Lactate Influx through the Endothelial Cell Monocarboxylate Transporter MCT1 Supports an NF-κB/IL-8 Pathway that Drives Tumor Angiogenesis

Frédérique Végran¹, Romain Boidot¹, Carine Michiels², Pierre Sonveaux¹, and Olivier Feron¹

Abstract
Lactate generated from pyruvate fuels production of intracellular NAD⁺ as an end result of the glycolytic process in tumors. Elevated lactate concentration represents a good indicator of the metabolic adaptation of tumors and is actually correlated to clinical outcome in a variety of human cancers. In this study, we investigated whether lactate could directly modulate the endothelial phenotype and thereby tumor vascular morphogenesis and perfusion. We found that lactate could enter endothelial cells through the monocarboxylate transporter MCT-1, trigger the phosphorylation/degradation of IκBα, and then stimulate an autocrine NF-κB/IL-8 (CXCL8) pathway driving cell migration and tube formation. These effects were prevented by 2-oxoglutarate and reactive oxygen species (ROS) inhibitors, pointing to a role for prolyl-hydroxylase and ROS in the integration of lactate signaling in endothelial cells. PHD2 silencing in endothelial cells recapitulated the proangiogenic effects of lactate, whereas a blocking IL-8 antibody or IL-8–targeting siRNA prevented them. Finally, we documented in mouse xenograft models of human colorectal and breast cancer that lactate release from tumor cells through the MCT4 (and not MCT1) transporter is sufficient to stimulate IL-8–dependent angiogenesis and tumor growth. In conclusion, our findings establish a signaling role for lactate in endothelial cells and they identify the lactate/NF-κB/IL-8 pathway as an important link between tumor metabolism and angiogenesis. Cancer Res; 71(7); 2550–60. ©2011 AACR.

Introduction
Tumor cells fuel their metabolism with a variety of nutrients including glucose, glutamine, and lactate to meet the bioenergetic and biosynthetic demands of proliferation (1, 2). Glycolysis coupling to the tricarboxylic acid (TCA) cycle and oxidative phosphorylations, is actually far to be the unique source of energy. Hypoxia and oncogenic mutations drive glucose to pyruvate transformation and then pyruvate to lactate conversion ensuring the regeneration of the NAD⁺ pool necessary to support a high glycolytic flux to fulfil the bioenergetic and biosynthetic demands of proliferation (1, 2). Glycerol, which is the source of energy. Hypoxia and oncogenic mutations drive glucose to pyruvate transformation and then pyruvate to lactate conversion ensuring the regeneration of the NAD⁺ pool necessary to support a high glycolytic flux to fulfil the bioenergetic and biosynthetic demands of proliferation (1, 2). Glycerol, which is an additional source of NAD⁺, can be generated from pyruvate, and then stimulate an autocrine NF-κB/IL-8 (CXCL8) pathway driving cell migration and tube formation. These effects were prevented by 2-oxoglutarate and reactive oxygen species (ROS) inhibitors, pointing to a role for prolyl-hydroxylase and ROS in the integration of lactate signaling in endothelial cells. PHD2 silencing in endothelial cells recapitulated the proangiogenic effects of lactate, whereas a blocking IL-8 antibody or IL-8–targeting siRNA prevented them. Finally, we documented in mouse xenograft models of human colorectal and breast cancer that lactate release from tumor cells through the MCT4 (and not MCT1) transporter is sufficient to stimulate IL-8–dependent angiogenesis and tumor growth. In conclusion, our findings establish a signaling role for lactate in endothelial cells and they identify the lactate/NF-κB/IL-8 pathway as an important link between tumor metabolism and angiogenesis. Cancer Res; 71(7); 2550–60. ©2011 AACR.

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proposed to prevent maturation of neoformed tubes (21). Because of the known HIF-mediated regulation of VEGF (22) and the capacity of glycolytic carboxylate intermediates including pyruvate and lactate to stimulate hypoxia-inducible factor (HIF) expression (23, 24), the effects of lactate on VEGF (and angiogenesis in general) are thought to be HIF-1α-dependent (16).

