CXCR4 Activation Defines a New Subgroup of Sonic Hedgehog–Driven Medulloblastoma

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Abstract

Medulloblastoma prognosis tends to be poor, despite aggressive therapy, but defining molecular subgroups may identify patients who could benefit from targeted therapies. This study used human gene array and associated clinical data to identify a new molecular subgroup of medulloblastoma characterized by coactivation of the Sonic hedgehog (SHH) and CXCR4 pathways. SHH–CXCR4 tumors were more common in the youngest patients where they were associated with desmoplastic histology. In contrast to tumors activating SHH but not CXCR4, coactivated tumors exhibited greater expression of Math1 and cyclin D1. Treatment with the CXCR4 antagonist AMD3100 inhibited cyclin D1 expression and maximal tumor growth in vivo. Mechanistic investigations revealed that SHH activation stimulated CXCR4 cell surface localization and effector signaling activity, whereas SHH absence caused CXCR4 to assume an intracellular localization. Taken together, our findings define a new medulloblastoma subgroup characterized by a functional interaction between the SHH and CXCR4 pathways, and they provide a rationale to clinically evaluate combined inhibition of SHH and CXCR4 for medulloblastoma treatment. Cancer Res; 72(1); 122–32. ©2011 AACR.
AMD3465 blocked the intracranial growth of xenografts of the desmoplastic medulloblastoma-derived cell line Daoy, suggesting that CXCR4 activation may also be important for maximal SHH-driven medulloblastoma growth (21).

Herein we report that overexpression of CXCR4 occurs in the SHH subgroup of medulloblastoma. Moreover, in vitro as well as in vivo data indicate that the SHH pathway enhances CXCR4 surface localization and signaling, resulting in a progrowth transcriptional response, maximal tumor growth, and a sensitivity to the antitumor effects of AMD3100. On the basis of these findings, we propose an additional molecular subgroup of medulloblastoma in which the SHH and CXCR4 pathways synergize to promote tumor growth, and for which combined SHH and CXCR4 antagonism may provide a therapeutic benefit.

Materials and Methods

Animals were used in accordance with an established Animal Studies Protocol approved by The Washington University School of Medicine Animal Studies Committee. Chemicals were obtained from Sigma unless otherwise noted.

Cell culture

Primary ND2:SmoA1 medulloblastoma isolates. Tumors were isolated from SmoA1 mice by surgical dissection. Meninges were removed and the tissue was dissociated with Accutase enzyme cell detachment medium (LifeSpan) containing 125 units/mL DNase (3× for 15 minutes). Cells were strained through a 70-μm nylon mesh (Falcon) and plated onto poly-D-lysine–coated (20 μg/mL) plates (2 × 10^6/mL) in Dulbecco’s Modified Eagle’s Medium/F12 (Life Technologies) containing 2-hydroxypropyl-β-cyclodextrin (HPBCD) and washed with PBS. Activated Gαi was quantified by a cell growth assay using an anti-Gαi antibody.

Human medulloblastoma (Daoy) cells were obtained from American Tissue Culture Collection and used at low passage (<10) as previously described (21).

GNP cells. Primary cultures of purified GNPs were prepared from postnatal day 6 wild-type C57/BL6 mice as previously described (21).

Growth factor and drug treatments

In vitro treatments. SmoA1 tumor, GNPs, Daoy, 293T, or HOSX4 cells were serum starved (24 hours) and treated with 1 μg/mL CSCL12 (PeproTech). Pretreatments were with/without 1 μg/mL SHH (R&D) or 10 μmol/L cycloamine [in 2% sodium phosphate/citrate buffer (pH = 3) containing 2-hydroxypropyl-β-cyclodextrin (HPBCD)] for 12 hours prior to CSCL12 treatment.

AMD3100 treatment of ND2:SmoA1 flank xenografts. Subcutaneous flank implants of 4 × 10^6 SmoA1−/− medulloblastoma cells in Matrigel were made in nude mice (Jackson). Tumor growth was monitored using serial caliper-based volumetric measures. Mice harboring tumors that exhibited steady growth over a 4-week period and were approximately 10 mm × 10 mm × 10 mm underwent subcutaneous insertion of Alzet minipumps filled with AMD3100 (30 μg/mL; n = 7) or PBS (n = 5). Serial tumor volume measurements continued until mice were euthanized 11 days after initiating treatment. Tumors were isolated and fixed in For immunohistochemistry or lysed in TRIzol for RNA analysis.

cAMP measurement

Intracellular cAMP concentrations in SmoA1 medulloblastoma or Daoy cells pretreated with/without cycloamine (10 μmol/L, 12 hours) then CSCL12 (1 μg/mL) for varying times (0–20 minutes) were determined as described (22).

