Distinctive Features of the Differentiated Phenotype and Infiltration of Tumor-Reactive Lymphocytes in Clear Cell Renal Cell Carcinoma

Qiong J. Wang, Ken-ichi Hanada, Paul F. Robbins, Yong F. Li, and James C. Yang

Abstract

Clear cell renal cell carcinoma (RCC) is considered an immunogenic tumor, but it has been difficult to identify tumor-infiltrating lymphocytes (TIL) that show in vitro tumor recognition. We compared the characteristics of fresh RCC TIL to peripheral blood lymphocytes (PBL) or melanoma TIL. Our results showed that RCC TIL contained fewer CD27+ T cells, and fewer naive and central memory (CM) T cells, but more effector memory (EM) T cells than melanoma TIL or renal PBL. We hypothesized that factors in the RCC microenvironment were skewing TIL phenotype toward EM. One possibility was the expression of CD70 on nearly all human RCCs, but not melanomas. Differentiation of naïve T cells to EM cells only occurred from CD70 costimulation in concert with T-cell receptor (TCR) stimulation (signal one), suggesting that EM TIL responding to CD70 would be enriched for T cells reactive with local antigens, including those associated with RCC. Clonotypic analysis of TCRs in fresh RCCs showed that EM T cells were more clonally expanded than CM or naïve T cells, and the clonal expansion occurred at the tumor site as oligoclonal TCRs were distinct from PBL TCRs from the same patient. In addition, we found that 2 TCRs from the highly represented EM TIL clones, when reexpressed in fresh PBL, recognized an MHC-class II or MHC-class I-restricted antigens shared by multiple RCC lines. Our results suggest that RCC-reactive TIL do exist in situ, but may be difficult to recover and study because of proliferative exhaustion, driven by tumor-expressed CD70.

Introduction

Adoptively transferring tumor-infiltrating lymphocytes (TIL) expanded in vitro back into patients with metastatic melanoma can mediate durable and complete tumor regressions (1, 2). These melanoma TILs will show immunologic recognition of their autologous melanoma cells in 67% of patients (3). However, this unexplained ability to generate tumor-reactive TIL from melanoma does not extend to clear cell renal cell carcinoma (RCC) as only a few have been generated from RCC (4–6), despite the fact that both cancers can respond to a variety of immunotherapies. There are many potential reasons why tumor-reactive TIL might not expand and destroy the tumor they reside in, including immunosuppression by inhibitory receptors, cytokines, or by T-regulatory cells (7). Factors such as PD-1, CTLA-4, IL-10, and TGF-β have been thought to play a role in blunting antitumor reactivity (8–12). Yet none of these factors explains the dramatic functional differences seen between TIL expanded in vitro from melanomas versus RCC. With both cancers responding clinically to IL-2, ipilimumab, and anti–PD-1 antibody (13–16), it would be hard to suggest that RCC-reactive TIL or T-cells do not exist in patients with RCC. We considered whether the failure to show tumor-reactive TIL in RCC might be an artifact of the need to expand them in vitro for study and began looking at fresh RCC TIL. In support of this, Dietrich and colleagues found that in vitro lymphocyte culture of RCC TIL had a negative impact on selecting T-cell repertoire, as some highly represented T-cell populations present in vivo disappeared after in vitro stimulation by T-cell receptor (TCR) analysis (17). Therefore, investigating phenotypes of T cells residing in renal tumors comparing with melanoma would help us to understand the differences between these 2 malignancies. Recently, CD70 (normally expressed by activated immune cells) has been identified as a diagnostic marker on RCC (18–19). CD70, a member of the TNF super family, has been implicated in T-cell survival and activation through interaction with its costimulatory receptor, CD27 (20). The role of CD70 in tumor immunology is controversial, conferring benefit when expressed by CD8+ TIL administered therapeutically to patients with melanoma (21), but perhaps mediating immune cell apoptosis and immune escape in glioblastoma and RCC (22–23). Most interestingly, in a CD70-transgenic murine model, Tesselaar and colleagues have shown that constitutive expression of CD70 by B cells resulted in exhaustion of the
naive T-cell pool, depletion of T cells from lymph nodes, and death from opportunistic infection (24). Therefore, we investigated activation and differentiation states of T cells residing in renal tumors comparing with melanoma, and the possible role of RCC-expressed CD70 in RCC TIL differentiation. We also investigated whether recognition of RCC-associated cognate antigens could be playing a role in driving differentiation of the most dominant RCC TIL clones. This possibility could lead to identifying RCC-reactive T-cell clones and new RCC-associated antigens.