As major metabolic pathways in tumors end up with lactate secretion and because this may occur under both hypoxic and nonhypoxic conditions (Warburg effect), we reasoned that lactate could act as a signaling molecule in endothelial cells through not only HIF-dependent but also -independent pathways. Furthermore, although hypoxia-independent expression of HIF-1 is possible in tumor cells in response to oncogenic or loss-of-function mutations pathways (22, 25), this is not occurring in genetically stable endothelial cells. Also, the paradigmatic HIF1-dependent angiogenic growth factor VEGF is produced to much larger extent by tumor cells or tumor-associated macrophages than by endothelial cells (10- to 50-fold more at the protein level, unpublished data). Thus, to examine a possible autocrine pathway in endothelial cells triggered by lactate arising from tumor cells, we first used a low density array strategy to identify possible actors. This initial exploration led us to document that lactate could promote a PHD2- and ROS-dependent NF-kB (26).

Materials and Methods

Cell culture and treatments

Human colorectal adenocarcinoma WiDr cells and MDA-MB231 breast cancer cells were obtained from the American Type Culture Collection Cell lines where they are regularly authenticated based on viability, recovery, growth, morphology, and isoenzymology. WiDr cells and MDA-MB231 cells were acquired in 2009 and 2010, respectively, stored according to the supplier’s instructions, and used within 6 months after resuscitation of frozen aliquots. Human umbilical vein endothelial cells (HUVEC) were used within 6 months of purchase from Sigma, acting as a distributor for ECACC (European Collection for Cell Cultures). HUVECs were routinely cultured in endothelial cell basal medium (Sigma) and used between passages 2 and 5. In some experiments, cells were treated with an anti-IL-8-neutralizing antibody (R&D systems), evodiamine (Calbiochem), or N-acetyl-L-cysteine (Sigma) in glucose- or lactate-containing medium.

Mouse models

For in vivo experiments, NMRI nude mice (Elevage Janvier) were injected subcutaneously with 400 μL Matrigel containing one of the following combinations: (i) $5 \times 10^5$ tumor cells (WiDr or MDA-MB231) mixed with $5 \times 10^4$ HUVEC cells, with or without IL-8-blocking antibody; (ii) $5 \times 10^5$ WiDr x shMCT1 or $5 \times 10^5$ WiDr shMCT4 cells mixed with $5 \times 10^4$ HUVEC cells; (iii) $5 \times 10^5$ MDA-MB231 or $5 \times 10^5$ MDA-MB231 shMCT4 cells mixed with $5 \times 10^4$ HUVEC cells. Tumor sizes were tracked with an electronic calliper, and tumors were collected after the animal sacrifice 14 days after injection. In some experiments, local tumor blood flow was measured with a Laser Doppler imager (Moor Instruments) on anesthetized mice as previously described (26).

Real-time PCR and silencing experiments

PCR amplification was performed in an IQ5 Thermocycler (Bio-Rad); primers sequences are described in Supplementary Data. For gene expression profiling, Taqman Angiogenesis Gene signature Arrays (Applied Biosystems Inc.) were used according to the manufacturer’s instructions. For siRNA silencing experiments, HUVEC cells were transfected (LipofectA-MINE 2000; Invitrogen) with specific siRNA (Qiagen), whereas for shRNA silencing experiments, BLOCK-iT H1 RNAi Entry Vector Kit (Invitrogen) was used. Sequences are described in Supplementary Data.

IL-8 ELISA, NF-κB luciferase reporter, and DNA binding assays

The extent of IL-8 production by HUVEC and tumor xenografts was determined in an ELISA assay (QuantiGlo Chemiluminescent Immunomassay; R&D systems). The luciferase reporter pNF-κB-Luc plasmid (Stratagene) was transfected 24 hours before cell treatments; a GFP-expressing vector was cotransfected to adjust for variations in transfection efficiencies. DNA binding of NF-κB subunit p65 was determined using a TransAM ELISA kit (Active Motif).

