Motility through a novel pathway mediated by CXCL14.

mitochondrial dysfunction and ROS stress promote cancer cell motility through a novel pathway mediated by CXCL14.

Introduction

Emerging lines of evidence suggest that cancer cells may have mitochondrial dysfunction and adapt aerobic glycolysis to generate ATP. The involvement of reactive oxygen species (ROS) signaling in tumor metastasis has also been recognized (1, 2). ROS are constantly generated during cellular metabolism, particularly through the mitochondrial respiration, and exert various physiological actions (3, 4). Oxidative stress is a state of redox imbalance caused by increased ROS generation, decreased antioxidant capacity, or both. Recent studies suggest that cancer cells, compared with the normal cells, are under increased oxidative stress associated with oncogenic transformation, alterations in metabolic activity, and increased generation of ROS (5–7). The increased ROS in cancer cells may in turn affect certain redox-sensitive molecules and further lead to significant consequences such as stimulation of cellular proliferation, cell differentiation, alterations in sensitivity to anticancer agents, promotion of mutations and genetic instability, and contributing to carcinogenesis (5, 8–11). The accumulation of certain genomic mutations enables cells to further acquire the ability to proliferate, generate a tumor, and eventually metastasize (12).

Mitochondria are major sites of ROS generation, which occurs mainly at complexes I and III of the respiratory chain. ROS production increases when electron transport function is compromised, leading to increased leakage of electrons, which react with oxygen to form superoxide. The molecular mechanisms involved in the mitochondrial dysfunction and increased ROS production are not well understood. Multiple pathways may converge in mitochondria to modify respiratory chain activity. Moreover, recent studies indicate that the production of ROS may be accompanied by changes in mitochondrial metabolism (13). The production of excess superoxide can cause oxidative DNA damage and genomic instability (14), transformation of cells in culture (8), and potentiate tumor progression (11). In fact, mitochondrial DNA (mtDNA) is highly susceptible to damage because it is not protected by histones and is directly exposed to ROS generated by the respiratory chain. Moreover, DNA repair capacity is less efficient in the mitochondria. These factors might explain why human cancers have high levels of mtDNA mutations.

Since Warburg proposed the cancer "respiration injury" theory (15), mitochondrial alterations in cancer cells have been intensively studied to understand their role in tumor development, ROS generation, and carcinogenesis (16, 17). Despite significant progress in this research area, little is known of the alterations in mitochondrial structure and function that contribute to cell mobility and metastasis, especially in breast cancer cells. Here we report that perturbation of mitochondrial respiration in breast cancer cells leads to redox alterations and more invasive behaviors, which seem to be mediated by elevated cytosolic calcium through a novel ROS→activator protein-1 (AP-1)→CXCL14 pathway.

Materials and Methods

Reagents. [3H]Deoxyglucose and [γ-32P]ATP were acquired from Amersham Pharmacia Biotech. Recombinant human active CXCL14 and CXCL14 antibody were obtained from R&D Systems. GRP78 and heat shock protein 60 antibodies were from Santa Cruz Biotechnology, Inc., and inositol 1,4,5-trisphosphate receptor (IP3R) antibody from BD Biosciences. SP600125 was acquired from AG Scientific, Inc. CM-H2DCF-DA, dihydroethidium (Het), Calcium Green-1, MitoTracker Green, Fluo-3, ER-Tracker Blue-White DPX, rhodamine-123, and ionomycin were purchased from Invitrogen/Molecular Probes. Phosphorothioate decoy oligo specific for AP-1 (5′-CCTCGGTGACTCACTGAGATTTAAAAAAAATCTCAGTGAGT-3′) and phosphorothioate scrambled oligo (5′-GCATGGACTGTATCGGAAAAACGATACAGTCCATGCCATTATA-3′) were synthesized by Sigma Genosys. The 2 bases at the 3′ and 5′ ends are phosphorothioates. Human CXCL14 cDNA clone was from Origene Technologies, Inc.

