Cardiac Inflammation after Local Irradiation Is Influenced by the Kallikrein-Kinin System

Vijayalakshmi Sridharan, Preeti Tripathi, Sunil K. Sharma, Eduardo G. Moros, Peter M. Corry, Benjamin J. Lieblong, Elena Kaschina, Thomas Unger, Christa Thöne-Reineke, Martin Hauer-Jensen, and Marjan Boerma

Abstract

Radiotherapy of intrathoracic and chest wall tumors may lead to exposure of the heart to ionizing radiation, resulting in radiation-induced heart diseases (RIHD). The main manifestations of RIHD become apparent many years after treatment and include cardiomyopathy and accelerated atherosclerosis. This study examines the role of the kallikrein-kinin system (KKS) in RIHD by investigating the cardiac radiation response in a kininogen-deficient Brown Norway Katholiek (BN/Ka) rat model. BN/Ka rats and wild-type Brown Norway (BN) rats were exposed to local heart irradiation with a single dose of 18 Gy or 24 Gy and were observed for 3 to 6 months. Examinations included in vivo and ex vivo cardiac function, histopathology, gene and protein expression measurements, and mitochondrial swelling assays. Upon local heart irradiation, changes in in vivo cardiac function were significantly less in BN/Ka rats. For instance, a single dose of 24 Gy caused a 35% increase in fractional shortening in BN rats compared with a 16% increase in BN/Ka rats. BN rats, but not BN/Ka rats, showed a 56% reduction in cardiac numbers of CD2-positive cells, and a 57% increase in CD68-positive cells, together with a 52% increase in phosphorylation of extracellular signal-regulated kinase 1/2 (Erk1/2). Local heart irradiation had similar effects on histopathology, mitochondrial changes, and left ventricular mRNA levels of NADPH oxidases in the two genotypes. These results suggest that the KKS plays a role in the effects of radiation on cardiac function and recruitment of inflammatory cells. The KKS may have these effects at least in part by altering Erk1/2 signaling.

Cancer Res; 72(19); 4984–92. ©2012 AACR.
in infarction (13). On the other hand, kinins are well known for their induction of nitric oxide and prostacyclin, mediating cardioprotection via vasodilation and inhibition of cardiac fibroblasts (14, 15).

Brown Norway Katholie (BN/Ka) rats are deficient in HK and LK due to a point mutation in the kininogen gene (16). BN/Ka rats have been used to study the role of the KKS in several models of cardiovascular disease (17–19). This study examined the role of the KKS in RIHD by investigating molecular, structural, and functional changes after local heart irradiation in BN/Ka and wild-type Brown Norway (BN) rats. Both clinical and preclinical studies have shown that heart irradiation alters functional effects of irradiation in the lung (20, 21). In turn, the effects of pathologic damage in the lung on function and structure of the heart have long been established. Because of known interactions between heart and lung, and potentially also the spinal cord, we used a new method of rat heart irradiation to limit radiation exposure of other tissues.

Materials and Methods

Kininogen-deficient animal model

BN rats were obtained from Harlan Laboratories (BN/Rijhsd colony). BN/Ka breeder rats were a kind gift from Drs. Elena Kaschina and Thomas Unger (Charité University, Berlin, Germany) (23). Animals were housed 2 to 3 per cage in our Division of Laboratory Medicine on a 12:12 light–dark cycle with free access to food and water. All procedures in this study were approved by the Instrumental Animal Care and Use Committee of the University of Arkansas for Medical Sciences.

Sequencing of kininogen

The sequence of the kininogen gene was assessed as described before (17). Total liver RNA was isolated with Ultrasep RNA reagent (Biotex Laboratories), treated with the Turbo DNA-free Kit, and used for cDNA synthesis with the High Capacity cDNA Reverse Transcription Kit (Applied Biosystems). The kininogen cDNA was amplified with the primers: 5'-ACGAGTACCCTGCTGGG-3' and 5'-TGTTTGCACAATGGAGTGA-3' in a touchdown PCR protocol: 95°C for 2 minutes, 30 cycles: 95°C for 30 seconds, 65°C (reduced by 0.5°C in each following cycle), 72°C for 40 seconds, 10 cycles: 95°C for 30 seconds, 55°C for 30 seconds, 72°C for 40 seconds, final extension at 72°C for 3 minutes. PCR products were separated in a 2% agarose gel, extracted with a QiAquick Gel Extraction Kit (Qiagen), and sequenced in both directions with a 3100 Genetic Analyzer (Applied Biosystems). At least 6 rats of each experimental group were genotyped. All of the examined BN/Ka rats carried the G to A point mutation at the expected position, whereas all of the BN rats carried the wild-type genotype.

