Bcl9/Bcl9l Are Critical for Wnt-Mediated Regulation of Stem Cell Traits in Colon Epithelium and Adenocarcinomas

Jürgen Deka1,2, Norbert Wiedemann1, Pascale Anderle4, Fabienne Murphy-Seiler1, Jennyfer Bultinck1, Sven Eyckerman1, Jean-Christophe Stehle3, Sylvie André1, Nathalie Vilain1, Olav Zilian1, Sylvie Robine5, Mauro Delorenzi6, Konrad Basler7, and Michel Aguet1,2

Abstract

Canonical Wnt signaling plays a critical role in stem cell maintenance in epithelial homeostasis and carcinogenesis. Here, we show that in the mouse this role is critically mediated by Bcl9/Bcl9l, the mammalian homologues of Legless, which in Drosophila is required for Armadillo/β-catenin signaling. Conditional ablation of Bcl9/Bcl9l in the intestinal epithelium, where the essential role of Wnt signaling in epithelial homeostasis and stem cell maintenance is well documented, resulted in decreased expression of intestinal stem cell markers and impaired regeneration of ulcerated colon epithelium. Adenocarcinomas with aberrant Wnt signaling arose with similar incidence in wild-type and mutant mice. However, transcriptional profiles were vastly different: Whereas wild-type tumors displayed characteristics of epithelial-mesenchymal transition (EMT) and stem cell–like properties, these properties were largely abrogated in mutant tumors. These findings reveal an essential role for Bcl9/Bcl9l in regulating a subset of Wnt target genes involved in controlling EMT and stem cell–related features and suggest that targeting the Bcl9/Bcl9l arm of Wnt signaling in Wnt-activated cancers might attenuate these traits, which are associated with tumor invasion, metastasis, and resistance to therapy. Cancer Res; 70(16); 6619–28. ©2010 AACR

Introduction

Canonical Wnt signaling regulates critical processes during embryonic development and adult tissue renewal, and aberrant activation of this pathway is associated with colorectal cancer and other cancers (1). Wnt-induced transcriptional regulation is mediated through a complex of Lef1/Tcf transcription factors and stabilized β-catenin that interacts with numerous cofactors (2). This complex includes Legless (Lgs; ref. 3), which tethers Pygopus to the most NH2-terminal repeat of Armadillo (Arm)/β-catenin and in Drosophila is required for the transcriptional activity of the complex (3, 4). In mammalian cells, BCL9 proteins contribute to, but may not be obligatory for, β-catenin–mediated transcription (5, 6). BCL9L mRNA knockdown in a human colon cancer cell line induced an epithelial phenotype that was associated with the translocation of β-catenin from the nucleus to the cell membrane, suggesting that BCL9L might regulate the switch between the adhesive and transcriptional functions of β-catenin (5). BCL9L upregulation has been associated with breast (7) and colon cancers (8–10), and BCL9L has recently been reported to enhance β-catenin–mediated transcription and increase the proliferation as well as the metastatic potential of tumor cells (9).

To investigate the role of Bcl9 proteins in mice, we generated loxP alleles of the murine lgs orthologues, Bcl9 and Bcl9l, which were crossed into transgenic strains expressing Cre recombinase either constitutively or inducibly under the control of the villin promoter (11). Thereby, deletion of Bcl9 and Bcl9l was targeted selectively to the intestinal epithelium, for which the essential role of Wnt signaling for epithelial homeostasis is well documented (12).

Here, we report that ablation of Bcl9/Bcl9l in the intestinal epithelium selectively abrogated the expression of a subset of Wnt-regulated genes, which are critical for controlling stem cell–like properties, both in normal colon epithelium and in a colon adenocarcinoma model.