Cell culture and migration/invasion assays. MCF7 and its subclones were cultured in DMEM-F12 medium (Cellgro, Mediatech, Inc.) supplemented with 10% fetal bovine serum and 2 mM/L glutamine at 37°C with
5% CO2. Cell migration and invasion assays were done using chambers with 8-µm-porosity polycarbonate filter membranes according to the manufacturer’s instructions (Becton Dickinson Labware). Cells that invaded to the lower surface were stained with Fisher Diagnostics Hema3 Staining System (Thermo Fisher Scientific, Inc.) and photographed under a microscope (Eclipse TS100, Nikon) at ×20 magnification.

Chromatin immunoprecipitation assay. The chromatin immunoprecipitation assay kit from Upstate was used for this study. Protein-DNA complexes were immunoprecipitated with AP-1 antibody (Santa Cruz Biotechnology). Gel Shift Assay System (Promega), according to the manufacturer’s instructions, with human recombinant AP-1 (c-jun) and 5'-GGCGACGGGCGGGCTGGGAAG-3' (forward) and 5'-GGAGACGCCACC-CAGCTCTG-3' (reverse) to amplify region −323/−83 of the CXCL14 promoter. Amplification of soluble DNA before immunoprecipitation was used as an input control. All chromatin immunoprecipitation experiments were repeated at least three times.

Measurements of respiration, glucose uptake, lactate production, ROS, mitochondrial mass, mitochondrial membrane potential, and calcium. Oxygen consumption, cellular glucose uptake, lactate production, mitochondria mass, ROS content, and mitochondrial transmembrane potential (ΔΨm) were measured as previously described (18, 19). To measure cellular Ca2+, cells were stained with 3 µmol/L Calcium Green-1 in serum-free medium for up to 1 h and analyzed using a FACScan flow cytometer (Becton Dickinson Bioscience). Cells were loaded for 1 h with 5 µmol/L Fluo-3-AM (1 × 106 cells/mL) in Ca2+-free HBSS buffer containing 1% bovine serum albumin. Then, cells were washed twice with HBSS and resuspended in HBSS containing 2 µmol/L/ Ca2+. Ca2+ influx was equipped with a 350-nm excitation laser and a 530-nm centered band-pass filter, as previously described (20).

Nuclear extract and gel shift assay. EMSA analysis was done using the Gel Shift Assay System (Promega), according to the manufacturer’s instructions, with human recombinant AP-1 (c-jun) and 5 µg of nuclear extracts from MCF7 cells, and subclones were prepared as previously described (21).

RNA isolation and reverse transcription-PCR analyses. RNA was isolated from MCF7 cells and its subclones using the RNeasy kit and reverse transcribed using the Omniscript RT kit (Qiagen) according to the manufacturer’s instructions. The amplification primers for CXCL14 were 5'-GTCAAATGCAAGTGCTCCC-3' (forward) and 5'-TTCTTCGTAGACCCCTGCACGCTG-3' (reverse) to amplify region −323/−83 of the CXCL14 promoter. Amplification of soluble DNA before immunoprecipitation was used as an input control. All chromatin immunoprecipitation experiments were repeated at least three times.

Subcellular fractionation and microsome Ca2+ release assay. Mitochondria and ER were isolated from cells as previously described (21). Microsomes from MCF7 cells and subclones (H and P clones) using 3 µmol/L CM-H2DCF-DA measured by flow cytometry. C, comparison of oxygen consumption (left), glucose uptake (center), and lactate production (right) in MCF7 cells and subclones. Columns, mean of three independent experiments; bars, SD. D, comparison of mitochondrial mass (left), mitochondrial transmembrane potential (ΔΨm, center), and cell growth (right) of MCF7 cells and subclones. Results of a representative experiment (n = 3).
measured by flow cytometry and triggered by the addition of 2 μmol/L thapsigargin after 10 s. To ensure proper loading, Ca2+ influx from each cell line was stimulated with 2 μmol/L ionomycin. Analysis was done using a FACScan flow cytometer (Becton Dickinson), and the kinetic analysis of Fluo-3 versus time as the mean value of individual cells was done using FlowJo software (Tree Star, Inc.).

**Animal studies.** Six-week-old female BABL/c nude mice were inoculated on the mammary fat pads with MCF7 or its subclones. In both cases, 2 × 10⁶ cells were combined with 100 μL of DMEM-F12 and Matrigel (1:1; BD Biosciences). Mice were inspected daily, and their well-being and body weight monitored. Moribund animals were euthanized as mandated by the Institutional Animal Care and Use Committee (IACUC) protocol, and the time of death was recorded. Tumor tissues from representative mice were sectioned, embedded in paraffin, and stained with H&E for histopathologic evaluation.