Local heart irradiation

Rats of 250 to 300 g were exposed to local heart irradiation with the small animal conformal radiation therapy device (SACRTD) developed at our institution. The SACRTD has a 225 kVp X-ray source (GE Isovolt Titan 225) mounted on a custom made "gantry," a stage on a robotic-arm positioning system (Viper s650 Adept Technology), and a flat panel digital X-ray detector of 200 μm resolution (XR6 0820 CM3 Perkin Elmer). For the purpose of local heart irradiation, a brass and aluminum collimating assembly produced a field of 19 mm diameter at the isocenter.

The dose rate at the isocenter was measured using a pinpoint ion chamber (PTW N301013, ADCL calibrated for 225 kV) following the TG-61 protocol of the American Association of Physicists in Medicine. In addition, dosimetry was carried out with Gafchromic EBT-2 film (Ashland Specialty Ingredients). A set of films was calibrated by exposing it to known doses on a Gamma Knife (Co-60) system, and the films were analyzed according to Devic and colleagues (24). A calibration curve was also drawn by exposing films with the SACRTD 225 kV X-ray beam. The films were energy independent and could be used for measurements of dose in the range used in our experiments. To measure relative depth dose, 11 pieces of film were placed in between 11 slabs of solid water phantom, each of 5 mm thickness. The top of the phantom was placed at the isocenter, perpendicular to the beam, and the films were exposed to 5 Gy at the isocenter (225 kV, 13 mA).

For local heart irradiation, rats were anesthetized with 3% isoflurane and placed horizontally in a styrofoam holder. The X-ray source was tilted horizontally and a digital X-ray image was acquired with the detector (65 kV, 5 mA). The heart was localized and the gantry was tilted vertically for irradiation. The heart was irradiated at 225 kV, 13 mA, (0.5 mm Cu-filtration) resulting in 1.92 Gy/min at 1 cm tissue depth. The hearts were exposed in three 19-mm-diameter beams of 6 Gy or 8 Gy each. An angle of 30° between the beams (one vertical, one beam –30° from vertical, and one beam +30° from vertical) was established by tilting the platform.

Rats were observed for 3 months or 6 months after irradiation to determine cardiac function, structure, and molecular changes as described below.

Echocardiography

A Vevo 2100 high-resolution in vivo microimaging system (VisualSonics) with the MS250 MicroScan microimaging system (VisualSonics) was used for echocardiography. Animals were anesthetized with 2% isoflurane and hair was removed from the chest with clippers and a depilatory cream. Short axis M-mode recordings at the midleft ventricular level were used to obtain echocardiographic parameters with the Vevo 2100 cardiac analysis software: thickness of the left ventricular anterior wall (LVAW), posterior wall (LV PW), inner diameter (LVID), volume, ejection fraction (EF), fractional shortening (FS), and stroke volume. B-mode recordings were used for strain analysis of 3 consecutive cardiac cycles using the Vevoscan software package (VisualSonics; 25). Recordings of the short axis were used to determine radial and circumferential velocity,
displacement, peak strain, and peak strain rate, and the long axis was used for longitudinal measurements.

**Ex vivo-perfused rat heart preparations**

Langendorff-type *ex vivo*-perfused rat heart studies were conducted as described before (26). In short, rats were anesthetized with 3% isoflurane; hearts were isolated and immediately perfused via the aorta with an oxygenated Krebs–Henseleit solution (37°C) at a flow rate of 10 mL/g heart/min. The ventricles were paced with electrodes positioned on the interventricular septum to obtain a heart rate of 250 beats/min. Both atria were removed and a fluid-filled balloon connected to a pressure transducer (model PT300, Grass Technologies) was placed in the left ventricle to measure pressures at balloon volumes between 80 μL and 300 μL. Coronary pressure was monitored continuously with a second pressure transducer.

After Langendorff studies, the hearts were weighed and processed for histology and immunohistochemistry (IHC).

**Histology and IHC**

Hearts were fixed in methanol Carnoy's solution (60% methanol, 30% chloroform, 10% acetic acid) and embedded in paraffin. For both histology and IHC, 5 μm sections were deparaffinized and rehydrated.

