advancements in knowledge from more than a century of researching metastasis, the molecular mechanisms are still not completely understood. Several reports have described the involvement of a recently discovered class of noncoding regulatory RNA, termed miRNA, in the regulation of cancer (oncomir; refs. 1, 2). More recently, a specialized family of miRNA, which we call metastamir, have been shown to have pro- and antimetastatic effects. miRNA were originally discovered because of their roles in controlling the timing of Caenorhabditis elegans larval development. Less than a decade later they were identified in plant and mammalian cells (see ref. 3 for review). Typically, pri-miRNA are transcribed by RNA polymerase II before capping, polyadenylation, and maturation of a hairpin loop structure by a type III ribonuclease (Drosha) into pre-miRNA. Following export into the cytoplasm, the pre-miRNA become associated with several ribonucleoproteins of the RNA-induced silencing complex (RISC), including Dicer and Argonaut family members from which a mature miRNA are formed. miRNA complement the 3′-UTR of mRNA in order to impair translation or alter message stability. Because of their small size, miRNA are predicted to be promiscuous and may have several hundred mRNA targets, meaning that a single miRNA can, by itself, impact the expression of hundreds of proteins (see ref. 3 for review).

Requests for reprints: Danny R. Welch, Department of Pathology, 1670 University Boulevard, room VH-G019, Birmingham, AL 35294-0019. Phone: 205-934-2961; Fax: 205-975-1126; E-mail: DanWelch@uab.edu.

Copyright © 2009 American Association for Cancer Research. DOI: 10.1158/0008-5472.CAN-09-2111

Published OnlineFirst September 22, 2009; DOI: 10.1158/0008-5472.CAN-09-2111

Metastasis involves multiple steps and multiple genes in which neoplastic cells dissociate from the primary tumor, enter body cavities, or, more commonly, circulatory systems (lymphatics or blood vasculature), survive during transport until they arrest at discontinuous sites, exit the circulation, and proliferate at ectopic sites (colonization) in response to local growth factors (4). The process is extremely inefficient (of the -4 million cells entering the vascular compartment per gram of tumor per day, much less than 0.01% develop macroscopic masses elsewhere; ref. 4). The inefficiency is perhaps because every step in the metastatic cascade is selective and rate-limiting (i.e., failure to complete any step precludes subsequent steps). Each step in metastasis requires coordinated temporal expression of genes and spatio-temporal expression of proteins.

Examination of mRNA expression patterns has yielded sometimes conflicting results related to roles in metastasis, prompting some to question even the existence of metastasis-regulatory genes. Yet, multiple laboratories, using several different human and rodent model systems, showed the existence of gene products that affect metastasis without promoting or inhibiting tumorigenicity at orthotopic sites (5). So, although tumor formation is prerequisite to metastasis, tumorigenicity and metastasis are distinct phenotypes, the latter requiring genetic changes superimposed upon those needed to make the tumor. These considerations led us and others to predict the existence of metastamir.

The invitation to write this mini-review was prompted by our discovery that the miR-146 family of miRNA could profoundly inhibit invasion and metastasis of MDA-MB-231 human breast carcinoma cells. In that report, we further showed that miR-146a/b was downstream of the BRMS1 metastasis suppressor and intermediate to BRMS1-regulated genes (6). Concurrently, we have shown that BRMS1 coordinately regulates entire families of metastamir, up-regulating metastasis-suppressing miRNA and down-regulating metastasis-promoting miRNA (7).

Those findings, coupled with an explosion of articles describing miRNA and metastasis-associated steps compelled us to expand the focus of this mini-review to consider the state of the field. To date, 11 miRNA have been shown to promote or inhibit metastasis in experimental models (Fig. 1), and the number is likely to grow even further because more than 20 additional miRNA have been shown to impact critical steps in the metastatic cascade, such as epithelial-mesenchymal transition (EMT), apoptosis, and angiogenesis (Fig. 1). Furthermore, several clinical studies have identified correlations between miRNA expression and recurrence, development of metastases and/or survival (for a recent review, see ref. 8). Therefore, our goal is to focus on the evidence for metastamir, the implications of their existence, and some technical and theoretical considerations that emerge from their discovery.