Materials and Methods

Generation of Bcl9+/−/Bcl9l+/− mice

For the generation of conditional Bcl9+/−/Bcl9l+/− mice, exon V of Bcl9 and exons VI and VII of Bcl9l were flanked with loxP sites in adjacent introns. Simultaneous deletion of both

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genes in the intestinal epithelium was achieved by crossing with villin-Cre-ER<sup>T2</sup> or villin-Cre transgenic mice, resulting in vil-Cre-ER<sup>T2</sup>-Bcl9<sup>loxP/loxP</sup>/Bcl9l<sup>loxP/loxP</sup> (on tamoxifen induction: vil-Cre-ER<sup>T2</sup>-Bcl9<sup>−/−</sup>/Bcl9l<sup>−/−</sup>) and vil-Cre-Bcl9<sup>−/−</sup>/Bcl9l<sup>−/−</sup> lines. More detailed information is available in Supplementary Materials and Methods.

**Mouse experimentation**

Mouse experiments were performed in accordance with Swiss guidelines and approved by the Veterinarian Office of Vaud, Switzerland. The protocols for gene ablation in vil-Cre-ER<sup>T2</sup>-Bcl9<sup>loxP/loxP</sup>/Bcl9l<sup>loxP/loxP</sup> mice and the induction of ulcerative colitis and colon chemical carcinogenesis are provided in Supplementary Materials and Methods.

**Laser dissection microscopy**

Samples of colon tissue were washed in chilled PBS, embedded in optimal cutting temperature compound, and air-dried. Frozen sections (14 μm) were mounted on Leica membranes (Leica Microsystems) for laser dissection microscopy, fixed in ethanol, and colored with Mayer’s hematoxylin. Membranes were rinsed in water and ethanol and air-dried. Samples were prepared using a laser-dissecting microscope (MMI-Nikon, Leica Microsystems). Total RNA (20 μL) was extracted using the PicoPure RNA Isolation Kit (MDS Analytical Technologies).

**Comparative gene expression analysis**

RNA quality was assessed using the Agilent 2100 Bioanalyzer (Agilent Biotechnologies) and RNA was quantified with a NanoDrop spectrophotometer (Witec AG). Total RNA (30 ng) was amplified using the NuGEN WT-Ovation Pico kit and the cDNA was hybridized to Affymetrix Mouse Genome 430 2.0 arrays. Robust multiarray averaging and quantile normalization were used to quantify gene expression (R, Bioconductor, RMA package). Significant differences were identified by applying a moderated t test approach (R, Bioconductor, LIMMA package; ref. 13). The gene expression microarray data reported herein were deposited in the Gene Expression Omnibus repository (accession no. GSE1603; see Supplementary Data for details). If not indicated otherwise, a threshold of an adjusted <i>P</i> ≤ 0.05 (Benjamini-Hochberg FDR correction) was used to identify significant changes between wild-type and mutant samples.

Genes with significantly different expression in wild-type as compared with Bcl9<sup>−/−</sup>/Bcl9l<sup>−/−</sup> tumors were analyzed with the functional annotation chart tool in DAVID (14). Gene ontology (GO) terms related to “biological processes” were used. Fisher’s exact test was applied to calculate significance. Terms with an adjusted <i>P</i> < 0.05 (Benjamini-Hochberg FDR correction) were defined as significant.

Gene set enrichment analysis (GSEA) was carried out according to Subramanian and colleagues (15). <i>P</i> values were computed using a bootstrap distribution created by resampling gene sets of the same cardinality. This is a heuristic approach to identifying gene sets and biological functions that, given the samples used, seem to be most related to the observed effects. Its <i>P</i> values could overestimate the appropriate level of confidence in relation to the extent of the biological variability that was sampled and are better considered as comparative scores between gene sets. Lists of direct Wnt targets (n = 24), genes involved in epithelial-mesenchymal transition (EMT; n = 13), and genes enriched in intestinal stem cells marked by the expression of Lgr5 were generated manually (Fig. 4A). Heat maps were generated using Cluster 3.0 and TreeView (ref. 16; http://rana.lbl.gov/EisenSoftware.htm). As a similarity measurement and clustering method, centered correlation and average linkage clustering, respectively, were applied. The Davies-Bouldin index was determined as described (17), whereby intercluster and intraclass differences were calculated using the average of all pairwise comparisons. The Davies-Bouldin index is a ratio that compares the average distance within a group of samples to that between groups. A value of 2 indicates no difference and smaller values indicate better discrimination with a theoretical minimum of 0. All configurations tested were obtained by cutting the hierarchical tree into two clusters that always corresponded to the five mutant and the five wild-type samples.

