Inactivating All Three Rb Family Pocket Proteins Is Insufficient to Initiate Cervical Cancer

Myeong-Kyun Shin1, Julien Sage2, and Paul F. Lambert1

Abstract

Human papillomavirus-16 (HPV-16) is associated etiologically with many human cervical cancers. It encodes 3 oncogenes E5, E6, and E7. Of these oncogenes, E7 has been found to be the dominant driver of cervical cancer in mice. More than 100 cellular proteins have been reported to associate with HPV-16 E7, which is thought to dysregulate the cell cycle in part by binding and inducing the degradation of pRb and its related pocket protein family members, p107 and p130. The ability of E7 to inactivate the pRb family correlates with its ability to induce head and neck cancers in mice. We previously showed that the inactivation of pRb is itself not sufficient to recapitulate the oncogenic properties of E7 in cervical carcinogenesis. In this study, we evaluated mice that were deficient in multiple pocket proteins, including mice that lacked pRb, p107, and p130. Strikingly, combined loss of two or all 3 pocket proteins resulted in development of high-grade cervical intraepithelial neoplasia, but not frank cervical carcinoma. These findings strongly argue that the oncogenic properties of HPV-16 E7 in human cervical carcinogenesis may involve disruption of E7 binding proteins beyond simply the pRb family members. Cancer Res; 72(20); 5418–27. ©2012 AACR.

Introduction

Cervical cancer remains a major woman’s health problem worldwide (1, 2). Infections by a subset of human papillomavirus (HPV) are the primary cause of cervical cancer and, among this ‘high-risk’ subset of human mucosal HPV’s, HPV type 16 (HPV-16) is responsible for approximately 60% of cervical cancers (3). HPV-16 encodes three major oncogenes; E5, E6, and E7 (4, 5). Of these, E6 and E7 are commonly found expressed in HPV-associated cervical cancers (6). E7 was previously identified as the predominant oncogene in terms of its ability to induce cervical cancer in transgenic mouse models (7, 8).

In tandem affinity purification/mass spec analyses, HPV-16 E7 protein has been found associated with more than 100 different cellular proteins (9). Through these associations, E7 has been implicated in dysregulating a wide range of cellular processes, including gene transcription, DNA synthesis, protein degradation, epigenetic reprogramming, genomic integrity and cellular metabolism (10). Of these cellular targets of E7, the best known are the tumor suppressor pRb and its related pocket protein family members, p107 and p130 (11). pRb is an important regulator of the cell cycle in the transition point from the G1 phase to the S phase at least in part because it can bind to and inactivate a family of transcription factors called E2Fs. In response to mitogenic stimuli, pRb is posttranslationally modified via phosphorylation by cyclin/cdk complexes, resulting in its release from, and consequent activation of these E2F transcription factors that are key regulators of expression of genes involved in cell cycle (12). E7’s binding to pRb leads to the inactivation of pRb and its degradation through proteasome-dependent degradation (13) resulting in the activation of the E2F transcription factors. Inactivation of pRb by E7 also is thought to contribute to alterations in differentiation, DNA damage responses, centrosome synthesis, and tumorigenesis (14, 15). In various human cancers, genetic or epigenetic inactivation of RB has been reported; these observations and the analysis of genetically engineered mice have identified pRb as a major tumor suppressor (12). These findings support the hypothesis that pRb inactivation by E7 is an important contributor to the oncogenic potential of HPV-16 E7 in cervical carcinogenesis. However, in our prior studies, we discovered that genetic inactivation of pRb is not sufficient to account for E7’s ability to induce cervical cancer in mice (16). This observation led us to the present studies directed at asking what other cellular targets of E7 contribute to cervical carcinogenesis.

Global gene expression analysis of HPV-associated cancers indicated that many of the cell cycle regulatory genes induced in HPV-positive cancers are E2F-responsive genes (17). For this reason we focused our attention on other cellular targets of E7 that are involved in regulating E2Fs, specifically other members of the pocket protein family, p107 and p130 (18). p107 and p130 have similarities with pRb, both in their overall structure, their ability to bind and inactivate E2Fs, as well as sequence
homology in a large C-terminal domain known to mediate their interaction with viral oncoproteins such as SV-40 T antigen, adenovirus E1A, and HPV E7 (19–22). Functional overlap among the pocket proteins has been documented among the pocket protein family members (23). Despite these similarities to pRb, the tumor suppressive activity of p107 and p130 remains largely questioned, as genetic and epigenetic alterations of these genes are not commonly found in human cancers (12, 24). Nevertheless, the concept that p107 and/or p130 function as tumor suppressors in the context of cervical carcinogenesis has remained a popular hypothesis because HPV-16 E7 is able to bind all three pocket proteins. Furthermore, multiple studies in mice have shown that p107 as well as p130 can function as tumor suppressors in different tissue contexts (25–29), including our own studies in the context of head and neck carcinogenesis (30).

