

## Upregulation of miR-196a and *HOTAIR* Drive Malignant Character in Gastrointestinal Stromal Tumors

Takeshi Niinuma<sup>1</sup>, Hiromu Suzuki<sup>1,3</sup>, Masanori Nojima<sup>4</sup>, Katsuhiko Noshō<sup>1</sup>, Hiroyuki Yamamoto<sup>1</sup>, Hiroyuki Takamaru<sup>1</sup>, Eiichiro Yamamoto<sup>3</sup>, Reo Maruyama<sup>3</sup>, Takayuki Nobuoka<sup>2</sup>, Yasuaki Miyazaki<sup>9</sup>, Toshiro Nishida<sup>9,10</sup>, Takeo Bamba<sup>11</sup>, Tatsuo Kanda<sup>11</sup>, Yoichi Ajioka<sup>12</sup>, Takahiro Taguchi<sup>13</sup>, Satoshi Okahara<sup>7</sup>, Hiroaki Takahashi<sup>7</sup>, Yasunori Nishida<sup>8</sup>, Masao Hosokawa<sup>8</sup>, Tadashi Hasegawa<sup>5</sup>, Takashi Tokino<sup>6</sup>, Koichi Hirata<sup>2</sup>, Kohzoh Imai<sup>14</sup>, Minoru Toyota<sup>3</sup>, and Yasuhisa Shinomura<sup>1</sup>

### Abstract

Large intergenic noncoding RNAs (lincRNA) have been less studied than miRNAs in cancer, although both offer considerable theranostic potential. In this study, we identified frequent upregulation of miR-196a and lincRNA *HOTAIR* in high-risk gastrointestinal stromal tumors (GIST). Overexpression of miR-196a was associated with high-risk grade, metastasis and poor survival among GIST specimens. miR-196a genes are located within the *HOX* gene clusters and microarray expression analysis revealed that the *HOXC* and *HOTAIR* gene were also coordinately upregulated in GISTs which overexpress miR-196a. In like manner, overexpression of *HOTAIR* was also strongly associated with high-risk grade and metastasis among GIST specimens. RNA interference-mediated knockdown of *HOTAIR* altered the expression of reported *HOTAIR* target genes and suppressed GIST cell invasiveness. These findings reveal concurrent overexpression of *HOX* genes with noncoding RNAs in human cancer in this setting, revealing miR-196a and *HOTAIR* as potentially useful biomarkers and therapeutic targets in malignant GISTs. *Cancer Res*; 72(5); 1126–36. ©2012 AACR.

### Introduction

Gastrointestinal stromal tumors (GIST) are the most common mesenchymal tumors of the gastrointestinal tract (1–3). GISTs arise predominantly in the stomach (60%) and small intestine (25%) but also occur in colon and rectum (5%), esophagus (2%), and other organs (3). Immunohistochemically, GISTs are positive for KIT and CD34 and are negative or

variably positive for other neural and smooth muscle cell markers. The expression of KIT and CD34 is a characteristic feature of the intestinal cells of Cajal (ICC), which are located in the intestinal wall and regulate gastrointestinal motility. GISTs are thus thought to originate from ICCs or ICC precursors. Activating *KIT* mutations have been identified in 80% to 90% of GISTs, and mutation of the platelet-derived growth factor receptor alpha gene (*PDGFRA*) is observed in approximately 5% of GISTs (1–3). In that context, imatinib mesylate (formerly STI571) was developed as a tyrosine kinase inhibitor and has been shown to inhibit the activities of BCR-ABL, KIT, and PDGFR. Imatinib mesylate is currently being used for the treatment of both chronic myeloid leukemia and metastatic GISTs.

Predicting the biologic potential of GISTs is often difficult, and considerable effort has been made to define the variables that could enable more accurate identification of tumors with malignant potential. In most classification systems, the key prognostic factors for estimating malignant potential are tumor size and mitotic rate, and to a more variable degree, the proliferation index or tumor site (4). Other potential and promising markers of GIST malignancy are molecular alterations. As mentioned, a large majority of GISTs exhibit activating *KIT* or *PDGFRA* mutations. By itself, however, mutation status does not fully explain the diverse biology of GISTs, and it is believed that additional molecular alterations are required for the progression of high-risk GISTs. For instance, expression of CD26 (encoded by *DPP4*) is strongly associated with poor survival among patients with gastric GISTs, suggesting its involvement in the malignant progression of the disease (5).

**Authors' Affiliations:** First Departments of <sup>1</sup>Internal Medicine and <sup>2</sup>Surgery, Departments of <sup>3</sup>Molecular Biology, <sup>4</sup>Public Health, <sup>5</sup>Surgical Pathology, <sup>6</sup>Medical Genome Science, Research Institute for Frontier Medicine, Sapporo Medical University School of Medicine; Departments of <sup>7</sup>Gastroenterology and <sup>8</sup>Surgery, Keiyukai Sapporo Hospital, Sapporo; <sup>9</sup>Department of Surgery, Osaka University Graduate School of Medicine; <sup>10</sup>Department of Surgery, Osaka Police Hospital, Osaka; Divisions of <sup>11</sup>Digestive and General Surgery and <sup>12</sup>Molecular and Diagnostic Pathology, Niigata University Graduate School of Medical and Dental Sciences, Niigata; <sup>13</sup>Division of Human Health and Medical Science, Graduate School of Kuroshio Science, Kochi University, Nankoku; and <sup>14</sup>Division of Novel Therapy for Cancer, The Advanced Clinical Research Center, The Institute of Medical Science, The University of Tokyo, Tokyo, Japan

**Note:** Supplementary data for this article are available at Cancer Research Online (<http://cancerres.aacrjournals.org/>).

T. Niinuma and H. Suzuki contributed equally to this work.

**Corresponding Authors:** Hiromu Suzuki, Department of Molecular Biology, Sapporo Medical University; S1, W17, Chuo-Ku, Sapporo 060-8556, Japan. Phone: 81-11-611-2111; Fax: 81-11-622-1918; E-mail: [hsuzuki@sapmed.ac.jp](mailto:hsuzuki@sapmed.ac.jp); and Yasuhisa Shinomura, First Department of Internal Medicine, Sapporo Medical University; S1, W16, Chuo-Ku, Sapporo 060-8543, Japan. Fax: 81-11-611-2282; E-mail: [shinomura@sapmed.ac.jp](mailto:shinomura@sapmed.ac.jp)

doi: 10.1158/0008-5472.CAN-11-1803

©2012 American Association for Cancer Research.