The significance of quantitative real-time PCR (qRT-PCR) data was determined using Student’s <i>t</i> test (<i>P</i> ≤ 0.05).

**Standard protocols and reagents**

Quantitative RT-PCR, <i>in situ</i> hybridization, and immunohistochemistry were carried out according to standard protocols. Details and reagents are provided in Supplementary Materials and Methods.

**Results**

**Ablation of Bcl9/Bcl9l results in decreased intestinal stem cell marker expression and impaired epithelial regeneration**

Despite the rather low overall sequence identity, mouse Bcl9 and Bcl9l share seven highly conserved domains (18), suggesting that they might be functionally redundant. As both genes were expressed in the small intestine and colon, with Bcl9 generally expressed at an approximately 2- to 4-fold higher level than Bcl9l (data not shown), all loss-of-function experiments presented herein were carried out on a compound Bcl9/Bcl9l mutant background. Conditional loss-of-function alleles of Bcl9 and Bcl9l (GenBank accession nos. AY296061 and AY296058) were generated by flanking exons encoding the functionally essential β-catenin interaction domain with loxp sites (Supplementary Fig. S1A). Mouse strains with compound inducible or constitutive deletion of both genes confined to the intestinal epithelium were obtained through crossing with villin-Cre-ER<sup>T2</sup> or villin-Cre transgenic mice, respectively (11), resulting in vil-Cre-ER<sup>T2</sup>-Bcl9<sup>loxP/loxP</sup>/Bcl9l<sup>loxP/loxP</sup> (on tamoxifen induction: vil-Cre-ER<sup>T2</sup>-Bcl9<sup>−/−</sup>/Bcl9l<sup>−/−</sup>) and vil-Cre-Bcl9<sup>−/−</sup>/Bcl9l<sup>−/−</sup> lines. Unless stated otherwise, experiments reported herein were carried out initially on a villin-Cre-ER<sup>T2</sup> background and reproduced using the constitutive villin-Cre transgene. Bcl9l<sup>loxP/loxP</sup>/Bcl9<sup>loxP/loxP</sup> littermates lacking the Cre transgene served as controls.
Deletion of Bcl9/Bcl9l proved highly efficient in both strains (Supplementary Fig. S1B–D). Induced ablation of Bcl9/Bcl9l in the villin-Cre-ERT2 strain assessed 23 weeks after tamoxifen administration proved similarly efficient as in mice with constitutively active Cre, suggesting that the deletion was unaffected by epithelial renewal and therefore had encompassed the epithelial progenitor cell compartment.

Neither induced nor constitutive ablation of Bcl9/Bcl9l in the intestinal epithelium resulted in any overt anomalies, and all intestine-specific cell lineages were equally present in mutant and control mice (Supplementary Fig. S2).