Reactive oxygen species measurements and lactate measurements

Intracellular reactive oxygen species (ROS) were assessed using oxidation-sensitive probe 2′,7′-dichlorofluorescein diacetate (DCFH-DA; Sigma). Lactate concentration was determined via the lactate dehydrogenase (LDH)-catalyzed oxidation of lactate into pyruvate in the presence of NAD$^+$ (6).

Endothelial cell migration and tube formation assays

HUVEC cell mobility and migration were evaluated in a modified Boyden chamber assay using transwell filters inserts with 8-μm pore size (Greiner Bio-one). The formation of capillary-like endothelial tubes was determined in an in vitro assay by plating endothelial cells on growth factor-reduced Matrigel (BD Pharmingen), as previously reported (27, 28).

Immunostaining and immunoblotting

Tumors were cryosectioned and sections were probed with a rat monoclonal antibody against CD31 (BD Pharmingen) or rabbit polyclonal antibodies against MCT1 (Millipore) and MCT4 (Polyepide) followed by a secondary antibody coupled to an Alexa Fluor-568 as previously described (6, 29). For immunoblotting, cells extracts were separated on SDS-PAGE and transferred onto PVDF membranes before incubation with the MCT1 (Millipore), IκBα (Millipore) and phospho-serine 32 and36 IκBα (Cell Signaling) antibodies; gel loading was normalized with a beta-actin antibody (Sigma).
Statistics

Results are expressed as mean ± SEM. Student's t test, 1-way ANOVA (Tukey's post-hoc test), were used where indicated. *, $P < 0.05$ or **, $P < 0.01$ was considered statistically significant in the different experiments.

Results

MCT1-driven lactate uptake in endothelial cells promotes IL-8 expression

We first determined the capacity of lactate to be taken up by endothelial cells and to influence cell proliferation. We found that independent of the presence or the absence of glucose, the addition of 10 mmol/L lactate in the culture medium led to a net increase in intracellular lactate concentration reaching 6-fold the level of lactate observed in the presence of glucose-containing (lactate-free) medium (Fig. 1A). This elevated level of cytosolic lactate was reached after 1 hour of exposure to extracellular lactate and then maintained itself around the newly setup basal value, independently of the lactate concentration in the medium as long as established between 1 and 25 mmol/L (not shown). Although no sign of cell death (i.e., cell detachment) could be observed, the presence of lactate in the medium (instead of glucose) reduced endothelial cell proliferation rate by half when compared with glucose-containing medium (Fig. 1B). The presence of lactate together with glucose in the culture medium however did not alter endothelial cell proliferation (Fig. 1B). We then examined the influence of lactate on the endothelial expression profile of angiogenesis-related genes using dedicated TaqMan arrays on 96-well microfluidic cards. HUVEC were exposed to 10 mmol/L lactate for 6 hours to detect early changes in gene expression. We found that 7 transcripts were upregulated more than 2-fold in response to lactate and among them, IL-8/CXCL8 mRNA expression was the most responsive with an approximately 8-fold increase (vs. control HUVEC maintained in glucose-containing medium; Table 1). The increase in IL-8 gene expression was confirmed by qPCR analysis using (other) specific primers (Fig. 1C) and further validated at the protein level in a dedicated ELISA (Fig. 1D). To document a direct relationship between lactate influx and IL-8, we used a dedicated siRNA to silence the expression of the major
monocarboxylate transporter MCT1 in HUVEC (unpublished data). A downregulation of more than 90% of MCT-1 expression (vs. control siRNA conditions) was confirmed at the mRNA (Fig. 1E) and protein (Fig. 1F) levels. MCT1 silencing prevented lactate entry into endothelial cells (Fig. 1A) and lactate-induced IL-8 mRNA and protein expressions (Fig. 1C and D); MCT1 silencing reduced endothelial cell proliferation to a larger extent than lactate conditions (Fig. 1B). Of note, the ELISA test also revealed that hypoxia (1% O2) was not able to induce IL-8 in our experimental set-up (Fig. 1D).

### Table 1. Influence of lactate (10 mmol/L) on the mRNA expression profile of endothelial cells using TaqMan Gene Signature Array

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<th>Gene</th>
<th>Mean</th>
<th>SEM</th>
<th>Gene</th>
<th>Mean</th>
<th>SEM</th>
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<td>GAPDH</td>
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NOTE: Data (mean ± SEM; n = 3) are presented as fold-change versus control condition (glucose-containing medium).