For determination of collagen, sections were incubated in Picrosirius red (American MasterTech) with Fast Green (0.01% w/v; Fisher Scientific) for 2 hours. Sections were analyzed with an Axioskop transmitted light microscope (Carl Zeiss) with a chilled color camera (Leica). Picrosirius red/Fast Green staining was quantified as the area stained positive with Picrosirius red divided by the total area.

For determination of mast cell numbers, sections were deparaffinized and rehydrated. For both histology and IHC, 5 μm sections were incubated in methanol Carnoy's solution (60% methanol, 30% chloroform, 10% acetic acid) and embedded in paraffin. For both histology and IHC, 5 μm sections were deparaffinized and rehydrated.

Rats were anesthetized with 3% isoflurane; hearts were isolated and snap-frozen in liquid nitrogen. Frozen samples from the left ventricle were homogenized in Ultraspec RNA reagent (Biotecx Laboratories). After treatment with RQ-DNase 1 (Promega) and RNase at 37°C for 30 minutes, followed by DNase inactivation at 75°C for 10 minutes, cDNA was synthesized using the High Capacity cDNA Archive Kit (Applied Biosystems). Steady-state mRNA levels were measured with real-time quantitative PCR (TaqMan) using the 7500 Fast Real-Time PCR System and the following predesigned assays for rat: B1 receptor (Rn02064589_s1), B2 receptor (Rn00597384_m1), NADPH oxidase (NOX1 (Rn00586652_m1), NOX2 (Rn00576710_m1), NOX4 (Rn00585380_m1), and p22phox (CYBA, Rn00577357_m1; all Applied Biosystems). Relative mRNA levels were calculated with the ΔΔCt method, using 18S rRNA as a normalizer.

**Western blots**

Left ventricular tissue was homogenized in radioimmunoprecipitation assay buffer with inhibitors of proteases (10 μL/mL) and phosphatases (10 μL/mL, Sigma Aldrich) and centrifuged at 20,000 g for 15 minutes. Supernatant protein amounts were determined with a bichinchoninic acid assay (Sigma-Aldrich). A total of 50 μg protein in Laemmli sample buffer containing β-mercaptoethanol (1:20 vol/vol) was boiled for 2 to 3 minutes, separated in Any kD Mini-Protean polyacrylamide gels (Bio-Rad) at 100 Volts and transferred to polyvinylidene difluoride membranes at 20 Volts overnight at 4°C.

Membranes were incubated in TBS containing 0.05% Tween-20 and 5% dry powdered milk, followed by rabbit antibodies against the following: phospho-Akt, phospho-Erk1/2, pan Akt, Erk1/2 (at 1:10,000), phospho-e-Jun (Ser63), phospho-c-Jun (Ser73), c-Jun (at 1:1,000), and horseradish peroxidase (HRP)-conjugated mouse anti-rabbit at 1:4,000 (for Akt and Erk1/2) or 1:10,000 (for c-Jun; all Cell Signaling Technology). Protein loading was determined with mouse anti-glyceraldehyde-3-phosphate dehydrogenase (1:2,000, Santa Cruz), followed by HRP-conjugated goat anti-mouse with a Teflon pestle. The homogenate was centrifuged at 700 g for 10 minutes at 4°C. The supernatant was removed and centrifuged at 12,500 g for 30 minutes to obtain the mitochondrial pellet. Pellets were resuspended in a 10 mmol/L HEPES buffer containing 395 mmol/L sucrose and 0.1 mmol/L EGTA, washed twice, and immediately analyzed.

Mitochondrial permeability transition pore opening was measured by Ca2+ induced swelling, indicated by a decrease in absorbance at 540 nm. Isolated mitochondria were suspended in swelling buffer containing 120 mmol/L KCl, 10 mmol/L Tris HCl, and 5 mmol/L KH2PO4, to a final concentration of 150 μg/mL, and immediately exposed to vehicle, 250 μmol/L CaCl2 or 250 μmol/L CaCl2 in combination with 2 μmol/L cyclosporin A (Csa) as an inhibitor of transition pore opening. Optical density at 540 nm (OD540) was measured with a Synergy 4 microplate reader (BioTek), immediately before the assay and every 2 minutes thereafter for a total of 20 minutes.