Canonical Wnt signaling as revealed by expression of the direct target gene Axin2 (19) did not seem to be significantly altered in small intestinal epithelium and was only mildly reduced in the colon epithelium of mutant mice (Supplementary Fig. S1D). To assess Wnt signaling more comprehensively, EDTA-dissociated colon epithelial cells (20) from three pools each of wild-type and Bcl9/Bcl9l–mutant mice were subjected to an exploratory comparative gene expression profiling. Using an adjusted P < 0.05 as a threshold (see Materials and Methods), no significant gene expression changes were observed between wild-type and mutant colon epithelium. Notably, Wnt target gene expression (http://www.stanford.edu/~rnusse/wntwindow.html) did not prove to be significantly affected in Bcl9/Bcl9l–mutant colon epithelium, strikingly, with the exception of the intestinal stem cell marker Lgr5 (21), which was reduced 3.4-fold (P = 2E−5, not adjusted for multiple testing). A GSEA (15) was therefore carried out with a recently reported set of genes with ≥2-fold enriched expression in Lgr5-positive stem cells isolated from the small intestine (22); results revealed a weak but significant enrichment in wild-type versus Bcl9/Bcl9l–mutant colon epithelium (P = 0.019). The expression of genes enriched in this analysis was further assessed by qRT-PCR (Fig. 1A). Expression of the other genes coexpressed selectively in Lgr5-positive small intestine stem cells was significantly reduced in Bcl9/Bcl9l–mutant epithelium and impaired regeneration of Bcl9/Bcl9l–mutant epithelium following dextran sodium sulfate (DSS) treatment (Fig. 1B). H&E staining and analysis of Axin2 expression by in situ hybridization on sections of mouse colon showing representative DSS-induced lesions 6 d after DSS administration was terminated. Bar, 150 μm.
diminished in the absence of Bcl9/Bcl9l. Interestingly, expression of achaete scute-like 2 (Ascl2), which has recently been reported to control intestinal stem cell fate (22), did not prove to be significantly different between wild-type and mutant colon epithelium (Supplementary Fig. S4).

To investigate the regenerative capacity of Bcl9−/−/Bcl9l−/− intestinal epithelium, mice were challenged by p.o. administration for 7 days of the irritant dextran sulfate sodium (DSS), which has been described to provoke severe colitis with extensive ulcerations (25). Animals were monitored for weight loss and sacrificed when the loss reached 20% (Fig. 1B). Whereas the majority of control mice recovered 5 to 7 days after DSS administration was discontinued and had lost less than 20% body weight, the majority of mutant mice crossed the 20% weight loss percentile. At this stage, mutant mice showed considerably more extended ulcerative lesions within the colon epithelium. Whereas control lesions underwent visible re-epithelialization with the formation of new crypts at wound borders, the large open wounds of mutant mice displayed no evidence of ongoing epithelial regeneration (Fig. 1C). Like in mutant colon epithelium, Axin2 expression seemed slightly reduced at mutant wound borders, this difference did not prove to be significant, however, when quantified by qRT-PCR (data not shown). Thus, ablation of Bcl9/Bcl9l resulted in reduced expression of intestinal stem cell markers in Bcl9/Bcl9l-mutant colon epithelium and was associated with impaired epithelial regeneration capacity.

**Bcl9−/−/Bcl9l−/−** adenocarcinomas show impaired expression of selected Wnt target genes

To investigate the effect of Bcl9/Bcl9l ablation on tumorigenesis, 6- to 8-week-old mice were exposed first to a single dose of dimethylhydrazine, which is metabolized in the liver to carcinogenic azoxymethane, followed by 7 days of p.o. administration of DSS in the drinking water. This regimen results in the emergence of dysplastic adenomas, which progress to differentiated adenocarcinomas that are morphologically similar to human colorectal adenocarcinoma and typically harbor β-catenin–stabilizing mutations of glycogen synthase kinase-3β phosphorylation sites (24). Accordingly, these tumors presented hallmarks of active Wnt signaling such as accumulation of nuclear β-catenin and expression of Wnt target genes (Fig. 2).

Vil-Cre-ERloxPlox-Bcl9−/−/Bcl9l−/− and vil-Cre-Bcl9−/−/Bcl9l−/− mice developed tumors in the distal colon at a slightly higher frequency than their littermate controls (not reproducibly significant; Fig. 2A). Tumors from mutant mice were generally of smaller size (mean tumor diameter: wild-type tumors, 4.4 ± 1.7 mm, n = 58; Bcl9−/−/Bcl9l−/− tumors, 2.4 ± 1.0 mm, n = 110; two-sample t test P = 5.1E−12). Bromodeoxyuridine incorporation, however, did not prove to be significantly different (data not shown). To rule out that tumors emerged from residual wild-type epithelial cells in otherwise mutant epithelium, cDNA was obtained from laser capture microdissected tumor samples, and expression of wild-type Bcl9 and Bcl9l transcripts assessed by qRT-PCR (Fig. 2B). The residual level of wild-type transcripts in tumors from mutant mice was incompatible with heterozygous escapee tumor cells and likely due to stromal components. Wild-type and mutant tumors presented as exophytic, low-grade adenocarcinomas (Fig. 2C) and were indistinguishable on H&E staining.