**Statistical analysis.** Student’s t test was used to evaluate the statistical differences between the experimental values of two samples being compared. P < 0.05 was considered statistically significant.

**Results**

**Generation and characterization of MCF7 subclones with aggressive cellular behaviors.** To study the role of mitochondrial dysfunction in invasion, we developed an experimental system using human breast cancer MCF7 cells, which can form localized tumors in nude mice with estrogen supplement but are weakly metastatic. Because electron leakage from the mitochondrial respiratory chain is a major source of ROS, we hypothesized that a disruption of mitochondrial respiration would increase ROS, which may promote genetic instability and the emergence of more aggressive malignant cells. As illustrated in Fig. 1A, we used rotenone, an inhibitor of the electron transport complex I known to promote superoxide generation (18), to induce ROS stress in MCF7 cells. After three cycles of rotenone treatment, the cells were plated in drug-free medium to allow the formation of colonies. Among 16 clones picked, two clones (H and P) with high ROS and active cell migration were selected for further studies. Figure 1B shows the representative ROS levels. Interestingly, the increased ROS in these clones persisted for at least 6 months without rotenone, likely reflecting persistent mitochondrial dysfunction. In fact, both H and P clones still exhibited reduced mitochondrial respiration, as indicated by a decrease in oxygen consumption (Fig. 1C) and a compensatory increase in glycolysis, as indicated by higher glucose uptake and lactate production (Fig. 1C). Analysis of cellular ATP showed that both clones had similar cellular ATP compared with the parental MCF7 cells (Supplementary Fig. S1), suggesting that the up-regulation of glycolysis was sufficient to compensate the decreased ATP generation in the mitochondria. Consistently, the subclones exhibited a slight increase in their mitochondrial mass with similar transmembrane potential (Fig. 1D). The subclones showed similar or slightly higher proliferation (Fig. 1D, right). These results are consistent with the...
previous observations that increased mitochondrial mass and mtDNA contents are early molecular events in response to endogenous oxidative stress (24).

Because ROS can alter gene expression (10, 25) and stimulate cell invasiveness (26), we tested the motility and invasive potential of the subclones. A wound healing assay showed that H and P clones exhibited a higher capacity to migrate and fill the gap in culture compared with MCF7 (Fig. 2A). To evaluate whether the increase in ROS in H and P clones was essential for this process, cells were pretreated with the antioxidant N-acetyl cysteine (NAC) before the wound healing assay. NAC effectively inhibited cell motility (Fig. 2A). We also isolated several clones from MCF7 cells without treatment with rotenone as control. No significant change in motility was observed (Supplementary Fig. S2). Subclones H and P exhibited significantly greater ability than the parental MCF7 cells to form tumors (Supplementary Fig. S3). The ability of clones P and H and the parental MCF7 cells to migrate through a layer of matrix gel and pass membrane pores (Fig. 2B and C). Furthermore, colony formation assay in soft agar showed similar or slightly higher anchorage-independent colony-forming rates (number of colonies per well) in the subclones, but the size of each colony was smaller than those of MCF7 cells (Supplementary Fig. S3).

We then tested the ability of clones P and H and the parental MCF7 cells to form tumors in vivo. Parental MCF7 cells and P and H clones were inoculated on the mammary fat pads of 6-week-old BAMBL/c nude mice. Both the P and H clones formed palpable tumors in the absence of exogenous estrogen, whereas the parental MCF7 cells exhibited poor ability to form tumors under these conditions. Three of the five mice inoculated with P clone cells exhibited extensive metastasis to liver, lung, and spleen. The histologic morphology of the primary and metastatic tumors is shown in Fig. 2D. No metastasis was detected in the mice inoculated with H clone cells by the time when the primary tumor reached the maximal size requiring euthanasia, as mandated by the IACUC-approved protocol. No metastasis was observed in mice inoculated with the parental MCF7 cells.