Western blot analyses

Whole-cell lysates. SDS-PAGE of cell lysates (40 μg/lane) was carried out as described (21).

Gαi activation assays. Activated Gαi pull-down assays (NewEast Biosciences) followed the manufacturer’s protocol. In brief, serum-starved cells (48 hours) were stimulated with CSCL12 (1 μg/mL, 10 minutes). Cell lysates were incubated for 1 hour with an anti-active Gαi antibody and Protein A-agarose beads. Activated Gαi was quantified by Western blot analysis using an anti-Gαi antibody.

Cell surface protein purification. Daoy cells (10 × 10^5) were pretreated with cycloamine or vehicle for 12 hours followed by CSCL12 (1 μg/mL, 30 minutes) or vehicle. The Cell Surface Protein Isolation Kit (Pierce) was used as described (23).

The following antibodies were used in Western blot analyses: anti-CXCR4 (Leinco Tech Inc. (1:1,000)), anti-Akt, anti-phospho-Akt (Ser473), anti-ERK, and anti-phospho-ERK (Cell Signaling; 1:1,000); anti-β-actin (Sigma; 1:10,000). Odyssey V3.0 (LI-COR) image acquisition/analysis system was used for detection and quantification of fluorescent bands. Normalization was to Actin or total content of specific proteins as indicated. Data are expressed as percentage of control (mean ± SEM). Statistical analysis was carried out by one-way ANOVA. P values are reported in legends.

Measurement of changes in intracellular calcium

Postnatal day 6 GNPs were cultured on poly-D-lysine–coated glass bottom culture dishes (MatTek Corporation) and pretreated with/without SHH (1 μg/mL) for 12 hours. Daoy cells were pretreated with cycloamine (10 μmol/L) or vehicle for 12 hours. Calcium flux was measured as described (17). Changes in cytosolic-free calcium were determined by monitoring excitation fluorescence using a dual-wavelength excitation source fluorimeter (Fura-2 filter, TILL microscope, TILL photonics GmbH Lochemer Schlag 19 Germany) in response to sequential excitation at 340 nm and 380 nm. Data are presented as the relative ratio of fluorescence at 340 and 380 nm.

Cell proliferation assay

A total of 1 × 10^5 SmoA1 tumor cells cultured in poly-D-lysine–coated 96-well flat-bottom plates were treated with AMD3100 (2.5 ng/mL) or CSCL12 (1 μg/mL) as indicated. Proliferation was determined per manufacturer’s instructions by the CellTiter 96 AQueous One Cell Proliferation Assay System (Promega) and a μQuant microplate Reader (Bio-Tek Instruments).
Reverse transcription PCR

Complementary DNA and quantitative PCR was carried out with RNA isolated from flank tumors as described (22). Reaction conditions included: 95°C (10 minutes), followed by 40 cycles of 95°C (30 seconds) and various annealing temperatures (1 minute), a final extension at 72°C (1 minute), followed by a melting curve from 55 to 95°C. No-template controls were negative. All reactions were normalized to a glyceraldehyde-3-phosphate dehydrogenase (GAPDH) control on the same plate. Fold change was calculated by the formula \(2^{(-\Delta\Delta C_t)}\).

Immunohistochemistry

Human medulloblastoma specimens were obtained from the Department of Pathology at Washington University (St. Louis, MO) per an Institutional Review Board–approved protocol for the use of archived pathologic specimens. Staining was accomplished as described (24) with the addition of Tyramide Signal Amplification (Perkin Elmer). Antibodies used and dilutions; CXCR4 clone 12G5 at 1:6,500 (R&D Systems), CXCR7 clone 11G8 at 1:1,000 (kind gift from Mark Penfold, Chemocentryx), CXCL12 at 1:500 (PeproTech), and Mouse IgG 1 μg/mL (Jackson Immunoresearch).