Nuclear accumulation of β-catenin was indicative of Wnt pathway activation and expression of Axin2 provided evidence for Wnt signaling activity, both in wild-type and mutant tumors, whereby Axin2 expression seemed to be reduced in the absence of Bcl9/Bcl9l (Fig. 2C).

To assess at a genome-wide level to what extent loss of Bcl9/Bcl9l function affected gene expression, individual cRNA probes were prepared from microdissected tumor epithelium of five randomly chosen tumors (two wild-type and three vil-Cre-Bcl9−/−/Bcl9l−/− littermates) and subjected to comparative gene expression profiling. Despite the close morphologic resemblance, transcriptomes from Bcl9−/−/Bcl9l−/− tumors proved vastly different from wild-type tumors.

Unsupervised hierarchical clustering with all probe sets formed two well-separated clusters according to the genotypes (Fig. 3A). Based on adjusted P ≤ 0.05 as a threshold (see Materials and Methods), 746 genes were expressed at a higher level and 946 at a lower level in wild-type than in Bcl9−/−/Bcl9l−/− tumors (Supplementary Table S1). Analysis of GO terms contained within the domain “biological processes” (14) sorted according to their overrepresentation (P < 0.05) revealed a marked enrichment in wild-type tumors of processes relating to development (Supplementary Table S2). GO terms relating to Wnt signaling were the only pathway-associated terms enriched in wild-type samples. In Bcl9−/−/Bcl9l−/− tumor samples, GO terms related to immune response were predominantly enriched (Supplementary Table S2), mainly due to the higher expression of epithelial chemokines and genes expressed in T and B cells (Supplementary Table S1). Preliminary evidence obtained from microdissected normal colon epithelium indicated that expression of chemoattractants in mutant tumor samples was closer to that of wild-type or mutant epithelium than to that of wild-type tumors (data not shown).

To assess to what extent canonical Wnt signaling was affected in Bcl9/Bcl9l–mutant tumors, a set of 24 Wnt target genes relevant to human colon and/or other cancers, with a proven direct transcriptional control through TCF binding sites, was assembled from the Wnt home page and published reports [http://www.stanford.edu/~russwe/wntwindow.html; Apcdd1 (ref. 25), Prox1 (ref. 26), Vim (ref. 27), S100a4 (ref. 28), Fscn1 (ref. 29), and T (ref. 30)]. This set was used for hierarchical clustering and GSEA (Fig. 3B) and revealed a significantly enriched expression in wild-type compared with Bcl9−/−/Bcl9l−/− tumors (P = 4.3E−05). Accordingly, intercluster separation assessed by the Davies-Bouldin index (17) proved stronger with this 24 Wnt target gene set than with the set of the 24 genes varying the most across all samples. These data unequivocally showed the relatedness of Bcl9/Bcl9l to the Wnt pathway. A subset of the Wnt target genes was downregulated in Bcl9−/−/Bcl9l−/− tumors as compared with wild-type tumors (Fig. 3B). The expression of all 24 genes contained in the Wnt target gene set was verified by qRT-PCR (Fig. 3C).
Bcl9−/−/Bcl9l−/− adenocarcinomas show reduced expression of EMT and stem cell–associated markers

The subset of Wnt target genes with reduced expression in Bcl9−/−/Bcl9l−/− tumors included genes relating to mesenchymal phenotypes [vimentin (31); T (brachyury; ref. 30)]. To address to what extent the processes relating to EMT, known to be mediated in part by Wnt signaling (32), were more generally affected in the absence of Bcl9/Bcl9l, a set of 13 genes described to relate to EMT was assembled (32, 33). Clustering the tumor samples with this set revealed a stronger separation between the two genotype clusters than using the 13 genes varying the most across all samples (Fig. 3B). Verification by qRT-PCR confirmed that a number of EMT-inducing transcription factors present in wild-type tumors were in part strongly downregulated in Bcl9−/−/Bcl9l−/− tumors, consistent with the marked reduction of the mesenchymal marker proteins vimentin, T (brachyury), and fibronectin 1 (Fig. 3C).