In this study, we made use of genetically engineered mice to determine whether the combined loss of function of multiple pocket proteins is sufficient to account for the oncogenic properties of HPV-16 E7 in cervical carcinogenesis. Our results show that the combinatorial inactivation of two or all three pocket proteins is not sufficient to induce cervical cancer but is sufficient to induce high grade cervical intraepithelial neoplasia (CIN), the precursor to cervical cancer. These findings indicate that other cellular target(s) of E7 must contribute to late stages in cervical carcinogenesis.

Materials and Methods

Mice

K14E7 mice have been described previously (8). Rbf/flp107−/−, Rbf/flp130−/−, and Rbf/flp130−/−p107−/− mice were previously described (31). KRT14-cre/Esrl (K14CreErtm) was obtained from Jackson Laboratory. K14CreRbf/flp130−/− mice have been described previously (16). Details on the nature of the genetic crosses made in this study are provided under the Supplementary Data section. For the irradiation studies, mice were exposed to 0 or 12 Gy ionizing radiation from a 137Cs source 24 hours before mice were injected IP with bromodeoxyuridine (BrdUrd; 10 μL per g body weight of 12.5 mg/mL solution). These mice were then sacrificed 1 hour later and tissues harvested. All mice were bred and maintained in the American Association for Accreditation of Laboratory Animal Care–approved McArdle Laboratory Cancer Center Animal Care Facility and were managed in accordance with an approved animal protocol.

Inducible Cre-mediated recombination

To generate the Rb-nulligenic state in the cervical stratified epithelia of K14CreErtm Rbf/flp107−/− mice, 6-week-old mice were intraperitoneally injected with tamoxifen [4 mg of tamoxifen dissolved in corn oil (Fisher Scientific Inc.) per day] for 5 consecutive days. To generate the Rb-nulligenic and p130-nulligenic states in the cervical stratified epithelia of K14CreErtmRbf/flp130−/−p107−/− mice, 6-week-old mice 4-hydroxytamoxifen [4-OHT; 0.05 mg of 4-hydroxytamoxifen dissolved in corn oil (Fisher Scientific Inc.) per day] was applied topically to the vaginal cavity for 5 consecutive days. We did not observe Cre activation by estrogen pellet in estrogen-treated K14CreErtmRbf/flp107−/− mice (data not shown) consistent with the fact that the Cre-Ertm gene product is selectively responsive to tamoxifen, not estrogen.

Immunohistochemistry

All of immunohistochemistry was conducted as previously described (30, 32). Briefly, the primary antibodies were used in different conditions as follows: anti-pRb (1:25, in 5% horse serum, BD Biosciences), anti-p107 (1:125 in 5% nonfat milk/5% horse serum, Santa Cruz Biotechnology), anti-p130 (1:100 in 5% nonfat milk/5% horse serum, BD Biosciences), anti-BrdUrd (1:50 in 5% horse serum, Calbiochem), anti-Mcm7 (1:200 in 5% nonfat milk/5% horse serum, LabVision Neomarkers).

Statistical analysis

A 2-sided Fisher’s exact test was used to determine the significance of differences in the incidence of cervical between each mouse group. To determine the significance of differences in the severity of disease and DNA synthesis level between each mouse group, a 2-sided Wilcoxon rank sum test was conducted.

