Enhancement of Cancer Vaccine Therapy by Systemic Delivery of a Tumor-Targeting Salmonella-Based STAT3 shRNA Suppresses the Growth of Established Melanoma Tumors

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Abstract

Cancer vaccine therapies have only achieved limited success when focusing on effector immunity with the goal of eliciting robust tumor-specific T-cell responses. More recently, there is an emerging understanding that effective immunity can only be achieved by coordinate disruption of tumor-derived immunosuppression. Toward that goal, we have developed a potent Salmonella-based vaccine expressing codon-optimized survivin (CO-SVN), referred to as 3342Max. When used alone as a therapeutic vaccine, 3342Max can attenuate growth of aggressive murine melanomas overexpressing SVN. However, under more immunosuppressive conditions, such as those associated with larger tumor volumes, we found that the vaccine was ineffective. Vaccine efficacy could be rescued if tumor-bearing mice were treated initially with \textit{Salmonella} encoding a short hairpin RNA (shRNA) targeting the tolerogenic molecule STAT3 (YS1646-shSTAT3). In vaccinated mice, silencing STAT3 increased the proliferation and granzyme B levels of intratumoral CD4\textsuperscript{+} and CD8\textsuperscript{+} T cells. The combined strategy also increased apoptosis in tumors of treated mice, enhancing tumor-specific killing of tumor targets. Interestingly, mice treated with YS1646-shSTAT3 or 3342Max alone were similarly unsuccessful in rejecting established tumors, whereas the combined regimen was highly potent. Our findings establish that a combined strategy of silencing immunosuppressive molecules followed by vaccination can act synergistically to attenuate tumor growth, and they offer a novel translational direction to improve tumor immunotherapy. \textit{Cancer Res}; 71(12); 4183–91. ©2011 AACR.

Introduction

Survivin (SVN) is a member of the inhibitor of apoptosis protein (IAP) family whose function is involved in prolonging cell survival and cell-cycle control (1, 2). SVN is an ideal tumor-associated antigen (TAA) for therapeutic vaccination because it is overexpressed by essentially all solid tumors and is poorly expressed in normal adult tissues (3). Increased expression of SVN is also observed in endothelial cells during angiogenesis, thereby serving as an additional target for therapy (4). In animal tumor models, downregulation or inactivation of SVN has already been shown to inhibit tumor growth (5–7). Therefore, strategies to boost tumor-specific responses, such as using adjuvants or immunogenic vectors, will be critical to the success of therapeutic vaccination (8, 9).

Even when favorable vaccination conditions are discovered that promote robust tumor-specific immunity, these responses can eventually be compromised by expanding numbers of intratumoral T-regulatory cells (Tregs) and myeloid-derived suppressor cells (10–13). STAT3 has been recognized as an oncogenic transcription factor in myeloid and tumor cells that, when activated, inhibits production of immunostimulatory molecules and promotes expression of immunosuppressive molecules (14–16). A promising approach to inactivating STAT3 is the use of siRNA or short hairpin RNA (shRNA), usually administered intratumorally, alone or conjugated to molecules that target specific cell populations (17, 18). Advances in the generation of attenuated enteric bacterial vectors, such as \textit{Salmonella typhimurium}, facilitate the highly translational tumor-specific delivery of antigens or plasmids (19–21). The vector itself acts as an adjuvant to elicit innate immunity and to aid in generation of adaptive immunity against recombinant antigen. The most common \textit{Salmonella} vaccines employ \textit{Salmonella} pathogenicity 1 (SPI1) type 3

Notes:

Supplementary data for this article are available at Cancer Research Online (http://cancerres.aacrjournals.org/).

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secretion systems (T3SS), which only produce recombinant antigen in a defined time frame as the pathogen penetrates the host cell (22). More advanced vaccine designs utilize SPI2 T3SS, which switches on recombinant antigen production when the *Salmonella* organisms have entered the host cell, allowing for extended antigen production (23). Numerous studies have documented strains that colonize hypoxic regions of solid tumors weeks after i.v. injection, with no detectable bacteria in peripheral organs, making it an ideal delivery vehicle for targeting shRNA therapeutics into solid tumors (24–26).