ROS promotes CXCL14 expression in MCF7 subclone cells through activation of AP-1. To elucidate the mechanisms by which mitochondrial dysfunction alters cell migration/invasion, we performed microarray analysis of gene expression in several clones of cells with increased ROS. We observed that the expression of CXCL14, a novel chemokine, was significantly elevated in all clones tested in comparison with MCF7 cells. The overexpression of CXCL14 in clones P and H was confirmed by reverse transcription-PCR (RT-PCR) and Western blotting (Fig. 3A). The ability of CXCL14 to stimulate cell migration was then tested using the wound healing assay. Figure 3B shows that recombinant CXCL14 protein (0.57 ng/mL) significantly enhanced MCF7 cell migration. Furthermore, conditioned medium from clones P and H also promoted cell migration (Fig. 3B). Ectopic overexpression of CXCL14 in MCF7 cells by transfection also rendered the cells highly migratory (Fig. 3C). To further evaluate the relationship between the induction of CXCL14 and increased migration and invasion, we used a decoy strategy using a synthetic double-stranded phosphorothioate oligonucleotide, which contained a hairpin structure resembling the CXCL14 promoter region and functioned as a cis-element to suppress CXCL14 expression. Transfection of H clone with the decoy DNA efficiently suppressed CXCL14 transcription and inhibited cellular migration, as evidenced by the reduced numbers of cells migrating through the membrane (Fig. 3D).

We then used an antioxidant to investigate whether ROS regulate CXCL14 expression. NAC partially suppressed the expression of
CXCL14 in clone P (Fig. 4A, bottom) and clone H (data not shown), suggesting that CXCL14 expression might be regulated by ROS. Consistently, hydrogen peroxide (H2O2) promoted CXCL14 expression in a time- and dose-dependent manner (Fig. 4A, top). Interestingly, another clone with slower proliferation rate (clone D) also exhibited increase ROS generation, higher CXCL14 expression, and active cell motility (Supplementary Fig. S4). Thus, the increase in cell motility was correlated with ROS generation and CXCL14 expression and was not due to active proliferation.

To further explore the mechanism by which ROS regulate CXCL14 expression, we performed a gene database analysis and identified a potential AP-1 binding sequence within the putative promoter region of CXCL14. EMSA assay using a radioactively labeled oligonucleotide containing the AP-1 consensus sequence showed that both subclones (H and P) had an increase in AP-1 activity compared with the parental cells (Fig. 4B, left). Furthermore, nuclear extract from MCF7 cells exposed to H2O2 for 20 minutes showed a significant increase in AP-1 DNA binding (Fig. 4B, right), suggesting a role of ROS in regulating CXCL14 expression through AP-1. To test this possibility, the CXCL14-overexpressing cells (clone H) were incubated with the c-Jun-NH2-kinase (JNK) inhibitor SP600125 to block c-Jun (a component of AP-1) activation and then tested if this would cause a decrease of CXCL14 transcription. Indeed, SP600125 inhibited AP-1 DNA binding and reduced CXCL14 expression at the mRNA and protein levels (Fig. 4C). Chromatin immunoprecipitation was then used to analyze if AP-1 binds to CXCL14 promoter. As shown in Fig. 4D, after protein-DNA cross-linking, immunoprecipitation with AP-1 antibody pulled down the CXCL14 promoter DNA sequence, which was detected by PCR using specific primers spanning the −323 to −83 region of the human CXCL14 promoter. These results show that AP-1 binds to CXCL14 promoter.

