**Priority Report**

**Functional MicroRNA Is Transferred between Glioma Cells**

Mark Katakowski¹, Benjamin Buller¹,², Xinli Wang¹, Thomas Rogers¹, and Michael Chopp¹,²

**Abstract**

MicroRNAs (miRNA) are single-stranded 17- to 27-nucleotide RNA molecules that regulate gene expression by posttranscriptional silencing of target mRNAs. Here, we transformed rat 9L gliosarcoma cells to express cel-miR-67, a miRNA that lacks homology in rat. Coculture of these cells with cells that expressed a luciferase reporter that contained a complementary sequence to cel-miR-67 resulted in significant suppression of luciferase expression. This effect was also observed in the U87-MG human glioma cell line. Moreover, luciferase suppression was inhibited by the addition of carbenoxolone to cocultures, suggesting that gap junction communication regulates intercellular transfer of miRNA. Finally, in situ hybridization revealed the presence of cel-miR-67 in cel-miR-67–null 9L cells after coculture with cel-miR-67–expressing cells. Our data show that miRNA transcribed in glioma cells can be transferred to adjacent cells and induces targeted inhibition of protein expression in the acceptor cells. These findings reveal a novel mechanism of targeted intercellular protein regulation between brain tumor cells. Cancer Res; 70(21); 8259–63. ©2010 AACR.

**Introduction**

MicroRNAs (miRNA) are single-stranded 17- to 27-nucleotide RNA molecules that regulate gene expression by posttranscriptional silencing of target mRNAs through complementary binding (1). Each miRNA can affect a number of mRNAs and, depending on its targets, has the potential to function as an oncogene and/or tumor suppressor (2). Recent studies indicate that some tumors, including gliomas, can secrete microvesicles that contain RNA, including miRNA (3). Other cells treated with purified tumor-shed microvesicles take up miRNAs contained within (3, 4). It has also been shown that cardiac myocytes exchange RNA in a process mediated by gap junctions (5). However, it is unknown whether miRNA secreted by tumors, either by microvesicles or otherwise, can regulate protein expression in neighboring cells by inherent mechanisms. Here, we show that miRNAs transcribed in rat gliosarcoma cells are transferred to adjacent cells and that this transferred miRNA induces targeted inhibition of protein expression in the acceptor cells.

**Materials and Methods**

**Cell culture**

The 9L rat gliosarcoma, U87-MG human glioblastoma, and U251 human glioblastoma cell lines were obtained from the American Type Culture Collection (ATCC) in 2003, 2007, and 2000, respectively. Cells were resuscitated and cultured in accordance with ATCC guidelines for less than 4 months before use in these experiments. ATCC performs authentication of all cell lines through short tandem repeat profiling (human cells), karyotyping, and cytochrome c oxidase I testing; we did not reauthenticate these lines in our laboratory. Cells were maintained at 37°C in DMEM containing 10% fetal bovine serum as previously described (6).

**Cell cocultures**

Cells were cultured for 24 hours in 96-well plates. 9L cells were added at 2:1 ratio of *C. elegans* miRNA-expressing cells (1.4 × 10⁴) to luciferase-expressing cells (7 × 10³). For *in situ* hybridization experiments, 4 × 10⁴ *C. elegans* miRNA-expressing cells were cocultured for 24 hours with 2 × 10⁶ enhanced green fluorescent protein (eGFP)–expressing cells in eight-well glass chamber slides.

**Cell transfection**

Cel-miR-67 pMIR-report luciferase and control plasmids (Signosis) and cel-miR-67 and cel-miR-239 expression plasmids (GenScript) were used. Transfection was carried out for 12 hours using Lipofectamine 2000 (Invitrogen) with 4 μg DNA per transfection. Twenty-four hours after transfection, cells were washed three times in PBS, collected by trypsinization, and resuspended in culture medium before use in experiments. Stably transfected cells were isolated under puromycin selection, tested for miRNA expression, and subsequently used in experiments. We verified that only 9L-mir67 cells expressed cel-miR-67 using a cel-miR-67 TaqMan miRNA assay (CT value = 28.7; Applied Biosystems).

**Luciferase assays**

Luciferase activity was determined using a Luciferase Assay System kit (Promega). Luminescence was determined...
on a Fusion plate reader (Perkin-Elmer). For all wells, each experimental group (e.g., 9L-miR-67) was the result of one transfection per experiment. Each luciferase experiment was performed four times, with six or more replicates per group.

Inhibition of gap junctions
To inhibit the function of gap junctions in 9L cells, we incubated the cells with 150 μmol/L carbenoxolone for 7 hours. Carbenoxolone is a broad-spectrum gap junction antagonist, and incubation for 7 hours in 150 μmol/L has been shown to disrupt the function of gap junctions (7).