Recent evidence suggests that EMT may be associated with the induction of stem cell–like properties (32, 34). A GSEA was therefore carried out with the same set of 50 genes with ≥2-fold enriched expression in Lgr5-positive intestinal stem cells that was previously used to probe the transcriptomes of wild-type and Bcl9−/−/Bcl9l−/− colon epithelium (ref. 22; Fig. 1A). This analysis revealed a highly significant enrichment for expression of this gene set in the wild-type...
tumor samples (P = 1E–6), and accordingly, intercluster separation was superior to the separation observed with the control set of 50 genes varying the most across all samples (Fig. 3B). Expression of genes contained in this stem cell–associated gene set that were ≥2-fold decreased in mutant tumors was validated by qRT–PCR (Fig. 3C).

For selected genes, expression differences between wild-type and Bcl9−/−/Bcl9l−/− tumor samples were confirmed by immunohistochemistry. Vimentin, whose expression can be directly regulated by β-catenin/Tcf (27), is one of the most reliable mesenchymal markers. The difference in vimentin expression in wild-type compared with Bcl9−/−/Bcl9l−/− tumors, however, was clearly attributed to its loss in mutant epithelial cells (Fig. 4A, c versus d, e versus f) and not to a lower stroma content. Higher magnification revealed that wild-type tumor cells typically expressed apical cytokeratin and basal vimentin intermediate filaments (Fig. 4A, g). This coexistence of epithelial and mesenchymal traits is characteristic of a metastable state in between the epithelial and mesenchymal extremes (31). In contrast, the vast majority of mutant tumor epithelium was constituted of vimentin-negative cells, which stained homogeneously for cytokeratin and EpCAM, indicative of a more epithelial state (Fig. 4A, h).

To investigate to what extent stem cell marker expression might coincide with this metastable cell state, expression of Sox6, the only expressionaly Lgr5-associated marker amenable to immunohistochemistry, was visualized. In wild-type tumors, Sox6 expression was confined to areas of strongest vimentin staining, whereas it was hardly detectable in mutant tumors (Fig. 4B, a versus b).

EMT occurs during development and marks a key step in tumor progression toward metastasis (33). An initial step during EMT, when epithelial cells acquire mesenchymal traits and become more motile, is the breakdown of the basement membrane (35). Immunohistochemical staining of the basement membrane component laminin in wild-type and Bcl9/Bcl9l−/− mutant colon tumors was consistent with the EMT state of wild-type tumors. Thus, virtually no basement membrane laminin was observed in vimentin-positive wild-type tumor epithelium (Fig. 4B, c). In contrast, the epithelium in Bcl9−/−/Bcl9l−/− tumors seemed to be aligned on a contiguous intact laminin membrane (Fig. 4B, d). Concordant observations were made with immunohistochemical staining of collagen IV (data not shown).

Recently, the Prox1 homeobox transcription factor was associated with the acquisition of malignant traits in a similar mouse colon carcinoma model and shown to be directly regulated by β-catenin/Tcf signaling in colorectal cancer (26). The marked difference in Prox1 expression between wild-type and Bcl9−/−/Bcl9l−/− adenocarcinomas (Fig. 3C) was confirmed by immunohistochemical staining (Fig. 4B, e versus f), showing that Prox1 protein expression was largely confined to vimentin-expressing cells and, like vimentin, was expressed only in rare small foci of mutant epithelium (Fig. 4B, g versus h). It is noteworthy that this concomitant expression of vimentin and Prox1 could be observed already in small wild-type tumor lesions but was barely discernible in similar-stage lesions of mutant mice (Supplementary Fig. S3).

