

## Pivotal Role of Innate and Adaptive Immunity in Anthracycline Chemotherapy of Established Tumors

Stephen R. Mattarollo<sup>1,3</sup>, Sherene Loi<sup>4</sup>, Helene Duret<sup>1</sup>, Yuting Ma<sup>5</sup>, Laurence Zitvogel<sup>5,6,7</sup>, and Mark J. Smyth<sup>1,2</sup>

### Abstract

We show, in a series of established experimental breast adenocarcinomas and fibrosarcomas induced by carcinogen *de novo* in mice, that the therapeutic efficacy of doxorubicin treatment is dependent on CD8 T cells and IFN- $\gamma$  production. Doxorubicin treatment enhances tumor antigen-specific proliferation of CD8 T cells in tumor-draining lymph nodes and promotes tumor infiltration of activated, IFN- $\gamma$ -producing CD8 T cells. Optimal doxorubicin treatment outcome also requires both interleukin (IL)-1 $\beta$  and IL-17 cytokines, as blockade of IL-1 $\beta$ /IL-1R or IL-17A/IL-17R $\alpha$  signaling abrogated the therapeutic effect. IL-23p19 had no observed role. The presence of  $\gamma\delta$  T cells, but not  $\text{J}\alpha 18^+$  natural killer T cells, at the time of doxorubicin treatment was also important. In tumor samples taken from breast cancer patients prior to treatment with anthracycline chemotherapy, a correlation between CD8 $\alpha$ , CD8 $\beta$ , and IFN- $\gamma$  gene expression levels and clinical response was observed, supporting their role in the therapeutic efficacy of anthracyclines in humans. Overall, these data strongly support the pivotal contribution of both innate and adaptive immunity in treatment outcomes of anthracycline chemotherapy. *Cancer Res*; 71(14); 4809–20. ©2011 AACR.

### Introduction

Chemotherapy treatment in cancer has long been associated with development of systemic immunosuppression (1, 2). However, more recent studies have reported augmentation of antitumor immune responses by chemotherapy (reviewed in ref. 3). The immunostimulatory properties of chemotherapy have been associated with increased expression of "stress-like" molecules by damaged or dying tumor cells, such as killer cell lectin-like receptor sub-family K ligands, Fas, and tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) death receptors and HSPs, that render cells receptive to direct attack by cells of the innate immune system; including natural killer (NK) cells, natural killer T cells (NKT cells), and  $\gamma\delta$  T cells (reviewed in ref. 4). Until recently, the mechanisms by which chemotherapy treatment facilitated an adaptive, tumor-

specific immune response were largely uncharacterized. Certain alarmins or danger-associated molecular patterns are expressed or released during tumor cell death induced by immunogenic chemotherapeutic agents, including anthracyclines. These include translocation of the endoplasmic reticulum resident calreticulin complex to the plasma membrane, providing an "eat-me" signal (5–7) and release of the nuclear alarmin HMGB1 to engage TLR-4 on dendritic cells (DC; refs. 8, 9). In addition, we showed recently that dying tumor cells release ATP, which then acts on P2X7 purinergic receptors from DCs and triggers the NOD-like receptor family, pyrin domain-containing-3 protein (NLRP3)-dependent caspase-1 activation complex (inflammasome), allowing for the secretion of proinflammatory interleukin (IL)-1 $\beta$ . Accordingly, anticancer chemotherapy turned out to be inefficient against tumors established in purinergic receptor *P2rx7*<sup>-</sup> or *Nlrp3*<sup>-</sup> or *Casp1*-deficient hosts (10).

Anthracyclines have been used in anticancer treatment for more than 40 years and now have a major role in the management of leukemia, lymphoma, uterine, ovarian, sarcoma, and breast malignancies. Doxorubicin became the most widely used anthracycline because of lower toxicity and potent antitumor activity against solid tumors. Although there were earlier reports evidencing the immune system contribution to doxorubicin-mediated antitumor effects (11, 12), Maccubbin and colleagues were the first to show that doxorubicin was an effective immunomodulator capable of boosting CTL responses (13). A multitude of studies investigating combination treatment of doxorubicin and immunotherapies followed; however, the underlying mechanisms of how immunogenic cell death caused by doxorubicin links to induction of a cytotoxic CD8 T-cell response were largely unknown.

**Authors' Affiliations:** <sup>1</sup>Cancer Immunology Program, Peter MacCallum Cancer Centre, East Melbourne; <sup>2</sup>Department of Pathology, University of Melbourne, Melbourne, Victoria; <sup>3</sup>The University of Queensland Diamantina Institute, Princess Alexandra Hospital, Woolloongabba, Queensland, Australia; <sup>4</sup>Breast Cancer Translational Research Laboratory JC Heuson, Jules Bordet Institute, Brussels, Belgium; <sup>5</sup>INSERM U1015 and <sup>6</sup>Center of Clinical Investigations in Biotherapies of Cancer CICBT 507, Institut Gustave Roussy, Villejuif; and <sup>7</sup>Faculté de Médecine de l'Université Paris-Sud XI, Le Kremlin-Bicêtre, France

**Note:** Supplementary data for this article are available at Cancer Research Online (<http://cancerres.aacrjournals.org/>).

**Corresponding Author:** Mark J. Smyth, Cancer Immunology Program, Peter MacCallum Cancer Centre, Locked Bag 1, A'Beckett Street, East Melbourne, Victoria 8006, Australia. Phone: 61-396-563-728; Fax: 61-396-561-411; E-mail: mark.smyth@petermac.org

doi: 10.1158/0008-5472.CAN-11-0753

©2011 American Association for Cancer Research.

Using experimental and carcinogen-induced mouse models of breast cancer and fibrosarcoma, respectively, here we show key cytokines and immune cell populations required for the antitumor therapeutic efficacy of doxorubicin. We show that CD8 T cells and IFN- $\gamma$  are critical effectors of IL-1 $\beta$ - and IL-17-dependent signaling in response to doxorubicin-treated breast tumors and sarcomas, and corroborating this, we also report that CD8 $\alpha$ , CD8 $\beta$ , and IFN- $\gamma$  gene expression levels in breast cancer patients treated with anthracycline chemotherapy correlate with treatment response.

## Materials and Methods

### Mice

Inbred wild-type (WT) C57BL/6 and BALB/c mice, and OT-I mice carrying a MHC class I-restricted transgenic T-cell receptor (TCR) for the ovalbumin (OVA<sub>257-264</sub>; SIINFEKL) peptide were obtained from Walter and Eliza Hall Institute (WEHI; Parkville, Australia). C57BL/6 gene-targeted knock-out IFN- $\gamma$ <sup>-/-</sup>, IL-1R<sup>-/-</sup>, IL-17A<sup>-/-</sup>, TCR $\delta$ <sup>-/-</sup>, J $\alpha$ 18<sup>-/-</sup>, IL-23p19<sup>-/-</sup>, and IL-12p35<sup>-/-</sup> mice were bred and maintained at the Peter MacCallum Cancer Center (Peter Mac; East Melbourne, Australia) as previously described (14, 15). TCR $\delta$ <sup>-/-</sup> mice were obtained from I. Frazer (University of Queensland, Australia) and IL-17A<sup>-/-</sup> mice were kindly provided by Y. Iwakura (University of Tokyo, Japan). Mice of 6 to 14 weeks of age were used in all experiments, and all procedures were approved by the Peter Mac Animal Ethics Committee.

### Tumor models

**Experimental.** C57BL/6-derived AT3 (kindly provided by Trina Stewart, Peter Mac) and EO771 (16) and BALB/c-derived H2N100 mammary adenocarcinoma (17) lines, and BALB/c-derived MCA2 (derived in-house from mice with an MCA-induced tumor) and C57BL/6-derived MCA205 (provided by L.Z.) fibrosarcoma lines, were all maintained in RPMI 1640 supplemented with 10% fetal calf serum (FCS). The OVA-expressing AT3 line (AT3<sup>OVA</sup>) was generated by retrovirally infecting the AT3 parental line with pMIG/MSCV-IRES-eGFP plasmid encoding membrane-bound OVA. To examine s.c. tumor growth, WT or gene-targeted mice were inoculated s.c. on the hind flank with the indicated number of cells and tumor size monitored. Once tumors were established (0.2–0.5 cm<sup>2</sup>) mice received a single intratumoral (i.t.) or i.v. treatment of doxorubicin hydrochloride (DOX; Sandoz) or equivalent volume of PBS. Some mice received control Ig (cIg; Mac-4 or agp3), anti-IFN- $\gamma$  (R4-6A2), anti-CD8 $\alpha$  (53.6.72), anti-CD8 $\beta$  (53.5.8), anti-IL-1 $\beta$  (B122.10), anti-IL-17A (M210), anti-IL-17R $\alpha$  (M751), or anti-IL-23p19 (16E5) monoclonal antibody (mAb) at the time of PBS/doxorubicin treatment, as indicated in the legends to neutralize or deplete cell subsets. Anti-agp3, anti-IL-23p19, anti-IL-17A, and anti-IL-17R $\alpha$  mAb were kindly provided by AMGEN Inc.

**3-Methylcholanthrene-induced carcinogenesis.** Groups of male BALB/c WT mice were inoculated s.c. in the hind flank with 400  $\mu$ g of 3-methylcholanthrene (MCA; Sigma-Aldrich in 0.1 mL of corn oil as described; ref. 18). When sarcomas had

established (palpable tumor 0.2–0.45 cm<sup>2</sup>) mice received i.t. doxorubicin or PBS once a week for 2 weeks. Some mice received cIg, anti-CD8 $\alpha$ , anti-IFN- $\gamma$ , anti-IL-1 $\beta$ , anti-IL-23p19, or anti-IL-17R $\alpha$  on days -1, 0, and weekly thereafter for 6 weeks relative to initial PBS/doxorubicin treatment. Development of fibrosarcomas was monitored weekly over the course of 250 days.