In addition, we recently showed that hypomethylation of repetitive DNA elements is predominantly observed in malignant GISTs, and that global hypomethylation correlates with increased chromosomal aberration (6).

miRNAs are a class of small noncoding RNAs that regulate gene expression by inducing translational inhibition or direct degradation of target mRNAs through base pairing to partially complementary sites (7). miRNAs are highly conserved among species and play critical roles in a variety of biologic processes, including development, differentiation, cell proliferation, and apoptosis. Consistent with their role in these processes, a number of studies have shown widespread alteration of miRNA expression patterns in cancer (8, 9). It has also been shown that in cancer global miRNA expression profiles, as well as expression of specific miRNAs, correlate with disease prognosis and clinical outcome (10). To date, however, only a few groups have studied miRNA expression in GISTs (11, 12), and no specific miRNAs that could serve as prognostic markers have yet been identified.

In this study, we investigated the global pattern of miRNA expression in GISTs. Our aim was to evaluate the contribution made by miRNAs to the malignant potential of GISTs and to identify predictive biomarkers. We determined that upregulation of miR-196a is strongly associated with high risk and poor prognosis in GIST patients. Furthermore, we provide evidence that overexpression of miR-196a is accompanied by upregulation of *HOXC* cluster genes and a metastasis-associated noncoding RNA in GISTs.

## Materials and Methods

### Tumor samples

A total of 56 fresh frozen GIST specimens were obtained from Sapporo Medical University Hospital, Keiyukai Sapporo Hospital, and Osaka University Hospital, as described (6). In addition, formalin-fixed paraffin-embedded (FFPE) tissue sections of 100 GIST specimens were obtained from Niigata University Hospital. Informed consent was obtained from all patients before collection of the specimens, and this study was approved by the respective Institutional Review Boards. Risk grade was assessed according to the risk definition system proposed by Fletcher and colleagues (4). Tumors that were less than 2 cm in diameter with a mitotic count of less than 5/50 high-power fields (HPF) were categorized as very low risk. Tumors that were 2 to 5 cm in diameter with a mitotic count of less than 5/50 HPF were considered to be low risk. Tumors that were less than 5 cm in diameter with a mitotic count of 6 to 10/50 HPF, or were 5 to 10 cm with a mitotic count of less than 5/50 HPF were considered to be intermediate risk. Tumors that were more than 5 cm in diameter with a mitotic count of more than 5/50 HPF, more than 10 cm in diameter with any mitotic count, or any size with a mitotic count of more than 10/50 HPF were considered to be high risk. Total RNA was extracted from fresh frozen tissue specimens using a mirVana miRNA Isolation Kit (Ambion). Total RNA was extracted from FFPE tissue specimens using a RecoverAll Total Nucleic Acid Isolation Kit for FFPE (Ambion). Tumor tissues were reviewed by

pathologists and were macrodissected; laser capture microdissection was not carried out in this study.

### miRNA microarray analysis

One-color microarray-based miRNA expression analysis was carried out according to the manufacturer's instructions (Agilent Technologies). Briefly, 100 ng of total RNA from fresh frozen GIST tissues was labeled using miRNA Labeling Reagent (Agilent Technologies), after which the labeled RNA was hybridized to a Human miRNA Microarray V3 (Rel 12.0, G4470C; Agilent Technologies), which covers 859 human miRNAs and 80 viral miRNAs. The microarray data were analyzed using GeneSpring GX version 11 (Agilent Technologies). The normalized microarray data were then compared with the TaqMan assay results using GraphPad PRISM version 5 (GraphPad Software Inc.). The Gene Expression Omnibus accession number for the miRNA microarray data is GSE31741.

### Quantitative RT-PCR of miRNA

miR-196a expression was analyzed using TaqMan microRNA Assays (Applied Biosystems). Briefly, 5 ng of total RNA were reverse transcribed using specific stem-loop RT primers, after which they were amplified and detected using PCR with specific primers and TaqMan probes. The PCR was run in triplicate using a 7500 Fast Real-Time PCR System (Applied Biosystems), and SDS v1.4 software (Applied Biosystems) was used for comparative  $\Delta C_t$  analysis. U6 snRNA (RNU6B; Applied Biosystems) served as an endogenous control.

### Gene expression microarray analysis

One-color microarray-based gene expression analysis was carried out according to the manufacturer's instructions (Agilent Technologies). Briefly, 700 ng of total RNA were amplified and labeled using a Quick Amp Labeling Kit One-Color (Agilent Technologies), after which the synthesized cRNA was hybridized to the Whole Human Genome Oligo DNA microarray, which includes 41,000 probe sets covering 19,416 genes (G4112F; Agilent Technologies). The microarray data were analyzed using GeneSpring GX version 11 (Agilent Technologies). The Gene Expression Omnibus accession numbers for the microarray data are GSE31802 and GSE32064.

### Quantitative RT-PCR of *HOTAIR*

Single-stranded cDNA was prepared using SuperScript III reverse transcriptase (Invitrogen). Quantitative reverse transcriptase PCR (RT-PCR) of *HOTAIR* was carried out using a TaqMan Gene Expression Assay (Assay ID, Hs03296631\_m1; Applied Biosystems) and a 7500 Fast Real-Time PCR System (Applied Biosystems). *GAPDH* (Assay ID, Hs99999905\_m1; Applied Biosystems) served as an endogenous control.

### DNA copy number and chromatin signature analysis

DNA copy number was analyzed using array-based comparative genome hybridization (CGH) as described previously (6). Trimethylated Histone H3 lysine 4 (H3K4me3) was analyzed using chromatin immunoprecipitation (ChIP) as described previously (13, 14). Details of the experimental procedures are provided in the Supplementary Methods.

### Transfection of miRNA inhibitors and siRNA molecules

GIST-T1 cells were described previously (15). For inhibition of miR-196a, cells ( $3 \times 10^5$  cells in 6-well plates) were transfected with 100 pmol of Anti-miR miRNA Inhibitors (Ambion) or Anti-miR miRNA Inhibitors Negative Control #1 (Ambion) using Lipofectamine2000 (Invitrogen). For RNA interference (RNAi)-mediated knockdown of *HOTAIR*, 3 different Stealth siRNAs against *HOTAIR* were generated by Invitrogen, after which a mixture of the 3 was used for transfection. Cells ( $3 \times 10^5$  cells in 6-well plates) were transfected with 100 pmol of siRNA or with Stealth RNAi Negative Control Medium GC (Invitrogen) using Lipofectamine2000 (Invitrogen). Total RNA extraction, cell viability assays, and Matrigel invasion assays were carried out 48 hours after transfection as described in the Supplementary Methods.