Abbreviation: n.d. not detected.

Lactate stimulates the NF-κB/IL-8 pathway in endothelial cells

Lactate-Induced IL-8 Pathway in Endothelial Cells

IL-8 induction by lactate depends on the NF-κB pathway

The known regulation of IL-8 expression by NF-κB led us to examine whether this transcription factor could mediate the lactate effects. As a first insight on the influence of lactate on NF-κB activity, we performed a TransAM assay. This experiment confirmed a significant increase in DNA binding of the p65 subunit of the NF-κB transcription complex in lactate-exposed endothelial cells (Fig. 2A). We then used a NF-κB reporter assay by transfecting HUVEC with a NF-κB–responsive luciferase plasmid (together with a GFP-encoding plasmid for normalization). We found that lactate dramatically stimulated luciferase activity (17-fold increase vs. control glucose medium, P < 0.01; Fig. 2B). Also, as observed for the effects of lactate on IL-8 expression (Fig. 1C and D), MCT-1 silencing blunted the increase in NF-κB reporter activity (Fig. 2B).

The recent demonstration of an autocrine IL-8–induced activation of NF-κB in endothelial cells (30) led us to verify that in our model, IL-8 was acting downstream of the lactate-induced NF-κB activity and not the converse. We therefore repeated the aforementioned experiments in the presence of an anti-IL-8–blocking antibody (10 μg/mL; Fig. 2C) or using endothelial cells wherein IL-8 expression was silenced using dedicated siRNA (Fig. 2D); a downregulation of more than 90% (P < 0.01, n = 3) of IL-8 expression (vs. control siRNA conditions) was confirmed by qPCR. We found that independently of the mode of IL-8 inhibition, lactate strongly stimulated NF-κB activity in the luciferase reporter assays (Fig. 2C and D). The use of evodiamine (300 nmol/L) as a pharmacological inhibitor of NF-κB (acting by blocking IκBα kinase activation; ref. 31) further confirmed the stimulatory effect of lactate on this transcription factor in our experimental model (Fig. 2C).

Lactate stimulates the NF-κB/IL-8 pathway in a ROS- and IκBα-dependent manner

To further dissect the pathway linking lactate stimulation and NF-κB/IL-8 induction, we examined the role of ROS usually described as key triggers of NF-κB activity. We first treated endothelial cells with the ROS scavenger N-acetylcysteine (NAC) and found that in these conditions, both lactate-induced NF-κB activity (Fig. 3A) and IL-8 expression (Fig. 3B) were completely abrogated, as determined using the luciferase reporter assay and qPCR, respectively. The use of the ROS fluorescent dye 2′,7′-dichlorofluorescin diacetate confirmed that the exposure of endothelial cells to 10 mmol/L lactate led to a rapid increase in ROS production (vs. control glucose conditions; Fig. 3C). Importantly, a similar increase in ROS production was observed when cells were exposed to both glucose and lactate (Fig. 3C). Because ROS-mediated activation of NF-κB is classically proposed to be dependent on IκBα phosphorylation/degradation (which favors the release of active NF-κB subunits; 32), we then examined the effect of lactate on IκBα serine 32 and 36 phosphorylations. We found that lactate supplementation (alone or together with glucose)
led to a net increase in IκBα phosphorylation, a process that we also found to be associated with IκBα degradation, as documented by immunoblotting (Fig. 3D).