Materials and Methods

Fresh tumor digests and tumor lines

Primary or stage IV metastatic tumors surgically resected from patients with clear cell RCC were enzymatically digested with 0.1% collagenase type IV, 0.01% hyaluronidase type V, and 30 U/mL deoxyribonuclease type IV (Sigma Chemical) in RPMI-1640 (Life Technologies) at room temperature for 3 hours. In total, tumors from 16 patients were processed, including 8 primary tumors and 8 metastatic tumors. The cells were filtered through 100-μm nylon mesh, and separated by density gradient centrifugation using Lymphocyte Separation Medium (Organon Teknica). After digestion, cells were suspended in 90% of human serum (Valley Biomedical) with 10% dimethyl sulfoxide (DMSO) and frozen at −70 °C. Tumor lines from patients with RCC were established and maintained in Dulbecco’s Modified Eagle Medium (DMEM; Life Technologies) including 10% FBS (Life Technologies), 10% tryptose phosphate (Sigma), 1 × insulin-transferrin selenium (Life Technologies), and 1 × serum pyruvate (Life Technologies). For T-cell phenotypic analyses, fresh tumor digests were thawed, resuspended in RPMI-1640 with 10% human serum, and cultured overnight at 37 °C in 5% CO₂. Nonadherent cells were then harvested for staining with corresponding antibodies. For non-T-cell analyses, the fresh tumor digests were thawed, blocked with purified IgG, and labeled with corresponding antibodies immediately.

Similarly, procured fresh tumor digests of melanoma and melanoma lines were obtained from Surgery Branch Laboratories (National Cancer Institute, Bethesda, MD), and the tumor lines were maintained in RPMI-1640 with 10% FBS. All melanomas used in this study are stage IV metastatic melanomas, including lung, liver, or kidney metastases.

All cell lines included in the study were generated at Surgery Branch, NCI, and tested and authenticated by HLA genotyping. The cell lines were routinely confirmed with their HLA typing and antigen expression by flow cytometry and coculture assays, respectively.

Antibodies

For immunophenotypic analyses, monoclonal antibodies (MoAb) including FITC-labeled anti-human IgG isotypes, CD8 (clone SK-1), CD16 (clone 3G8), CD19 (clone HB19), CD69 (clone L78), or Lin 1; PE-labeled anti-human IgG isotypes, CD62L (clone Dre56), CD70 (Ki-24), or CD25 (clone 2A3); PE-cy7-labeled anti-human IgG isotypes or CD3 (clone SK7); APC-labeled anti-human IgG isotypes, CD45RO (clone UCHL1), CD28 (clone CD28.2), CD11c (S-HCL-3), or CD14 (clone MφP9); and APC-cy7-labeled anti-human IgG isotypes or CD4 (clone SK3) were purchased from BD Pharmingen. APC-labeled anti-human CD27 (M-T271) Ab was purchased from eBiosciences.

The effect of plate-bound human CD70

Purified human CD70 fused with murine CD8 alpha (CD70-muCD8, Ancell Corporation; 0.03–10 μg/mL) or controls were plated onto a 96-well flat-bottom plate that was precoated with anti-mouse CD8 alpha or PBS alone (Ancell; 10 μg/mL) overnight at 4 °C. The plate was blocked with PBS containing 1% bovine serum albumin (Sigma) for 30 minutes at room temperature, and washed with PBS 3 times before adding T cells.