### Statistical analysis

All gene expression levels were log transformed for subsequent statistical analysis because the distribution of expression data seemed to follow a log-normal distribution. Geometric means were therefore calculated as summary statistics for expression levels. Comparisons of continuous variables were made using *t* tests or one-way ANOVA with post hoc multiple comparisons (Games–Howell test). Pearson's correlation coefficients were calculated to describe the strength of the correlation between 2 variables. Comparisons of categorical variables were made using Fisher exact test. To assess the association between prognostic factors and gene expression levels, logistic or Cox regression analyses were carried out. For these regression analyses, the most optimal cutoff points were employed to calculate ORs and HRs, with or without adjustment for clinical factors. Kaplan–Meier curves were plotted to compare 2 groups stratified by gene expression status. All statistical analyses were done using SPSS Statistics 18 (IBM Corporation).

## Results

### Detection of upregulated miR-196a expression in high-risk GISTs

To examine the miRNA expression signature in GISTs, we carried out miRNA microarray analysis with 32 fresh frozen GIST specimens (10 low-risk, 8 intermediate-risk, and 14 high-risk GISTs). The clinicopathologic features of the 32 patients are listed in Supplementary Table S1. Of 939 probe sets, 470 were excluded because of the absence of a detectable signal in any of the samples tested. Unsupervised hierarchical clustering using the remaining 469 probe sets revealed that GISTs in which there was abundant expression of miRNAs encoded on chromosome 14q32.31 form a separate cluster (Supplementary Figs. S1 and S2). Moreover, by comparing the miRNA expression profiles with array CGH results, we found that this cluster is enriched in tumors without 14q loss. These results are consistent with recent reports showing an inverse relationship between 14q loss and expression of miRNAs located on 14q in GISTs (11, 12), which is indicative of the reliability of our microarray analysis. We next carried out a scatter plot analysis and found that miR-196a is markedly upregulated in high-risk

GISTs, as compared with low- or intermediate-risk GISTs (Fig. 1A). As shown in Fig. 1B, miR-196a was undetectable in all but one of the low- and intermediate-risk GISTs tested, whereas it was upregulated in more than half of the high-risk tumors. The elevated expression of miR-196a was observed in both gastric and small intestinal GISTs (Supplementary Table S1).

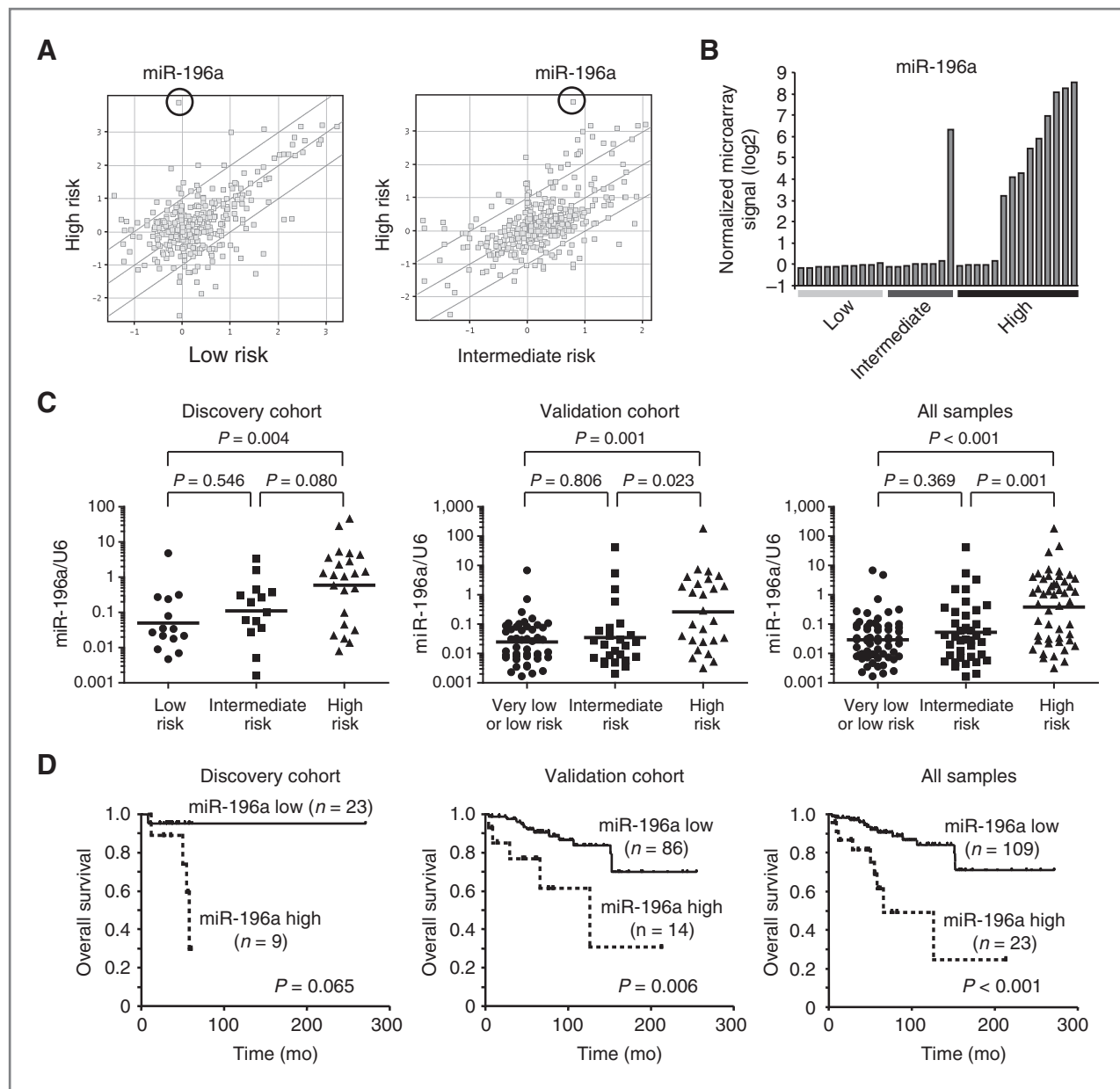
### Upregulation of miR-196a is associated with GIST malignancy

To assess the clinical importance of miR-196a upregulation in GISTs, we next carried out TaqMan assay with 56 fresh frozen GIST specimens (discovery cohort), including the 32 specimens initially analyzed by microarray. The clinicopathologic features of the patients are summarized in Table 1. The TaqMan assay results were highly consistent with the microarray data, though the TaqMan assay did reveal low levels of miR-196a expression in samples in which there was no detectable signal from the microarray (Supplementary Fig. S3).