**RNA isolation and real-time PCR**

Rats were anesthetized with 3% isoflurane, hearts were isolated and snap-frozen in liquid nitrogen. Frozen samples from the left ventricle were homogenized in Ultraspec RNA reagent (Biotecx Laboratories). After treatment with RQ-DNase 1 (Promega) at 37°C for 30 minutes, followed by DNase inactivation at 75°C for 10 minutes, cDNA was synthesized using the High Capacity cDNA Archive Kit (Applied Biosystems). Steady-state mRNA levels were measured with real-time quantitative PCR (TaqMan) using the 7500 Fast Real-Time PCR System and the following predesigned assays for rat: B1 receptor (Rn02064589_s1), B2 receptor (Rn00597384_m1), NADPH oxidase (NOX1 (Rn00586652_m1), NOX2 (Rn00576710_m1), NOX4 (Rn00585380_m1), and p22phox (CYBA, Rn00577357_m1; all Applied Biosystems). Relative mRNA levels were calculated with the ΔΔCt method, using 18S rRNA as a normalizer.
1:4000, Jackson ImmunoResearch). Antibody binding was visualized with ECL Plus Detection reagent (GE Healthcare Life Sciences) on CL-Xposure Film (Thermo Scientific). Films were scanned with an Alphalager gel documentation system (ProteinSimple) and bands were quantified with ImageJ.

**Statistical analysis**

Data were evaluated with the software package NCSS 2007 (NCSS). Dose dependencies were tested with linear regression. Data from Langendorff-perfused heart preparations and mitochondrial swelling assays were tested with repeated measures ANOVA. All other data were analyzed with 2-way ANOVA, followed by Newman–Keuls individual comparisons. The criterion for significance was a $P < 0.05$. Data are reported as average ± SEM.

**Results**

This study investigated the role of the KKS in RIHD by comparing cardiac radiation injury in kininogen-deficient BN/Ka rats with injury in wild-type BN rats. In vivo cardiac function was measured with echocardiography at 3 months and 6 months after a single dose of 18 Gy or 24 Gy. All parameters and analyses of dose dependencies are shown in Supplementary Tables S1–S6. Effects of radiation on M-mode parameters were more severe at 3 months after irradiation compared with 6 months (Supplementary Data S1). At 3 months, linear regression revealed a dose-dependent decrease in systolic LVID and volume in BN rats, together with a dose-dependent increase in systolic LVAW and LVPW thickness, EF, and FS (Supplementary Data S2). On the other hand, stroke volume and cardiac output were not altered by irradiation. In BN/Ka rats, there was only a dose-dependent increase in systolic LVPW thickness, and borderline significance in EF and FS. In a direct comparison of the 2 genotypes, more severe changes were found in BN/Ka rats with injury in wild-type BN rats.

In vivo cardiac function was measured with echocardiography at 3 months and 6 months after irradiation both BN and BN/Ka rats showed dose-dependent increases in systolic radial velocity, displacement and strain, and circumferential velocity, strain, and strain rate (Supplementary Data S4). In diastole, both genotypes showed dose-dependent changes in radial velocity, circumferential velocity, and circumferential strain rate (Supplementary Data S6). In a direct comparison of the 2 genotypes, 24 Gy caused more severe changes in BN in the following parameters in systole: circumferential velocity ($P < 0.03$), strain ($P < 0.003$), strain rate ($P < 0.003$), and radial velocity ($P < 0.04$), displacement ($P < 0.03$), and strain rate ($P < 0.001$). In addition, BN rats showed larger changes in diastolic circumferential velocity ($P < 0.03$), strain rate ($P < 0.004$), and radial velocity ($P < 0.03$).

In accordance with the echocardiography results at 6 months after irradiation, no significant effects of radiation on ex vivo cardiac Langendorff parameters were found in hearts isolated at this time point (data not shown). Heart to body weight ratios were not altered at 6 months after 18 Gy, but were significantly reduced from 3.0 ± 0.1 in sham-irradiated animals to 2.6 ± 0.1 and 2.7 ± 0.1 in BN and BN/Ka rats exposed to 24 Gy ($P < 0.05$).