In this report, we show a novel strategy utilizing 2 therapeutic agents delivered systemically that are inadequate to control tumor growth as single agents but succeed as a combined therapy. Specifically, attenuated *S. typhimurium* organisms carrying either a STAT3-specific shRNA plasmid (YS1646-shSTAT3) or an SVN expression plasmid (3342Max) were administered consecutively and observed to function synergistically leading to effective tumor rejection. The combined approach improves the prospects for successful vaccination against cancer by altering the tumor microenvironment to be less antagonistic to tumor-infiltrating T cells such as those stimulated by vaccine-encoded TAA.

### Materials and Methods

**Animals, tumor lines, and bacterial strains**

C57BL/6 mice (The Jackson Laboratory, 6–8 weeks) were obtained from breeding colonies housed at the City of Hope (COH) Animal Research Center. The B16F10 murine melanoma line was a kind gift from Drs. Hua Yu and Marcin Kortylewski (COH). Animals, tumor lines, and bacterial strains

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**Salmonella SPI2 expression vectors, shRNA plasmids, and generation of recombinant *Salmonella***

pWSK29 constructs containing the SPI2 expression cassette for LisA (2810) or SVN (3342) are described elsewhere (23). For construction of pWSK29 encoding *Salmonella* codon-optimized survivin (CO-SVN), 2810 was digested with *XbaI*/EcoRV, and the gel-purified pWSK29 backbone was used to clone the CO-SVN gene (Genscript) engineered with *XbaI*/EcoRV sites for in-frame fusion with the *sseF* gene. shRNA constructs against STAT3 (Origene) were tested for silencing by stable transfection of B16F10 cells followed by Western blot analysis using polyclonal rabbit antibody against STAT3 (Santa Cruz Biotech). The pGFP-V-RS vector containing the 29-mer shRNA sequence ACCTGAAAGAAGGTTCAATCGTGTGAGCA (ID: G556360) exhibited more than 70% STAT3 knockdown and was selected for generation of recombinant YS1646. SPI2 expression vectors and shRNA plasmids were electroporated into MVP728 or YS1646, respectively, with a BTX600 electroporator (BTX).

**Western blot analysis**

Western blot analysis for *Salmonella* expression of SVN was carried out as described previously (23). Briefly, 3342 and 3342Max were grown overnight in MOPS-based media (Sigma) at 37°C containing either low phosphate (113 μmol/L) to induce SPI2 expression or high phosphate (25 mmol/L). Bacterial pellets were boiled in SDS loading buffer and equal amounts of lysate were loaded. Blots were probed using a monoclonal rabbit antibody (ab76424) against SVN (Abcam).

**Tumor challenge, vaccination, and shRNA therapy**

For tumor challenge, 10⁵ B16F10 cells were injected s.c. into C57BL/6 mice. Tumor growth was monitored daily or every other day using a caliper. For testing vaccination alone, MVP728 carrying 2810, 3342, or 3342Max were administered by gavage twice, 4 days apart, when tumors reached 3.5 to 4 mm in diameter at 10⁶ colony-forming units (CFU). For combined therapy, PBS, YS1646-STAT3, or YS1646-scrambled was first injected at 10⁷ CFU in C57BL/6 mice when tumor volumes were 50 mm³ or above (7–8 mm in diameter) followed by gavage with PBS or 10⁷ CFU MVP728-2810 or MVP728-3342Max.

**Quantitative PCR for detection of STAT3 levels**

Mice bearing B16F10 tumors (≥50 mm³) were i.v. injected with 10⁷ CFU of PBS, YS1646-shSTAT3, or YS1646-scrambled twice, 4 days apart. At days 3, 7, and 10, mice (n = 3) were sacrificed and RNA was extracted from tumor homogenates for generation of single-stranded cDNA (Fermentas). To quantitate STAT3 levels, SYBR Green quantitative PCR (qPCR) analysis (BD Biosciences) using primers specific for STAT3 (forward: 5’-CATGGGCTATAAGCATCATGGTGCGAC-3’, reverse: 5’-AGGGCTCGACAGCCTTACCGGTATTTC-3’) was carried out using GAPDH (glyceraldehyde-3-phosphate dehydrogenase; forward: 5’-CAAGGTATCCATGACACTTTG-3’, reverse: 5’-GTCACCACACCTTGTTGCTTAG-3’) for normalization.