Human tissue samples, RNA preparation

Primary medulloblastoma \(n = 112\) and normal cerebella \((n = 9, \text{adult } n = 5)\) were obtained in accordance with the Hospital for Sick Children (Toronto, Canada) Research Ethics Board and profiled on Affymetrix GeneChip Human Exon 1.0ST arrays by the Toronto Centre for Applied Genomics (TCAG) as published (4). Molecular subgroups were established as previously described for 103 of 112 samples (4).

CXCR4-high versus -low medulloblastoma class rationale

Samples were divided into CXCR4-high or -low expressers on the basis of their expression patterns relative to average fetal cerebellar levels in a relaxed and stringent analysis. Tumors with more than 1.5-fold (relaxed) and more than 2-fold (stringent) expression levels were classified as CXCR4-high expressers and those with less than –1.5-fold (relaxed) or less than –2-fold (stringent) as CXCR4-low expressers.

Molecular analysis of CXCR4-high versus -low tumors

To detect gene expression changes associated with CXCR4-high versus -low expression, Gene Set Enrichment Analysis (GSEA; ref. 25) was used. As previously discussed (4), a comparative enrichment score for ranked genes in CXCR4-high versus -low expressers was calculated using 1,000 permutations permitting the detection of statistically significant genes and pathways associating with each phenotype.

Results

CXCR4 is overexpressed in the SHH subgroup of medulloblastoma

Gene expression profiling has established 4 molecular subgroups of medulloblastoma: WNT, SHH, group C, and group D, with distinct demographics, clinical presentation, transcriptional profiles, genetic abnormalities, and outcome (4). The transcriptional profile of the SHH subgroup resembles an early developmental stage of cerebellar GNPs when their proliferation is driven by SHH (26). During early development, SHH-induced GNP proliferation is modulated by multiple factors including CXCR4, a G-protein–coupled receptor (GPCR), also implicated in medulloblastoma growth (18, 21). We evaluated CXCR4 expression in normal human fetal and adult cerebellum, and across medulloblastoma subtypes. CXCR4 expression was high in the fetal cerebellum and declined significantly with age. A broad range of CXCR4 expression levels were evident in medulloblastoma with Group C and D tumors exhibiting low level, adult cerebellum-like CXCR4 expression, and WNT and SHH subtype tumors exhibiting high level, more fetal cerebellum-like levels of expression (Fig. 1A). While CXCR4 was present in both the WNT and SHH subtype tumors, only the SHH subtype tumors exhibited CXCR4 overexpression relative to the normally high levels found in the fetal cerebellum.

The SHH medulloblastomas could be further categorized into CXCR4-high and -low expressing subgroups (Fig. 1B). The majority of SHH medulloblastomas belonged to the CXCR4-high subgroup (~80%). The level of CXCR4 expression was correlated with other prognostically important features of disease such as histology and age. Desmoplastic tumors were exclusively in the CXCR4-high subgroup. Anaplastic large cell tumors occurred at a rate of only 4% in the CXCR4-high, compared with 25% in the CXCR4-low subgroup. In addition, nearly half of the CXCR4-high tumors occurred in infants, whereas only 13% of CXCR-low tumors occurred in this age group.

While rare mutations in CXCR4 have been described in medulloblastoma (27), CXCR4 activation appears to be ligand dependent. The CXCR4 ligand, CXCL12 was detected in genomic profiling of the SHH subgroup, and as previously described (18, 21), was primarily limited to the endothelium of tumor-associated blood vessels (Fig. 1C). Recently, a second CXCL12 receptor, CXCR7, was identified and shown to be expressed and functional in certain brain tumors such as glioblastoma (28). To determine whether CXCR7 might also mediate CXCL12 effects in the SHH subgroup of medulloblastoma we analyzed genomic data as well as tissue sections for evidence of CXCR7 expression. Little to no CXCR7 mRNA expression was detected in microarray analyses. Immunohistochemical staining of tumor sections from 3 separate human desmoplastic medulloblastoma specimens revealed CXCR7 expression primarily limited to endothelial cells (Fig. 1C). This was in contrast to CXCR4 expression, which as previously described, was evident at high levels throughout the specimens (18, 21). Thus CXCR4 appears to be the sole mediator of CXCL12 effects on medulloblastoma.