 Peripheral blood lymphocytes (PBL) from patients with RCC were thawed and cultured in RPMI-1640 with 10% human serum overnight. Nonadherent cells were harvested the next day, labeled with fluorescence-conjugated antibodies (Abs) CD3, CD62L, and CD45RO. Cells were incubated for 30 minutes at 4 °C, washed with PBS containing 1% bovine serum albumin, and sorted for CD3+ CD62L+ CD45RO− (naïve) or CD3+ CD62L+ CD45RO+ [central memory (CM)] subpopulations on FACS Aria (BD Biosciences). The purity of sorted populations was confirmed by analyzing a small portion of cells on FACS Canto II (BD Biosciences). After sorting, cells were plated at 2 × 10⁵ cells per well and cultured for 5 days. On day 5, cells were labeled with fluorescence-conjugated Abs CD3, CD62L, and CD45RO, and analyzed on FACS Canto II.

Clonotypic analysis of renal fresh tumor digests and peripheral blood

Effector memory (EM; CD3+ CD62L− CD45RO+), CM, or naïve T-cell subpopulations were sorted from renal fresh tumor digests or PBL as described above. Total RNA from each subpopulation was purified with an RNeasy mini kit (Qiagen). 5’ RACE reaction was carried out by the SMARTer RACE cDNA Amplification Kit (Clontech) following the manufacturer’s instructions. The RACE cDNAs (~800 bp) were obtained with primers complementary to the constant region of TCR α or β chain and then inserted into the pCR2.1 vector by TA cloning (Life Technologies). Primers for the TCR α or β chain were synthesized by Life Technologies, and their sequences were 5’-GCCACAGCACTGTTGCTCTTGAAGTCC or 5’-CAGGCCAGTATCTGGAGTCATTGAG, respectively. After TA cloning, 96 colonies were picked from each 5’ RACE product of both TCR α and β chains and their variable regions and complementarity determining region 3 (CDR3) were sequenced.

Retroviral production and transduction of anti-CD3-stimulated PBL

cDNAs encoding selected full-length TCR α chains and β chains were cloned into the pMSGV1 plasmid, which is a derivative of the murine stem cell virus-based splice-gag vector (pMSGV), as described in previous publications with some modifications (25). Briefly, full-length TCR α and β chain cDNAs were amplified by PCR using the pairs appropriate to corresponding sequences of each TCR α chain and β chain with a P2A sequence used as the spacer in between.
To produce retrovirus, 293gp cells were transfected with 9 μg of pMSGV1-TCR and 3 μg of plasmid RD114 using Lipofectamine 2000 (Life Technologies; 60 μL). Two days later, the supernatants were harvested and used to transduce anti-CD3–stimulated PBL. PBL from allogeneic donors were stimulated with soluble OKT-3 (50 ng/mL) and IL-2 (300 IU/mL) for 2 days before transduction was carried out. The stimulated cells were added to 24-well plates initially coated with Retronectin (Takara) and subsequently precoated with retrovirus by spinoculation (2,000 × g, 32°C, 2 hours) at 5 × 10⁷/mL. The plates were then centrifuged at 1,000 × g for 10 minutes and incubated overnight at 37°C in a 5% CO₂ incubator. This procedure was repeated the next day and cells were split as necessary to maintain cell density between 0.5 and 1 × 10⁶ cells/mL. Transduction efficiency was determined by analyzing β3 expression of retrovirally transduced cells if the antibody was available.

Cytokine release and blocking assay
Retrovirally transduced cells (1 × 10⁶) were cocultured with 5 × 10⁵ autologous renal tumor lines with or without transduction with the class II transcriptional activator (CIITA) at 37°C, 5% CO₂ overnight and tested for IFN-γ secretion. In some experiments, HLA-matched allogeneic renal tumor lines were included in the assay.