**Immunofluorescence staining**

For detection of intracellular SVN expression from recombinant *Salmonella*, RAW264.7 macrophages seeded on coverslips were infected for 30 minutes at a multiplicity of infection (MOI) of 10 with wild-type MVP728, 3342, or 3342Max. Cells were incubated overnight in DMEM-10 containing 10 μg/mL gentamicin. Cells were fixed/permeabilized with 1:1 acetone/methanol and stained with conjugated antibodies fluorescein isothiocyanate (FITC)-LPS (Santa Cruz Biotech) and phycoerythrin (PE)-HA (Covance) overnight at 4°C followed by 4’6-diamidino-2-phenylindole (DAPI). Cells were imaged on an Axiovert 200 using live imaging software (Axiovision). Images shown are representative of cells observed within multiple fields.

**Flow cytometry**

Conjugated monoclonal antibodies (mAb) directed to PE-Cy7-CD8, PerCP-CD45, and PE-phospho-STAT3 were purchased from BD Pharmingen and mAbs to APC-Cy7-CD4, APC-F4/80, FITC-Ki-67, PE-granzyme B, and FITC-Annexin
V were purchased from eBioscience. Intracellular phospho-STAT3, granzyme B, Ki-67, and Annexin V staining were carried out following the manufacturer’s protocol (eBioscience). Samples were run on a FACS-Canto (Becton Dickinson) and analyzed using FlowJo software (TreeStar).

Cytotoxicity assay
Cytotoxicity against B16F10 melanoma cells in treated mice was determined using a standard 51Cr release assay (27). Briefly, effectors were derived from spleens of B16F10-bearing C57BL6 mice (n = 4) and i.v. injected with PBS, 107 CFU of YS1646-shSTAT3 or YS1646-scrambled followed by gavage with PBS or 107 CFU of 3342Max or 2810 4 days later. Mice were sacrificed about 1 week postgavage and splenocytes were cocultured with RMA-S cells loaded with human SVN library (27). Effectors were then cocultured for 4 hours with 5,000 51Cr-loaded B16F10 targets in 96-well plates at ratios of 100:1, 20:1, and 4:1 (in triplicate). Radioactivity released into the supernatant was measured using a Cobra Quantum gamma counter (PerkinElmer). Percent specific lysis = (experimental release–spontaneous release)/(maximum release–spontaneous release) × 100.

Statistical analysis
Statistical significance for comparisons among 2 or more groups was calculated with the GraphPad Prism Software v4.03 using the Student t test or 1-way ANOVA, respectively. A value of P < 0.05 was considered statistically significant. All experiments were typically carried out at least in duplicate, and all data are presented as mean ± SEM. P-values are as shown in Figs. 2–5: *, P < 0.05; **, P < 0.01; and ***, P < 0.001.

Results

Construction and evaluation of SVN expression vectors
Previous work using the MVP728 bacterial vector transformed with plasmid 3342, which expresses SVN, showed partial success in rejecting murine models of colon carcinoma and glioblastoma (23). We found that SVN expression from 3342 was suboptimal when compared with LisA expression from 2810 (data not shown). We hypothesized that codon optimization of SVN to Salmonella-preferred codons would allow for increased stability and protein expression leading to greater antitumor effects (28, 29). To test this hypothesis, a S. typhimurium CO-SVN sequence was generated using an online algorithm (30) and then synthesized (Genscript). As shown in Fig. 1A, the low copy plasmid pWSK29 was engineered to encode the SPI2 chaperone protein sscB (31) and sseF fused to LisA (2810), SVN (3342), or CO-SVN (3342Max) for expression and secretion by MVP728. Ultimately, expression of these genes would be regulated by the SPI2-specific promoter for sseA.