**In vivo CD8 T-cell assays and flow cytometry.** OT-I T cells were purified from spleens and lymph nodes of OT-I transgenic mice, with positive selection using a CD8 T-cell isolation kit (Miltenyi Biotec). CD8 T-cell purity was always found to be greater than 90%. Purified OT-I cells were labeled with 2.5  $\mu$ mol/L carboxyfluorescein succinimidyl ester (CFSE) and injected i.v. ( $2 \times 10^6$ ) into the tail vein of C57BL/6 mice harboring established, PBS/doxorubicin-treated AT3<sup>OVA</sup> or AT3-empty vector (AT3<sup>EV</sup>) tumors. Five days later, tumor-draining lymph nodes (TDLN) were harvested and CFSE dilution in CD8<sup>+</sup>KbOVA<sub>257-264</sub>-tetramer<sup>+</sup> cells was assessed by flow cytometry. Indices of proliferation were generated using FlowJo software (Tree Star). For tumor-infiltrating lymphocyte (TIL) analysis, AT3-OVA tumors were excised and single-cell suspensions created by a combination of collagenase type 4 (Worthington Biochemical Corp.) and DNase I (Roche) enzymatic digestion at 37°C, and mechanical disruption. OT-I T cells were identified by antibody staining with anti-CD45.2 (104; eBioscience), anti-CD8 $\alpha$  (Ly-2; eBioscience), and KbOVA<sub>257-264</sub>-tetramer (provided by A. Brooks). Dead cells were excluded by addition of 1  $\mu$ g/mL of fluorogold (Sigma-Aldrich) in the final wash. Cells were acquired on an LSR-II (BD) or Canto II (BD) flow cytometer and analysis was done using FlowJo software. For intracellular IFN- $\gamma$  analysis, tumor cell suspensions were cultured overnight in RPMI 1640 supplemented with 10% FCS in the presence of 0.1  $\mu$ mol/L OVA<sub>257-264</sub> (SIINFEKL) peptide (Auspep Pty) and 5 ng/mL recombinant mouse IL-2 (BD Biosciences). Monensin (BioLegend) was added to the cells in the final 4 hours of culture to inhibit cytokine release from the Golgi/endoplasmic reticulum complex. Permeabilization and fixation of cells were conducted using the BD Cytotfix/Cytoperm kit according to the manufacturers' instructions (BD Biosciences) before staining with anti-IFN- $\gamma$  mAb (XMG1.2; eBioscience).

### Gene expression analysis

Two cohorts of breast cancer gene expression data sets were used for these analyses. Complete clinical data are included in the Supplementary Information (Supplementary Table S1). The first comprised early-stage breast cancer patients who had received no systemic treatment to determine the association of the genes with clinical outcome without anthracycline (i.e., association with prognosis). The 4 data sets used (NKI; ref. 19; VDX; ref. 20; MAINZ; ref. 21; and TRANSBIG; ref. 22) have been previously described. Clinical endpoint used for the analyses was the first distant metastatic event. Tumor biopsy samples taken for gene expression profiling were collected at surgery. Gene expression data sets were retrieved from public databases or authors' Web sites. We used normalized data (log<sup>2</sup> intensity in single-channel platforms or log<sup>2</sup> ratio in dual-channel platforms).

Hybridization probes were mapped to the Entrez GeneID as in Shi and colleagues, 23, using RefSeq and Entrez database version 2007.01.21. When multiple probes were mapped to the same GeneID, the one with the highest variance in a particular data set was selected to represent the GeneID. Each gene analyzed was then scaled such that quartiles 2.5% and 97.5% are equal to  $-1$  and  $+1$ , respectively. This scaling is robust to outliers and hence allows combination of the microarray data from each data set. This meta-analytic technique has been previously described (24–26). Univariate Cox regression analysis was used to determine prognostic significance with clinical outcome of the genes as a continuous measurement and were calculated stratified by data set.

The second cohort of breast cancer patients was treated with anthracycline chemotherapy to determine the association of immune genes with clinical response to doxorubicin. These women received epirubicin as single-agent chemotherapy (100 mg/m<sup>2</sup>) for 4 cycles prior to surgery in the setting of a neoadjuvant clinical trial (clinical trials.gov NCT00162812; ref. 27). Epirubicin is an anthracycline chemotherapy favored in clinical practice due to its better toxicity profile than doxorubicin. Tumor biopsy samples were taken using a core biopsy 14- to 16-G needle prior to chemotherapy. Clinical endpoint used was pathologic complete response (pCR) documented at surgery, which is an accepted surrogate for disease-free and overall survival in breast cancer. Genes were correlated with pCR as a continuous variable (i.e., to determine whether higher expression correlated with a higher chance of obtained pCR), using a receiver operating characteristic (ROC), and measured using an area under the curve (AUC). Microarray analysis was done using Affymetrix GeneChips.

### Statistical analysis

**Mouse.** Statistical analyses were done using GraphPad Prism software. Significant differences between groups were assessed by a 2-tailed *t* test or Mann–Whitney *U* test, as indicated. Values of  $P < 0.05$  were considered significant.

**Human.** Statistical analysis using gene expression data was done with R version 2.5.1 and BioConductor version 1.8. R code used is available as "genefu" from the comprehensive R archive network. The remaining statistical analyses were conducted on SPSS 18.0 (SPSS, Inc.). Results were not corrected for multiple testing, as the analyses were considered hypothesis generating.

## Results

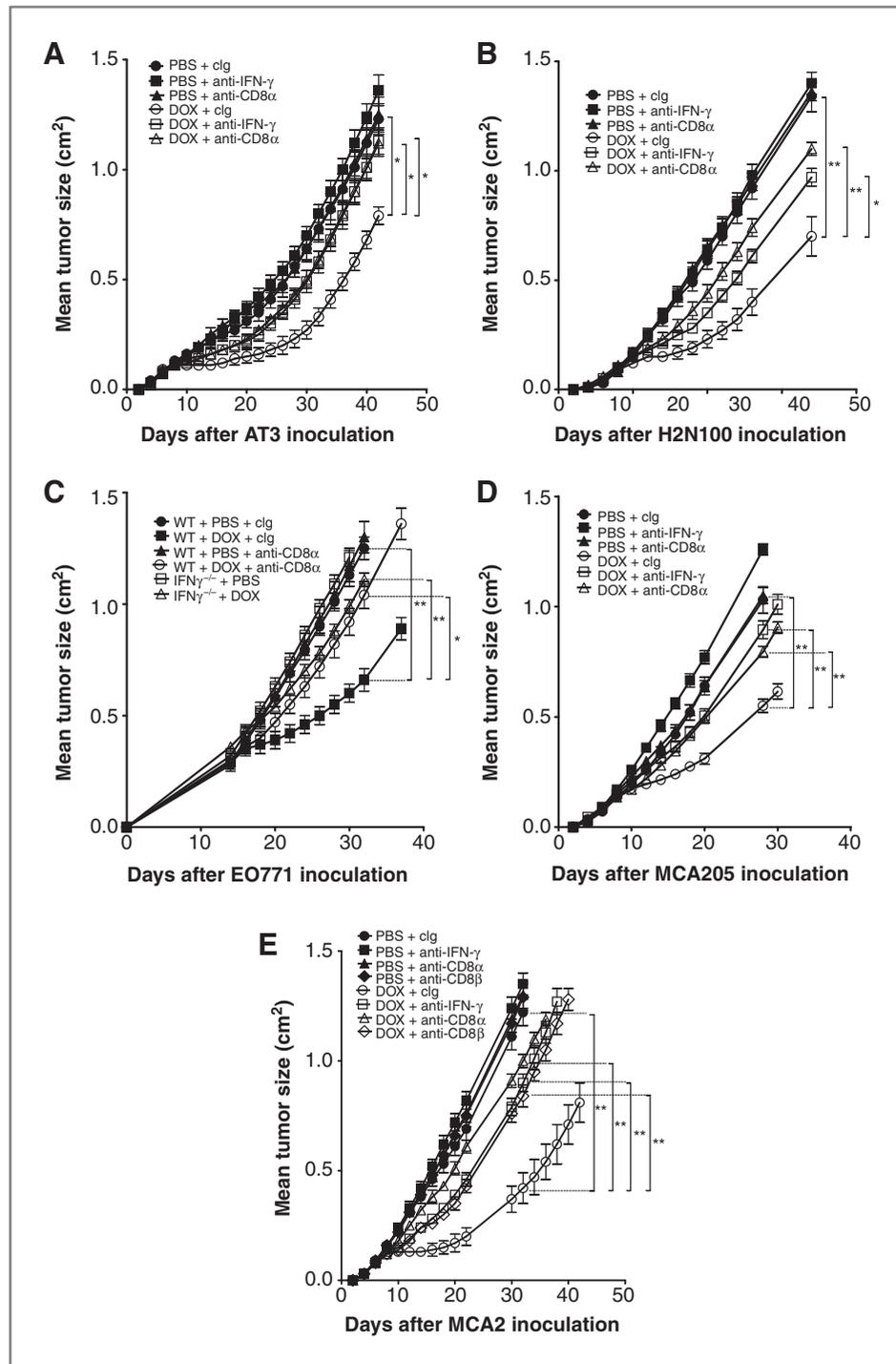
### The efficacy of doxorubicin therapy in established s.c. tumors requires CD8<sup>+</sup> cells and IFN- $\gamma$