2-oxoglutarate inhibits while PHD2-silencing mimics the effects of lactate on the NF-κB/IL-8 signaling pathway

A prolyl hydroxylase (PHD)-dependent activation of NF-κB activity was previously reported to occur through the IκBα pathway (33–35). Considering that PHD activity requires 2-oxoglutarate as cosubstrate and may thereby be influenced by other carboxylate metabolic intermediates (see Introduction), we examined whether in endothelial cells, lactate competition with 2-oxoglutarate could account for the observed increase in NF-κB/IL-8 expression. We found that 2-oxoglutarate dose-dependently prevented lactate-induced NF-κB activity (Fig. 4A), as determined in a luciferase reporter assay. We also found that PHD2-silencing in endothelial cells led to a net increase in both NF-κB activity (Fig. 4B) and IL-8 expression (Fig. 4C) under basal conditions (i.e., in lactate-free glucose-containing medium) and failed to further stimulate the NF-κB/IL-8 expression pathway in response to lactate (Fig. 4B and C). Downregulation of more than 90% of PHD-2 expression (vs. control siRNA conditions) was confirmed by qPCR (Fig. 4D). We also documented that lactate-induced IL-8 expression was strongly inhibited by 2-oxoglutarate and diphenylene iodonium chloride (DPI), a potent inhibitor of NADPH oxidase (a major enzymatic source of ROS in endothelial cells; Fig. 4C). Interestingly, PHD2-silencing failed to stimulate IL-8 expression in glucose-containing medium when DPI was present (Fig. 4C), suggesting that both PHD2 inhibition and ROS production are required to promote the NF-κB pathway in endothelial cells.

MCT4-driven release of lactate from cancer cells stimulates IL-8 production together with tumor angiogenesis and perfusion

To examine the NF-κB/IL-8–driven effects of lactate in vivo, we used tumor cells genetically modified for their capacity to handle lactate. Accordingly, we established clones from the WiDr human colorectal cancer cell line selected for stable expression of shRNA directed against MCT1 or MCT4. We found that WiDr cells express both types of MCT transporters and that the use of dedicated shRNA led to the silencing of MCT1 expression (91.2%/C66% downregulation; Fig. 5A) and MCT4 (96.5%/C69% downregulation) (Fig. 5B), respectively. We found that although MCT1-silencing did not prevent the efflux of lactate induced under hypoxia (Fig. 5A, bottom), MCT4-silencing had a dramatic effect on the capacity of WiDr cells to release lactate in the same hypoxic conditions (Fig. 5B, bottom). Measurements of cell proliferation rate in vitro however did not reveal any difference between shMCT1 and shMCT4 WiDr cell clones (Fig. 5C). Of note, the extents of lactate release in shMCT1 WiDr cells and sham-transfected WiDr were not significantly different, in agreement with a major role of this transporter in the influx—and not the efflux—of lactate in this cell type (6).
To validate the functional consequences of a deficit in lactate release, we first used the conditioned medium (CM) collected from either shMCT1 or shMCT4 WiDr cells to treat endothelial cells. We found that IL-8 expression (as determined by qPCR) was stimulated by CM collected from hypoxic shMCT1 cells but not altered by the exposure to CM from hypoxic shMCT4 cells (Fig. 5D). In another set of experiments, we injected s.c. plugs of Matrigel containing $5 \times 10^5$ HUVEC together with either $5 \times 10^5$ WiDr shMCT1 cells or $5 \times 10^5$ WiDr shMCT4 cells. We found that tumor growth from the plug containing shMCT4 WiDr cells was reduced (vs. shMCT1 cells containing plug; Fig. 5E). We also verified that tumor lactate (Fig. 5F) and IL-8 (Fig. 5G) contents were reduced in shMCT4-expressing tumors (vs. shMCT1 tumors). Importantly, this was associated with a dramatic inhibition of angiogenesis as determined by CD31 immunostaining of vascular structures, together with a reduction in tumor blood flow as measured by laser Doppler imaging (Fig. 5H, see bottom for quantification). We repeated these experiments using a MDA-MB231 breast cancer cell clone wherein MCT4 was also silenced and found a similar phenotype (i.e., reduced CD31-labeled microvascular density and tumor perfusion; see Fig. 5I). For both tumors, the use of a CD31 antibody selective for mouse tissues (36) did, however, not lead to any specific staining, indicating that the vascular signal was of human origin and thus corresponded to HUVEC.