For the blocking assay, RCC cells (5 × 10⁴ cells in 100 μL culture medium) were incubated with each blocking mAb at a concentration of 10 μg/mL for 30 minutes at 37°C in a flat-bottom 96-well plate. T cells (1 × 10⁵ cells/well) were then added and incubated with target cells overnight at 37°C. The supernatants were harvested and assayed for IFN-γ concentration by ELISA.

Statistical analysis
The 2-tailed unpaired t test with Welch correction was used for comparing T-cell population phenotypes and lineage 1-negative cell populations in renal and melanoma tumors.

Results
Phenotypic differences between fresh tumor digests from RCC and melanomas
To understand the differences between renal tumors and melanomas, we compared the phenotypes of immune cells residing in fresh tumor digests from these 2 malignancies. No differences were observed in the percentages of T cells (CD8 or CD4 subpopulations), natural killer (NK) cells, or B cells when comparing renal tumors with melanomas. Interestingly, renal fresh tumors contained a significantly lower percentage of CD27-expressing T cells than melanomas and PBL from patients with RCC (Fig. 1A). Moreover, when analyzing the differentiation and activation states of T cells residing in tumors by expression of CD62L and CD45RO (Fig. 1B), melanoma fresh tumor digests contained significantly more naïve and CM subpopulations than renal fresh tumor digests, as shown by higher percentages of CD62L-CD45RO+ T cells and CD62L+CD45RO- T cells, respectively. In contrast, renal tumors contained significantly more EM (CD62L-CD45RO-) cells than melanomas. The differences were observed in both CD8 and CD4 T-cell subpopulations. No differences were observed between primary and metastatic renal cancers. In addition, PBL from patients with RCC exhibited a less differentiated T-cell phenotype than renal tumors, as shown by a higher percentage of naïve and CM T cells and a lower percentage of EM T cells. Furthermore, a higher percentage of T cells from renal tumors expressed a late-differentiation marker, CD57, than those from melanomas (data not shown). Our results show that T cells that reside in renal tumors are more terminally differentiated.

CD70 expression on renal tumors versus melanomas and its effect on naïve T cells
Because renal fresh tumor digests contain fewer CD27-expressing T cells, we further analyzed CD70 expression in renal tumors compared with melanoma. RCC-resident B-cell and NK populations all contained a higher percentage of CD70⁺ cells than melanomas (data not shown). However, the most striking difference was the consistent presence of CD70-expressing cells in Lineage 1-negative (Lin⁻) cell populations (i.e., nonimmune cells) in renal fresh tumor digests compared with melanoma (Fig. 2A). Further analysis of CD70 expression on our renal cancer and melanoma cell lines confirmed previous studies in which renal tumors expressed CD70 on their surface (Fig. 2B; ref. 18), whereas melanomas did not. The correlation between CD70 expression in Lin⁻ populations and T-cell phenotypes was also analyzed and the data suggest that the higher percentage of EM and lower percentage of CM T cells were associated with higher level of CD70 expression (Supplementary Fig. S1). We further tested the effect of CD70 using anti-CD3 antibody and purified human recombinant CD70 protein coated onto plastic plates. As shown in Fig. 2C and D, naïve human T cells differentiated when they received both a TCR signal and costimulation by CD70 as shown by increased EM and CM T cells and diminished naïve T cells. This effect was CD70 dose dependent. These data suggested that the tonic costimulation by RCC-expressed CD70 would have maximal effect on T-cell clones only when T cells were engaging their cognate antigen in the RCC tumor microenvironment. Therefore, T-cell clonotypes reacting with RCC-associated antigens might be those with the most differentiated phenotype.