SHH pathway activation is required for CXCR4 signaling in medulloblastoma

Understanding how the SHH and CXCR4 pathways interact could elucidate new mechanisms of medulloblastoma genesis and identify novel therapeutic targets. A comparison of CXCR4 activation between postnatal day 6 cerebellar...
GNPs and Daoy cells revealed that Daoy cells, which possess constitutive activation of the SHH pathway exhibit significantly stronger responses to CXCL12 than GNPs (21). We postulated that among the differences between GNPs and Daoy cells were the degree of SHH pathway activation and that SHH would potentiate CXCR4 function. To test this hypothesis we evaluated whether blockade of SHH signaling in medulloblastoma cells abrogated the effects of CXCL12. For these studies we used primary tumor cells derived from spontaneous medulloblastoma that arise in ND2:SmoA1 mice which possess an activating mutation in smo, constitutive SHH pathway signaling, and a high incidence of medulloblastoma (29). We treated dissociated SmoA1 tumor cells in vitro for 12 hours with the SHH pathway antagonist cyclopamine or vehicle and measured the effects on CXCL12-induced cAMP suppression. CXCL12 stimulation resulted in a significant reduction of intracellular cAMP levels, which was completely blocked by pretreatment with cyclopamine, but not its inactive stereo-isomer, tomatidine (Fig. 2A). The effect of SHH had specificity for CXCR4 as cyclopamine pretreatment had no effect on cAMP elevation in response to PACAP, the ligand for the Gtαi-coupled GPCR, PAC1 (Fig. 2B; ref. 30).

The earliest stages of CXCR4 signaling involve Gtαi activation. Thus, we examined the effect of cyclopamine on CXCR4-Gtαi coupling. Intracellular Gtαi activation was measured by immunoprecipitation of the GTP-bound (activated) form of Gtαi. Acute stimulation of SmoA1 tumor cells with CXCL12 (1 μg/mL, 10 minutes) resulted in a significant increase in Gtαi activation, which was blocked by cyclopamine pretreatment (Fig. 2C and D). These data indicated that CXCL12-induced cAMP suppression in SmoA1 cells was a direct consequence of CXCL12-induced Gtαi activation and that the SHH pathway is required for this action. Together, these data support the hypothesis that mutational activation of SHH pathway sensitizes CXCR4 function.

We used Daoy cells to further explore the molecular basis for SHH effects on CXCR4 signaling. Similar to SmoA1 cells, CXCL12-induced activation of Gtαi and its downstream effectors in Daoy cells was dependent upon activation of the SHH pathway (Supplementary Fig. S1A and S1B). CXCL12-induced cAMP suppression (Supplementary Fig. S1C), F-actin polymerization (Supplementary Fig. S1D), calcium mobilization (Supplementary Fig. S1E and S1F), and extracellular signal-regulated kinase (ERK) 1/2 and Akt activation (Supplementary Fig. S1G and S1H). Inhibition of CXCR4 function was not the result of changes in total CXCR4 expression which was unaffected by cyclopamine (Supplementary Fig. S1G).
To address the possibility that cyclopamine might have direct off-target effects on CXCR4, we treated HOSX4 cells with CXCL12 in the presence and absence of cyclopamine. HOSX4 cells are an osteosarcoma cell line that overexpresses CXCR4 but does not express components of the SHH pathway such as Ptc and Smo. In these cells, CXCL12-induced ERK1/2 phosphorylation was unaffected by cyclopamine (Supplementary Fig. S3A and S3B). Thus, CXCR4 signaling is not always dependent upon SHH action and cyclopamine has no direct effects on CXCR4.

**CXCR4 function is SHH dependent in GNP s**

To determine whether regulation of CXCR4 by the SHH pathway might be limited to transformed cells, we cultured normal GNP s in the presence or absence of SHH for 12 hours and measured CXCL12-stimulated calcium flux. In the absence of continuous SHH treatment, SHH pathway activation is downregulated in GNP s by 12 hours postisolation. This is evident in mRNA abundance of the SHH transcriptional target Gli1 (Supplementary Fig. S3A). CXCL12 treatment of GNP s in the absence of SHH had little to no effect on calcium flux (Fig. 3A). In contrast, pretreatment of GNP s with SHH for 12 hours sensitized them to CXCL12 treatment, resulting in substantial CXCL12-induced calcium flux (Fig. 3A and B). Thus, SHH pathway activation can promote CXCR4 pathway responsiveness even in normal, nonneoplastic cells.