In situ hybridization and immunostaining
Probes against rno-miR-21 and cel-miR-67 (Exiqon) were hybridized to miRNAs according to a published protocol that was modified for adherent cells (8). Cells were counterstained with a monoclonal antibody against eGFP (Aves Labs). The probe for rno-miR-21 was used as a positive hybridization control because miR-21 is highly expressed in gliomas (9).

Statistical analysis
Data are shown as mean ± SD. P values were calculated using one-way ANOVA or Student’s t test.

Results
Cell-miR-67 is not expressed in rat, mouse, or human and, for this reason, is often used as an inert miRNA negative control in experiments using mammalian cells (10). Using real-time reverse transcription-PCR with a cel-miR-67 specific primer, we confirmed that 9L rat gliosarcoma cells did not express cel-miR-67 miRNA.

To test whether functional miRNA can be transferred from one tumor cell to another, we cocultured 9L cells transfected with an expression vector for cel-miR-67 (9L-mir67) or cel-miR-239 (9L-mir239) or an empty expression vector (9L-mirNeg), with 9L cells transfected with a luciferase reporter encoding for mRNA with a 3′ untranslated region containing a complementary sequence to cel-miR-67 (9L-Luc67). When 9L-mir67 cells were cocultured for 24 hours with 9L-Luc67 cells, we observed a 25% signal attenuation compared with the 9L-mirNeg control group. However, when 9L-mir239 cells were cocultured with 9L-Luc67 cells, luminescence was not significantly altered compared with 9L-mirNeg/9L-Luc67 control. The luciferase activity detected in the 9L-mir67/9L-Luc67 coculture cells was also significantly less than that detected in the 9L-mir239/9L-Luc67 group (Fig. 1A). These data show that the downregulation of luciferase in the 9L-Luc67 cells was due to the presence of cel-miR-67, produced by the 9L-mir67 cells with which they were cocultured.

We next posited that intercellular regulation of luciferase might be dependent on the expression of cel-miR-67 DNA that had not incorporated into the genome. To test this hypothesis, we established, through puromycin selection, 9L cell lines that stably expressed either cel-miR-67 (9L-mir67s) or cel-miR-239 (9L-mir239s), or 9L cells transfected with a luciferase reporter encoding for mRNA with a 3′ untranslated region containing a complementary sequence to cel-miR-67 (9L-Luc67). When 9L-mir67s cells were cocultured for 24 hours with 9L-Luc67 cells, we observed a 13% signal attenuation compared with the 9L-mir239s/9L-Luc67 control group. These data show that cells that have incorporated cel-miR-67 into their genome downregulate luciferase expression in 9L-Luc67 cells when cultured together.

Figure 1. Cel-miR-67–expressing 9L gliosarcoma cells suppress luciferase expression in, and transfer cel-miR-67 to, neighboring cel-miR-67–negative cells by a gap junction–dependent mechanism. A, luciferase detected in 9L-Luc cells cocultured with 9L cells transfected with an empty expression vector (9L-mirNeg) or a vector encoding for cel-miR-67 (9L-mir67) or cel-miR-239 (9L-mir239). n = 4, P = 0.004 (ANOVA). B, luciferase detected in 9L-Luc67 cells cocultured with 9L cells stably expressing cel-miR-67 (9L-mir67s), cel-miR-239 (9L-mir239s), or empty expression vector (9L-mirNegs). n = 4, P = 0.006 (ANOVA). C, luciferase detected in 9L-Luc67 cells cocultured with 9L-mir67s, 9L-mir239s, or 9L-mirNegs in the presence of 150 μmol/L carbenoxolone, a gap junction antagonist. n = 4, P = 0.459 (ANOVA). Bars, SD. Post hoc multiple comparisons are two-tailed t tests.
As mentioned above, there is evidence that transfer of RNA between cardiac myocytes occurs and that it is mediated by gap junctions (5). To determine if transfer of functional miRNA between brain tumor cells is regulated by gap junctions, we inhibited them by incubating the cells with carbadoxolone, a broad-spectrum connexin channel antagonist. The addition of carbadoxolone to 9L cocultures significantly blocked the effect of cel-miR-67-expressing cells on luciferase expression in Luc67-expressing cells (Fig. 1C). These findings indicate that gap junction intercellular communication significantly mediates the transfer of functional miRNA between 9L cells.

To confirm that cel-miR-67 was present in cocultured 9L acceptor cells that did not express cel-mir67, we cocultured 9L-mir67s cells with 9L gliosarcoma cells that express eGFP (9L-eGFP). We used in situ hybridization to visualize cel-miR-67. We observed colocalized green fluorescent protein (GFP) and cel-miR-67 signals in 9L-mir67/9L-eGFP coculture.