Clonal diversity of T-cell subpopulations in renal tumors
To avoid the effects of differential clonal expansion in vitro and best reflect the in situ situation, we examined the clonal diversity in RCC TIL subpopulations by randomly selecting and sequencing TCR α and β chains from fresh TIL. The most overrepresented TCRs were then reexpressed in PBL and tested for reactivity with RCC from which the TCRs were obtained. EM, CM, and naïve T-cell populations were FACS sorted from freshly dispersed RCCs, RNA extracted, and cDNAs were synthesized and sequenced using primers complementary to the constant regions of either the α or β chains. Approximately, 70 α chain and 70 β chain sequences were analyzed from each T-cell subpopulation. Figure 3 shows representative results from one RCC in which freshly sorted EM TIL contained oligoclonal TCRs as shown...
by 3 highly represented TCR α chains (TRAV14*03TRAJ3*01, TRAV13-1*01TRAJ0*01, and TRAV38-1*01TRAJ23*01) and 3 dominant TCR β chains (TRBV13*01TRBD2*01TRBJ2-1*01, TRBV9*01TRBJ2-7*01, and TRBV27*01TRBD1*01TRBJ2-5*01). CM T cells showed less pronounced oligoclonality (2 TCR α chains and 1 TCR β chain) and naïve T cells had a diversified repertoire. Interestingly, the specific oligoclonal TCRs found in EM TIL were never found in either the CM or naïve T-cell populations. In total, renal fresh tumor digests from 4 patients with RCC were analyzed and 3 showed distinct oligoclonality in their sorted EM cells. We used the same methods to examine the diversity of EM T cells in the peripheral blood and renal tumors of the same patient. As shown in Fig. 4, the diversity and clonotypes of TCR α (Fig. 4A) and β chains (Fig. 4B) were totally different between the RCC TIL and the PBL. Clonal expansion not only occurred more extensively and more frequently in EM T cells from fresh tumor digests than from PBL, but the overrepresented TCR α or β chains present in the renal fresh tumor digest were not found in the patient’s PBL. Conversely, the only oligoclonal TCR α chain in renal PBL was not present in the renal fresh tumor digest. Therefore, oligoclonal EM T cells from renal fresh tumor digests were distinct from PBL. This supports the hypothesis that the oligoclonality of EM T cells may be the product of the highly stimulatory renal tumor microenvironment.

**Functional analysis of oligoclonal EM TCRs**

The presence of overrepresented oligoclonal EM TCRs allowed us to isolate full-length TCR α and β chains, pair them in all possible combinations in bicistronic retroviral vectors,
and test their reactivity against renal tumors by transducing allogeneic PBLs. All 3 patients with RCC with oligoclonal TCRs in their EM populations were examined and their tumor reactivity was tested. Testing only a total of 8 TCR α chains and 8 TCR β chains from these 3 patients, tumor-reactive TCRs were identified in 2 of them. Tumor reactivity of TCRs from patient RV was shown in Fig. 5. Nine retroviruses representing all the possible pairing of 3 TCR α chains and 3 β chains, and used to transduce to anti-CD3-stimulated allogeneic PBL. One TCR, consisting of TRAV24 and TRBV24, could confer reactivity against CIITA-transduced, HLA-class II expressing autologous tumor cells (RCC/CIITA) but not parental RCC line (Fig. 5A). This reactivity could be blocked by anti-HLA DR antibodies, but not anti-HLA DR antibodies (Fig. 5B). When it was tested against multiple renal tumor lines, TRAV24/TRBV24-transduced PBL recognized 2 of 4 HLA-DRB1*01-matched tumor lines when induced to express HLA class II by CIITA transduction (RCC #2 and RCC #12; Fig. 5C).