Lactate-dependent induction of IL-8 expression supports angiogenesis and tumor growth

To dissect the NF-κB/IL-8–driven proangiogenic effects of lactate, we first measured the migration of HUVEC in a modified Boyden chamber. Migration was determined after 24 hours by counting DAPI-stained cells on the bottom face of the filter. To first validate the IL-8 capacity to drive cell migration in this assay, recombinant IL-8 protein was added to glucose-containing medium in the upper part of the well. A 10-fold increase in the number of migrated endothelial cells was observed (vs. untreated conditions; Fig. 6A). Replacement of glucose by lactate (10 mmol/L) in the medium of the upper chamber led to a similar stimulation of endothelial cell migration. This promigratory effect of lactate was IL-8–dependent, as migration was completely blocked by either a dedicated IL-8 siRNA cell pretreatment or the presence of a blocking anti-IL-8 antibody in the upper well compartment (Fig. 6A). The use of NAC and evodiamine similarly reduced the capacity of lactate to promote the motility of endothelial cells (Fig. 6A).

We next examined how interfering with lactate cellular uptake and consecutive increase in IL-8 expression could influence the capacity of endothelial cells to form tubes when plated on Matrigel. We first showed that lactate exposure stimulates the formation of an endothelial network in these experimental conditions (Fig. 6B). We then documented that siRNA directed against either MCT-1 or IL-8 prevented...
lactate-driven formation of tubes (Fig. 6B and C, respectively, for quantification). Importantly, the addition of recombinant IL-8 to IL-8–siRNA–treated endothelial cells rescued tubulogenesis (Fig. 6B). As observed in Figure 6A, the use of NAC and evodiamine confirmed a role for ROS/NF-κB in the lactate-dependent IL-8–driven formation of endothelial tubes (Fig. 6C). Also, in adequation with results of Figure 4A and B, 2-oxoglutarate blunted the tubulogenic response to lactate and PHD2 silencing in glucose-containing medium recapitulated the effects of lactate with a strong stimulation of tube formation (Fig. 6C).

Finally, to further validate the in vivo relevance of the lactate/IL-8 pathway in endothelial cells, we injected s.c. 2 plugs of Matrigel containing 5 × 10^5 wild-type WiDr cells and 5 × 10^4 HUVEC with or without IL-8-blocking antibody. In this tumor model of lactate-driven angiogenesis, we found that the presence of anti-IL-8–blocking antibodies within the Matrigel plug delayed tumor growth (Fig. 6D) and considerably reduced the development of the tumor vasculature, as revealed by anti-CD31 immunostaining (Fig. 6E, top: −75 ± 13% vs. control, P < 0.01, n = 5). Finally, to prove the role of MCT1 in vivo, we similarly co-injected WiDr cells and HUVEC pretreated with MCT1 siRNA. We found that tumor growth and development of the microvascular network (−66 ± 21% vs. control, P < 0.05, n = 5) were both inhibited in response to MCT1 silencing in endothelial cells (Fig. 6D and E, bottom).

Discussion

Major findings of this study are the identification of a lactate/NF-κB signaling pathway in endothelial cells, the existence of a lactate-driven, feed-forward IL-8 autocrine loop driving angiogenesis in tumors and the key roles of monocarboxylate transporters MCT1 and MCT4 in this lactate-based dialog between cancer cells and endothelial cells. Our data indeed support a model according to which lactate released from tumor cells through MCT4 may be taken up by endothelial cells via the MCT1 transporter and consecutively stimulate angiogenesis through NF-κB/IL-8 signaling (Supplementary Fig. S1); all these lactate-driven signaling events may importantly occur in the presence of glucose.

We identified key players acting upstream and downstream of this pathway in endothelial cells. We found that ROS were...
produced in endothelial cells in response to lactate and that ROS inhibitors prevented lactate-induced activation of NF-κB and IL-8 expression. Also, we showed that the PHD cosubstrate 2-oxoglutarate could prevent the effects of lactate on NF-κB activation and that conversely, PHD2 silencing recapitulated them in normal glucose conditions. We further documented the stimulatory effects of lactate on IκBα phosphorylation and consecutive degradation in endothelial cells, a process known to promote translocation of freed NF-κB activation into the nucleus for transcriptional regulation (32). Using siRNA directed against IL-8 (rescued or not with recombinant IL-8), we showed that this autocrine pathway...
accounts for most of the proangiogenic effects of lactate in endothelial cells. Importantly, we validated in vivo that the release of lactate by cancer cells can effectively contribute to the development of the tumor vasculature by stimulating the IL-8 pathway. A blocking antibody directed against IL-8 could indeed prevent the stimulation of angiogenesis (Fig. 6E) and reduce tumor growth (Fig. 6D) in a model where lactate efflux through the MCT4 transporter is a key trigger of the development of tumor vasculature (Fig. 5H and I).