Discovery of Metastamir

In retrospect, it was self-fulfilling that miRNA-regulating metastasis would be found because the process itself involves hundreds
of genes. To date, metastamir have typically been discovered using in vitro screens for steps in the metastatic cascade including cell growth, EMT, adhesion, migration, invasion, apoptosis, and/or angiogenesis. Most commonly, metastamir promoting cell migration and invasion have been described. Figure 1 shows a relatively current listing of metastamir impacting the cascade, but highlights (in red) those for which actual functional data have been collected for metastasis in vivo. The latter point is critical because it is not possible to study metastasis in vitro. Yet, at a conceptual level, it should be possible to design in vivo screens using miRNA or antagonomir (miRNA antagonists) libraries to discover metastasis-promoting or metastasis-suppressing metastamir. Antagonomir studies will depend upon yet-to-be-perfected technology to stably knock-down miRNA expression. Screening for metastamir in vivo would be cost-prohibitive in most laboratories; so, in vitro surrogates make economical sense.

Another interesting point is that the vast majority of metastamir have been identified in breast and/or mammary tumor cell lines. The reasons for this preponderance may be associated with funding levels or availability of robust metastasis models. Nonetheless, the number of metastamir identified in other tumor types is likely to expand in the near future. Until then, it is important not to summarily extrapolate function in breast tumors to all histotypes.

Metastasis-Suppressing Metastamir

**miR-335 and -206.** The first suppressing metastamir was identified in Joan Massague’s laboratory by Tavazoie and colleagues, who compared miRNA expression in metastatic variants derived from the human breast carcinoma cell line MDA-MB-231 (9). They identified six miRNA with a low relative expression in the metastatic cells. Three of these, miR-335, -126, and -206, suppressed metastasis in vivo, however, miR-126 also inhibited cell proliferation and tumorigenesis. Therefore, we did not include miR-126 in our list of metastasis suppressors. Both miR-335 and -206 inhibited invasion and migration in vitro. miR-335 targets SRY-box containing transcription factor (SOX4), receptor type tyrosine protein phosphatase (PTPRN2), c-Mer tyrosine kinase (MERTK), and possibly tenascin C (TNC). Additionally, inhibition of SOX4 or TNC by shRNA inhibited invasion in vitro and metastasis in vivo. Their findings elegantly show how a single miRNA could impact several downstream pathways by arborizing signaling pathway components. There was also a clinical association of miR-335 expression with metastasis-free survival in a set of 20 primary breast tumor samples.

**miR-146a/b.** Several groups have shown a role for miR-146 in inflammation through regulation of nuclear factor-κB (NFκB; ref. 10). Although miR-146a and b are encoded on different

---

**Figure 1.** Critical steps in metastasis altered by metastamir. Pro- and antimetastatic metastamir are listed with the steps in the metastatic cascade of which they affect. The metastamir that have been functionally tested for metastasis in vivo are highlighted in red.
chromosomes, their mature sequence differs by only two nucleotides at the 3’ region. So their mRNA targets are predicted to overlap significantly. Indeed, both miR-146a and b inhibit invasion and migration of breast cancer cells by down-regulating NF-κB by targeting IRAK1 and TRAF6 (11). These studies were extended in vivo by showing miR-146a and b suppressed metastasis that may involve targeting of EGF receptor (6) or ROCK1 (12), both of which are involved in promoting invasion and metastasis. miR-146a expression is inversely correlated with prostate cancer progression (12).

miR-31. Inhibition of any single step in metastasis results in metastasis suppression. Inhibition of multiple steps would therefore result in more robust inhibition of the metastatic process. miR-31 inhibits multiple steps of metastasis including invasion, anoikis, and colonization leading to a 95% reduction in lung metastasis in an orthotopic model of breast cancer (13). Clinically, miR-31 levels were lower in breast cancer patients with metastasis (n = 56 patients).

Metastasis-Promoting Metastamir

miR-10b. Ma, Weinberg, and colleagues were the first to discover a metastasimir (14). They hypothesized that certain miRNA could regulate specific stages of tumor progression and found that miR-10b was highly expressed only in metastatic breast cancer cell lines compared with primary human mammary epithelial or spontaneously immortalized cells. After showing that miR-10b enhanced migration and invasion in vitro and metastasis in vivo, they identified a pathway in which the prometastatic gene TWIST1 up-regulates miR-10b, which targets HOXD10 leading to an increase in RHOC. Additionally, RTQ with 23 primary breast tumors was used to show a general increase in miR-10b expression in patients with metastasis.