Both CD8 T cells and IFN- $\gamma$  are important in immunosurveillance of a range of malignancies (28–31). More recently, adaptive immunity and IFN- $\gamma$  have been implicated in the immunogenic effects of chemotherapy (10, 32), but CD8<sup>+</sup> T cells have not been previously directly shown to contribute to the antitumor activity of doxorubicin. Despite the fact that anthracyclines are commonly used in the clinical management of human breast cancer, very little data concerning the mechanism of action of doxorubicin against mammary carci-

nomas has been obtained in experimental models or from clinical samples. We assessed the requirement for CD8<sup>+</sup> T cells and IFN- $\gamma$  in the effectiveness of doxorubicin chemotherapy treatment against a variety of established breast cancers and fibrosarcomas. In WT mice, a localized (i.t.) treatment of doxorubicin was sufficient to suppress s.c. growth of AT3, H2N100, and EO771 mammary tumors and MCA205 fibrosarcomas (Fig. 1A–D). Depletion of CD8 $\alpha$ <sup>+</sup> cells or neutralization of IFN- $\gamma$  with mAb administration immediately prior to doxorubicin treatment severely abrogated the antitumor effect of the drug in the established tumor setting with significant loss of tumor growth suppression (Fig. 1A–D). Abrogation of doxorubicin efficacy was also observed in IFN- $\gamma$ <sup>-/-</sup> mice (Fig. 1C). To confirm that the loss of therapeutic effect of doxorubicin following anti-CD8 $\alpha$  mAb treatment was specifically due to depletion of CD8<sup>+</sup> T cells, we assessed an additional group of mice receiving anti-CD8 $\beta$  mAb prior to doxorubicin treatment of established MCA2 fibrosarcoma tumors. Depletion of CD8 T cells alone was sufficient to abrogate the antitumor effect of doxorubicin (Fig. 1E). In addition, mice challenged with MCA2 tumors were treated with i.v. delivery of doxorubicin to establish that systemic administration of doxorubicin was comparable with localized treatment at inducing CD8 T cell and IFN- $\gamma$ -mediated antitumor activity. For the first time, these studies validate across a series of experimental tumors the critical role of CD8<sup>+</sup> T cells and IFN- $\gamma$  *in vivo* in the mechanism of antitumor activity of doxorubicin.

### Carcinogen-induced tumors respond to doxorubicin in a CD8 T-cell- and IFN- $\gamma$ -dependent manner

The activity and mechanism of action of most cancer therapies, including doxorubicin, have rarely if ever been assessed in mouse models of *de novo* tumorigenesis. This type of model is more practical in assessing any agent's action in the context of a developing and progressing tumor, as opposed to a transplant setting. Recently, we have employed established fibrosarcomas induced by MCA in mice as a therapeutic model for various immune targets (33, 34) and to determine the importance of host IL-1 $\beta$  in the mechanism of action of doxorubicin (10). To date, however, the importance of many other host immune elements in doxorubicin antitumor activity has not been examined in a mouse model in which the tumor has arisen *de novo* in the host. Depletion of CD8 T cells following establishment of palpable sarcomas (0.20–0.45 cm<sup>2</sup>) did not significantly alter the growth kinetics of the established tumor (Fig. 2, left; Supplementary Fig. S4). Doxorubicin treatment (i.t.) of established MCA sarcomas achieved modest tumor growth suppression in most mice, and 4 of 30 (13%) showed a significant period of retarded tumor growth but eventually succumbed (Fig. 2A; Supplementary Fig. S4). This variable response was consistent with the vast heterogeneity between individual MCA-induced fibrosarcomas. Importantly, the significant therapeutic effect of doxorubicin in inhibiting fibrosarcoma growth across the whole cohort of control mAb-treated mice ( $P < 0.0001$ ; Supplementary Fig. S4) was abolished if mice received anti-CD8 $\alpha$  mAb or anti-IFN- $\gamma$  mAb during doxorubicin therapy (Fig. 2B and C;



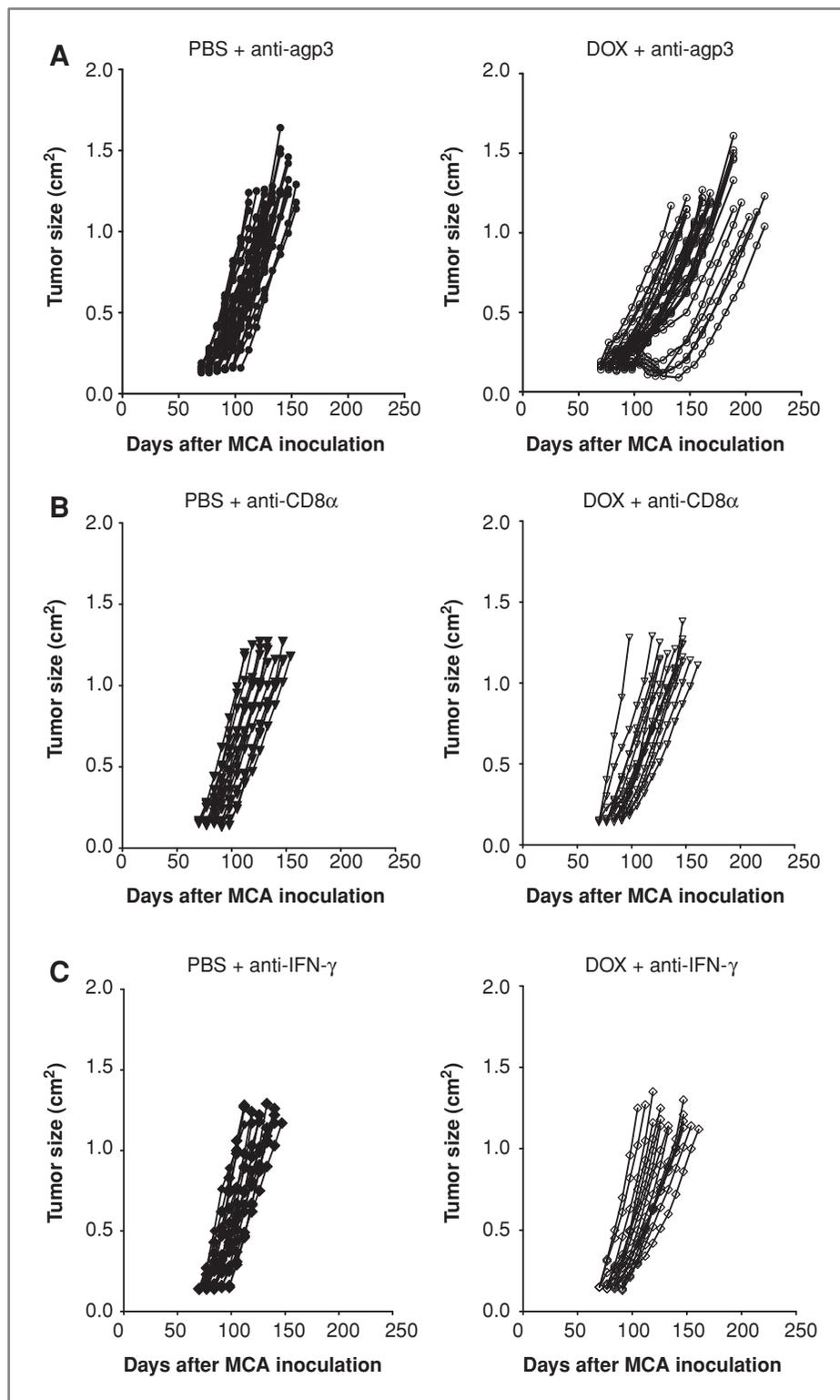
**Figure 1.** Doxorubicin therapy requires CD8<sup>+</sup> T cells and IFN- $\gamma$ . Groups of 5 syngeneic WT or gene-targeted mice as indicated were injected s.c. with  $5 \times 10^5$  AT3 mammary adenocarcinoma cells (A),  $5 \times 10^5$  H2N100 mammary adenocarcinoma cells (B),  $5 \times 10^5$  EO771 mammary adenocarcinoma cells (C),  $8 \times 10^5$  MCA205 fibrosarcoma cells (D), or  $1 \times 10^5$  MCA2 fibrosarcoma cells (E). Mice then received either i.t. PBS or doxorubicin (50  $\mu$ L, 2 mmol/L) on day 7 (A), day 8 (B), day 14 (C), and day 7 (D) after tumor inoculation or PBS or doxorubicin (2 mg/kg) i.v. on days 7 and 14 (E) after tumor inoculation. Some mice received control Ig (clg), anti-CD8 $\alpha$ , anti-CD8 $\beta$ , or anti-IFN- $\gamma$  (100  $\mu$ g i.p.) on day -1, 0, and weekly thereafter relative to initial PBS/doxorubicin treatment. Tumor size was measured as indicated. Data show means  $\pm$  standard errors of 5 mice per group. Data for AT3, H2N100, EO771 (A-C), and MCA2 (E) are representative of 2 independent experiments. Statistical analyses were conducted at the time point indicated on the figure using Mann-Whitney test (\*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ).

Supplementary Fig. S4), indicating that both CD8<sup>+</sup> T cells and IFN- $\gamma$  were critical for the antitumor effects of doxorubicin.