Collectively, these findings show that in this tumor model, Bcl9/Bcl9l control a subset of direct Wnt target genes that are involved in the regulation of EMT and stem cell traits.

Discussion

In Drosophila, Lgs loss-of-function alleles resulted in phenotypes characteristic of reduced Wnt pathway activity (3). In the mouse, within the context of the adult intestinal epithelium, ablating the Lgs homologues Bcl9/Bcl9l did not cause anomalies reminiscent of other mouse Wnt pathway mutants, which typically resulted in reduced epithelial proliferation and loss of crypts (36–39). Analysis of Bcl9/Bcl9l−/− mutant colon epithelial cells, however, revealed a marked reduction of markers associated with intestinal stem cells and regulated in part by Wnt signaling (ref. 22; Fig. 1A). The absence of any noticeable effect on epithelial homeostasis, the unaltered expression of the stem cell specifying transcription factor Ascl2 (ref. 22; Supplementary Fig. S4), and the similar incidence of chemically induced colon adenocarcinomas (Fig. 2A), which are likely derived from crypt stem cells (40), suggested that specification of intestinal stem cells was largely unaffected in the absence of Bcl9/Bcl9l. The reduced expression of stem cell markers associated with the regenerative deficiency observed in an ulcerative colitis model (Fig. 1) suggested, however, that their function was impaired.

A relationship between Bcl9/Bcl9l and Wnt-mediated regulation of stem cell traits was further revealed when Wnt target gene expression was comprehensively assessed in a mouse chemical carcinogenesis model of colon

Figure 3. Comparative gene expression profiling in chemically induced wild-type versus Bcl9/Bcl9l−/−–mutant adenocarcinomas. A, unsupervised hierarchical clustering of gene expression profiles from microdissected tumor epithelium of five randomly chosen tumors (two wild-type and three Bcl9/Bcl9l−/− mutant mice) using all gene probe sets. The dendrogram shows a clear separation of the two genotypes. B, unsupervised hierarchical clustering using a set of direct Wnt target genes (n = 24), genes involved in EMT (n = 13), and genes with ≥2-fold enriched expression in Lgr5-positive stem cells isolated from the small intestine (n = 50, ref. 22), WT, wild-type mice; KO, vil-Cre-Bcl9−/−/Bcl9l−/− mice. Heat maps indicate relative differences between single samples and average expression (log 2 scale). DBI, Davies-Bouldin index for intercluster separation (17). Sets of Wnt–, EMT–, and intestinal stem cell–related genes were compared with sets of the same size containing the genes with the highest variation across all samples (DBI in square brackets). A lower DBI value indicates a better intercluster separation (see Materials and Methods). C, expression of Wnt target genes listed in B assessed by qRT–PCR (n = 5 ± SE; P ≤ 0.05, except for CECPL; P > 0.05); expression of EMT–related genes listed in B assessed by qRT–PCR (n = 5 ± SE; P = 0.05, except for CTSG; P > 0.05); expression assessed by qRT–PCR of genes contained within the set enriched ≥2-fold in Lgr5-positive intestinal stem cells (22). For intestinal stem cell–related genes, only genes with significantly reduced expression in Bcl9−/−/Bcl9l−/− compared with wild-type tumors are shown (n = 5 ± SE; P ≤ 0.05). qRT–PCR validations were carried out with cDNA from the same tumor sections as in B.