Results

Conditional inactivation of both Rb and p107 or Rb and p130 in cervical epithelia

To study whether the combined loss of pRb and other pocket protein members, either p107 or p130, contribute to cervical carcinogenesis, we generated mice that were inactivated for both pRb and p130, or pRb and p107 in stratified cervical epithelia. To achieve knockout of pRb and p130, we crossed a conditional knockout allele of Rb with a germline knockout allele for p130. To knock out Rb in the cervical epithelium, we placed the Rbf/flp130−/−mice onto the K14Cre transgenic mouse background that expresses Cre recombinase in all stratified squamous epithelia of the adult mouse. The same strategy could not be used to generate mice knocked out both Rb and p107 in the cervix because K14CreRbf/flp107−/− mice display high rates of mortality at a very early age, which we and others ascribe to difficulty in the mice drinking water or eating food owing to hyperplasia leading to occlusion of the esophagus (30). Therefore, to generate mice inactivated for both pRb and p107 in cervical epithelium, we crossed Rbf/flp107−/− mice to K14CreErtm, a transgenic mouse strain that expresses a tamoxifen-inducible form of Cre in stratified squamous epithelia. We then administered tamoxifen to adult K14CreErtmRbf/flp107−/− mice, at 6 weeks of age, well after the age when mortality was observed in the K14CreRbf/flp107−/− mice. By this time K14 driven transgenes have become actively expressed in the cervical and vaginal epithelium. Before this age K14-driven transgenes are poorly expressed in the female reproductive tract (data not shown). This strategy eliminated mortality issues presumably because the adult esophagus is sufficiently large that the epithelial hyperplasia resulting from inactivation of both pRb and p107 does not occlude it. To verify the efficiency of Cre or CreErtm-induced disruption of pRb expression in cervical stratified epithelium of the K14CreRbf/flp130−/− and K14CreErtmRbf/flp107−/− mice, we conducted pRb, as well as p107 and p130 immunohistochemical staining on the female
reproductive tracts of these mice, as well as that of nontransgenic, \( Rbf^{+/+}/p130^{-/-} \), \( Rbf^{+/+}/p107^{-/-} \), \( K14CreRbf^{+/+} \), and \( K14E7 \) transgenic mice (Fig. 1). As expected, pRb staining in cervical epithelium was sharply reduced in the \( K14E7 \), \( K14CreRbf^{+/+} \), \( K14CreRbf^{+/+}/p130^{-/-} \), and the tamoxifen-treated \( K14CreERtnRbf^{+/+}/p107^{-/-} \) mice, whereas still observed in Cre-negative \( Rbf^{+/+} \) mice, such as \( Rbf^{+/+}/p130^{-/-} \), \( Rbf^{+/+}/p107^{-/-} \) mice (Fig. 1, left). These observations indicate that the floxed Rb allele in cervical stratified epithelium had undergone recombination by the K14 promoter-driven Cre or CreER transgenes. Likewise, we could not detect p107 in cervical stratified epithelium nulligenic for p107, such as \( Rbf^{+/+}/p107^{-/-} \) and tamoxifen-treated \( K14CreERtnRbf^{+/+}/p107^{-/-} \) mice, as well as \( K14E7 \) mice, but it was detected in the \( Rbf^{+/+}/p130^{-/-} \), \( K14CreRbf^{+/+} \), \( K14CreRbf^{+/+}/p130^{-/-} \), and nontransgenic mice (Fig. 1, middle). Levels of p107 were increased in both \( K14CreRbf^{+/+} \) and \( K14CreRbf^{+/+}/p130^{-/-} \) mice over that observed in Rb-sufficient nontransgenic control mice. Moreover, p107 positive-stained cells were more uniformly observed in the stratified epithelium of the \( K14CreRbf^{+/+}/p130^{-/-} \) mice compared with that of the \( K14CreRbf^{+/+} \) mice. These results indicate that compensatory increases in expression of p107 can arise in cervical epithelium when the tissue is deficient in the expression of pRb alone, and more so when it is deficient in expression of both pRb and p130. A similar trend was observed for p130. It was not detectable in cervical epithelium from both \( Rbf^{+/+}/p130^{-/-} \) and \( K14CreRbf^{+/+}/p130^{-/-} \) mice, which are harboring the \( p130 \) null allele (Fig. 1, right), but was increased in pRb-deficient tissue, although not nearly as strongly as was seen for p107. Taken together, these data indicate that the K14-driven Cre and CreERtn transgenes were effective at inducing recombination of the floxed Rb allele in the cervical epithelia within the female reproductive tract, and show that compensatory increases in expression of functional pocket proteins can arise in cervical stratified epithelium.