Lactate was previously documented to stimulate angiogenesis through activation of the VEGF/VEGFR2 signaling pathway (16–20). Although it may be difficult to discriminate between the role of hypoxia and lactate in triggering VEGF signaling in vivo, our finding of NF-κB acting as a lactate-responsive transcription factor unravels a pathway which may link tumor cell metabolism and angiogenesis independently of the O₂ conditions. An interesting parallel may be drawn with previous studies by Verma and colleagues who reported that pyruvate, lactate, and several TCA cycle intermediates can stabilize HIF-1 through inhibition of PHD enzymes (23, 24). These data are proposed to support the Warburg effect, that is, the capacity of nonhypoxic tumor cells to exploit glycolysis without a need for coupling to oxidative phosphorylations to support the high biosynthetic and energy demands of proliferating cells. Our study allows to extend this paradigm to tumor angiogenesis and supports a central role of lactate bridging avid glucose metabolism and key aspects of malignancy, namely tumor cell proliferation and angiogenesis (Supplementary Fig. S1).

Interestingly, reduction in PHD activity appears to be the common denominator in the stabilization of HIF and NF-κB observed in response to lactate. In particular, our data are in
line with the work of Cummins and colleagues (34) who previously showed that hypoxia may activate NF-κB through a decreased PHD1-dependent hydroxylation of IKKβ and consecutive phosphorylation-dependent degradation of IκBα. In a more recent study, Chan and colleagues, starting from the observation of a frequent loss of PHD2 expression in cancers, documented that the tumor suppressor gene potential of this hydroxylase was associated with an increased NF-κB activation in tumor cells (33). To reproduce the loss of PHD2 expression, these authors used PHD2-silenced cancer cells and documented that mouse injection with these modified tumor cells led to the development of more vascularized tumors. The effects of lactate observed in our study therefore mimic the effects of a downmodulation of PHD2, leading in fine to the activation of NF-κB–driven angiogenic cascades. Interestingly, we found that lactate-driven angiogenesis leads in vivo to a net increase in tumor perfusion indicating that the stimulated NF-κB/IL-8 pathway also contributes to the maturation of the tumor neovasculature. These data may be related to a recent study by Mazzzone and colleagues who documented that heterozygous deficiency of PHD2 normalized the tumor endothelial lining, resulting in improved tumor perfusion (37). The same authors further documented that haploinsufficiency of PHD2 restores tumor oxygenation and thus possibly induce a metabolic shift to a more oxidative, less malignant phenotype. Similarly, our data support the hypothesis that in response to lactate exposure, endothelial cells may exhibit alterations in their metabolic phenotype leading to increased cell survival in glucose-free conditions (Fig. 1B). The ROS production observed in endothelial cells in response to lactate could actually arise from the activation of the NAD(P)H oxidase by NADH accumulation resulting from the conversion of lactate into pyruvate, as we previously reported in tumor cells (6).

In conclusion, our study provides a new rationale for associating elevated lactate concentrations in tumors and negative outcomes for patients. These results further support the current enthusiasm for new cancer treatments targeting metabolic pathways (38). In particular, inhibitors of MCT1 and/or MCT4 bear the promise of interfering with the subtle exchanges of lactate between tumor cells and endothelial cells, and to indirectly attenuate the IL-8 effect on tumor vascular development and associated tumor burden.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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References

Lactate Influx through the Endothelial Cell Monocarboxylate Transporter MCT1 Supports an NF-κB/IL-8 Pathway that Drives Tumor Angiogenesis

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