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adenocarcinoma. Transcriptional profiles analyzed with a set of direct Wnt target genes unequivocally distinguished wild-type and Bcl9/–Bcl9l–mutant tumors. However, the highly significant intercluster separation proved to be due to the downregulation of only some direct Wnt target genes in Bcl9/–Bcl9l–mutant tumors (Fig. 3B). This subset contained genes that are associated with a mesenchymal phenotype [T (brachyury; ref. 30); vimentin (31)], intestinal stem cells (Lgr5; ref. 21), or colon cancer progression (Prox1; ref. 26). In addition to Wnt-regulated mesenchymal markers, the expression of genes characteristic of EMT, including Foxc2, which promotes mesenchymal differentiation during EMT (41), Hmga2, which regulates expression of Snai1 (42), and other EMT transcription factors including Snai2, Twist1, Zeb1/2, as well as the matrix glycoprotein fibronectin, was downregulated in mutant tumors (Fig. 3C). EMT has recently been associated with the generation of cancer cells with stem cell–like properties (34, 43). Strikingly, like in colon epithelium, a gene signature associated with Lgr5-positive intestinal stem cells (22) was strongly attenuated in Bcl9/–Bcl9l–mutant tumors and clearly distinguished wild-type from Bcl9/–Bcl9l–mutant tumors (Fig. 3B). Collectively, these observations indicated that Bcl9/–Bcl9l are critical for regulating a subset of Wnt target genes relevant to controlling EMT- and stem cell–associated traits. In contrast, Wnt-mediated transduction of proliferative signals remained by and large unaffected by the lack of Bcl9/–Bcl9l.

In Drosophila, Lgs (Bcl9/–Bcl9l) acts as a linker to recruit Pygo to the Arm (β-catenin) complex (3). Pygo is an essential transcriptional coactivator of this complex and has been suggested to exert its function through binding to histone H3 tails and histone decoding (44). Recently, Pygo2 has been
reported to control Wnt signaling–dependent expansion of mouse mammary progenitor cells (45). These findings are in line with our observations and suggest a more general role for Bcl9/Bcl9l and Pygo proteins in regulating stem cell–relevant Wnt target gene expression, possibly by maintaining chromatin in a permissive state (2).

Our observations on the role of Bcl9/Bcl9l may prove clinically relevant. EMT is implicated in tumor invasion and metastasis (33). There is growing evidence suggesting that certain chemotherapy-resistant tumor cells display EMT traits and stem cell properties (32, 46–48). In addition to the lack of EMT features and the loss of stem cell–like properties, Bcl9/Bcl9l–mutant tumors showed reduced expression of Wnt-regulated genes that have been directly associated with malignant tumor traits. Thus, Prox1 has been implicated in malignant progression of colon adenocarcinoma, and deletion of Prox1 in intestinal adenomas resulted in phenotypic changes similar to those described herein, such as the preservation of well-organized basement membranes around Prox1-deleted adenomas (26). It is therefore likely that some of the phenotypic changes caused by the loss of Bcl9/Bcl9l may be due to the downregulation of Prox1. Two other Bcl9/Bcl9l–dependent, direct Wnt target genes, Fascin1 (29), a key component of filopodia, and Left1 (49), have been associated with tumor invasion and metastasis in colon or lung cancer. Expression of Bambi, a newly described Wnt-regulated and Bcl9/Bcl9l–dependent transforming growth factor-β inhibitor, predictive of metastatic potential (50), also proved significantly downregulated in Bcl9/Bcl9l–mutant versus wild-type adenocarcinomas (33.6% of mRNA expression = 5; n = 5; P = 0.015). The colon adenocarcinoma model explored herein did not allow assessment of tumor invasiveness and dissemination due to locally advanced growth and precocious intestinal obstruction. Recently, however, RNA interference–mediated silencing of Bcl9 expression in a colon cancer cell line was reported to significantly reduce its metastatic potential (9). Collectively, the virtual abrogation in the absence of Bcl9/Bcl9l of numerous traits associated with malignant tumor progression strongly suggests that targeting Bcl9/Bcl9l function in colon cancers may result in attenuated malignancy, through reduction of their propensity to disseminate and through inhibition of stem cell–associated features, which may restore responsiveness to therapy (32).

These first time observations in loss-of-function mouse mutants indicate that Bcl9/Bcl9l control a subset of Wnt-regulated genes implicated in stem cell control. They warrant further studies to investigate underlying mechanisms involved in selective Wnt target gene regulation as well as to explore the Bcl9/Bcl9l–regulated Wnt subprogram with a view to therapeutically target stem cell–like traits in colon and other Wnt-activated cancers.

**Disclosure of Potential Conflicts of Interest**

No potential conflicts of interest were disclosed.

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**References**


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