Also consistent with the microarray results was the finding that miR-196a was markedly upregulated in high-risk GISTs, as compared with the other groups ( $P = 0.004$ , one-way ANOVA; Fig. 1C, Supplementary Table S2). In addition, logistic regression analysis revealed that the association between miR-196a upregulation and the high-risk category was highest when we employed a cutoff value of miR-196a/U6 0.4 or more (OR = 13.7; 95% CI: 3.4–54.6;  $P < 0.001$ ; Supplementary Table S3). Survival data were obtained for 32 patients, and Cox hazard analysis revealed the highest HR for patients with elevated miR-196a expression when a cutoff value of 1.4 was employed (Table 2). Kaplan–Meier analysis showed poor overall survival among patients with GISTs expressing high levels of miR-196a, though the effect was not statistically significant (Fig. 1D).

We next used TaqMan assay to analyze an independent validation cohort consisting of 100 FFPE GIST specimens (Table 1). Consistent with the findings summarized above, we observed that upregulation of miR-196a was associated with high-risk GISTs (Fig. 1C, Supplementary Tables S2 and S3). By using the same cutoff value (miR-196a/U6  $\geq 1.4$ ), Cox hazard analysis revealed an elevated HR for patients exhibiting high levels of miR-196a expression (Table 2), and Kaplan–Meier analysis showed shorter survival times for the same patients (Fig. 1D). These results confirmed the prognostic value of miR-196a expression in both fresh frozen and FFPE GIST specimens.

Finally, we combined the GIST samples in the discovery and validation cohorts to examine the clinicopathologic significance of miR-196a. Expression of miR-196a correlated positively with high-risk grade (Fig. 1C, Supplementary Tables S2 and S3), poor clinical outcome (Fig. 1D, Table 2), tumor size, mitotic count, and metastasis (Table 3). Interestingly, although expression of miR-196a was not associated with age or gender, it was strongly associated with tumor location (Table 3). The median level of miR-196a expression was lowest in specimens from esophageal GISTs and then increased as the GIST site moved from the oral side toward the anal side of the gastrointestinal tract ( $P < 0.001$ ; Table 3, Supplementary Fig. S4). Importantly, although the average level of miR-196a expression was higher in small intestine than in stomach, it was positively



**Figure 1.** Upregulation of miR-196a expression in GISTs is associated with a high-risk grade and poor prognosis. **A**, scatter plots analyses: plotting low-risk ( $n = 10$ ) versus high-risk GISTs ( $n = 14$ ; left) and intermediate-risk ( $n = 8$ ) versus high-risk GISTs ( $n = 14$ ; right) revealed overexpression of miR-196a in high-risk GISTs. Microarray data are normalized and log transformed (base 2). Expression of miR-196a is highlighted by a circle. **B**, levels of miR-196a expression obtained from microarray analysis of 32 GIST specimens. Risk categories are indicated below. **C**, comparison of miR-196a expression using TaqMan assay in low- ( $n = 14$ ), intermediate- ( $n = 14$ ), and high-risk GISTs ( $n = 23$ ) in a discovery cohort (left), very low- or low-risk ( $n = 46$ ), intermediate-risk ( $n = 25$ ), and high-risk GISTs ( $n = 26$ ) in a validation cohort (middle) and all GIST specimens (right). Results are normalized to internal U6 snRNA. **D**, Kaplan-Meier curves showing the effect of miR-196a expression (high, miR-196a/U6  $\geq 1.4$ ; low, miR-196a/U6  $< 1.4$ ) on overall survival in the discovery cohort (left), validation cohort (middle), and all GIST patients (right).

correlated with high-risk grade in both organs (Supplementary Table S4).

#### Concurrent upregulation of miR-196a and HOX cluster genes in GISTs

To analyze the relationship between miR-196a upregulation and the global gene expression profiles in GISTs, we selected

age-, gender-, and tumor location-matched GIST specimens showing either low ( $n = 7$ ; average miR-196a/U6 = 0.1) or high miR-196a expression ( $n = 7$ ; average miR-196a/U6 = 15.7) and subjected them to gene expression microarray analysis (Supplementary Table S5). We found that for 4,947 probe sets (corresponding to 3,206 unique genes), there was more than a 2-fold difference in expression between GISTs with miR-196a



**Table 1.** Clinical features of the GIST samples used in this study

Discovery cohort	
Age (y, median $\pm$ SD)	68.0 $\pm$ 15.2
Gender	
Male	32 (57.1%)
Female	24 (42.9%)
Tumor location	
Stomach	40 (71.4%)
Small intestine	14 (25.0%)
Omentum	1 (1.8%)
Colorectum	1 (1.8%)
Risk category (n = 51)	
Low	14 (27.5%)
Intermediate	14 (27.5%)
High	23 (45.0%)
Validation cohort	
Age (y, median $\pm$ SD)	64.0 $\pm$ 12.4
Gender	
Male	44 (44.0%)
Female	56 (56.0%)
Tumor location	
Esophagus	5 (5.2%)
Stomach	84 (84.0%)
Small intestine	8 (8.0%)
Colorectum	3 (3.0%)
Risk category (n = 97)	
Very low	1 (1.0%)
Low	45 (46.4%)
Intermediate	25 (25.8%)
High	26 (26.8%)

overexpression and those without it. Hierarchical clustering analysis using the 4,947 probe sets clearly distinguished between tumors on the basis of their miR-196a expression status (Fig. 2A), and Gene Ontology analysis suggested that genes related to "immune system," "plasma membrane," and "cell communication" are strongly overrepresented among the differentially expressed genes (Supplementary Table S6).

To further characterize the differentially expressed genes, we carried out a gene set analysis and obtained the highest enrichment score with the *HOX* gene set (Supplementary Table S7). We found miR-196a to be encoded at 2 paralogous loci, *miR-196a-1* and *miR-196a-2*, which are located within the *HOXB* and *HOXC* clusters, respectively (Fig. 2B; ref. 16). Hierarchical clustering analysis using the expression data for *HOXC* genes clearly differentiated the GIST samples into 2 groups, and we observed perfect correspondence between higher expression of multiple *HOXC* genes and upregulation of miR-196a (Fig. 2C). By contrast, genes in other *HOX* clusters did not show such obvious correlations with miR-196a (Fig. 2C, Supplementary Fig. S5). We next compared the microarray signal for each *HOX* gene with the miR-196a expression and found strong positive correlations between the expression levels of a number of *HOXC* genes and those of miR-196a (Fig. 3D, Supplementary Fig. S6). Notably, we also found that expression of *HOTAIR*, which encodes a large intergenic non-coding RNA (lincRNA) and is located in an antisense orientation relative to the *HOXC* genes, is concurrently upregulated with miR-196a (Fig. 2C and D). Levels of miR-196a expression also correlated moderately with those of the *HOXB* genes neighboring *miR-196a-1* (*HOXB13* and *HOXB9*), but the correlations were less significant than those with *HOXC* genes (Supplementary Fig. S7).