It is possible that SHH effects on GNP s were not the result of direct effects of SHH on CXCR4 but rather a consequence of SHH’s mitogenic effects and expansion of a CXCL12-responsive subpopulation of cells. To exclude this possibility, we carried out parallel studies in 293T cells. 293T cells exhibited basal and SHH-induced activation of the SHH pathway as evidenced by both SHH-stimulated and cyclopamine-inhibited Gli1 expression (Supplementary Fig. S3B–S3D). Neither SHH nor cyclopamine had any effect on 293T cell number (Supplementary Fig. S3E). Basal responsiveness of the CXCR4 pathway was also evident as CXCL12 treatment resulted in G0i activation, cAMP suppression and phosphorylation of ERK1/2 and Akt (Supplementary Fig. S4A–S4F). Consistent with SHH pathway–dependent activation of CXCR4 signaling, each of these CXCL12 effects was blocked by pretreatment with cyclopamine. Therefore, we conclude that the SHH pathway directly modulates CXCR4 signaling rather than merely expanding the relative abundance of a CXCL12-responsive subpopulation.

Together, these in vitro studies clearly indicate that crosstalk between the SHH and CXCR4 pathways, in both normal and neoplastic cells, can be an important regulator of CXCR4 signaling.

**SHH pathway regulates cell surface expression of CXCR4**

A potential mechanism for regulating CXCR4 coupling to G0i activation is through control of CXCR4 cell surface localization. To quantitate SHH pathway effects on CXCR4 surface localization, we carried out surface biotinylation and surface protein isolation on Daoy cells treated with CXCL12 and
cyclopamine. CXCL12 stimulation for 30 minutes led to significant loss of surface CXCR4 through internalization (Fig. 4A and B). Interestingly, exposure to cyclopamine for 12 hours also significantly reduced cell-surface CXCR4, which was further reduced upon subsequent CXCL12 treatment. 

Cell surface CXCR4 levels are determined by the balance between receptor internalization and insertion in the plasma membrane. These data suggest that cyclopamine might block the membrane insertion of CXCR4, leading to an overall reduction in surface receptor levels. Neither CXCL12 nor cyclopamine had any effect on the surface levels of PAC1, again suggesting that SHH effects have specificity for CXCR4 rather than for GPCRs in general. Taken together, these data suggest that SHH modulates CXCR4 signaling by regulating the cell surface fraction of CXCR4.

CXCR4 is necessary for maximal growth of SHH-driven medulloblastoma

CXCL12 has a positive effect on SHH-induced GNP proliferation (17). To determine whether crosstalk between the SHH and CXCR4 pathways similarly affected the growth of medulloblastoma cells, we measured proliferation of SmoA1 cells in response to CXCL12. Addition of CXCL12 to primary tumor cultures for 12 hours had a small but significant effect on cell proliferation. This effect was completely blocked by the CXCR4 antagonist AMD3100 suggesting that maximal tumor cell proliferation may require both the SHH and CXCR4 pathways (Fig. 5A).

To test whether CXCR4 enhanced medulloblastoma growth in vivo, we examined the effect of AMD3100 treatment on the growth of SmoA1 xenografts. Primary SmoA1 tumor cells were isolated and injected into the flanks of nude mice. Tumor volume was monitored over a 4-week period.
period and animals that exhibited steady tumor growth underwent implantation of subcutaneous osmotic infusion pumps filled with either PBS or AMD3100. Control animals (PBS group) all exhibited an increase in tumor volume over the 2-week experimental period. AMD3100 treatment led to significant inhibition of tumor growth indicating that CXCR4 potentiates SHH-driven medulloblastoma growth in vivo (Fig. 5B).

To further examine the importance of CXCR4 to medulloblastoma growth, we treated Ptc+/−/math1GFP mice with AMD3100. Medulloblastoma develops in about 15% of these mice with a peak incidence between 16 and 24 weeks (10, 15, 16, 31, 32). Six tumors developed in the PBS (n = 56) and 8 in the AMD3100 (n = 57) group. While not statistically different, the median time to tumor presentation was delayed to 14.9 weeks in the AMD3100 group compared with 10.6 weeks in the PBS group (Supplementary Fig. S5B).