Figure 2. Merged fluorescent and phase-contrast images reveal colocalization of cel-miR-67 in situ signal with eGFP 9L cells. A, no detection of cel-miR-67 in 9L-eGFP/9L-mir239 cocultured cells (negative control). B, detection of rno-miR-21 in 9L-eGFP/9L-mir67 cocultured cells (positive control). C and D, detection of colocalized cel-miR-67 and eGFP in 9L-eGFP/9L-mir67 cocultured cells. Red arrowheads indicate positive in situ signal. Bar, 25 μm.
(~75% of GFP-reactive cells were positive for cel-miR-67), but no cel-miR-67 in 9L-mir239/9L-eGFP controls (Fig. 2A–D). These results confirmed the presence of cel-miR-67 in 9L-eGFP acceptor cells. Thus, 9L gliosarcoma cells can transfer functional miRNA between cells, and this miRNA can regulate protein expression in the acceptor cells.

Finally, we tested whether miRNA could be transferred between other glioma cells. To this end, we transfected the human glioma cell lines U87-MG and U251 with the cel-mir67, cel-mir239, or Luc67 plasmid and performed the same coculture experiment. Here, we found that luciferase expression in the U87-MG cells was significantly reduced by coculture with cel-miR-67-expressing U87-MG cells (U87-mir67); however, no effect was observed in the U251 cocultures (Fig. 3A and B). These results indicate that miRNA transfer may occur in human as well as rat glioma cells; however, a lack of luciferase suppression in the U251 cocultures suggests that miRNA transfer may not occur or may be limited in some glioma cell types.

Discussion

Alterations in the expression of miRNA contribute to the pathogenesis of human cancers (11). Dysregulation of miRNAs promotes malignancy of glioblastoma and contributes to cell proliferation, invasion, and angiogenesis and glioma stem cell multipotency and survival (12, 13). Intercellular transfer of RNA was hypothesized as early as 1971 (14). Recent evidence indicates that gliomas can shed microvesicles that contain functional miRNAs, mRNAs, and receptors (3, 15). These microvesicles have been detected in biological fluids including blood, urine, and cerebral spinal fluid (3, 16). Proteins, RNAs, and miRNAs transported by microvesicles are now believed to play a critical role in tumor invasion and metastases (17). Previous studies showed that cells exposed to tumor-shed microvesicles take up and incorporate proteins and nucleic acids contained within (3, 4, 15). These prior experiments provided compelling evidence that intercellular protein regulation through transferred miRNA is possible, if not likely. However, using a vector encoding for alien miRNA, we provide direct evidence that gliosarcomas exchange functional miRNA, and importantly, that this transferred miRNA leads to significant alterations in protein expression in the acceptor cells.

Previously, Valiunas and colleagues showed that oligonucleotides the size of siRNA are permeable to gap junctions (18). More recently, it was shown that cardiac myocytes exchange small RNAs through a gap junction–dependent mechanism (5). These studies are important because they suggest a route of intercellular RNA transfer independent of microvesicles. Our findings indicate that gap junctions mediate the transfer of miRNA between 9L cells. However, it remains to be determined whether miRNAs are transported through these intercellular channels directly or if gap junctions influence processes such as microvesicle release. It is also possible that carbenoxolone affects mechanisms other than gap junctions, and future experiments targeting specific connexins with siRNA could test for this.

An obvious next step will be to determine if tumors can manipulate protein expression in neighboring nontumor cells, which, if true, may contribute to recruitment or transformation of nontumor cells. Interestingly, miRNA transfer either does not occur in U251 cells or elicits an effect below the sensitivity of our assay. This finding raises the possibility that intercellular miRNA transfer varies between glioma cell types. Of note, it has been reported that connexin 43 is highly expressed in cultured U87-MG cells, but not in U251 cells (19). As gap junctions contribute to miRNA transfer, it would be interesting to test if the difference we observed was due to
the differential connexin 43 expression between the two cell lines. Furthermore, we have not established whether cell-to-cell contact is necessary for miRNA transfer or effective suppression of protein expression in the acceptor cells. Nevertheless, this study shows direct and targeted regulation of protein expression between brain tumor cells. These findings have wide-ranging implications for our understanding of tumorigenesis and progression, as well as for the development of miRNA-based antitumor therapies.

Acknowledgments

We thank Zheng Gang Zhang and Feng Jiang for comments and reagents, and Ann Hozeska-Solgot for image analysis.

Grant Support

NIH grant R01 CA129446 and a Henry Ford Hospital Research Proposal Development Program Grant (M. Katakowski).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received 02/18/2010; revised 08/24/2010; accepted 08/27/2010; published OnlineFirst 09/14/2010.

References

Functional MicroRNA Is Transferred between Glioma Cells

Mark Katakowski, Benjamin Buller, Xinli Wang, et al.

Cancer Res 2010;70:8259-8263. Published OnlineFirst September 14, 2010.

Updated version
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-10-0604

Cited articles
This article cites 19 articles, 4 of which you can access for free at:
http://cancerres.aacrjournals.org/content/70/21/8259.full.html#ref-list-1

Citing articles
This article has been cited by 13 HighWire-hosted articles. Access the articles at:
/content/70/21/8259.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.