Interestingly, the BRMS1 metastasis suppressor that regulates miR-146a/b also regulates TWIST, miR-10b, and RHOC expression (7). Whether the regulation of these genes by BRMS1 is direct or indirect is still not known. Regardless, the data all point to common pathways impacted by these metastasis regulatory molecules. Perhaps hopefully, with additional experimentation, we will be able to find a point of convergence for several signaling cascades important in metastasis.

miR-373 and -520c. Huang, Agami, and colleagues transduced nonmetastatic MCF7 human breast cancer cells with an miRNA expression library and screened the transductants using a transwll migration assay (15). Both miR-373 and -520c promoted migration and were subsequently found to increase in vivo metastasis at least in part by targeting the adhesion molecule CD44. Clinically, miR-373 expression was higher in lymphnode metastasis compared with the primary tumors from 11 pairs of matched samples.

miR-21, -143, and -182. Invasion and migration are increased whereas apoptosis is decreased by miR-21 expression in multiple model systems (breast cancer, colon cancer, and glioma; refs. 16–18). miR-21 was found to target TPM1 (tropomyosin 1), programmed cell death 4 (PDCD4), and regulators of matrix metalloproteinases. miR-143 and miR-182 promoted hepatocellular carcinoma and melanoma, respectively (19, 20). miR-143 is up-regulated by NF-κB and decreases adhesion. miR-182’s effects can be reversed by re-expression of microphthalmia-associated transcription factor M (MITF) or FOXO3. miRNA-182 is part of a cluster (miR-183-96-182). Many miRNA are encoded as genetically linked clusters and are expressed as a single pri-miRNA. As a result it is not always possible to distinguish biological effects that are the result of a single miRNA or the collective actions of multiple miRNA.

Metastamir Pathways, Concepts, and Future Directions

Although metastamir have only been recognized for slightly more than 2 years, the explosive rate of discovery of this important family of molecules is impressive. Metastamir are components of complex pathways and are often expressed downstream of pro- or antimetastatic signals, including pathways regulated by NF-κB, EGF, TWIST, BRMS1, ZEB1/2, and HIF1α (8). Unfortunately, the mechanisms by which miRNA are regulated still remain relatively ill-defined (21), and will be the subject of intense future investigation.

Interestingly, positive and negative feedback loops have been found whereby the upstream effectors are themselves targets of the miRNA that they regulate. This implies an important role for metastamir in modulating key signaling pathways involved in metastasis. Because of their position as nodes within these pathways and their promiscuity with regard to downstream targets, each metastamir can (and probably does) multiply pro- and antimetastatic signaling events. It is likely that metastamir regulation of these signaling events are context dependent, relying on micro-environmental cues in both directions. We predict that yet-to-be-discovered cofactors will lead to specificity of miRNA effects on selected pathways; however, their existence is speculation at this time.

We find ourselves in the midst of a revolution with regard to the biochemical and molecular regulation of cancer metastasis. Old notions of identity equating tumorigenicity with metastasis have to be discarded. There are clear distinctions between the phenotypes; biologically, biochemically, and genetically. Understanding the interrelationships between regulatory genes and gene products (proteins and noncoding RNA) and how these are modulated by the micro-environmental context is beginning to unravel the complex tapestry that is cancer metastasis.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

Received 6/9/09; revised 7/9/09; accepted 7/13/09; published OnlineFirst 9/22/09.

Grant support: U.S. Public Health Service (USPHS) grants CA87728 (D.R. Welch) and F32CA113037 (D.R. Hurst); predoctoral fellowship from the U.S. Army Medical Research and Materiel Command W81XWH-08-1-0786 (M.D. Edmonds); and National Foundation for Cancer Research, Center for Metastasis Research (D.R. Welch).

References

Metastamir: The Field of Metastasis-Regulatory microRNA Is Spreading

Douglas R. Hurst, Mick D. Edmonds and Danny R. Welch


Updated version
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-09-2111

Cited articles
This article cites 21 articles, 4 of which you can access for free at:
http://cancerres.aacrjournals.org/content/69/19/7495.full.html#ref-list-1

Citing articles
This article has been cited by 32 HighWire-hosted articles. Access the articles at:
/content/69/19/7495.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.