#### Doxorubicin treatment enhances antigen-specific CD8 T-cell proliferation *in vivo*

We next investigated in more detail the antigen-driven responses of CD8 T cells following chemotherapy in breast

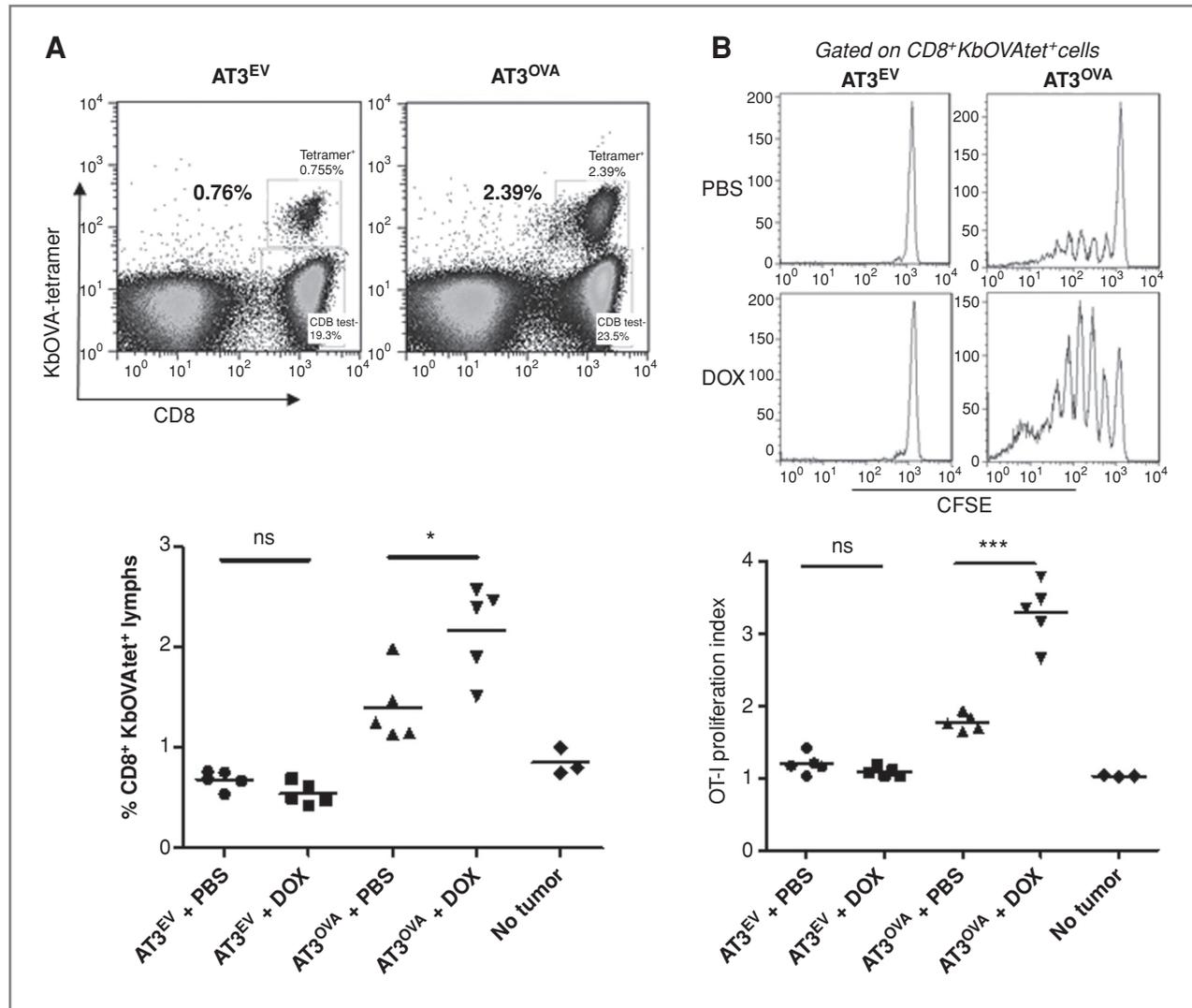
cancer. Immunocompetent B6 mice showed considerable resistance to challenge with s.c. AT3<sup>OVA</sup> tumors in comparison with AT3<sup>EV</sup> control tumors (Supplementary Fig. S1). Next, we adoptively transferred naive OVA-specific CD8 T cells (OT-I cells) 1 day after mice harboring established AT3<sup>OVA</sup> or AT3<sup>EV</sup> were treated with doxorubicin. Not surprisingly, the proportion of OT-I cells recovered from



**Figure 2.** Tumors induced *de novo* by carcinogen respond to doxorubicin in a CD8<sup>+</sup> T cell- and IFN- $\gamma$ -dependent manner. Groups of 15 to 30 male BALB/c WT mice were injected s.c. on the flank with 400  $\mu$ g MCA on day 0. When sarcomas had established (the second week of palpable tumor 0.20–0.45 cm<sup>2</sup>), BALB/c mice received either i.t. PBS (left) or doxorubicin (50  $\mu$ L, 2 mmol/L; right) once a week for 2 weeks. Some mice received (A) control Ig (anti-agg3), (B) anti-CD8 $\alpha$ , or (C) anti-IFN- $\gamma$  (100–500  $\mu$ g i.p.) on days –1, 0, and weekly thereafter for 6 weeks relative to initial PBS/doxorubicin treatment. Mice were then monitored for tumor development over 250 days and recorded as the growth curves (tumor size in cm<sup>2</sup>) of individual mice with tumor in each group.

TDLN after 5 days was significantly higher in mice with AT3<sup>OVA</sup> tumors than in those inoculated with AT3<sup>EV</sup> tumors. More importantly, the proportion of OT-I cells in TDLN was enhanced following doxorubicin treatment, however, only in

the context of cognate antigen recognition (AT3<sup>OVA</sup> setting; Fig. 3A). Furthermore, *in vivo* proliferation of OT-I cells in the presence of OVA, as measured by CFSE dilution, was significantly increased after doxorubicin treatment (Fig. 3B). Overall,



**Figure 3.** Doxorubicin treatment enhances CD8 T-cell proliferation to cognate tumor antigen. C57BL/6 mice were inoculated s.c. with either AT3<sup>EV</sup> ( $1 \times 10^6$ ) or AT3<sup>OVA</sup> ( $1 \times 10^6$ ) tumors. After 30 days, groups of mice ( $n = 5$ ) were i.t. treated with doxorubicin (1 mmol/L) or PBS, and the following day all mice received i.v. transfer of purified, CFSE-labeled CD8<sup>+</sup> OT-I T cells ( $2 \times 10^6$ ). Five days after OT-I cell transfer, TDLNs were excised and fluorescence-activated cell-sorting (FACS) analyses on KbOVA-tetramer-reactive CD8 T cells were conducted. **A**, representative FACS plots of CD8<sup>+</sup>KbOVA-tetramer<sup>+</sup> (CD8<sup>+</sup>KbOVA-tet<sup>+</sup>) cells in the DLN of mice harboring AT3<sup>EV</sup> or AT3<sup>OVA</sup> tumors. Data shown are the percentage of OT-I cells within the lymphocyte population in individual mice with the indicated tumors and treatment. \*,  $P < 0.02$ ,  $t$  test; ns, not significant. **B**, FACS histograms showing representative CFSE dilution within the gated CD8<sup>+</sup>KbOVA-tetramer<sup>+</sup> transferred OT-I cell population in DLN of mice harboring AT3<sup>EV</sup> or AT3<sup>OVA</sup> tumors after prior treatment with PBS (top) or doxorubicin (bottom). OT-I cell proliferation indices with means are shown in the underlying graph. \*\*\*,  $P < 0.0001$ ,  $t$  test; ns, not significant. All graphs are representative of 3 independent experiments.

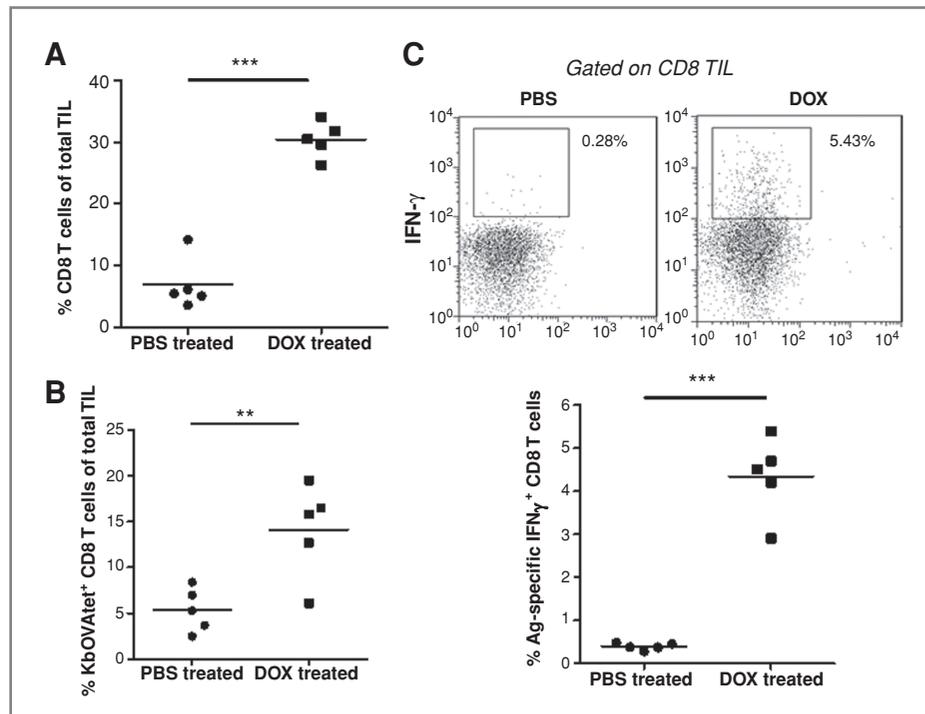
localized doxorubicin treatment of established tumors enhances the accumulation and proliferation of CD8 T cells responding to cognate tumor antigen in secondary lymphoid organs.