Coactivation of SHH and CXCR4 pathways results in a unique expression profile in medulloblastoma

The correlation between CXCR4 expression and histologic subtype of medulloblastoma, together with the impact of CXCR4 activation on model medulloblastoma growth, suggested that coactivation of the SHH and CXCR4 pathways might result in a unique progrowth phenotype. We determined that CXCR4-high and -low medulloblastoma possessed unique transcriptional profiles and could be distinguished by gene array analysis (Fig. 6). Prominent among those genes with differential expression was Atoh1 (Math1), a HLH transcription factor known to regulate GNP
coregulated along with CXCR4, we isolated SmoA1 tumor cells from flank tumors treated with AMD3100 or PBS and measured mRNA abundance by quantitative PCR (Supplementary Table S1). Four genes appeared to be regulated by CXCR4 (Table 1). Remarkably, CXCR4 significantly suppressed the expression of PPP2R2C (~60%), the regulatory subunit of the phosphatase PP2A and elevated the expression of cyclin D1 (~20%). These changes in gene expression could enhance SHH-driven growth by promoting cell-cycle progression (cyclin D1) and blocking differentiation via PP2A-mediated dephosphorylation of S6 kinase (PPP2R2C; ref. 33). Together, these data suggest a model in which functional interaction between the SHH and CXCR4 pathways results in a unique progrowth phenotype in medulloblastoma.

**Discussion**

Medulloblastoma, the most common malignant pediatric brain tumor, arises from multiple cerebellar cell lineages and can be driven by several genetic abnormalities. Recent gene expression profiling and immunohistochemical studies have elucidated 4 distinct molecular subgroups with unique expression profiles (3, 4, 6). Three subgroups are characterized by abnormal activation of the Wnt, SHH, or Myc pathways. The fourth subgroup includes tumors whose molecular foundations are not yet specified.

Clinically, Wnt tumors carry the best prognosis and those medulloblastoma with MYC alterations, the worst (34). SHH tumors exhibit an intermediate outcome (1). Identifying the pathways altered in each of these tumor subgroups is necessary for the development of novel and molecularly targeted therapies and in addition, promises to improve outcome through molecular stratification of patients for clinical trials. In this regard, identification of a medulloblastoma as belonging to the SHH subgroup may not provide complete characterization. SHH pathway activation alone does not determine medulloblastoma biology. SHH medulloblastoma can include low-risk desmoplastic tumors or more aggressive anaplastic disease. Hence, secondary

### Table 1. Genes that are significantly altered after CXCR4 inhibition of SmoA1 flank tumors: CXCR4-regulated genes

<table>
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<th>Sample</th>
<th>mRNA</th>
<th>( C_t ) (primer)</th>
<th>( C_t ) (GAPDH)</th>
<th>Fold change ( \left[ 2^{-\Delta \Delta C_t} \right] )</th>
<th>( P ) (t test)</th>
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pathways, acting in concert with SHH must also determine biology within the SHH subtype. In the current study, we show that the level of CXCR4 expression further divides the SHH group of medulloblastoma into relevant subsets. We identified 2 medulloblastoma populations with either high or low CXCR4 expression. These 2 subsets of SHH-driven medulloblastoma possess distinct molecular, epidemiologic, and histologic profiles. CXCR4-high tumors are predominantly either desmoplastic or classic histology and occur most frequently in the youngest patients. In contrast, CXCR4-low tumors more commonly occur in older patients and exhibit classic or large cell anaplastic histology. Thus, CXCR4 expression may distinguish populations of SHH medulloblastoma with different prognoses.

The impact of CXCR4 in the SHH subgroup of medulloblastomas appears to be a consequence of unique molecular interactions that occur between these pathways. Our data suggest that SHH regulates CXCR4 activity at one of the earliest stages of its signaling. While SHH did not affect overall CXCR4 levels, prolonged inhibition of the SHH pathway reduced the cell surface fraction of CXCR4 and abrogated cellular responses to CXCL12. Cell surface localization of CXCR4, like other GPCRs, is regulated through desensitization, involving receptor phosphorylation and arrestin-mediated internalization, followed by either degradation or recycling to the cell surface (35–37). SHH could potentially affect any of these processes. Recent studies have highlighted the complexity of such heterologous regulation of GPCR internalization/recycling. Yu and colleagues showed that ligand-induced activation of Neurokinin 1 receptors (NK1R) produced a nonreciprocal inhibition of ligand-induced mu-opioid receptor endocytosis as a consequence of NK1R-dependent sequestration of arrestins on endosome membranes (38). In addition, nerve growth factor (NGF) has been shown to modulate the recycling of delta-opioid receptors to the surface membrane of central synaptic terminals (39).