Suprabasal DNA synthesis in cervical stratified squamous epithelium deficient in pocket proteins

HPV-16 E7 confers acute changes to the cervical epithelium including the reprogramming suprabasal cells to support DNA synthesis, and disrupting DNA damage-induced cell-cycle
arrest, activities that may contribute to the accumulation of mutations and carcinogenesis (10). As previously described (16), pRb inactivation alone was not sufficient to cause these same phenotypes in cervical epithelium. We asked if the combinatorial inactivation of pRb and p130, or pRb and p107 could cause induction of DNA synthesis in suprabasal cells. To address this, we analyzed by immunohistochemistry (16, 32) the frequency of BrdUrd-positive suprabasal cells in sections of tissue from mice injected with this nucleoside analog 1 hour before sacrifice. In Rb−/−p130−/−, tamoxifen-treated Rb−/−p107−/−, and K14CreRb−/− mice, there was no significant induction of suprabasal DNA synthesis in cervical stratified epithelia compared with nontransgenic mice (Fig. 2A and B), indicating that inactivation of any one pocket protein is not able to induce a suprabasal DNA synthesis in cervical stratified epithelia. The data obtained with the K14CreRb−/− mice was consistent with our prior study (16). In contrast, in K14CreRb−/−p130−/− mice, we observed a significant increase in suprabasal DNA synthesis compared with either Rb−/−p130−/− or K14CreRb−/− mice although this induction of suprabasal DNA synthesis was significantly lower than that observed in K14E7 mice (Fig. 2A). Similarly, we observed an increase of suprabasal DNA synthesis in tamoxifen-treated K14CreERtmRb−/−p107−/− mice compared with either nontransgenic or tamoxifen-treated Rb−/−p107−/− mice, but the frequency of BrdUrd-positive cells was less than that observed in K14E7 mice (Fig. 2B). These observations show that the combined loss of pRb and either p107 or p130 partially accounts for the DNA synthesis in suprabasal compartment induced by E7.

Abrogation of DNA damage response in irradiated cervical epithelia deficient for pRb and p130 or pRb and p107

One of the hallmarks of E7 is its ability to abrogate DNA damage response in epithelia including the cervical epithelium (16). To determine if the combinatorial inactivation of pRb and either p107 or p130 is sufficient to account for HPV-16 E7’s ability to inhibit DNA damage response, mice were exposed to...
ionizing radiation from a $^{137}$Cs source, injected with BrdUrd 24 hours later, and sacrificed 1 hour after injection. The prevailing effect of ionizing radiation on cervical epithelia is an arrest in DNA synthesis within the normally proliferating basal layer of the cervical epithelium, which is maximally observed at 24 hours postirradiation. We scored the frequency of BrdUrd-positive basal cells at this time point. In the cervical epithelia of the different mouse strains. There was a significant radiation-induced arrest in DNA synthesis after irradiation in nontransgenic mice, but this effect of radiation was absent in the $K14E7$ mice (Fig. 2C and D), consistent with prior studies showing that E7 abrogates this DNA damage response (16). In mice singly deficient for 1 pocket protein, we also observed a significant decrease in DNA synthesis after irradiation, indicating that inactivation of individual pocket proteins does not inhibit DNA damage responses. For the $K14CreRbf/f$ mice, this result was consistent with our prior study (16). However, in both $K14CreRbf^f$/$p130^{--/-}$ and tamoxifen-treated $K14CreErtmRbf^f/p107^{--/-}$ mice, we saw an abrogation of the DNA damage response much like that seen in $K14E7$ mice. Thus, combined loss of pRb and either p107 or p130 is sufficient to recapitulate E7’s ability to inhibit DNA damage response in cervix, supporting the premise that there is some degree of functional overlap among pocket proteins in the context of radiation-induced cessation in cellular DNA synthesis.