We determined whether CO actually increased SVN expression by growing the recombinant Salmonella in SPI2-inducing conditions (ref. 32; Fig. 1B). Under noninducing conditions (PCN – P), we found no significant expression of SVN. Surprisingly, under inducing conditions (PCN + P), we observed much greater SVN expression from 3342Max than the nonoptimized 3342. To further evaluate SVN expression and secretion by 3342Max, we infected RAW264.7 murine macrophages with either 3342 or 3342Max to determine intracellular expression of SVN by immunofluorescence. As shown in Fig. 1C (HA-survivin panel), we observed greater SVN expression (characterized by more foci) than 3342. As expected, mAb staining for both the lipopolysaccharide (LPS; LPS-St panel) and HA (HA-survivin panel) localized to the cytoplasm and overlapped in the Merge panel, independent of the nuclear DAPI staining. These data suggest that

Figure 1. Construction and validation of SVN expression vectors. A, the expression vectors 2810, 3342, and 3342Max were constructed to encode HA-tagged LisA, SVN, or SVN codon optimized for Salmonella (CO-SVN), respectively, using the low copy plasmid backbone pWSK29. Each of these proteins was fused to the SPI2 protein sseF and its expression is dependent on the SPI2 promoter sseA. Each construct was then electroporated into MVP728 (23). B, SVN expression from MVP728 harboring 3342 and 3342Max (Max) constructs was detected by Western blotting of bacterial lysates cultured in inducing conditions (low phosphate media, PCN – P) or noninducing conditions (high phosphate media, PCN + P). Fusion protein was detected using anti-SVN antibody. C, RAW264.7 cells infected with MVP728 alone, MVP728-3342, or MVP728-3342Max were fixed and permeabilized and then stained with the conjugated antibodies LPS-FITC, HA-PE, and the nuclear stain DAPI. Cells were imaged under 100 × oil immersion using an Axiovert 200. Scale bars, 5 μm.
optimization of SVN tailored to preferred Salmonella codons greatly improves recombinant antigen expression.

**CO-SVN enhances suppression of tumor growth**

We next evaluated whether enhanced expression of SVN by 3342Max translated into a more efficacious vaccine using a B16F10 melanoma, which naturally overexpresses SVN (inset of Fig. 2A). Subcutaneously injected tumor cells were allowed to grow until a palpable tumor was present, generally 3.5 to 4 mm in diameter (<10 mm³). Mice were then gavaged twice (4 days apart) with either PBS or MVP728 harboring 2810, 3342, or 3342Max constructs. As shown in Fig. 2A, 3342Max vaccination was superior to all other experimental treatments in attenuating tumor growth (P < 0.01). We then determined the lymphocyte subsets that were most responsible for the attenuation by carrying out in vivo antibody depletions of CD8⁺, CD4⁺, or natural killer cell (NK) populations in tumor-bearing mice vaccinated with 3342Max as in Fig. 2A. We observed that depletion of CD8⁺ T lymphocytes resulted in significant loss of tumor growth control with an intermediate effect of NK depletion, which has been described previously (17). These data suggest that vaccination with 3342Max elicits superior CD8⁺ T-cell responses that limit tumor growth, likely a result of enhanced SVN expression.

**Attenuation of STAT3 mRNA levels using shRNA**

When s.c. B16F10 tumors were grown to larger volumes before treatment (7–8 mm in diameter, ≥50 mm³), we discovered that 3342Max vaccination had no efficacy to attenuate growth (data not shown). Because we showed that 3342Max works efficiently in less demanding circumstances (Fig. 2), we presumed that failure under more stringent conditions was likely the result of greater levels of tumor-derived immunosuppression (33). To determine whether we could rescue the efficacy of the vaccine, we sought to manipulate the tumor microenvironment by silencing the tolerogenic molecule STAT3 (15, 34, 35). We chose to inactivate STAT3 mRNA expression using an shRNA expression plasmid carried by the tumor-targeting *Salmonella* strain YS1646 (36). We first tested several commercially available shRNA plasmids (Origene) to silence the expression of STAT3 in stably transfected B16F10 tumor lines. As shown in Fig. 3A, shSTAT3#60 showed dramatic silencing (>70%) of endogenous STAT3 when compared with scrambled shRNA control plasmid. Other shSTAT3 plasmids had intermediate to no effect on endogenous STAT3 expression.