CXCL14 induced an increase in cellular Ca2+ through interaction with IP3R. Because Ca2+ plays an essential role in cell motility and the ER is a major regulator of cellular Ca2+ homeostasis (27), we tested if CXCL14 affects ER function and alters cytosolic Ca2+ in the subclones. Using subcellular fractionations, we found that CXCL14 was significantly higher in the ER compartments of clones H and P than MCF7 cells (Fig. 5A). Interestingly, analysis of cytosolic Ca2+ revealed that clones H and P had significantly higher cytosolic Ca2+ contents than the parental MCF7 cells (Fig. 5B, left), whereas the ER Ca2+ levels were significantly lower in clones H and P (Fig. 5B, right). This suggests a possible "leakage" of Ca2+ from the ER to the cytosol associated...
with the localization of CXCL14 to ER. Transfection of MCF7 cells with CXCL14 caused an increase in cytosolic Ca\(^{2+}\) (Fig. 5C). Because the IP\(_3\)R of ER plays an important role in Ca\(^{2+}\) signaling via interaction with inositol 1,4,5-trisphosphate (IP\(_3\); ref. 28), we examined whether CXCL14 binds to IP\(_3\)R and disturbs its function. Immunoprecipitation with IP\(_3\)R antibody and Western blotting of the pulled-down products using anti-CXCL14 showed a physical association between CXCL14 and IP\(_3\)R (Fig. 5D). We reasoned that if the binding of CXCL14 to IP\(_3\)R caused a leak of Ca\(^{2+}\) from the ER to the cytosol, the amount of Ca\(^{2+}\) stored in the ER would be lower in the subclones than in the parental MCF7 cells and that a release of ER Ca\(^{2+}\) into the cytosol triggered by thapsigargin would cause less increase of cytosolic Ca\(^{2+}\) in the subclones. As shown in Fig. 6A, thapsigargin caused a rapid increase of cytosolic Ca\(^{2+}\) in MCF7 cells and a substantially smaller Ca\(^{2+}\) increase in the P clone cells, suggesting lower ER Ca\(^{2+}\) storage in P cells. The effect of thapsigargin on Ca\(^{2+}\) levels in these cells was further investigated in the presence of 100 μmol/L 2-aminoethoxydiethyl borate (2-APB), a compound that blocks receptor-induced Ca\(^{2+}\) store emptying (29). Preincubation of MCF7 cells with 2-APB almost completely suppressed thapsigargin-induced Ca\(^{2+}\) release in MCF7 cells, but this effect was substantially attenuated in clone P (Fig. 6A). Thus, CXCL14 might interact with the 2-APB–sensitive domain of IP\(_3\)R, leading to Ca\(^{2+}\) leakage and a blunted response to thapsigargin. To directly test the role of CXCL14 on Ca\(^{2+}\) flux regulation in the ER, we isolated the ER fraction from MCF7 cells and measured Ca\(^{2+}\) release from the ER microsomes in the presence and absence of purified CXCL14 protein. As shown in Fig. 6B to D, CXCL14 induced Ca\(^{2+}\) release from the ER microsomes in a time- and dose-dependent manner. As a positive control, Ca\(^{2+}\) release was triggered by 10 μmol/L IP\(_3\) to activate IP\(_3\)R or by 10 μmol/L thapsigargin to inhibit the ER Ca\(^{2+}\) pump. As expected, IP\(_3\) and thapsigargin caused a dramatic Ca\(^{2+}\) release, whereas CXCL14 induced a moderate but consistent Ca\(^{2+}\) release (Fig. 6D). This may explain why overexpression of CXCL14 led to a moderate and sustained increase in cytosolic Ca\(^{2+}\) without causing cytotoxicity.

**Discussion**

Breast cancer metastasis represents a major challenge in clinical treatment of this disease, and the detailed mechanisms that regulate breast cancer cell motility and metastasis remain to be an important research area. It is known that mitochondria are major sites of ROS generation in the cells and that ROS can function as signaling molecules to stimulate cell proliferation and promote cell motility (1). Interestingly, mitochondria dysfunction has been shown to contribute to tumor progression by enhancing the metastatic potential of tumor cells (30). However, the mechanism by which ROS stimulates cell motility is still unclear. The present study provides several lines of evidence suggesting that increased expression of CXCL14 may play an important role in this process. (a) Both clones H and P, which were isolated after chemical stimulation of cell motility using recombinant CXCL14 protein, conditional medium, or overexpression of CXCL14 through gene transfection. (d) CXCL14 induced Ca\(^{2+}\) release from the ER through binding to IP\(_3\)R, leading to increased cytosolic Ca\(^{2+}\). (e) ROS seem to induce CXCL14 expression through activation of the
AP-1 pathway, as shown by specific decoy DNA, JNK inhibitor, and antioxidant.