**Contribution of combinatorial inactivation for either pRb and p107 or pRb and p130 on cervical carcinogenesis**

To determine if combined loss of pRb and either p107 or p130 is also able to recapitulate the oncogenic potential of E7 in cervical carcinogenesis, we exposed mice of the different genotypes described above to exogenous estrogen for 6 months. Exogenous estrogen sufficient to induce continuous estrus acts as a cocarcinogen in the mouse cervix, leading to high incidence of progressive neoplastic disease culminating in cervical cancer in HPV-16 transgenic mice. E7 transgenic mice are particularly sensitive to estrogen induced cervical cancers (8). After treatment with estrogen for 6 months, we evaluated the incidence of cervical cancer and precancerous cervical lesions in cohorts of mice of the following genotypes: non-transgenic, $Rbf^f/p130^{--/-}$ tamoxifen-treated $Rbf^f/p107^{--/-}$, $K14CreRbf^f$, $K14CreRbf^f/p130^{--/-}$, tamoxifen-treated $K14CreErtmRbf^f/p107^{--/-}$, and $K14E7$ mice. Because mice deficient for pRb and p130 were generated on one mixed genetic background, and mice deficient for pRb and p107 were generated on a slightly different genetic background (see Supplementary data), separate sets of positive and negative control groups (e.g., nontransgenic, E7 transgenic, and mice disrupted for individual pocket proteins) were generated on each of these 2 genetic backgrounds, and the histopathology results for each genetic background are reported in separate tables (Tables 1 and 2). Frequencies of cervical cancer and overall severity of disease between the negative controls (i.e., comparing nontransgens in Table 1 to nontransgens in Table 2 $P = 1.1$) and between the positive controls (i.e., comparing $K14E7$ in Table 1 to $K14E7$ in Table 2, $P = 0.7722$, 0.9921) on the 2 genetic backgrounds were not significantly different (see Tables 1 and 2), indicating that these 2 genetic backgrounds had little effect on susceptibility to cervical carcinogenesis. Consistent with our prior studies (8, 16), nearly all $K14E7$ mice developed severe cervical dysplasia and/or invasive cervical cancers, whereas most nontransgenic mice failed to develop high-grade cervical dysplasia or cancer (Tables 1 and 2). Neither high-grade dysplasia nor cervical cancer was observed in estrogen treated $Rbf^f/p130^{--/-}$, $K14CreRbf^f$ (Table 1), and tamoxifen-treated $Rbf^f/p107^{--/-}$ mice (Table 2), indicating that inactivation of single pocket proteins is not sufficient to induce either severe CIN or cancer. Interestingly, in the $K14CreRbf^f/p130^{--/-}$ and tamoxifen-treated $K14CreErtmRbf^f/p107^{--/-}$ mice, we also failed to see a significant increase in frequencies of cancer compared with that observed in the single inactivation of pocket proteins (Tables 1 and 2), despite the fact that pRb was absent in virtually all cells in these tissues (Fig. 1), indicative of efficient Cre recombination.

<table>
<thead>
<tr>
<th>Table 1. Histopathology summary in cervix for the both pRb and p130 conditionally deficient mice treated with estrogen for 6 months</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Genotype</strong>$^a$</td>
</tr>
<tr>
<td></td>
</tr>
<tr>
<td>NTG (n = 22)</td>
</tr>
<tr>
<td>$K14E7^{Rbf}$ (n = 32)</td>
</tr>
<tr>
<td>$K14CreRbf^f$ (n = 42)</td>
</tr>
<tr>
<td>$Rbf^f/p130^{--/-}$ (n = 33)</td>
</tr>
<tr>
<td>$K14CreRbf^f/p130^{--/-}$ (n = 29)$^{b,c,d}$</td>
</tr>
</tbody>
</table>

$^a$All mice were on the same FVB/129C57J mixed genetic background (see Materials and Method for details on breeding scheme).  
$^bP = 0.014$ comparing the incidence of cervical cancer in $K14CreRbf^f/p130^{--/-}$ versus $K14E7$.  
$^cP = 1.7 \times 10^{-10}$ comparing the severity of cervical disease in $K14CreRbf^f/p130^{--/-}$ versus $K14E7$.  
$^dP = 8.2 \times 10^{-6}$, $7.7 \times 10^{-7}$, $1.3 \times 10^{-7}$ comparing the severity of cervical disease in $K14CreRbf^f/p130^{--/-}$ versus NTG, $Rbf^f/p130^{--/-}$, $K14CreRbf^f$, respectively.
The severity of cervical disease, which takes into account the contribution of the inactivation for 3 pocket proteins, pRb, p107, and p130 on cervical carcinogenesis

In K14CreER\textsuperscript{Rb}\textsuperscript{+/−}p130\textsuperscript{−/−} mice, only one out of 29 mice developed invasive cervical cancer (Table 1); this incidence of cervical cancer was not significantly different compared with that in either Rb\textsuperscript{−/−}p130\textsuperscript{−/−} (P = 0.4677), K14CreRb\textsuperscript{−/−} (P = 0.4065), or the nontransgenic (P = 1) mice, whereas it was significantly lower than that seen in the K14E7 mice (P = 0.014). Treatment of mice with estrogen induces not only vaginal neoplasia (Table 2) but also cervical cancers consistent with the role of high-risk HPVs in vaginal cancer in women. We therefore evaluated the susceptibility of the above-described cohorts of mice injected for 1 or 2 pocket proteins to vaginal neoplastic disease. A similar result was observed as seen for the cervix: inactivation of two pocket proteins was not sufficient to induce vaginal neoplasia efficiently as seen in K14E7 transgenic mice (Supplementary Tables 2 and 3).