**Targeted silencing of STAT3 combined with 3342Max results in significant suppression of tumor growth in a more advanced melanoma tumor model**

We next generated YS1646 carrying the shSTAT3#60 plasmid (YS1646-shSTAT3) to test whether systemic delivery of *Salmonella* by i.v. could silence STAT3 expression *in situ* in the tumor. Mice bearing s.c. B16F10 tumors (≥50 mm³) were injected twice i.v. with 10⁷ CFU of YS1646-shSTAT3, YS1646-scrambled, or PBS 4 days apart. Posttreatment, no significant attenuation of tumor growth was observed for mice treated with YS1646-shSTAT3 alone compared with control groups (data not shown). These same results were also observed in less stringent conditions where initial tumor volumes were less than 10 mm³ (data not shown). Nonetheless, tumors were isolated, homogenized, and total RNA was extracted for quantitative PCR (qPCR). Surprisingly, there was significant silencing of STAT3 3 days after YS1646-shSTAT3 administration compared with mice administered YS1646-scrambled or PBS (Fig. 3B). STAT3 silencing continued to increase on day 7 of the shSTAT3 group, which is consistent with the continued effectiveness of the therapeutic strategy (see below). On day 10, STAT3 silencing moderated but was still lower than the control groups. Confirmation that YS1646-shSTAT3 succeeded in specifically silencing STAT3 mRNA but failed to reject tumors as a single agent motivated us to combine delivery of shSTAT3 and 3342Max vaccination in mice with significantly larger B16F10 tumors. Therefore, mice bearing B16F10 tumors of size 50 mm³ or greater were i.v. injected.
Day 7

Day 10

12

8

4

16

vaccination is a powerful synergistic approach to attenuate
results suggest that combining shSTAT3 therapy and SVN
tumor growth significantly better than control groups. These
3342Max rescues the activity of the vaccination to attenuate
þ
2810, or PBS. As shown in Fig. 3C, the combination of shSTAT3
3, 7, or 10 after first injection and tumor lysates were subjected to RNA
expression in B16F10 tumor following i.v. injection of YS1646-shSTAT3.
Western blotting of STAT3 protein expression from B16F10 lysates after
stable transfection of shRNA constructs (58–61) with potential for silencing
3) were sacrificed on days
61) with potential for silencing
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with 10^7 CFU of YS1646-shSTAT3, YS1646-scrambled, or PBS.
Four days later, mice were gavaged with 10^7 CFU of 3342Max,
2810, or PBS. As shown in Fig. 3C, the combination of shSTAT3 + 3342Max rescues the activity of the vaccination to attenuate
tumor growth significantly better than control groups. These
results suggest that combining shSTAT3 therapy and SVN vaccination is a powerful synergistic approach to attenuate
tumor growth.

Figure 3. Targeted silencing of STAT3 using YS1646-shSTAT3 results in significant suppression of tumor growth when combined with 3342Max. A, Western blotting of STAT3 protein expression from B16F10 lysates after stable transfection of shRNA constructs (58–61) with potential for silencing
3) were sacrificed on days
was used as a loading control. B, silencing of STAT3 expression in B16F10 tumor following i.v. injection of YS1646-shSTAT3. Mice bearing palpable B16F10 tumors were i.v. injected with 10^7 CFU of YS1646-shSTAT3 twice, 4 days apart. Mice (n = 3) were sacrificied on days
3, 7, or 10 after first injection and tumor lysates were subjected to RNA extraction for qPCR analysis of STAT3 transcripts. C, YS1646-shSTAT3 rescues antitumor effects of MVP728-3342Max in B16F10 model. C57BL/6 mice bearing B16F10 tumors (≥50 mm³) were treated with PBS, YS1646-shSTAT3, or YS1646-scrambled by i.v. injection. Four days following treatment, mice were vaccinated with PBS, MVP728-2810, or MVP728-3342Max and then monitored for tumor growth.