The similarity between the expression patterns of *HOXC* genes and those of the noncoding RNAs encoded in the same locus is indicative of a common regulatory mechanism involved in the activation of these genes in GISTs. However, array CGH analysis of 27 GIST specimens failed to detect either gain or loss in any *HOX* loci, irrespective of *miR-196a* or *HOX* gene expression, which makes it unlikely that genomic amplification is the cause of their overexpression (Supplementary Fig. S8).

#### Upregulation of *HOTAIR* is associated with GIST malignancy

A recent study showed that *HOTAIR* is overexpressed in primary breast cancer and is associated with metastasis (17). To examine its clinical significance in GISTs, we carried out TaqMan assays of *HOTAIR* with the discovery cohort samples. We found that the microarray signals for *HOTAIR* were highly

**Table 2.** miR-196a expression is associated with poor clinical outcome in GIST patients

	Outcome						
	miR-196a/U6	Survival	Death	HR (95% CI)	P	HR <sup>a</sup> (95% CI)	P
Discovery cohort	<1.4	22	1				
	$\geq$ 1.4	5	4	6.3 (0.7–57.5)	0.104	32.9 (2.0–551.3)	0.015
Validation cohort	<1.4	73	13				
	$\geq$ 1.4	9	5	3.9 (1.4–11.1)	0.011	8.4 (2.6–26.9)	<0.001
All samples	<1.4	95	14				
	$\geq$ 1.4	14	9	4.9 (2.1–11.7)	<0.001	9.1 (3.5–23.7)	<0.001

<sup>a</sup>Age and gender adjusted HR.

**Table 3.** Correlation between miR-196a expression and clinicopathologic features of GISTs

	<i>n</i>	miR-196a/U6		<i>P</i>			
		Geometric mean	95% CI				
Age (y)							
<65	76	0.093	0.051–0.168	0.581			
≥65	81	0.074	0.044–0.127				
Gender							
M	76	0.104	0.059–0.186	0.297			
F	80	0.069	0.040–0.119				
Location							
Esophagus	5	0.019	0.001–0.417	Ref			
Stomach	124	0.061	0.039–0.094	0.741	Ref		
Small intestine	22	0.395	0.163–0.957	0.161	0.002	Ref	
Colorectum	4	4.936	2.564–9.502	0.023	<0.001	<0.001	Ref
				<0.001			
Tumor size (cm)							
≤5.0	70	0.045	0.026–0.075	0.016			
>5.0	81	0.118	0.066–0.210				
Mitotic count (/50 HPF)							
≤5	105	0.036	0.025–0.053	<0.001			
>5	35	0.539	0.215–1.353				
Metastasis							
Yes	28	0.747	0.307–1.819	<0.001			
No	108	0.041	0.027–0.063				

consistent with the TaqMan assay results (Supplementary Fig. S9). *HOTAIR* was upregulated exclusively in high-risk GISTs, as compared with low- or intermediate-risk GISTs ( $P = 0.018$ ; Fig. 3A), and its expression correlated positively with the expression of miR-196a (Fig. 3B) and *HOXC* genes (Fig. 3C, Supplementary Fig. S10). In addition, logistic regression analysis revealed that high levels of *HOTAIR* expression in GISTs ( $HOTAIR/GAPDH \geq 0.0002$ ) were strongly associated with metastasis (age and gender adjusted OR = 8.2; 95% CI: 1.4–48.4;  $P = 0.021$ ). Cox hazard analysis suggested an elevated HR for patients with high *HOTAIR* expression (Table 4), and Kaplan–Meier analysis showed poor overall survival for the same patients, though the effect was not statistically significant (Fig. 3D). We also tried to analyze *HOTAIR* expression in the FFPE specimens; however, we failed to detect expression of either *HOTAIR* or *GAPDH* in these samples, most likely due to an inadequate quality of the RNA.