Contribution of the inactivation for 3 pocket proteins, pRb, p107, and p130 on cervical carcinogenesis

To learn if inactivation for all 3 pocket proteins is able to recapitulate the oncogenic properties of E7, we treated K14CreER\textsuperscript{Rb}\textsuperscript{−/−}p130\textsuperscript{−/−} mice topically with low doses of 4-OHT to recombine the floxed alleles of pRb and p130 selectively in the lower female reproductive tract. This topical treatment was necessary because systemic delivery of even low doses of tamoxifen to K14CreER\textsuperscript{Rb}\textsuperscript{−/−}p130\textsuperscript{−/−} mice led to high morbidity/mortality (data not shown). Topical treatment with 4-OHT led to efficient disruption of expression of pRb and p130 throughout the lower reproductive tract, including the vagina and the majority of the cervix without causing morbidity/mortality. In the cervical septum, which is higher up
in the female reproductive tract and consequently more difficult to expose to 4-OHT, the pattern of loss of pRb and p130 expression was mosaic (approximately 50% of cells losing expression, compared with 100% in all other areas of the cervix). Surprisingly, when these mice were treated for 6 months with estrogen, none of 4-OHT-treated K14CreERtmRbf/fp130f/fp107/C0 mice developed cervical cancer, although a high percentage of developed high-grade dysplasia (CIN3; Table 2). A similar result was observed in the vagina (Supplementary Table S3). Thus, inactivation of all 3 pocket proteins is not sufficient to account for HPV-16 E7's potent ability to induce cancers in the lower female reproductive tract pointing to the likely importance of other cellular targets of E7 in mediating oncogenic potential. That we observed a high incidence of CIN3 and VIN3 shows that disruption of all 3 pocket proteins drives precancerous neoplastic development in the lower female reproductive tract (Table 2, Supplementary Table S3).

Expression of MCM7 in cervical stratified epithelium from the mice deficient for pRb and p107, pRb and p130, and all 3 pocket proteins

MCM7 is a potent biomarker for the detection and diagnosis of progressive cervical disease in HPV-16 transgenic mice and cervical cancer in women (33). MCM7, a component of a cellular DNA helicase (34), is upregulated by E2F-transcription factors (35) that are negatively regulated by pocket proteins (36). MCM7 induction by E7 correlates at least in part with pRb inactivation (16). We conducted MCM7 immunohistochemical staining on the cervical epithelium from mice deficient for multiple pocket proteins (Fig. 3). Consistent with our previous observations, the expression of MCM7 was robustly upregulated throughout the cervical stratified epithelium in K14E7 mice, whereas expression of MCM7 in nontransgenic mice was only detected in basal cells. MCM7 staining in cervical epithelium of both Rb<sup>fl</sup>/p107<sup>−/−</sup> and Rb<sup>fl</sup>/p130<sup>−/−</sup> mice also was mainly restricted to basal cells with some of positive staining in parabasal cells. MCM7 staining was more strongly upregulated in cervical epithelium of K14CreRb<sup>fl</sup>/p130<sup>0/−</sup> mice compared with that detected in either Rb<sup>fl/p107<sup>−/−</sup> or Rb<sup>fl</sup>/p130<sup>−/−</sup> mice, but not as strong and even throughout the whole epithelium compared with that seen in K14E7 mice. MCM7 staining in the K14CreERtmRb<sup>fl</sup>/p107<sup>−/−</sup> and K14CreRb<sup>fl</sup>/p130<sup>0/−</sup> mice was more abundant than seen in K14CreRb<sup>fl</sup>/p107<sup>−/−</sup> mice, particularly for the K14CreERtmRb<sup>fl</sup>/p130<sup>0/−</sup> mice. MCM7 staining in the K14CreERtmRb<sup>fl</sup>/p107<sup>−/−</sup> mice was most comparable to that seen in K14E7 mice. These observations indicate that each pocket protein can partially regulate the expression of MCM7, but inactivation of single or combinations of 2 pocket proteins is not sufficient to recapitulate fully the induction of MCM7 expression seen with HPV-16 E7.