CXCL14 (also known as BRAK or MIP-2γ) is a chemokine with yet unknown function (31, 32). This molecule was first identified by differential display using normal epithelial cells and head and neck squamous carcinoma cells and was shown to be expressed in normal epithelial tissues and in various tumors of epithelial origin with heterogeneous expression levels (31, 33). The receptor for CXCL14 has not yet been identified, and conflicting biological functions of this molecule are described in the literature (34–37). CXCL14 is associated with the development and progression of breast cancer, and its gene expression level is correlated with the clinical metastases time of breast cancer patients (38). Moreover, CXCL14 seems to increase the invasiveness of pancreatic cancer cells by activation of the nuclear factor-κB pathway (39). Our study suggests that in breast cancer cells, CXCL14 promotes cell motility through its ability to induce release of Ca^{2+} to the cytosol.

CXCL14 seems to provide a molecular link between ROS stress and increased cell motility. DNA analysis revealed that the CXCL14 putative promoter region contains a binding site for AP-1, a transcription regulatory complex known to be redox sensitive. We showed that inhibition of JNK by SP600125 suppressed AP-1 DNA binding activity and reduced CXCL14 expression. The AP-1 decoy also suppressed CXCL14 expression and inhibited cell motility. Thus, activation of AP-1 is a likely mechanism for ROS-induced CXCL14 up-regulation. Interestingly, metastatic breast carcinoma cells (estrogen receptor negative) have higher AP-1 DNA binding activity than do less metastatic carcinomas (estrogen receptor positive), and expression of Fra-1 correlates with AP-1 activity and invasiveness in these cells (40). Several AP-1–responsive genes with roles in facilitating the invasion of malignant cells have been described (41, 42), and our study suggests that CXCL14 is another important AP-1 effector. Our observations that mitochondrial dysfunction induces ROS-mediated AP-1 activation and CXCL14 up-regulation is consistent with a recent study that mitochondrial oxidative stress through overproduction of H_{2}O_{2} induced the Jun kinase pathway (13).

Calcium plays an essential role in the assembly of actin filaments and cytoskeleton reorganization and, thus, affects cell motility and migration. We found that cytosolic Ca^{2+} levels in both clones H and P were elevated. This increase in cytosolic Ca^{2+} is likely due to the release of Ca^{2+} from the ER. We showed that CXCL14 was present in the ER, physically interacted with IP_{3}R, and directly induced Ca^{2+} release from isolated ER microsomes. Furthermore, the IP_{3}R

Figure 6. CXCL14-induced Ca^{2+} release in microsomes and its inhibition by 2-APB in clone P in vitro. A, CXCL14 prevented the inhibition of IP_{3}R by 2-APB in MCF7 subclones. Cells were loaded with 5 μmol/L Fluo-3 for 1 h. MCF7 (red curve) and clone P (green curve) were treated with 2 μmol/L thapsigargin (TG) to deplete intracellular stores and activate capacitative calcium entry. MCF7 (brown curve) and clone P (blue curve) were treated with 100 μmol/L 2-APB for 10 min before adding 2 μmol/L thapsigargin. The results of a representative recording of the free Ca^{2+} measured by Fluo-3 fluorescence using flow cytometry as described in Materials and Methods are shown (n = 3). B, CXCL14 induced Ca^{2+} release from microsomes in a time-dependent manner. Microsomes were incubated with 1 μg CXCL14 protein, and at the indicated time, they were spun down and the supernatant containing Ca^{2+} was kept to further evaluate Ca^{2+} released. C, CXCL14 induced Ca^{2+} release from microsomes in a dose-dependent manner. Microsomes were incubated with up to 1 μg CXCL14 protein for 2 h and then were spun down and the supernatant was kept to evaluate Ca^{2+} released. D, comparison of Ca^{2+} released from microsomes induced by IP_{3}R, thapsigargin, or CXCL14. Microsomes were incubated with 0.5 or 1 μg of CXCL14 protein, 10 μmol/L IP_{3} or 10 μmol/L thapsigargin for 1 h and then spun down, and the supernatant was used to evaluate Ca^{2+} released. Columns, mean of at least three independent experiments; bars, SD.
inhibitor 2-APB was significantly less effective in preventing thapsigargin-induced Ca\textsuperscript{2+} mobilization in CXCL14-overexpressing clone P cells than in the parental MCF7 cells. Together, these observations suggest that CXCL14 induces cytosolic Ca\textsuperscript{2+} increase by mobilization of Ca\textsuperscript{2+} from the ER through binding to IP\textsubscript{3}R, and that CXCL14 might attenuate the function of 2-APB–sensitive IP\textsubscript{3}R channels. In future study, it would be interesting to further determine the detailed molecular interaction between CXCL14 and IP\textsubscript{3}R through systemic deletion of the CXCL14 peptide segments to identify the interface responsible for binding, and use specific antibodies to show the colocalization of the two molecules on the ER membranes.