#### Reduced expression of miR-196a and *HOTAIR* target genes in GISTs

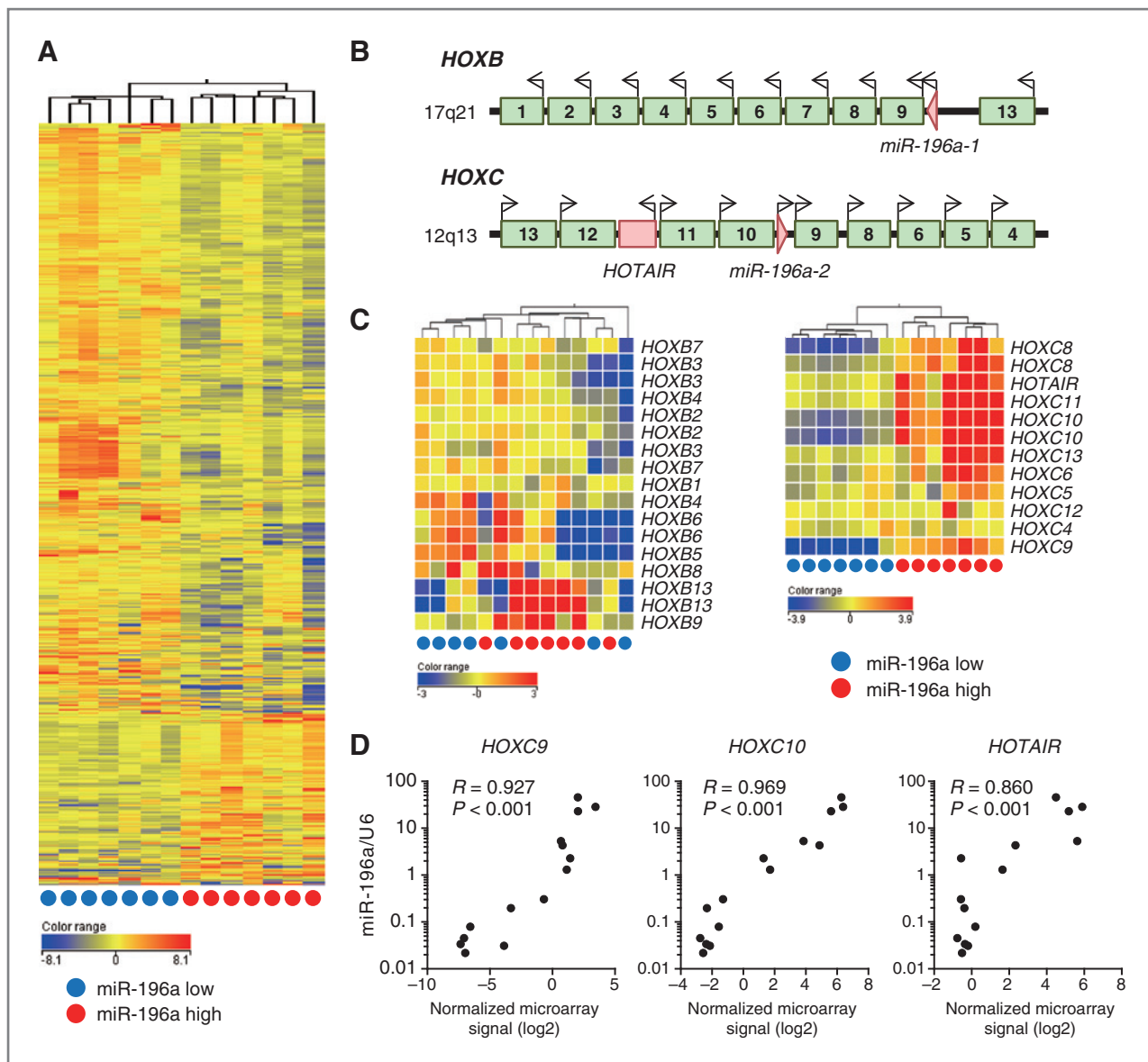
To examine the functional role of miR-196a in GISTs, we interrogated our gene expression microarray data for miR-196a target genes computationally predicted by TargetScan. Of the 2,248 genes whose expression was reduced in GISTs overexpressing miR-196a, 95 corresponded to predicted targets (Supplementary Fig. S11, Supplementary Table S8). This gene list included *ANXA1* (Annexin A1), which is an experimentally validated miR-196a target gene (18). In addition, expression of

several *HOX* genes, including *HOXB8*, was reduced in GISTs overexpressing miR-196a, which is consistent with an earlier finding of miR-196a-directed cleavage of *HOXB8* mRNA (Supplementary Fig. S11; ref. 19).

In normal human fibroblasts, *HOTAIR* represses *HOXD* gene expression by interacting with polycomb repressive complex 2 (PRC2; ref. 20). In breast cancer cells, overexpression of *HOTAIR* was shown to recruit PRC2 to more than 800 gene promoters, leading to histone H3K27 methylation and epigenetic silencing of the target genes (17). We therefore examined our microarray data for the reported *HOTAIR*-induced PRC2 target genes. Among 14 GISTs analyzed with the microarray, all 7 tumors strongly expressing miR-196a showed elevated *HOTAIR* expression (average  $HOTAIR/GAPDH = 0.00254$ ), whereas all tumors only weakly expressing miR-196a showed little or no *HOTAIR* expression (average  $HOTAIR/GAPDH = 0.00001$ ). We found that expression of 144 *HOTAIR* target genes was reduced in GISTs overexpressing *HOTAIR* (Supplementary Fig. S11, Supplementary Table S9). These results indicated that overexpression of miR-196a and *HOTAIR* may contribute to the malignant progression of GISTs by modulating expression of their target genes.

#### Inhibition of miR-196a and *HOTAIR* suppresses GIST cell invasion

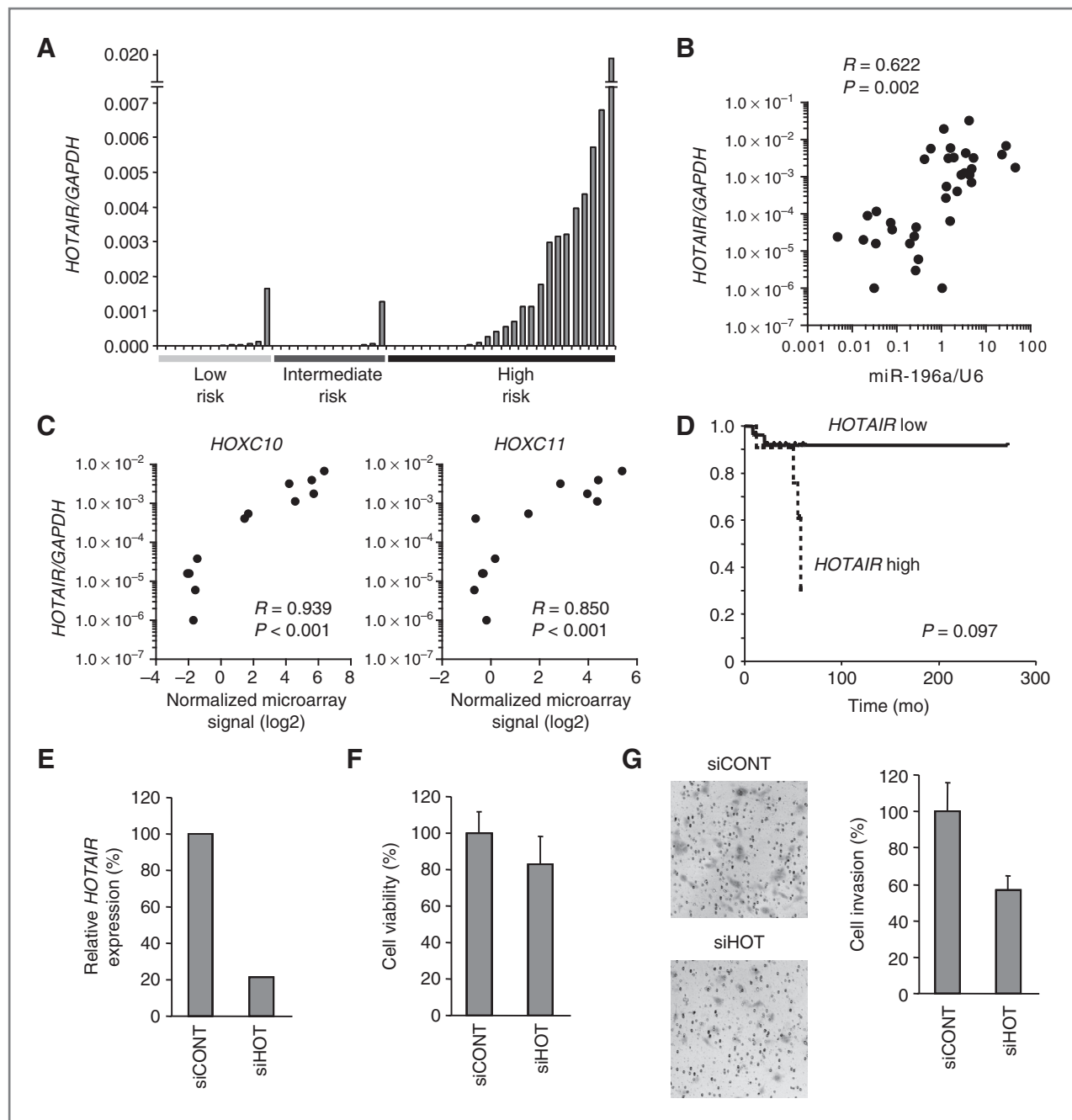
We next utilized a cultured GIST cell line to determine whether upregulation of miR-196a and *HOTAIR* is responsible for the malignant potential of GISTs. We found that both