Because most severe cardiac function changes were seen at 3 months after a single dose of 24 Gy, left ventricular molecular changes were examined at this time point. Local heart irradiation caused a significant increase in gene expression of the kinin B2 receptor in both BN/Ka and BN rats (Table 1). B1 receptor transcripts could not be detected in sham-irradiated or irradiated hearts (data not shown). Intracellular signaling upon kinin receptor activation involves Akt and Erk1/2. Local heart irradiation did not alter the left ventricular levels of total Erk1/2 or Akt. In addition, radiation-induced Akt phosphorylation was not statistically significant. On the other hand, a significant increase in phosphorylation of Erk1/2 and the Erk1/2 target c-Jun was observed in BN rats (Fig. 1 and Supplementary Figs. S7 and S8). No increased phosphorylation of Erk1/2 or c-Jun was found in BN/Ka rats.

Bradykinin may modify cardiac function and tissue remodeling by altering expression and activity of Nox enzyme complexes. We examined left ventricular mRNA levels of the main membrane components of the Nox complexes that are expressed in the heart: p22phox, Nox1, Nox2, and Nox4. Nox 1 transcript levels were around the detection limit of the assay (data not shown), while p22phox, Nox2, and Nox4 mRNAs could

**Table 1.** Left ventricular mRNA values of the B2 receptor and of membrane components of Nox complexes at 3 months after 24 Gy or 0 Gy, relative to BN after 0 Gy (average ± SEM; $n = 6$)

<table>
<thead>
<tr>
<th></th>
<th>BN</th>
<th>BN/Ka</th>
</tr>
</thead>
<tbody>
<tr>
<td>0 Gy</td>
<td>24 Gy</td>
<td></td>
</tr>
<tr>
<td>B2 receptor</td>
<td>1.11 ± 0.22</td>
<td>2.24 ± 0.37 *</td>
</tr>
<tr>
<td>Nox2</td>
<td>1.03 ± 0.10</td>
<td>1.50 ± 0.22 *</td>
</tr>
<tr>
<td>Nox4</td>
<td>1.06 ± 0.19</td>
<td>2.34 ± 0.43 *</td>
</tr>
<tr>
<td>p22phox</td>
<td>1.01 ± 0.07</td>
<td>1.15 ± 0.09</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>0 Gy</th>
<th>24 Gy</th>
</tr>
</thead>
<tbody>
<tr>
<td>BN</td>
<td>0.99 ± 0.24</td>
<td>2.26 ± 0.61 *</td>
</tr>
<tr>
<td>BN/Ka</td>
<td>0.99 ± 0.07</td>
<td>1.64 ± 0.25 *</td>
</tr>
</tbody>
</table>

*Significant difference with 0 Gy ($P < 0.05$).
be detected in all samples. Local heart irradiation caused significant increases in Nox2 and Nox4 mRNA. No differences were found between BN/Ka rats and BN rats (Table 1).

Mitochondrial membrane integrity is important for cardiac function. Because bradykinin may alter the mitochondrial membrane, we investigated swelling in response to CaCl2 in mitochondria isolated from BN and BN/Ka hearts. Enhanced swelling was observed in mitochondria isolated at 3 months after local heart irradiation, as shown by a reduction in their OD540 (Fig. 2). The reduction in OD540 was inhibited with CsA, confirming that swelling was caused by enhanced mitochondrial transition pore opening. There was no difference between mitochondria isolated from irradiated BN hearts and mitochondria from irradiated BN/Ka hearts.

Histopathologic changes at 6 months after irradiation included vacuolar degeneration of cardiomyocytes after 18 Gy and 24 Gy and local areas of severe cardiomyocyte degeneration and interstitial fibrosis after 24 Gy (Fig. 3). Because of the local nature of the fibrosis, the total left ventricular area of interstitial collagen was not significantly altered by radiation exposure. In addition, no statistically significant differences were found between BN and BN/Ka rats (Fig. 4).

The KKS modulates inflammation. We therefore determined the numbers of various inflammatory cells at 6 months after (sham)-irradiation (Fig. 5). Radiation caused a dose-dependent reduction in the number of CD2-positive cells [T cells and natural killer (NK) cells] in BN rats, but not in BN/Ka rats. In addition, radiation at a dose of 24 Gy caused a significant increase in the number of CD68-positive cells (monocytes and macrophages) in BN rats, but not in BN/Ka rats. Radiation at a dose of 24 Gy caused a significant increase in the number of mast cells in both genotypes.