Although a high level of cytosolic Ca\textsuperscript{2+} can stimulate mitochondrial respiration in cells with competent mitochondrial, this may not happen in cells with compromised mitochondrial function. In our experimental system, the Ca\textsuperscript{2+} increase seemed to be a consequence of mitochondria dysfunction initially induced by rotenone. It is unclear if the increased cytosolic Ca\textsuperscript{2+} may play a role in up-regulation of glycolysis. Interestingly, most glycolytic enzymes bind to cytoskeleton, and it is possible that increased cytosolic Ca\textsuperscript{2+} might promote the assembly of cytoskeleton-bound glycolytic enzymes leading to high glycolytic activity.

Of note, the aggressive malignant cell behaviors of the clones H and P were stable after the cells were cultured in medium free of rotenone, suggesting that a possible initial ROS increase induced by rotenone might have induced mutations in mtDNA or and nuclear DNA. We have attempted to identify mtDNA mutations by direct sequencing of mtDNA for these two subclones in comparison with the parental MCF7 cells. No mtDNA mutation was detected. It is possible that in clones H and P, mutation(s) might have occurred in the nuclear DNA at a gene that affects mitochondrial function leading to decreased respiration and increased glycolysis, which seemed to affect cell proliferation and colony formation in a complex manner: high colony number but smaller colony sizes. Although the exact reason for such complex growth alterations remain unclear, this phenomenon may explain why the subclones showed similar overall cell growth curves compared with the parental MCF7 cells, as measured by direct counting of cell number (Fig. 1D, right).

In conclusion, our findings suggest that mitochondrial dysfunction in human breast cancer cells can promote cell motility and invasive behaviors through the AP-1/CXCL14/calcium pathway. The increased ROS in cancer may promote cellular proliferation and migration leading to more aggressive malignant behaviors and metastasis. It should be pointed out, however, that because elevated ROS may also cause cellular damage and induce senescence, only those cancer cells capable of mobilizing their antioxidant systems and other regulatory mechanisms to effectively counteract the adverse effects of ROS can reestablish redox balance and emerge as highly malignant clones. In fact, we observed that the cancer cells with high ROS generation in long-term culture tended to rebalance their ROS generation and elimination, leading to a steady redox state that favored their continuous proliferation in culture. Thus, ROS signaling and its roles in cancer development and metastasis are extremely complex biological processes. Understanding these mechanisms is essential for developing new therapeutic strategies to effectively inhibit cancer growth and prevent metastasis.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

Received 8/28/2008; revised 11/13/2008; accepted 12/1/2008; published OnlineFirst 3/10/09.

Grant support: National Cancer Institute grants CA53563, CA109041, CA104028, CA116032, and CA16672.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

We thank Wendy D. Schober and K. Dwyer for flow cytometry analysis and Kenneth Dunner, Jr. for assistance in the transmission electron microscopy studies.

References


Downloaded from cancerres.aacrjournals.org on April 14, 2017 © 2009 American Association for Cancer Research.
Mitochondrial Dysfunction and Reactive Oxygen Species Imbalance Promote Breast Cancer Cell Motility through a CXCL14-Mediated Mechanism

Helene Pelicano, Weiqin Lu, Yan Zhou, et al.


Updated version
Access the most recent version of this article at: doi:10.1158/0008-5472.CAN-08-3359

Supplementary Material
Access the most recent supplemental material at: http://cancerres.aacrjournals.org/content/suppl/2009/03/05/0008-5472.CAN-08-3359.DC1

Cited articles
This article cites 40 articles, 15 of which you can access for free at: http://cancerres.aacrjournals.org/content/69/6/2375.full.html#ref-list-1

Citing articles
This article has been cited by 7 HighWire-hosted articles. Access the articles at: /content/69/6/2375.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.