**Figure 2.** GIST gene expression signatures reveal a strong correlation between the *miR-196a* and *HOXC* genes. **A**, heat map of the gene expression signatures correlated with *miR-196a* expression. Rows represent probe sets and columns represent patients. A total of 4,947 probe sets differentially expressed (>2-fold change) between GISTs with ( $n = 7$ ) and without ( $n = 7$ ) *miR-196a* overexpression were selected, after which hierarchical clustering was carried out. The *miR-196a* expression status is indicated below. **B**, schematic representations of the *miR-196a* family locations within the *HOX* gene clusters. **C**, hierarchical clustering analysis using *HOXB* (left) and *HOXC* (right) expression data. *miR-196a* expression status is indicated below. **D**, correlations between the expression levels of *miR-196a* and *HOXC* genes or *HOTAIR*. Expression of *miR-196a* was analyzed using TaqMan assay and was normalized to internal U6 snRNA. Microarray data for *HOXC* and *HOTAIR* were normalized and log transformed (base 2). The Pearson correlation coefficients and  $P$  values are shown.

*miR-196a* and *HOTAIR* are expressed in GIST-T1 cells (Supplementary Fig. S12). We then carried out cell viability and Matrigel invasion assays after transfecting GIST-T1 cells with an anti-*miR-196a* inhibitor molecule. Gene expression microarray analysis revealed that a number of predicted *miR-196a* target genes, including *ANXA1* and *HOXA5*, were upregulated by inhibition of *miR-196a* (Supplementary Table S10), and although we observed no effects on cell viability, inhibition of *miR-196a* moderately suppressed cell invasion (Supplementary Fig. S13). We next disrupted *HOTAIR* expression by

transfecting the cells with siRNAs targeting it (Fig. 3E). Although knockdown of *HOTAIR* did not significantly affect cell viability, it suppressed the invasiveness of GIST-T1 cells (Fig. 3E and F). Moreover, gene expression microarray analysis revealed that a number of reported *HOTAIR* target genes, including *PCHD10*, *SEMA6A*, and *STK17B*, were upregulated upon knockdown of *HOTAIR* (Supplementary Table S11). In all, we found that 1,424 genes were upregulated by siHOT (>2-fold), and Gene Ontology analysis revealed enrichment of genes related to "nucleus," "chromosome," and "membrane-bounded



**Figure 3.** Upregulation of *HOTAIR* in malignant GISTs. **A**, TaqMan assay of *HOTAIR* in a panel of GIST specimens ( $n = 52$ ). Results are normalized to internal GAPDH expression. Risk categories are indicated below. **B**, correlation between levels of *HOTAIR* and miR-196a expression detected using TaqMan assay. The Pearson correlation coefficient and  $P$  value are shown. **C**, correlations between levels of *HOTAIR* expression detected using TaqMan assay and those of *HOXC* genes detected from microarrays. The Pearson correlation coefficients and  $P$  values are shown. **D**, Kaplan-Meier curves showing the effect of *HOTAIR* expression (high, *HOTAIR*/GAPDH  $\geq 0.0002$ ; low, *HOTAIR*/GAPDH  $< 0.0002$ ) on overall survival among GIST patients. **E**, TaqMan assay for *HOTAIR* in GIST-T1 cells transfected with control siRNA (siCONT) or siRNA targeting *HOTAIR* (siHOT). **F**, cell viability assay using GIST-T1 cells transfected with siCONT or siHOT. Cell viabilities were determined 48 hours after transfection. Shown are the means of 8 replications; error bars represent SDs. **G**, Matrigel invasion assay using GIST-T1 cells transfected with siCONT or siHOT. Shown on the right are the means of 3 random microscopic fields per membrane; error bars represent SDs.

organelle" (Supplementary Tables S12 and S13). These results suggested that *HOTAIR* may modulate transcription of a large number of genes and may have previously unidentified roles in GIST cells.

Finally, we sought to clarify the biologic relationship between miR-196a, *HOTAIR*, and *HOXC* genes. We first tested whether upregulation of miR-196a is a downstream effect of *HOTAIR* dysregulation, or vice versa. We found that inhibition



**Table 4.** HOTAIR expression is associated with poor clinical outcome in GIST patients

HOTAIR/GAPDH	Outcome					
	Survival	Death	HR (95% CI)	P	HR <sup>a</sup> (95% CI)	P
<0.0002	26	2				
≥0.0002	7	4	3.8 (0.7–21.2)	0.123	9.0 (1.2–68.9)	0.034

<sup>a</sup>Age and gender adjusted HR.

of miR-196a had no effect on *HOTAIR* expression in GIST-T1 cells, nor did knockdown of *HOTAIR* affect miR-196a expression. This suggested that overexpression of miR-196a or *HOTAIR* is not a simple downstream effect of their dysregulation (Supplementary Fig. S12). By contrast, analysis of the chromatin status in GIST-T1 cells using ChIP-PCR revealed enrichment of trimethylated histone H3 lysine 4 (H3K4me3), a hallmark of active gene transcription, at the transcription start sites of multiple *HOXC* genes and *HOTAIR* (Supplementary Fig. S14). Moreover, we found concurrent overexpression of miR-196a, *HOTAIR*, and *HOXC* genes in other cancer cells, including the KatoIII gastric cancer cell line. By carrying out high-resolution ChIP-seq analysis with the KatoIII cells, we observed significant enrichment of H3K4me3 over a wide range (more than 50 kb) of the *HOXC* cluster, which suggested that an epigenetic mechanism is involved in the dysregulation of this genomic region (Supplementary Fig. S15).