#### Tumor-infiltrating CD8 T cells and IFN- $\gamma$ production are enhanced by doxorubicin treatment

We harvested AT3<sup>OVA</sup> tumors from mice that had received i.t. doxorubicin or PBS treatment and naive OT-I T cell adoptive transfer to determine whether localized doxorubicin administration increased the proportion and function of CD8

TILs. A significant increase in the proportion of total CD8 T cells amongst AT3<sup>OVA</sup> TILs was observed following doxorubicin treatment (Fig. 4A), and a similar increase was observed for infiltrating Ag-specific OT-I cells (Fig. 4B). Doxorubicin treatment also induced Ag-specific IFN- $\gamma$  production from CD8 TILs, observed after *in vitro* restimulation with cognate antigen (Fig. 4C). No cytokine was detected from CD8 TILs in the absence of restimulation (data not shown). These experiments are amongst the first to directly isolate and show the increase in the proportion of tumor-localized and functional CD8<sup>+</sup> T cells following treatment with doxorubicin.

**Figure 4.** Local doxorubicin treatment enhances numbers of CD8 TILs and IFN- $\gamma$  production. C57BL/6 mice were inoculated s. c. with AT3<sup>OVA</sup> ( $1 \times 10^6$ ) tumors. After 42 days, groups of mice ( $n = 5$ ) were treated i.t. with doxorubicin (1 mmol/L) or PBS, and the following day received i.v. transfer of purified, CD8<sup>+</sup> OT-I T cells ( $2 \times 10^6$ ). Seven days after OT-I cell transfer, tumors were excised and FACS analyses on CD8<sup>+</sup> TILs were conducted. CD8 T cells (A) and KbOVA-tetramer-reactive CD8 T cells (B) as a percentage of total TILs for individual mice treated with PBS or doxorubicin are shown. C, antigen-specific IFN- $\gamma$  cytokine production from CD8 TILs measured by intracellular staining after overnight *in vitro* stimulation of AT3<sup>OVA</sup> tumor cell suspensions with OVA peptide + IL-2. \*\*,  $P < 0.01$ ; \*\*\*,  $P < 0.0001$ , *t* test. Data are representative of 3 independent experiments.



#### Doxorubicin treatment requires IL-1 $\beta$ and IL-17 but not IL-23p19

We have recently shown that IL-1 $\beta$  production is pivotal for a robust antitumor response to chemotherapy, by facilitating the priming of tumor-specific CD8 T cells (10). We now have extended these findings in a range of mammary adenocarcinomas to show that IL-1 $\beta$ /IL-1R signaling is also critical to the therapeutic outcome of doxorubicin treatment (Fig. 5). Because IL-1 $\beta$  is known to regulate IL-17 responses (35–37), we utilized both IL-17A<sup>-/-</sup> mice and an anti-IL-17R $\alpha$  mAb to also show a critical role for IL-17 in doxorubicin treatment of transplantable mammary tumors (Fig. 5). In concert with the mammary carcinoma data, a critical role for IL-1 $\beta$  and IL-17A in doxorubicin antitumor activity was also illustrated using fibrosarcomas generated *de novo* by MCA (Fig. 6B and D; Supplementary Fig. S4). Interestingly, the absence of IL-1 or IL-17 signaling had little or no direct effect on the growth of untreated established tumors. In contrast IL-23, another immunoregulatory cytokine known to cooperate with IL-1 $\beta$  in the regulation of IL-17 responses (35–37), was dispensable in these models, as no alteration in doxorubicin efficacy was observed in IL-23p19<sup>-/-</sup> mice or by neutralizing IL-23 in WT mice using an anti-IL-23 mAb (Fig. 6C; Supplementary Figs. S2 and S4). In addition, by comparison, the antitumor efficacy of doxorubicin in transplanted tumors was partially reduced in the absence of the IL-12p35 subunit (Supplementary Fig. S2).

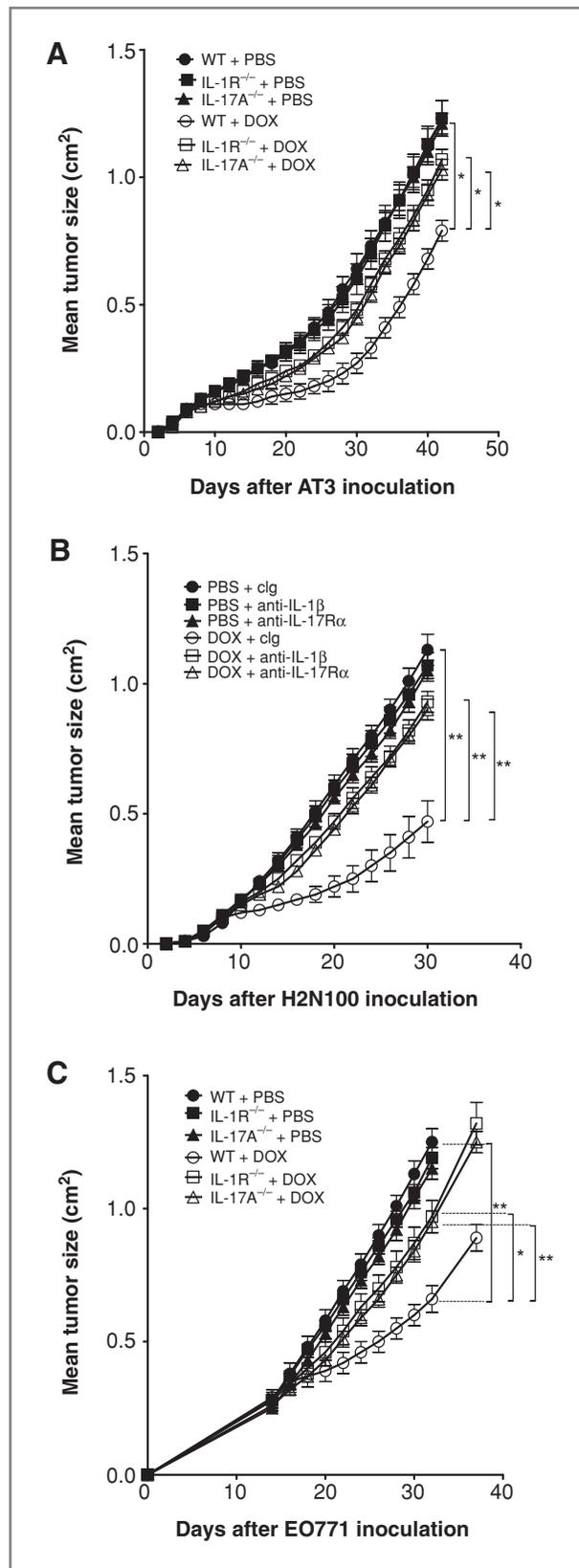
#### Doxorubicin therapy requires $\gamma\delta$ T cells but not type I NKT cells

Both NKT cells and  $\gamma\delta$  T cells have been implicated in innate immunosurveillance of tumors (38, 39) and are also a

potent early source of IL-17 in response to IL-1 $\beta$  cytokine stimulation (35, 36). We assessed the requirement for these innate lymphocytes in doxorubicin therapy of established AT3 mammary tumors and MCA205 fibrosarcomas (Supplementary Fig. S3). The absence of type I J $\alpha$ 18<sup>+</sup> NKT cells in TCRJ $\alpha$ 18<sup>-/-</sup> mice did not perturb the antitumor effects of doxorubicin; however, deficiency of  $\gamma\delta$  T cells in TCR $\delta$ <sup>-/-</sup> mice resulted in significant inhibition of doxorubicin activity in both AT3 and MCA205 tumors. The loss of NKT cells or  $\gamma\delta$  T cells did not alter the outgrowth of these tumors in the absence of therapy (Supplementary Fig. S3).

#### Clinical relevance using gene expression data from human breast cancer patients

We went on to explore whether these genes were important for therapeutic efficacy of anthracycline treatment in breast cancer patients, using publicly available gene expression data sets. The first cohort consisted of patients diagnosed at multiple different hospitals who had their tumor biopsy samples taken at surgery. The samples examined here are representative of the global breast cancer population, as there was a mixture of different tumor sizes, nodal status, and estrogen receptor (ER) expression (Supplementary Table S1). As these women received no systemic treatment, the true prognostic effects of these genes could be examined without treatment confounders. Therefore, we could examine the prognostic significance of *CD8A*, *CD8B*, *IFN- $\gamma$* , *IL1B*, *IL17A*, and *IL23p19* genes independent of anthracycline therapy in patients who had received no systemic treatment (refs. 19–22; Table 1). As observed, only high IFN- $\gamma$  levels were associated with a good prognosis in these women, that is, high levels were associated with fewer long-term breast cancer relapses.