Decreased phosho-STAT3 levels are observed in tumor
cells following shSTAT3 and 3342Max treatment

We hypothesized that the success of the combined treatment was, in part, due to suppression of phospho-STAT3 levels in specific immune populations. Therefore, we used flow cytometry to determine the levels of activated STAT3 in specific immune subsets present in the tumor following treatment. We found no significant changes in phospho-STAT3 levels for CD4^+ , CD8^+ , CD11c^+ , or Gr1^+ CD11b^+ cells in all treatment groups (Supplementary Fig. S1). However, we did observe significantly decreased phospho-STAT3 levels (P < 0.05) in F4/80^+ macrophages for the shSTAT3 + 3342Max–treated group (Fig. 4A). Surprisingly, no significant decreases of phospo-STAT3 were observed for the shSTAT3 + 2810 group. These results suggest that only the shSTAT3 + 3342Max treatment is able to prevent activation of STAT3 in the F4/80^+ subset, likely a result of early STAT3 silencing followed by tumor growth control, whereas shSTAT3 + 2810 is unable to do so regardless of early STAT3 silencing due to uncontrolled tumor growth.

Combined shSTAT3 and 3342Max administration enhances infiltration of T lymphocytes

Because it was known that ablation of STAT3 increases intratumoral immune function (14, 17, 35), we first examined the frequency and functional status of intratumoral CD4^+ and CD8^+ T cells in vaccinated mice. The percentage of B16F10 intratumoral CD4^+ and CD8^+ T cells was statistically greater in mice treated with shSTAT3 + 3342Max than in the scrambled + 3342Max or shSTAT3 + 2810 treatment groups (Fig. 4B). We next evaluated the proliferative index of these intratumoral CD4^+ and CD8^+ T cells by determining Ki-67^+ expression levels. Both CD4^+ and CD8^+ populations expressed higher levels of Ki-67^+ in the shSTAT3 + 3342Max group than in control groups (Fig. 4C and D). The markedly higher proliferation potential suggests that the combined shRNA and vaccination treatments allow for a proliferative expansion of intratumoral T cells, and the increased frequency may therefore not solely be explained by a redistribution of existing T cells from other sites.

YS1646-shSTAT3 enhances tumor-specific cytotoxic responses and tumor cell apoptosis

We addressed tumor cell death by evaluating the extent of apoptosis using Annexin V staining of gated CD45^− cells, mainly tumor cells (37), from all of the treatment groups. The CD45^− cells revealed significantly higher apoptotic frequencies in mice treated with shSTAT3 + 3342Max than the control groups (Fig. 5A). The increased apoptosis of tumor cells could either be explained by the cytotoxic activity of immune cells or possibly by a shSTAT3-based mechanism to enhance apoptotic signal transduction. To address immune
Based on the results presented, it is evident that targeted silencing of STAT3 using YS1646-shSTAT3 significantly suppresses tumor growth when combined with 3342Max. Flow cytometry analysis revealed a notable decrease in phospho-STAT3 levels in F4/80^+ macrophages following combined treatment, indicating a potential mechanism to prevent STAT3 activation. Furthermore, the frequency and functional status of intratumoral CD4^+ and CD8^+ T cells were enhanced, suggesting improved infiltration of T lymphocytes. The combined therapy also showed a marked increase in Ki-67^+ expression levels within CD4^+ and CD8^+ T cells, indicating better proliferation. Additionally, the increased apoptosis of tumor cells in the combined treatment group points to a possible immune-mediated mechanism or shSTAT3-based enhancement of apoptosis. These findings collectively support the synergistic therapeutic potential of combining shSTAT3 and 3342Max for the effective treatment of B16F10 tumor models.
expressing granzyme B in the mouse group treated with shSTAT3 + 3342Max was dramatically higher than both control groups. These results suggested a potential cytotoxic mechanism for tumor growth control, which we further assessed using a direct in vitro cytotoxicity assay.