Figure 2. CD70 expression on tumors and the effect of purified CD70 on naïve CD8⁺ or CD4⁺ T cells. A, CD70 expression on nonimmune cells in renal fresh tumor digests compared to melanoma. Fresh tumor digests were labeled with fluorescence-conjugated Lineage 1 cocktail antibodies (Lin 1) and CD70, and then analyzed on FACS Canto II. Lin 1⁻ cells represent nonimmune cells in the fresh tumor digests. FACs plots represented CD70 expression in RCC or melanoma. B, CD70 expression on renal tumor lines and melanoma lines. Tumor lines established from 10 patients with RCC, and 10 patients with melanoma were thawed and grown to their confluence. The tumor cells were then harvested and labeled either with PE-conjugated anti-CD70 Abs (filled histogram) or isotype control (open histogram), and analyzed on FACS Canto II. C, the effect of purified CD70 on naïve CD8⁺ or CD4⁺ T cells. Peripheral blood mononuclear cells were thawed and cultured in RPMI with 10% human serum overnight, and sorted for CD8⁺/CD62L⁻/CD45RO⁻ or CD4⁺/CD62L⁻/CD45RO⁻ naïve T-cell populations on FACS Aria. Sorted naïve CD8⁺ or CD4⁺ T cells (2 × 10⁵/well) were added to the 96-well plate that were precoated with anti-human CD3 (10 μg/mL) and/or purified human CD70 protein (10 μg/mL) and cultured in RPMI + 10% human serum + IL-2 (50 cu/mL) for 5 days. Five days later, T cells were labeled with fluorescence-conjugated Abs, CD3, CD8, CD4, CD62L, and CD45RO, and analyzed on FACS Canto II. D, the dose-dependent effect of purified CD70 on naïve CD8⁺ or CD4⁺ T cells. Sorted naïve CD8⁺ or CD4⁺ T cells were cultured with different doses of purified human CD70 protein (0–10 μg/mL) in the presence or absence of anti-CD3 Ab (10 μg/mL). The experiments in C and D were conducted multiple times and gave similar results.
No tumor recognition was detected when tumor lines did not express HLA class II. In addition, an HLA-DRB1*01/C8*01 tumor line (RCC #13) was not recognized by TRAV24/TRBV24-transduced PBL. Furthermore, control PBL transduced with GFP did not recognize any of these tumors. These results suggested that the reactivity of TRAV24/TRBV24-transduced PBL was restricted by HLA-DRB1*01 and recognized a shared antigen expressed by 3 of 5 HLA-DRB1*01+ renal tumors.

Figure 3. TCR clonotypes in EM, CM, or naïve T cells from renal fresh tumor digests. Fresh tumor digests from patient DS with RCC were thawed and cultured in RPMI+10% human serum overnight. The nonadherent cells were stained with fluorescence-labeled CD3, CD62L, and CD45RO, and FACS-sorted to EM (CD62L−CD45RO−), CM (CD62L+CD45RO+), and naïve cell (CD62L+CD45RO+) subpopulations. Approximately, 70 sequences of TCR α chains and β chains in each population were evaluable after 5’RACE using gene-specific primers. Each pile chart shows the composition of TCR α or β chains. Each sector represents a single α or β chain with a unique CDR3.

Figure 4. Frequencies of EM TCR α chains (A) and β chains (B) from fresh tumor digests compared with PBL from patient RV with RCC. Each single bar represents the frequency of a single variable TCR α or β chain. The gray bar within each bar represents the frequency of the dominant single TCR α or β chain with a unique CDR3.
Using the same α/β TCR pairing approach, we also identified a tumor-reactive EM TCR in patient DS with RCC. As shown in Fig. 6A, allogeneic PBL were retrovirally transduced with 9 TCRs, and one TCR (TRAV14/TRBV13) could confer low-level reactivity against the autologous tumor line. This reactivity could be blocked by HLA-class I Ab, but not HLA-class II or HLA-DR (Fig. 6B), and it recognized 2 HLA-A3-matched renal tumor lines (Fig. 6C), suggesting the reactivity could be HLA-A3 restricted. Because of the availability of the antibody against TRBV13 (Vβ23), we were able to fluorescence-sort Vβ23+ cells from RCC TIL and analyze the sequences of α chains (Table 1). Among 68 sequences analyzed, 44 had the same variable region TRAV14 and 43 of them shared the identical CDR3 region with our reconstructed candidate TCR. These results showed that the tumor-reactive EM TCR (TRAV14/TRBV13) was a naturally occurring TCR.