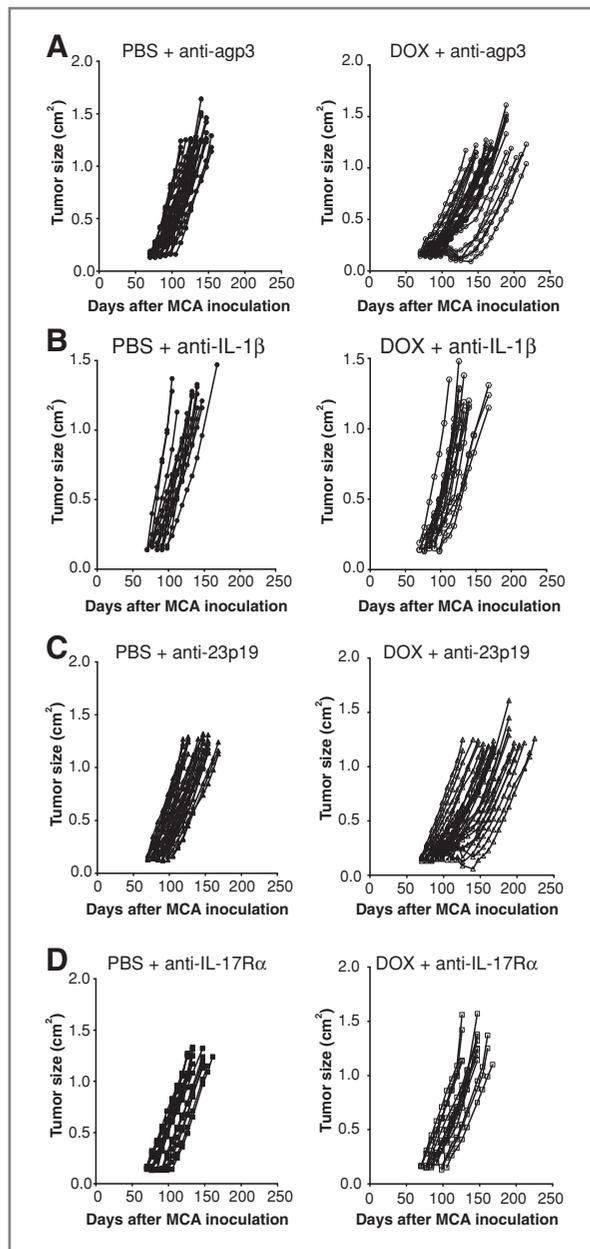


We then went on to examine these genes in the context of treatment with anthracycline chemotherapy. The second cohort consisted of women enrolled into a breast cancer clinical trial specifically designed to investigate biomarkers to anthracycline chemotherapy. As a consequence, all of the women in this study were negative for expression of ER (as this is the more chemoresponsive breast cancer population) and all received single-agent epirubicin, an anthracycline-type chemotherapy commonly used in clinical practice. Tumor biopsy samples were taken with core biopsy needles and were subject to gene expression profiling prior to chemotherapy (27). In these breast cancer patients, increasing levels of IFN- $\gamma$ , as well as CD8 $\alpha$  and CD8 $\beta$  were associated with a better response to therapy, as measured by the amount of tumor left at surgical resection after 4 cycles of therapy. Notably, the complete disappearance of invasive tumor after neoadjuvant chemotherapy was associated with an excellent survival from breast cancer and is an accepted surrogate for long-term survival from breast cancer (40). There was no significance for the other genes in these cohorts. Overall, these data support further the role of IFN- $\gamma$ , CD8 $\alpha$ , and CD8 $\beta$  in the clinical outcomes in breast cancer patients who have received anthracycline-type chemotherapy.

## Discussion

Anthracyclines are front-line chemotherapeutic agents in the treatment of breast cancers and sarcomas. In this study, we have provided an extensive amount of new data exploring the mechanism of action of doxorubicin in a series of transplantable mammary carcinomas and fibrosarcomas generated *de novo* by MCA. The new data in AT3, H2N100, and EO771 mammary tumor models showed the critical role of host CD8<sup>+</sup> T cells and IFN- $\gamma$ , additionally supported by novel human clinical data in 2 different cohorts of women with breast cancer. Our study is the first to describe the specific importance of CD8<sup>+</sup> T cells *in vivo*, as opposed to broadly assessing adaptive immunity by employing *RAG*-deficient or nude mice that are deficient in many immune components including all T-cell subsets and B cells. Furthermore, analysis included CD8 $\alpha$ - and CD8 $\beta$ -specific depletions to specifically define the importance of CD8<sup>+</sup> T cells. Doxorubicin treatment enhanced the proliferation of CD8 T cells in the tumor DLN in a cognate antigen-specific manner. Moreover, localized doxorubicin treatment increased the proportions of CD8

**Figure 5.** Doxorubicin therapy requires IL-1R and IL-17A. Groups of 5 syngeneic WT or gene-targeted mice as indicated were injected s.c. with  $5 \times 10^5$  AT3 mammary adenocarcinoma cells (A),  $5 \times 10^5$  H2N100 mammary adenocarcinoma cells (B), or  $5 \times 10^5$  EO771 mammary adenocarcinoma cells (C). Mice then received either i.t. PBS or doxorubicin (50  $\mu$ L, 2 mmol/L) on day 7 (A), day 8 (B), and day 14 (C) after tumor inoculation. Some mice received control Ig (clg), anti-IL-1 $\beta$ , or anti-IL-17R $\alpha$  (500  $\mu$ g i.p.) on days 7, 8, 15, and 22 relative to tumor inoculation. Tumor size was measured as indicated. Data show means  $\pm$  standard errors of 5 mice per group, representative of 2 independent experiments. Statistical analyses were conducted at the time point indicated on the figure by using Mann-Whitney test (\*,  $P < 0.05$ ; \*\*,  $P < 0.01$ ).



**Figure 6.** Tumors induced *de novo* by carcinogen respond to doxorubicin in an IL-1 $\beta$ - and IL-17-dependent manner. Groups of 15 to 30 male BALB/c WT mice were injected s.c. on the flank with 400  $\mu$ g MCA on day 0. When sarcomas had established (the second week of palpable tumor 0.20–0.45 cm<sup>2</sup>) BALB/c mice received either i.t. PBS (left) or doxorubicin (50  $\mu$ L, 2 mmol/L; right) once a week for 2 weeks. Some mice received (A) control Ig (anti-aggp3), (B) anti-IL-1 $\beta$ , (C) anti-IL-23p19, or (D) anti-IL-17R $\alpha$  (100–500  $\mu$ g i.p.) on days –1, 0, and weekly thereafter for 6 weeks relative to initial PBS/doxorubicin treatment. Mice were then monitored for tumor development over 250 days and recorded as the growth curves (tumor size in cm<sup>2</sup>) of individual mice with tumor in each group. Control Ig graphs in (A) are reproduced from Fig. 2A.

T cells infiltrating the tumor and enhanced tumor antigen-specific IFN- $\gamma$  production from these CD8 TILs. Combined, these data are amongst the strongest to indicate that tumor cell

death associated with doxorubicin treatment enhances the generation and functional activation of tumor-reactive CD8 T cells that are required for the antitumor activity of doxorubicin.

We also completed the most extensive characterization of mechanism of action of doxorubicin in tumors established *de novo* by carcinogen, including a comparative evaluation of the role of CD8<sup>+</sup> T cells, IFN- $\gamma$ , IL-1 $\beta$ , IL-17A, and IL-23. These types of established tumors generated *de novo* are arguably more relevant than any short-term transplanted tumors that have been used to previously establish the principles that chemotherapy may be immunogenic. Our study has clearly shown that CD8<sup>+</sup> T cells, IFN- $\gamma$ , IL-1 $\beta$ , and IL-17A but not IL-23, were critical in the antitumor activity of doxorubicin. We further illustrated the key role of IL-1 $\beta$  and IL-17A, but not IL-23, in doxorubicin control of a series of mammary carcinomas. Our data extend our recently published data that illustrated the importance of IL-17A in the antitumor activity of doxorubicin against transplanted MCA205 tumors and tracked the generation of IL-17-producing  $\gamma\delta$ <sup>+</sup> T cells following doxorubicin treatment (41). We confirmed the role of  $\gamma\delta$ <sup>+</sup> T cells, but not type I NKT cells, in the mechanism of action of doxorubicin, using transplantable mammary carcinomas.

In human breast cancer patients, although IFN- $\gamma$  levels seemed relevant for patient clinical outcome independent of anthracycline, the importance of both CD8 T cells and IFN- $\gamma$  in therapeutic efficacy was supported by the observation that increasing levels of *CD8 $\alpha$* , *CD8 $\beta$* , and *IFN- $\gamma$*  gene expression correlated with a better response to anthracycline chemotherapy. Notably, here the tumor biopsy samples were taken prior to chemotherapy, supporting the concept that an intact immune system aids therapeutic response to anthracyclines. However, it would be most interesting to also see the relative changes in gene expression before and shortly after administration of therapy. Although the analyses in human breast cancer patients presented here are limited by the use of retrospective data sets, nonrandomized treatment cohorts, and patients treated with epirubicin rather than doxorubicin, which is a different type of anthracycline chemotherapy, it supports our laboratory observations and a recent study (42) of the clinical relevance of antitumor immunity for better clinical outcomes from anthracycline therapy in breast cancer.

It is likely that the context in which IL-17A is produced, particularly the constituents of the tumor microenvironment, will determine the outcome of IL-17 secretion. IL-17 production, particularly from innate T lymphocytes such as NKT cells and  $\gamma\delta$  T cells, is dependent on IL-1R signaling (36, 43) and has been shown to boost CD8 T-cell IFN- $\gamma$  responses (44). Given the requirement for IL-1 $\beta$  and CD8 T cells in the antitumor immune response following doxorubicin treatment, we assessed the role for IL-17 as an intermediary molecule in this pathway. IL-17A or IL-17R $\alpha$  blockade resulted in loss of doxorubicin efficacy, very similar to that of IL-1 $\beta$ /IL-1R blockade, indicating that IL-17 is probably a major downstream response cytokine to IL-1 $\beta$  secretion. Interestingly, IL-23, itself a modulator of antitumor immunity (45–48), but also an important cofactor along with IL-1 $\beta$  in the stimulation of IL-17 production from memory T cells (37), NKT cells (35), and  $\gamma\delta$  T cells (36), was dispensable in doxorubicin treatment

**Table 1.** Association of genes with clinical outcome in breast cancer patients treated with and without anthracycline chemotherapy<sup>a</sup>

Genes	Cohort 1 <sup>b</sup> HR (95% CI)	Cohort 2 <sup>c</sup> AUC (95% CI)
<i>IFN-γ</i>	0.8 (0.65–0.95), <i>P</i> = 0.031	0.69 (0.56–0.81), <i>P</i> = 0.016
<i>CD8A</i>	0.82 (0.5–1.3), <i>P</i> = 0.4	0.72 (0.59–0.84), <i>P</i> = 0.005
<i>CD8B</i>	0.87 (0.59–1.3), <i>P</i> = 0.21	0.65 (0.52–0.78), <i>P</i> = 0.049
<i>IL17A</i>	0.82 (0.67–1.1), <i>P</i> = 0.051	0.5, <i>P</i> = 0.44
<i>IL1B</i>	0.87 (0.7–1.06), <i>P</i> = 0.18	0.39, <i>P</i> = 0.19
<i>IL23</i>	0.88 (0.75–1.03), <i>P</i> = 0.13	0.56, <i>P</i> = 0.42

Abbreviation: pCR, pathologic complete response.