Discussion
This study examined the role of the KKS in RIHD, by comparing the cardiac radiation response in kininogen-deficient BN/Ka rats with the response in wild-type BN rats. Local heart irradiation caused more severe changes in 
\textit{in vivo} cardiac function and significantly altered the numbers of CD2-positive and CD68-positive cells in the wild-type BN rats only, suggesting a role for the KKS in cardiac function changes and recruitment of inflammatory cells in response to radiation. No obvious differences were found in myocardial degeneration and fibrosis, suggesting that the KKS may not play a significant role here.

BN and BN/Ka rats have been used to study the role of the KKS in other cardiovascular diseases. BN/Ka rats are more prone to the induction of aortic aneurysms, but do not differ from BN rats in atherosclerosis (17). In myocardial infarct models, BN/Ka rats have shown to be more sensitive (27), or did not differ from BN rats (19), and showed a reduced response to angiotensin converting enzyme (ACE) inhibition (19). Hence, current and previous studies have shown that the KKS plays a distinct role in different models of cardiovascular...
disease. This may depend, in part, on B1 and B2 receptor expression and function. Few studies have investigated the effects of radiation on cardiovascular bradykinin receptor expression and/or function. Whole body irradiation with a dose of 2 Gy enhanced immediate B1 receptor expression in the heart (28). Ex vivo beta and gamma irradiation of rabbit aorta induced B1 gene expression within hours after irradiation, and the response to B1 activation was enhanced (29). To our knowledge, this is the first study to examine left ventricular expression of bradykinin receptors several months after irradiation. We found an increase in left ventricular mRNA of the B2 receptor at 3 months after local heart irradiation, a time point at which in our rat model histopathologic changes become apparent. The B2 receptor is constitutively expressed in the heart, whereas the B1 receptor is expressed only in certain conditions of inflammation or injury (9). Although B2 expression was increased in both BN and BN/Ka rats, kinin signaling seemed to be impaired in BN/Ka rats, as shown by reduced phosphorylation of Erk1/2. Activation of Erk1/2 may have favorable effects in the heart, with its prosurvival and proangiogenic properties and promotion of cardiac contractility (30). Indeed, Erk1/2 activation is involved in beneficial cardiovascular effects of the KKS (31). Erk1/2 has a large array of target proteins, including many transcription factors. Together with c-Jun N-terminal kinase, Erk1/2 may activate c-Jun (30). As part of the activator protein-1 transcription complex, c-Jun may promote inflammation and the recruitment of macrophages (32). We found a radiation-induced increase in c-Jun phosphorylation in BN rats only, suggesting that this transcription factor may be part of the pathways by which the KKS regulates inflammation in the irradiated heart. To further determine mechanisms of action of the KKS, future studies may aim to identify protein phosphorylation patterns or transcription profiles induced by Erk1/2 in the wild-type BN rats.
In vivo analysis of cardiac function in this study included echocardiographic strain analysis, which is a method that gives direct measures of local contractility of the ventricular wall. This method is often considered more sensitive than conventional echocardiography. For instance, strain analysis shows early changes in those segments of the left ventricular wall that have been exposed to radiation during radiotherapy (33). It is also useful in the detection of local and global changes in the ventricular wall in small animal models (25). In our rat model, the increase in systolic strain, together with an increase in EF and FS, but in the absence of a change in stroke volume, may reflect efforts of the irradiated hearts to maintain their cardiac output. Changes in echocardiography parameters were most severe at 3 months after irradiation with 24 Gy. Although future studies will have to define the effects of fractionated irradiation in BN and BN/Ka rats, here, we set out to determine potential mechanisms by which the KKS may affect cardiac function at 3 months after 24 Gy.

Studies have shown that bradykinin is able to both upregulate and downregulate expression and/or activity of Nox enzyme complexes (34, 35). The main function of Nox enzymes is the production of reactive oxygen species (ROS) by reducing oxygen to superoxide. The membrane-bound subunits of the Nox complexes consist of p22phox and gp91phox. Of the known isoforms of gp91phox, Nox1, Nox2, and Nox4 are expressed in the heart (36). Several studies have shown that ionizing radiation-induced upregulation of Nox expression may cause prolonged production of ROS, contributing to normal tissue radiation injury (37). Nox enzymes also play various roles in cardiac health and disease. Nox2 has been implicated in endothelial dysfunction (38) and adverse cardiac remodeling (39). Nox4, on the other hand, may have some cardioprotective properties, for instance by promoting neovascularization (40). We found an upregulation of both Nox2 and Nox4 gene expression after local heart irradiation in the rat, independent of genotype. The exact role of Nox complexes in RIHD remains to be determined.