Discussion

The current study investigated the differences between RCC and melanoma TIL. By phenotypic analyses of the TIL from both malignancies, we found that they were significantly different in phenotype, with a much more differentiated profile in RCC TIL (i.e., more EM T cells, and less CM and naïve T cells) than in melanoma TIL or PBL from patients with RCC. T cells residing in renal tumors also contained significantly higher percentage of CD57-expressing T cells than melanoma (data not shown), suggesting that RCC TIL were mostly late in the differentiation sequence. This advanced state of differentiation could account for the failure of tumor-reactive RCC TIL to

Figure 5. Oligoclonal EM TCRs from patient RV with RCC could confer HLA-class II-restricted tumor reactivity. A, IFN-γ production by PBL transduced with oligoclonal EM TCRs. Allogeneic PBL were stimulated with anti-CD3 (50 ng/mL) for 2 days and transduced twice with retrovirus encoding oligoclonal EM TCRs at 0.5 × 10^5 cells/well in a 24-well plate. PBL retrovirally transduced with green fluorescence protein (GFP) was used as the control. Three days after transduction, transduced cells (1 × 10^5) were cocultured with 5 × 10^5 autologous tumors (RCC) or tumors transduced with CIITA (RCC/CITIA), which is a known transcriptional factor to upregulate HLA-class II on the cell surface as shown in Supplemental Fig. S2. After overnight incubation, the supernatant was harvested and IFN-γ production was measured. GFP-transduced and untransduced PBLs were used as controls. B, the reactivity of PBL transduced with TRAV24/TRBV24 was blocked with HLA-class II and HLA-DR Abs. PBL transduced with TRAV24/TRBV24 were cocultured overnight with autologous RCC/CITIA in the presence of anti-HLA class I, anti-HLA class II, or anti-HLA DR antibodies (10 µg/mL), and IFN-γ from the coculture supernatant was measured the next day. PBLs transduced with a HLA-class I-restricted TCR recognizing MART-1 and a HLA-class II-restricted TCR recognizing tyrosinase were used as controls. C, IFN-γ production of PBL transduced with TRAV24/TRBV24 against HLA-DRB1*01 matched or mismatched tumors.
expand in vitro. Our studies of fresh RCC also confirmed the published finding that the tumor cells themselves express CD70 (18–19), the member of tumor necrosis factor family. In a normal host, expression of CD70 is mainly restricted to lymphoid tissues such as active T cells, B cells, or dendritic cells, and the interaction between CD70 and its receptor CD27 can lead to different immune response outcomes depending on the timing, context, and intensity (26). Several studies have suggested that CD27–CD70 costimulation positively modulates T-cell activation, differentiation, and survival, and therefore maintains antigen-specific T-cell responses (27–29). Using CD27-deficient mice, Hendriks and colleagues showed that these mice infected with influenza virus had decreased numbers of effector T cells in the lung. They have also shown that costimulation of CD27–CD70 contributed to maturation of T cells and promoted CD8+ virus-specific T-cell responses in acute viral infection (27). Furthermore, CD27 promotes proliferation and survival of activated T cells (29). On the contrary, some groups have advocated that CD70 induces apoptosis in T cells, which serves as one of the immune escape mechanisms of CD70-expressing tumors such as RCC and glioblastoma (22–23). However, in our study, we did not observe an increase in steady-state apoptosis in RCC TIL (data not shown). Another situation where there is a negative impact of CD70 has been in chronic viral infection, with poor T-cell responses and prognosis being associated with CD70 expression in human immunodeficiency virus-positive patients (30). This closely reflects the CD70-transgenic mouse model, where immunosuppression and opportunistic infections are seen, associated with a terminally differentiated T-cell repertoire (24). In addition, when the CD70/CD27 pathway is blocked, viral control was improved in a mouse chronic lymphocytic choriomeningitis
most RCC-reactive T-cells during their first IL-2 therapy. We then relapsing virtually never respond to additional IL-2 therapy. This hypothesis was also attractive in light of the clinical observation that RCC patients responding to high-dose IL-2 for study, characterization, or adoptive therapy. Such clonotypic dominance was not seen in naïve or CM TIL from the same patients. The most overrepresented TCR-α and TCR-β chains in a patient’s TIL were then transduced in random pairs to produce putative α/β TCRs for testing against the autologous tumor line. Two of 3 patients with TIL subjected to this extensive process produced α/β TCR that recognized the autologous tumor. In one case where an antibody to the TCR-β chain was available, we were further able to show that our TCR pairing was likely the native pairing when sequencing showed that nearly all TIL sorted for the selected TCR-β contained the exact TCR-α we had selected with tumor recognition. Still, not all patients and overrepresented TCR species could be shown to react with autologous tumor. This could be because of technical limitations, or the TCR of some TIL may be recognizing antigens presented by antigen presenting cells rather than tumor itself. Yet in extensive studies of bulk RCC TIL, there are only rare instances of finding tumor reactive T-cells, so this experience in a small number of cases seems unusual.