<sup>a</sup>Association of genes with clinical outcome in breast cancer patients treated with and without anthracycline chemotherapy. Cohort 1 consisted of 1,062 patients from 4 gene expression data sets who received no systemic treatment after their surgery to determine the clinical outcome association independent of anthracycline. Seventy percent of tumors were ER-positive, 85% were node negative at diagnosis. Univariate HRs presented are stratified by data set. Genes are correlated as continuous variables. Here, increasing expression of IFN- $\gamma$  is correlated with a good prognosis or a longer time free from distant metastases. Cohort 2 consisted of 114 patients treated in a neoadjuvant trial of the anthracycline epirubicin given as a sole therapy prior to surgery. All patients were ER-negative with 86% of tumors greater than 2 cm and 54% with positive lymph nodes at study entry. Biopsies were taken prior to therapy. Here, increasing expression of IFN- $\gamma$ , CD8A, and CD8B is associated with a higher chance of a complete response from chemotherapy at surgery. In this second cohort, a pCR was strongly correlated with a better survival from breast cancer. (Full clinical information for cohorts 1 and 2 can be found in the Supplementary Table S1.)

<sup>b</sup>Endpoint: distant metastases; number of patients = 1,062; treatment = none.

<sup>c</sup>Endpoint: pCR (complete disappearance of invasive tumor at surgery after 4 cycles of epirubicin chemotherapy given at 100 mg/m<sup>2</sup>); number of patients = 114; treatment = epirubicin monotherapy.

of breast cancers and fibrosarcomas, with no alteration in therapeutic effect following IL-23 blockade. Notably, gene expression of *IL1B*, *IL17A*, or *IL23* was not associated with clinical outcomes to doxorubicin treatment in human breast cancer patients, but perhaps this was not surprising given that the tumor biopsy samples were taken prior to doxorubicin treatment, and these cytokines were likely to be induced following treatment. In a parallel study, we have very recently shown that IL-17 from  $\gamma\delta$  T cells is critical for the efficacy of chemotherapy and preceded the activation and polarization of CD8 T cells for IFN- $\gamma$  production (41). Also,  $\gamma\delta$  T cells that lacked IL-1R lost the capacity to amplify the action of chemotherapy, confirming the causal relationship with IL-1 $\beta$  production from adenomatous polyposis coli (*APC*) causing IL-17 production from  $\gamma\delta$  T cells and resulting in IFN- $\gamma$  production from tumor-specific CD8 T cells. The current study provides important additional data in mammary tumor transplant and *de novo* fibrosarcoma models to support a role for IL-17A in doxorubicin mechanism of action. In doing so, we also show the relevance of  $\gamma\delta$ <sup>+</sup> T cells but not of type I NKT cells.

Overall, our *in vivo* mouse data and human genetic analysis show the role of host CD8 T cells and IFN- $\gamma$  in doxorubicin mechanism of action. The study also strongly supports our most recent work on the role of IL-17A in the mechanism of action of chemotherapy (41), particularly with respect to novel data concerning doxorubicin and mammary carcinoma. In addition, it extensively characterizes mechanism of action of doxorubicin in tumors established *de novo* by carcinogen, including comparative evaluation of the role of CD8<sup>+</sup> T cells, IFN- $\gamma$ , IL-1 $\beta$ , IL-17, and IL-23. These types of established

tumors are arguably more relevant than any short-term transplanted tumors that have been used to previously establish the principle that chemotherapy may be immunogenic. The work corroborates recent studies showing that numbers of TILs in breast cancer are a predictor of response to neoadjuvant chemotherapy (49, 50). Similar outcomes in mouse sarcomas suggest more widespread applicability of these findings to other tumor cell and tissue types. Future work should address how to facilitate the potential development of sustained tumor-specific, adaptive immunity by anthracycline chemotherapy. This includes the generation of long-lived memory T cells and identification of the patients who are best candidates for such immunotherapeutic approaches, as enhancing tumor immunosurveillance by chemotherapy may allow extended immune-mediated control of the malignancy beyond the treatment period of the drug.

#### Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

#### Acknowledgments

The authors thank Qerime Mundrea and Ben Venville for maintenance of the mice, Nicole Haynes for generation of the AT3<sup>OVA</sup> cell line, Jennifer Towne (AMGEN, Inc.) for providing the anti-agp3, anti-IL-23, and anti-IL-17R $\alpha$  mAbs, and Benjamin Haibe-Kains for bioinformatics assistance.

#### Grant Support

This work was supported by The Victorian Cancer Agency, The Victorian Breast Cancer Consortium, and the Susan G. Komen Breast Cancer

Foundation. S.R. Mattarollo was supported by a Balzan Foundation Fellowship. S. Loi was supported by a National Health & Medical Research Council (NH&MRC) clinical fellowship. Y. Ma was supported by China Scholarship Council. L. Zitvogel was supported by LIGUE labellisee, INFLACARE FP7 EU grant, INCa, Fondation pour la Recherche Medicale, and Fondation de France. M.J. Smyth received support from an NH&MRC Australia Fellowship.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received March 7, 2011; revised April 27, 2011; accepted May 12, 2011; published OnlineFirst June 6, 2011.