Correlation of combinational inactivation of pocket proteins, arising from leaky CreERtm activity, with cervical dysplasia

Low level Cre activity was previously documented in the absence of tamoxifen induction in the skin of K14creERtm mice (37). Consistent with this observation, we observed a mosaic pattern of the expression of pRb (data not shown) and MCM7 (Fig. 4A) expression in cervical epithelium from 6-month estrogen-treated K14CreERtmRb<sup>fl</sup>/p107<sup>−/−</sup> mice that were not treated with tamoxifen. The areas with high MCM7-positive cells displayed moderate- to high-grade cervical dysplasia (Fig. 4C and 4F), whereas areas largely negative for MCM7 staining did not show this grade dysplasia. No cancers arose in these mice. The frequency of the dysplasia in the

Figure 3. Evaluation of MCM7 expression in cervical epithelium from estrogen-treated mice. Representative images stained with anti-MCM7 antibody. Brown, positive staining; blue, hematoxylin counterstain. Magnification, ×40; scale bar, 200 μm.
K14CreERtmRb<sup>flox</sup>/p107<sup>−/−</sup> mice that were not treated with tamoxifen (Supplementary Table S1) was not as high as in the same genotype treated with tamoxifen (Table 2). These findings provide further evidence for the leakiness of the K14creERtm allele, and further show that inactivation of multiple pocket proteins is sufficient to drive efficient development of dysplasia, but not cancer.

**Discussion**

**The consequence of inactivating all pocket proteins in cervical carcinogenesis**

In this study, we observed that the inactivation of 2 or even all 3 pocket proteins failed to result in the efficient development of invasive cervical cancer in estrogen-treated mice. However, inactivation of pRB and p107, or all 3 pocket proteins was sufficient to drive development of high-grade dysplasia (Tables 1 and 2). These findings provide evidence that p107, and perhaps p130 to a lesser degree, act as tumor suppressors in the mouse cervix in concert with pRB. Furthermore, the combined loss of 2 pocket proteins, either pRB/p107 or pRB/p130, recapitulated at least in part the acute phenotypes induced by E7 in cervical stratified epithelium (Fig. 2). We interpret these results to indicate that the acute phenotypes of E7 are driven by inactivation of pRB and at least 1 other pocket protein, and correlate with the development of precancerous lesions, but not cancer itself. These observations support a model in which inactivation of pocket proteins drive development of benign lesions, but other activities of E7 are necessary for malignant progression. Consistent with this hypothesis, HPV-16 E7 was observed to cause cervical cancer in mice expressing a mutant form of pRB that E7 could not bind (16). Cancers did not form in these same Rb mutant mice without the expression of E7, arguing that an Rb-independent activity (s) of E7 contributes to cervical carcinogenesis. We conclude that pocket proteins-independent activities of E7 contribute to cervical carcinogenesis.

The degree of inactivation for Rb family proteins may be subtly different between two groups: floxed Rb allele/germline Rb-null allele mice versus E7 transgenic mice. Previously, it has been shown that E7 cannot completely inactivate Rb family proteins; in contrast, Cre expression in Rb-floxed allele mice results in more than 90% loss of detectable pRB in cervical epithelium (16). We suspect that this difference does not significantly affect our results for 2 reasons: (i) all of Rb family proteins were scarcely detected in Rb-knockout allele mice as well as K14E7 mice (Figs. 1 and 2) acute phenotypes and biomarker studies indicate that the effect of inactivation for Rb-knockout allele equates to that observed in K14E7 mice (Figs. 2 and 3).

**Redundancy of pocket protein family members in cervix**

Studies have reported functional overlap between pocket protein family members in the context of development and cell-cycle regulation. This functional overlap may explain both the genetic redundancy and the functional compensation among pocket protein family members (23). We observed that the expressions of p107 and to a lesser degree p130 are increased in cervical epithelium deficient for pRB (Fig. 1). A compensatory induction of p107 in response to the inactivation of pRB has been reported in several other mouse tissues (28, 38, 39). It remains unclear whether compensatory induction in expression of pocket proteins in cervical epithelium necessarily informs us on functional compensation.

www.aacjrournals.org  Cancer Res; 72(20) October 15, 2012  5425
Role of other cellular target(s) of E7 in cervical carcinogenesis