Mitochondrial transition pore opening adversely affects cardiac function by causing ATP depletion, oxidative stress, dysregulation of Ca\(^{2+}\) homeostasis, and ultimately cardiac cell death (41). Local heart irradiation in this study caused enhanced mitochondrial transition pore activity, which may contribute to changes in cardiac function. The cause of mitochondrial transition pore activation after ionizing radiation is not known. Prolonged oxidative stress, as well as mitochondrial proteome changes may play a role (42, 43). Interestingly, bradykinin is known to alter the mitochondrial membrane and may prevent the opening of the mitochondrial transition pore (44). In this study, no differences were found between BN and BN/Ka after irradiation, suggesting that mitochondrial...
transition pore activity after the doses of radiation used in this study is not affected by the KKS.

The KKS is well known for its pro- and antiinflammatory effects. Accordingly, BN and BN/Ka have different plasma profiles of certain inflammatory cytokines (17). Moreover, many inflammatory cells express kinin receptors (45) and may therefore be directly affected by the KKS. We examined cardiac numbers of inflammatory cells at 6 months after local heart irradiation in BN and BN/Ka rats. A dose-dependent reduction in the number of CD2-positive cells (T cells and NK cells) was observed in BN rats, but not in BN/Ka rats. A single dose of 24 Gy caused an increase in the number of CD68-positive cells (monocytes and macrophages) in BN rats, but not in BN/Ka rats. These results show that the KKS affects the recruitment of various inflammatory cells. Inflammation plays a dual role in cardiac disease (46), and the exact role of the different subsets of inflammatory cells in RIHD needs to be determined.

Bradykinin can activate mast cells (47), and mast cell-derived enzymes may interact with mediators of the KKS (8). In previous studies we found that cardiac mast cell numbers increase after local heart irradiation in the rat, coinciding with cardiac radiation injury, and mast cells may play a predominantly protective role in RIHD in the rat (26). In the current study, both histopathologic changes and mast cell numbers were not altered by kininogen deficiency. These results suggest that there is a close correlation between myocardial injury and mast cell numbers, but that the KKS may not be involved in mast cell recruitment. Whether the KKS modifies the function of cardiac mast cells after irradiation is still unknown.

This study suggests that pharmacologic modification of the KKS may have beneficial effects on certain aspects of RIHD. Because of the dual role of the KKS in inflammation and cardioprotection, B1 and B2 receptor agonists and antagonists may each have positive and negative effects on cardiac remodeling and function. For instance, ACE inhibitors are considered to be cardioprotective in part by their inhibition of bradykinin breakdown (48). Although in experimental models of myocardial infarction a long-acting analog of bradykinin-inhibited changes in cardiac function (49), a B2 receptor antagonist inhibited myocardial fibrosis (13). Studies are required to determine the effects of receptor agonists and antagonists in experimental models of RIHD.

In conclusion, this study shows that the KKS is involved in cardiac function changes and myocardial inflammatory infiltration in response to local irradiation. The KKS may have these effects at least in part by altering Erk1/2 signaling. Future studies have to address the effects of fractionated irradiation, as well as pharmacologic modification of the KKS or its targets.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Conception and design: P. Corry, M. Boerma
Development of methodology: V. Sridharan, P. Tripathi, S.K. Sharma, E.G. Moros, P. Corry, M. Boerma
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): V. Sridharan, P. Tripathi, S.K. Sharma, E.G. Moros, B.J. Lieblong, T. Unger, C. Thoene-Reineke, M. Boerma
Analysis and interpretation of data (e.g., statistical analysis, bios-statistics, computational analysis): V. Sridharan, P. Tripathi, B.J. Lieblong, M. Boerma
Writing, review, and/or revision of the manuscript: V. Sridharan, S.K. Sharma, E.G. Moros, P. Corry, B.J. Lieblong, E. Kaschina, M. Hauer-Jensen, M. Boerma
Study supervision: E.G. Moros, M. Boerma

Acknowledgments
The authors acknowledge Dr. Sue A. Theus and Kimberly Henning for excellent support in animal care.

Grant Support
This work was supported by the NIH (CA148679, CA17382) and the American Cancer Society (RSG-10–125–01-CCE).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received May 9, 2012; revised July 19, 2012; accepted July 25, 2012; published OnlineFirst August 3, 2012.

References