Tumor-specific cytotoxicity contributes to control of established subcutaneous B16F10 tumors

We determined whether T cells obtained from B16F10 tumor–bearing mice treated with shSTAT3 + 3342Max possessed functional capacity to kill survivin-expressing tumor cells in vitro by conducting a chromium release assay (Fig. 5C). Splenocytes harvested from B16F10 tumor–bearing mice (n = 4) treated as in Fig. 5A were in vitro stimulated with a human SVN peptide library, then evaluated for in vitro cytotoxic recognition and killing of chromium-loaded B16F10 tumor targets. Mice treated with either scrambled + 3342Max or shSTAT3 + 2810 could not effectively kill B16F10 tumor cells. In contrast, splenocytes from all mice receiving shSTAT3 + 3342Max treatment were very effective at killing B16F10 tumor targets (0.001 < P < 0.01) at all effector ratios (Fig. 5D). These results further suggest that a potential mechanism of tumor growth attenuation is by tumor-specific T cells, stimulated through SVN vaccination or SVN peptide stimulation, directly killing tumor cells, though only when mice are pretreated with shSTAT3 and then vaccinated with 3342Max.

Discussion

The goal of these studies was to discover a translational approach that would provide durable control of solid tumor growth. Our initial hypothesis was that SVN as a ubiquitously expressed TAA would provide the widest versatility for vaccination. In contrast to Salmonella-based SVN vaccines used in previous studies, which have been relatively ineffective when used alone, the goal was to find a vaccine strategy that would not require additional cytokine or chemokine components for effectiveness (8, 9, 23). A simplified one-component regimen based on Salmonella delivery of SVN was the initial goal of this study. Our use of oral systemic administration of Salmonella transformed with SVN expression plasmids was similar to other reports describing Salmonella routes of administration. The advantage to this approach is that Salmonella species are efficiently recognized by antigen-processing macrophages in the gut or other mucosal sites (38, 39).

Initial trials using 3342 were only partially successful against small tumors that had just a few days to develop vascularization (Fig. 2).
levels of SVN from 3342 and bacterial LisA from 2810, we discovered significantly lower levels of SVN by Western blot analysis compared with the bacterial LisA protein under identical conditions (data not shown). Because the bacterial LisA protein was so heavily expressed in Salmonella, we theorized that changing the sequence of SVN from human to S. typhimurium–preferred codons might achieve the same goal. In preliminary experiments, we found a gradation of effectiveness against growth of established s.c. tumors dependent on expression levels (data not shown). The predictability of the increasing effectiveness to reject established tumors made it unnecessary to continue to simultaneously evaluate all forms, and the most effective form (3342Max) was exclusively used in all further comparisons. Ultimately, we found that control of B16F10 tumor growth using 3342Max vaccination only worked shortly after tumor challenge, when tumors became palpable. Upon treatment of mice with larger B16F10 tumors of volumes 50 mm³ or above, the vaccine was unable to attenuate tumor growth. This was not surprising, as immunosuppressive mechanisms likely became more established within the growing tumor, and the single-modality vaccination had no means to overcome them (11, 40).

Despite advances in therapeutic vaccines, there now exist numerous studies that support the idea of tumor-derived immunosuppression contributing to tumor evasion (41). These mechanisms include the secretion of TGF-β or interleukin (IL)-10 leading to T-helper 2 (Th2) cell polarization (42–44) or production of indoleamine 2,3-dioxygenase (IDO)