We previously described alterations in CXCR4 signaling as a consequence of reduced desensitization in astrocytes null for the tumor suppressor neurofibromin (Nf1; ref. 22). Loss of Nf1 resulted in hyperactivation of the RAS-MAP kinase pathway and resultant ERK-dependent inhibition of GRK2, a kinase involved in CXCR4 desensitization. GRK2 inhibition was correlated with reduced phosphorylation of CXCR4 and sustained CXCL12-induced suppression of cAMP. Interestingly, GRK2 and β-arrestin 2 also directly interact with Smo and regulate SHH-signaling (40, 41). Thus, activation of the SHH pathway might modulate CXCR4 signaling through regulation of the desensitization machinery, including GRK2 and β-arrestin 2.

The biologic significance of interactions between the SHH and CXCR4 pathways may lie in the unique transcriptional profile of the CXCR4-high tumors. PPP2R2C is the regulatory subunit of the phosphatase PP2A, which enhances SHH-driven proliferation by stabilizing N-myc (42). In addition, PP2A blocks differentiation within the granule lineage by inactivating S6 kinase (33). SHH directly induces expression of the catalytic subunit of PP2A and here we show that CXCR4 suppresses expression of the regulatory subunit of PP2A. Both effects could increase PP2A activity and promote proliferation.

Cyclin D1 is a regulator of cyclin-dependent kinases such as CDK4/CDK6, which are critical for G1 to S transition and cell-cycle progression. Induction of cyclin D1 expression is essential for SHH-induced proliferation (43, 44). CXCR4 enhanced cyclin D1 expression in the SHH-medulloblastomas and this is likely to be a critical component of CXCR4’s positive effects on tumor growth. Furthermore, CXCR4’s effect on cyclin D1 expression suggests that high CXCR4 expression and/or activation may feed back and augment SHH signaling.

Significantly, CXCR4 activation appeared to be necessary for maximal proliferation of SHH-driven tumors. In SmoAl tumor models, the CXCR4 antagonist AMD3100 significantly inhibited tumor growth. These data indicate that dual inhibition of SHH and the CXCR4 pathways may provide additional benefit in treating those SHH subtype medulloblastomas that also express CXCR4. A recent clinical report described a single patient with medulloblastoma whose tumor was initially sensitive to SHH antagonist therapy, but who subsequently experienced an aggressive recurrence because of mutation in Smo (45–47). The ability of AMD3100 to inhibit the growth of SmoAl-driven tumors suggests that perhaps CXCR4 antagonism might address this mechanism of resistance.

The interplay between the SHH and CXCR4 pathways could function in normal development and cancer biology in any circumstances where the elements of both pathways are coexpressed. In addition to medulloblastoma, components of the CXCR4 and SHH pathways are expressed in primary specimens of breast, pancreatic, and prostate cancers as well as in gliomas and rhabdomyosarcomas (24, 48–53). It will be important to determine whether coexpression of these pathways provides additional prognostic value and whether combined antagonism of these pathways may have added benefit in the treatment of these cancers. Defining how SHH and CXCR4 modulate each other’s signaling is likely to open up new avenues of inquiry in each of these fields and new approaches to treating any cancer in which both pathways are operative.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

The authors thank Nicole Warrington for technical assistance and critical reading of the manuscript. Calcium flux measurements were made through the Regional Center for Biodefense and Emerging Infectious Diseases Research Live Cell Microscopy Core Facility at the Washington University School of Medicine.

Grant Support

This work was supported by R01NS052323-04S1 (RW-Reya) and RO1CA118399 (J.B. Ruine). M.D. Taylor holds a CBHR Clinician-Scientist Phase II Award and is supported by grants from the NIH (RO1CA148699), and the Pediatric Brain Tumor foundation.

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Received May 17, 2011; revised October 29, 2011; accepted October 31, 2011; published OnlineFirst November 3, 2011.
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doi:10.1158/0008-5472.CAN-11-1701

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