In addition to possibly elucidating one reason why RCC TIL do not show tumor recognition after expansion for study, this approach may lead to a general technique for finding T-cell clones with tumor recognition from RCC and perhaps other tumors. Only melanoma routinely yields tumor reactive TIL, whereas all other cancers studied do so very infrequently. This does not illuminate the reasons for the singular behavior of melanoma, nor provide answers about other cancers that do not display CD70. Future studies will examine the phenotype of fresh TIL from other cancers, looking for evidence of advanced T-cell differentiation or alternate explanations. The ability to find individual T-cell clones with tumor reactivity can be exploited by cloning their TCR and retrovirally reengineering it into the T cells of other patients using widely available and efficient techniques. We do not yet know the antigens recognized by the T-cell clones we found. Future antigen identification efforts will be applied to receptors with the highest avidity, and broad and shared reactivity, preferably restricted by MHC class I. Additional studies attempting to correlate CD70 levels or TIL differentiation states in RCC with their response to immunotherapy would also be of use in corroborating the hypothesis proposed by these studies. In summary, these studies suggest a novel mechanism to explain how tumor-reactive TIL can reside in RCC, potentially participate in the strategy of rescuing their TCRs and reexpressing them in fresh T-cells to allow their characterization.

Several RCC TIL did indeed show individual T-cell clonotypes in their EM populations that constituted up to 19% of all the TCR randomly selected and cloned with 5’RACE. Such clonotypic dominance was not seen in naïve or CM TIL from the same patients. The most overrepresented TCR-α and TCR-β chains in a patient’s TIL were then transduced in random pairs to produce putative α/β TCRs for testing against the autologous tumor line. Two of 3 patients with TIL subjected to this extensive process produced α/β TCR that recognized the autologous tumor. In one case where an antibody to the TCR-β chain was available, we were further able to show that our TCR pairing was likely the native pairing when sequencing showed that nearly all TIL sorted for the selected TCR-β contained the exact TCR-α we had selected with tumor recognition. Still, not all patients and overrepresented TCR species could be shown to react with autologous tumor. This could be because of technical limitations, or the TCR of some TIL may be recognizing antigens presented by antigen presenting cells rather than tumor itself. Yet in extensive studies of bulk RCC TIL, there are only rare instances of finding tumor reactive T-cells, so this experience in a small number of cases seems unusual.

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### Table 1. 5’ RACE of TCR α chain in Vβ23+ EM populations from patient DS with RCC

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*82% of Vβ23+ EM populations was TRBV13, 88% of which shared the same CDR3 region.

*Total sequences analyzed for α chain were 68.*
Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors' Contributions
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Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): Q.J. Wang, J. Yang
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): Q.J. Wang, J. Yang
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Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): J. Yang
Study supervision: J. Yang

References

Acknowledgments
The authors thank Drs. Steven A Rosenberg, John Wunderlich, Shicheng Yang, and Jianping Huang for thoughtful discussions.

Grant Support
This research was supported by the Intramural Research Program of the NIH, National Cancer Institute, and Center for Cancer Research.

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Received February 17, 2012; revised September 17, 2012; accepted September 25, 2012; published online First October 15, 2012.


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