## References

- Rasmussen L, Arvin A. Chemotherapy-induced immunosuppression. *Environ Health Perspect* 1982;43:21–5.
- Steele TA. Chemotherapy-induced immunosuppression and reconstitution of immune function. *Leuk Res* 2002;26:411–4.
- Ma Y, Kepp O, Ghiringhelli F, Apetoh L, Aymeric L, Locher C, et al. Chemotherapy and radiotherapy: cryptic anticancer vaccines. *Semin Immunol* 2010;22:113–24.
- Haynes NM, van der Most RG, Lake RA, Smyth MJ. Immunogenic anti-cancer chemotherapy as an emerging concept. *Curr Opin Immunol* 2008;20:545–57.
- Obeid M, Tesniere A, Ghiringhelli F, Fimia GM, Apetoh L, Perfettini JL, et al. Calreticulin exposure dictates the immunogenicity of cancer cell death. *Nat Med* 2007;13:54–61.
- Panaretakis T, Joza N, Modjtahedi N, Tesniere A, Vitale I, Durchschlag M, et al. The co-translocation of ERp57 and calreticulin determines the immunogenicity of cell death. *Cell Death Differ* 2008;15:1499–509.
- Panaretakis T, Kepp O, Brockmeier U, Tesniere A, Bjorklund AC, Chapman DC, et al. Mechanisms of pre-apoptotic calreticulin exposure in immunogenic cell death. *EMBO J* 2009;28:578–90.
- Apetoh L, Ghiringhelli F, Tesniere A, Obeid M, Ortiz C, Criollo A, et al. Toll-like receptor 4-dependent contribution of the immune system to anticancer chemotherapy and radiotherapy. *Nat Med* 2007;13:1050–9.
- Guerriero JL, Ditsworth D, Catanzaro JM, Sabino G, Furie MB, Kew RR, et al. DNA alkylating therapy induces tumor regression through an HMGB1-mediated activation of innate immunity. *J Immunol* 2011;186:3517–26.
- Ghiringhelli F, Apetoh L, Tesniere A, Aymeric L, Ma Y, Ortiz C, et al. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1 $\beta$ -dependent adaptive immunity against tumors. *Nat Med* 2009;15:1170–8.
- Orsini F, Pavelic Z, Mihich E. Increased primary cell-mediated immunity in culture subsequent to adriamycin or daunorubicin treatment of spleen donor mice. *Cancer Res* 1977;37:1719–26.
- Schwartz HS, Grindey GB. Adriamycin and daunorubicin: a comparison of antitumor activities and tissue uptake in mice following immunosuppression. *Cancer Res* 1973;33:1837–44.
- Maccubbin DL, Cohen SA, Ehrke MJ. Indomethacin modulation of adriamycin-induced effects on multiple cytolytic effector functions. *Cancer Immunol Immunother* 1990;31:373–80.
- Smyth MJ, Thia KY, Street SE, Cretney E, Trapani JA, Taniguchi M, et al. Differential tumor surveillance by natural killer (NK) and NKT cells. *J Exp Med* 2000;191:661–8.
- Teng MW, Swann JB, von Scheidt B, Sharkey J, Zerafa N, McLaughlin N, et al. Multiple antitumor mechanisms downstream of prophylactic regulatory T-cell depletion. *Cancer Res* 2010;70:2665–74.
- Stagg J, Divisekera U, McLaughlin N, Sharkey J, Pommey S, Denoyer D, et al. Anti-CD73 antibody therapy inhibits breast tumor growth and metastasis. *Proc Natl Acad Sci U S A* 2010;107:1547–52.
- Stagg J, Sharkey J, Pommey S, Young R, Takeda K, Yagita H, et al. Antibodies targeted to TRAIL receptor-2 and ErbB-2 synergize *in vivo* and induce an antitumor immune response. *Proc Natl Acad Sci U S A* 2008;105:16254–9.
- Swann JB, Vesely MD, Silva A, Sharkey J, Akira S, Schreiber RD, et al. Demonstration of inflammation-induced cancer and cancer immunoevasion during primary tumorigenesis. *Proc Natl Acad Sci U S A* 2008;105:652–6.
- van de Vijver MJ, He YD, van't Veer LJ, Dai H, Hart AA, Voskuil DW, et al. A gene-expression signature as a predictor of survival in breast cancer. *N Engl J Med* 2002;347:1999–2009.
- Wang Y, Klijn JG, Zhang Y, Sieuwerts AM, Look MP, Yang F, et al. Gene-expression profiles to predict distant metastasis of lymph-node-negative primary breast cancer. *Lancet* 2005;365:671–9.
- Schmidt M, Bohm D, von Tonne C, Steiner E, Puhl A, Pilch H, et al. The humoral immune system has a key prognostic impact in node-negative breast cancer. *Cancer Res* 2008;68:5405–13.
- Desmedt C, Piette F, Loi S, Wang Y, Lallemand F, Haibe-Kains B, et al. Strong time dependence of the 76-gene prognostic signature for node-negative breast cancer patients in the TRANSBIG multicenter independent validation series. *Clin Cancer Res* 2007;13:3207–14.
- Shi L, Reid LH, Jones WD, Shippy R, Warrington JA, Baker SC, et al. The MicroArray Quality Control (MAQC) project shows inter- and intraplatform reproducibility of gene expression measurements. *Nat Biotechnol* 2006;24:1151–61.
- Haibe-Kains B, Desmedt C, Rothe F, Piccart M, Sotiriou C, Bontempi G. A fuzzy gene expression-based computational approach improves breast cancer prognostication. *Genome Biol* 2010;11:R18.
- Loi S, Haibe-Kains B, Majaj S, Lallemand F, Durbecq V, Larsimont D, et al. PIK3CA mutations associated with gene signature of low mTORC1 signaling and better outcomes in estrogen receptor-positive breast cancer. *Proc Natl Acad Sci U S A* 2010;107:10208–13.
- Wirapati P, Sotiriou C, Kunkel S, Farmer P, Pradervand S, Haibe-Kains B, et al. Meta-analysis of gene expression profiles in breast cancer: toward a unified understanding of breast cancer subtyping and prognosis signatures. *Breast Cancer Res* 2008;10:R65.
- Desmedt C, Di Leo A, de Azambuja E, Larsimont D, Haibe-Kains B, Sellselags J, et al. Multifactorial approach to predicting resistance to anthracyclines. *J Clin Oncol* 2011;29:1578–86.
- Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pages C, et al. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science* 2006;313:1960–4.
- Kaplan DH, Shankaran V, Dighe AS, Stockert E, Aguet M, Old LJ, et al. Demonstration of an interferon gamma-dependent tumor surveillance system in immunocompetent mice. *Proc Natl Acad Sci U S A* 1998;95:7556–61.
- Naito Y, Saito K, Shiiba K, Ohuchi A, Saigenji K, Nagura H, et al. CD8<sup>+</sup> T cells infiltrated within cancer cell nests as a prognostic factor in human colorectal cancer. *Cancer Res* 1998;58:3491–4.
- Street SE, Trapani JA, MacGregor D, Smyth MJ. Suppression of lymphoma and epithelial malignancies effected by interferon gamma. *J Exp Med* 2002;196:129–34.
- Casares N, Pequignot MO, Tesniere A, Ghiringhelli F, Roux S, Chaput N, et al. Caspase-dependent immunogenicity of doxorubicin-induced tumor cell death. *J Exp Med* 2005;202:1691–701.
- Teng MW, Andrews DM, McLaughlin N, von Scheidt B, Ngiew SF, Moller A, et al. IL-23 suppresses innate immune response independently of IL-17A during carcinogenesis and metastasis. *Proc Natl Acad Sci U S A* 2010;107:8328–33.
- Uno T, Takeda K, Kojima Y, Yoshizawa H, Akiba H, Mittler RS, et al. Eradication of established tumors in mice by a combination antibody-based therapy. *Nat Med* 2006;12:693–8.
- Rachitskaya AV, Hansen AM, Horai R, Li Z, Villasmil R, Luger D, et al. Cutting edge: NKT cells constitutively express IL-23 receptor and ROR $\gamma$  and rapidly produce IL-17 upon receptor ligation in an IL-6-independent fashion. *J Immunol* 2008;180:5167–71.
- Sutton CE, Lalor SJ, Sweeney CM, Brereton CF, Lavelle EC, Mills KH. Interleukin-1 and IL-23 induce innate IL-17 production from gamma-delta T cells, amplifying Th17 responses and autoimmunity. *Immunity* 2009;31:331–41.

37. van Beelen AJ, Zelinkova Z, Taanman-Kueter EW, Muller FJ, Hommes DW, Zaat SA, et al. Stimulation of the intracellular bacterial sensor NOD2 programs dendritic cells to promote interleukin-17 production in human memory T cells. *Immunity* 2007;27:660–9.
38. Crowe NY, Smyth MJ, Godfrey DI. A critical role for natural killer T cells in immunosurveillance of methylcholanthrene-induced sarcomas. *J Exp Med* 2002;196:119–27.
39. Gao Y, Yang W, Pan M, Scully E, Girardi M, Augenlicht LH, et al. Gamma delta T cells provide an early source of interferon gamma in tumor immunity. *J Exp Med* 2003;198:433–42.
40. Wolmark N, Wang J, Mamounas E, Bryant J, Fisher B. Preoperative chemotherapy in patients with operable breast cancer: nine-year results from National Surgical Adjuvant Breast and Bowel Project B-18. *J Natl Cancer Inst Monogr* 2001;96–102.
41. Ma Y, Aymeric L, Locher C, Matarollo SR, Delahaye NF, Pereira P, et al. Contribution of IL-17-producing {gamma}{delta} T cells to the efficacy of anticancer chemotherapy. *J Exp Med* 2011;208:491–503.
42. Ladoire S, Mignot G, Dabakuyo S, Arnould L, Apetoh L, Rebe C, et al. *In situ* immune response after neoadjuvant chemotherapy for breast cancer predicts survival. *J Pathol* 2011. [Epub ahead of print].
43. Doisne JM, Soulard V, Becourt C, Amniai L, Henrot P, Havenar-Daughton C, et al. Cutting edge: crucial role of IL-1 and IL-23 in the innate IL-17 response of peripheral lymph node NK1.1-invariant NKT cells to bacteria. *J Immunol* 2011;186:662–6.
44. Umemura M, Yahagi A, Hamada S, Begum MD, Watanabe H, Kawakami K, et al. IL-17-mediated regulation of innate and acquired immune response against pulmonary *Mycobacterium bovis* bacille Calmette-Guerin infection. *J Immunol* 2007;178:3786–96.
45. Teng MW, von Scheidt B, Duret H, Towne JE, Smyth MJ. Anti-IL-23 monoclonal antibody synergizes in combination with targeted therapies or IL-2 to suppress tumor growth and metastases. *Cancer Res* 2011;71:2077–86.
46. Kortylewski M, Xin H, Kujawski M, Lee H, Liu Y, Harris T, et al. Regulation of the IL-23 and IL-12 balance by Stat3 signaling in the tumor microenvironment. *Cancer Cell* 2009;15:114–23.
47. Stewart CA, Trinchieri G. Reinforcing suppression using regulators: a new link between STAT3, IL-23, and Tregs in tumor immunosuppression. *Cancer Cell* 2009;15:81–3.
48. Langowski JL, Zhang X, Wu L, Mattson JD, Chen T, Smith K, et al. IL-23 promotes tumour incidence and growth. *Nature* 2006;442:461–5.
49. Denkert C, Loibl S, Noske A, Roller M, Muller BM, Komor M, et al. Tumor-associated lymphocytes as an independent predictor of response to neoadjuvant chemotherapy in breast cancer. *J Clin Oncol* 2010;28:105–13.
50. Hornychova H, Melichar B, Tomsova M, Mergancova J, Urminska H, Ryska A. Tumor-infiltrating lymphocytes predict response to neoadjuvant chemotherapy in patients with breast carcinoma. *Cancer Invest* 2008;26:1024–31.

# Cancer Research

The Journal of Cancer Research (1916–1930) | The American Journal of Cancer (1931–1940)

## Pivotal Role of Innate and Adaptive Immunity in Anthracycline Chemotherapy of Established Tumors

Stephen R. Mattarollo, Sherene Loi, Helene Duret, et al.

*Cancer Res* 2011;71:4809-4820. Published OnlineFirst June 6, 2011.

**Updated version** Access the most recent version of this article at:  
doi:[10.1158/0008-5472.CAN-11-0753](https://doi.org/10.1158/0008-5472.CAN-11-0753)

**Supplementary Material** Access the most recent supplemental material at:  
<http://cancerres.aacrjournals.org/content/suppl/2011/06/06/0008-5472.CAN-11-0753.DC1>

**Cited articles** This article cites 49 articles, 26 of which you can access for free at:  
<http://cancerres.aacrjournals.org/content/71/14/4809.full#ref-list-1>

**Citing articles** This article has been cited by 5 HighWire-hosted articles. Access the articles at:  
<http://cancerres.aacrjournals.org/content/71/14/4809.full#related-urls>

**E-mail alerts** [Sign up to receive free email-alerts](#) related to this article or journal.

**Reprints and Subscriptions** To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at [pubs@aacr.org](mailto:pubs@aacr.org).

**Permissions** To request permission to re-use all or part of this article, use this link  
<http://cancerres.aacrjournals.org/content/71/14/4809>.  
Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.