Using tandem affinity purification/mass spec analyses, more than 100 different cellular factors have been found associated with E7 protein (9). Which of these contribute to E7-mediated cervical carcinogenesis remains largely unclear. We have previously shown a role of inactivation of p21, one such target of E7 and a known cellular tumor suppressor, in cervical carcinogenesis (32). Thus, there is at least 1 nonpocket protein target of E7 that contributes to E7’s oncogenic potential in this tissue. But, there are likely more. E7 can associate with Mi2β and histone deacetylase (40), and these interactions have led to the prediction that E7 can broadly dysregulate transcription of cellular genes through chromatin remodeling. More recently, HPV-16 E7 was discovered to associate with E2F6-containing polycomb transcriptional repressor complexes (41) that control the expression of a variety of genes through histone modification (42), and to induce the expression of histone demethylase (43). Aberrant DNA methylation patterns are known to be frequent events in cancers. DNA methylation is thought to be important in many processes including DNA repair, genome stability, and chromatin structure (44, 45). Recently, it was reported that E7 directly associates with and alters the activity of Dnmt1, a DNA methyltransferase (46). Dnmt1 is highly overexpressed in HPV-associated cancers compared with that normal tissue (17). We propose that E7’s modulation of the epigenome contributes to its role in cervical cancer in a manner that synergizes with its other capabilities of dysregulating the cell cycle. Epigenetic reprogramming is thought to be a relatively late step in neoplastic progression (10, 47). Consistent with this observation, the number of genes dysregulated in the progressive disease leading to cervical cancer most dramatically increases in the transition between CIN3 and cervical cancer (Ahlquist et al., in preparation).

Differences in the importance of pocket proteins in E7-driven carcinogenesis in different tissues

The findings of this study differ greatly from our prior results in the context of head and neck cancer, another cancer type caused by high risk HPV’s in which E7 is again the primary driver (48). Inactivation of pRb and p107 together were found to induce susceptibility to head and neck cancers as efficiently as did E7 (30). A less dramatic synergy was seen with the inactivation of both pRb and p130. This is in striking contrast to findings in the current study investigating cervical carcinogenesis, in which inactivation of all three, let alone 2 pocket proteins was unable to efficiently induce cervical cancer. What can explain this difference? One possibility is the manner in which cancer is induced in these different tissues. For cervical cancer, we use estrogen that likely acts primarily as a reversible promoter (49) through its activation of its receptor ERα (50). In contrast, in the head and neck mouse model we use a mutagen, 4-NQO, that likely is driving the accumulation of genetic changes in cells. Another possibility is that this difference in susceptibility reflects differences in the tissue type, not the cocarcinogen. For instance, p107 is able to act as a tumor suppressor in the context of both retinoblastoma and non-small cell lung cancer in an Rb-deficient background (25, 27). Whereas, p130 can act as a tumor suppressor in the context of small cell lung carcinoma as well as non-small cell lung carcinomas in an Rb/p53-deficient background (26, 28). These findings support our hypothesis that tumor susceptibility may differ depending on the tissue context.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Authors’ Contributions

Conception and design: M.-K. Shin, P.F. Lambert
Development of methodology: M.-K. Shin, J. Sage
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): P.F. Lambert
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): M.-K. Shin, P.F. Lambert
Writing, review, and/or revision of the manuscript: M.-K. Shin, J. Sage, P.F. Lambert
Study supervision: P.F. Lambert

Acknowledgments

The authors thank the members of the Lambert laboratory for helpful discussions. This work was supported by grants from the NIH CA098428 and CA022443.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received May 31, 2012; revised August 15, 2012; accepted August 20, 2012; published OnlineFirst August 1, 2012.

References

11. Munger K, Werness BA, Dyson N, Phelps WC, Harlow E, Howley PM. Complex formation of human papillomavirus E7 proteins with the
The Role of Pocket Proteins in Cervical Carcinogenesis

Inactivating All Three Rb Family Pocket Proteins Is Insufficient to Initiate Cervical Cancer

Myeong-Kyun Shin, Julien Sage and Paul F. Lambert


Updated version
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-12-2083

Supplementary Material
Access the most recent supplemental material at:
http://cancerres.aacrjournals.org/content/suppl/2012/08/30/0008-5472.CAN-12-2083.DC1

Cited articles
This article cites 49 articles, 26 of which you can access for free at:
http://cancerres.aacrjournals.org/content/72/20/5418.full.html#ref-list-1

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.