Figure 5. YS1646-shSTAT3 enhances tumor-specific cytotoxic responses. B16F10 tumor–bearing mice (n = 5) received combined treatment as described in Fig. 4. A, individual histograms of FITC-Annexin V–stained tumor homogenates for a representative mouse from each treatment group. B, tumor homogenates (n = 5) from each group were stained with FITC-Annexin V and analyzed by flow cytometry. MFI of Annexin V represents cells gated from total tumor CD45⁻ cells. Error bars represent SEM. C, tumor homogenates (used in A) were stained with PE-granzyme B and PeCy7-CD8 and then analyzed by flow cytometry. Data represent mean percentages of granzyme B⁻CD8⁺ cells of total CD8⁺ cells. D, splenocytes from mice in A (n = 4) were isolated to generate effectors for use in a chromium release assay against B16F10 targets. To generate effectors, splenocytes were incubated for 7 days with RMA-S cells initially loaded with total human SVN library (15 mers, overlapping by 11). Effectors were then incubated in a 4-hour ⁵¹Cr release assay with ⁵¹Cr-loaded B16F10 targets at E:T (effector:target) ratios of 100:1, 20:1, and 4:1, in triplicate. Percent specific lysis = (experimental release − spontaneous release)/(maximal release − spontaneous release) × 100.
by myeloid cells to induce the generation of Tregs and T-cell anergy (45, 46). The novel mechanisms by which STAT3 causes immunosuppression are just beginning to be unraveled (14, 15, 47). With its other roles in tumor progression, STAT3 has become a multifaceted target with the potential to attenuate tumor growth on its own or enhance the antitumor effects of any immunotherapy. Disrupting tumor-induced immunosuppression has generally been studied in the context of its effects that are independent of antigen-specific vaccination. Only rarely has the combination of disrupting immunosuppressive mechanisms within tumors been combined with vaccination to limit tumor growth. A more recent study has examined tumor-associated stromal cells expressing fibroblast activation protein-α (FAP) as a source of immunosuppression in a model of pancreatic ductal adenocarcinoma (48). Administration of a therapeutic vaccine in the absence of FAP-expressing stromal cells showed significant increases in hypoxia-induced tumor necrosis when compared with FAP+ mice. Similarly, we have also shown modest additive effects when vaccination is combined with the drug gemcitabine (27), whereas others reported increased anti-tumor responses by inhibiting the tolerogenic molecule IDO (49). These studies emphasize that successful outcomes of immunotherapy will likely require overcoming tumor-induced immunosuppression.

In several genetic models of conditional STAT3 deletion, subsequent immune enhancement enabled dramatic inhibitory effects on tumor growth. Mechanism-based studies revealed changes in cytokine profile, T-cell subsets, and signal transduction modifiers that all contributed to the blunting of tumor growth as a result of a reduction or elimination of STAT3 expression (14, 34, 35). These elegant studies have led to preliminary therapeutic strategies employing small molecule inhibitors and RNA interference (RNAi) by a variety of approaches that have in common direct intratumoral administration. These approaches have shown moderate efficacy, but in every case, there is tumor breakthrough within 20 to 25 days postadministration. An alternative strategy has been the approach of tumor-targeting Salmonella delivery of shRNA eukaryotic expression plasmids by i.v. injection. In contrast to our findings, others have found efficacy through intratumoral administration of shSTAT3 alone (18). Nonetheless, the growth attenuation was transient and its translational potential, as an intratumoral therapy, remains in doubt. Although systemic administration of YS1646-shSTAT3 may require more diligent efforts to determine its specific cell targets, this obstacle does not detract from the translational feasibility of the approach for treatment of solid tumors. Our Salmonella approach and work published by others was similar in that the STAT3-specific shRNA sequence only had a single target. In contrast, a CpG DNA chimera with an RNAi sequence that was administered intratumorally had multiple off-target sequences above 100 in the mouse genome that temper the interpretation of the results (17).

We show for the first time that an i.v. administered shRNA against STAT3 acts synergistically with an oral Salmonella-based vaccine against SVN in a therapeutic setting, resulting in suppression of s.c. B16F10 melanoma growth. We conclude that the in vivo suppression of B16F10 tumor growth is the result of increased tumor cell apoptosis, as determined by Annexin V staining, possibly caused by an increased level of tumor-specific CD8+ T cells within the tumor. We saw no changes in tumor-expressed SVN during the treatments (Supplementary Fig. S2) that might explain the increase in apoptosis or eventual escape from control (50). The higher Ki-67+ levels also indicated that these intratumoral T cells were actively proliferating, thereby supporting the notion that shRNA against STAT3 attenuated immunosuppression within the tumor microenvironment. Moreover, the fact that neither the vaccine nor shRNA against STAT3 alone was effective to control tumor growth suggests that the combined treatments acted synergistically. These data support that implementing successful immunotherapy may be futile without a receptive tumor microenvironment generated through additional modalities such as shRNA to inhibit immunosuppression.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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Enhancement of Cancer Vaccine Therapy by Systemic Delivery of a Tumor-Targeting *Salmonella*-Based STAT3 shRNA Suppresses the Growth of Established Melanoma Tumors

Edwin R. Manuel, Céline A. Blache, Rebecca Paquette, et al.

*Cancer Res* 2011;71:4183-4191. Published OnlineFirst April 28, 2011.

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