## Discussion

Although the results of recent studies suggest that the gene expression signatures of GISTs are predictive of malignancy and drug sensitivity of the tumors (5, 21), the clinical significance of the miRNA expression signature is not yet fully understood. In this study, we found that upregulation of miR-196a is strongly associated with a high-risk grade, metastasis, and poor prognosis in GIST patients. Furthermore, overexpression of miR-196a is accompanied by upregulation of multiple *HOXC* genes and the metastasis-related lincRNA *HOTAIR*. To our knowledge, this is the first article to show concurrent overexpression of collinear *HOX* genes and non-coding RNAs in human malignancy.

A number of studies have implicated miR-196a in malignancy, but its role may differ among tumor types. Upregulation of miR-196a is observed in esophageal adenocarcinomas and their precancerous lesions, Barrett's esophagus and dysplasia, which suggests miR-196a is a potential marker of the malignant progression of Barrett's esophagus (22). Strong expression of miR-196a is also associated with a poor prognosis in pancreatic adenocarcinoma and glioblastoma patients (23, 24). In addition, functional analysis showed that expression of miR-196a in esophageal, breast, and endometrial cancer cells promotes proliferation and suppresses apoptosis through downregulation of *ANXA1* (18). These results suggest that miR-196a contributes to oncogenesis in cancer. On the other hand, miR-196a is significantly downregulated in melanoma, and its

reexpression inhibited the invasive behavior of melanoma cells by targeting *HOXC8* (25). Similarly, miR-196a suppressed *HOXC8* and inhibited invasion and metastasis by breast cancer cells (26). Thus miR-196a seems to exert opposite effects in tumors of different origins.

The *HOX* genes are a highly conserved subgroup of the homeobox superfamily, and they play essential roles in a variety of biologic processes, including development, differentiation, apoptosis, and angiogenesis (27). In humans, 4 *HOX* clusters containing 39 *HOX* genes have been identified, and dysregulation of their expression is observed in various malignancies. Although the role of *HOX*s in cancer is not fully understood, its aberrant expression is thought to affect pathways that promote tumorigenesis and metastasis (27). For instance, *HOXC8* mRNA is overexpressed in prostate cancer cells and is associated with tumor cell proliferation and metastasis (28–30). In addition, *HOXC5* and *HOXC8* mRNAs are upregulated in cervical cancer cells (31), and one recent study suggested *HOXC10* plays a key role of in the progression and invasion in cervical cancer (32).

An association between miR-196a and *HOX* expression in cancer has also been reported. Reduced expression of miR-196a in malignant melanoma cells leads to upregulation of *HOXB7* and, in turn, activation of *BMP4*, a major modulator of migration (33). As mentioned above, miR-196a also inhibits invasion and metastasis by downregulating *HOXC8* in melanoma and breast cancer cells (26, 34). Taken together, these results suggest that miR-196a acts as a tumor suppressor by targeting *HOX* genes in these tumor types. By contrast, we show in this study that both the miR-196a and *HOXC* genes are concurrently upregulated in malignant GISTs. Our findings are reminiscent of an earlier report showing that the expression patterns of miRNAs embedded in *HOX* clusters are very similar to those of *HOX* genes during mammalian embryogenesis (35). Global gene expression analysis revealed that expression of multiple putative miR-196a targets, including *ANXA1*, is diminished in GISTs overexpressing miR-196a, whereas their expression is enhanced upon inhibition of miR-196a in cultured GIST-T1 cells. In addition, inhibition of miR-196a in GIST cells overexpressing that miRNA moderately suppressed cell invasion. Taken together, our results indicate that upregulation of *HOXC* genes along with miR-196a may contribute to the malignant potential of GIST.

*HOTAIR* is located within the *HOXC* cluster and encodes a lincRNA known to repress its target genes by directly interacting with histone modification complexes. Epigenetic gene

regulation is closely associated with histone modifications in which di- or trimethylation of histone H3 lysine 4 (H3K4me2 or H3K4me3) is enriched within active gene promoters. In addition, trimethylation of histone H3 lysine 27 (H3K27me3) is a marker of gene silencing. In normal adult fibroblasts, *HOTAIR* suppresses the *HOXD* locus by recruiting the PRC2 complex, which consists of the histone H3K27 methylase EZH2, SUZ12, and EED (20). It was also recently shown that *HOTAIR* serves as a scaffold for multiple repressor complexes, including PRC2 and LSD1/CoREST/REST (36). LSD1 is a demethylase that specifically mediates demethylation of H3K4, leading to repression of the target genes. *HOTAIR* is also strongly implicated in cancer metastasis. In breast cancer cells, *HOTAIR* induces retargeting of the PRC2 complex throughout the genome, which leads to the silencing of multiple tumor suppressor and metastasis suppressor genes (17). Overexpression of *HOTAIR* is also predictive of recurrence in hepatocellular carcinoma patients after liver transplantation (37). We observed that upregulation of *HOTAIR* is closely associated with GIST aggressiveness and metastasis. In addition, functional analysis using GIST-T1 cells showed that RNAi-mediated knockdown of *HOTAIR* suppressed cell invasion. These results strongly suggest that upregulation of *HOTAIR* is one of the mechanisms that promote aggressiveness in GISTs. Interestingly, depletion of *HOTAIR* induced a significant change in the gene expression profile in GIST cells, suggesting that *HOTAIR* may regulate a spectrum of genes other than the previously reported target genes. Further studies, including genome-wide histone modification analysis, may reveal as yet unidentified roles played by *HOTAIR* in the malignant progression of GISTs.

The mechanism underlying upregulation of *HOX* cluster genes and noncoding RNAs in GISTs is intriguing. Our array CGH analysis did not detect chromosomal aberrations in any *HOX* loci, making it unlikely that gene amplification is the cause of their overexpression. However, we found that the transcription start sites of multiple genes in the *HOXC* cluster are marked by an active histone mark, H3K4me3, in GIST-T1 cells. Moreover, high-resolution ChIP-seq analysis revealed

that, in cancer cells, the entire region is significantly enriched with H3K4me3, leading to overexpression of the affected genes. Our results are reminiscent of the recent finding that rearrangement of *MLL* in leukemia induces active histone modifications at the promoters of *HOXA* genes and miR-196b, resulting to their overexpression (38–40). Although such rearrangements are not known in GISTs, further study to clarify the involvement of epigenetic modifiers in malignant GISTs may lead to identification of new therapeutic targets.

Overall, our study has shown that noncoding RNAs encoded in the *HOXC* cluster could be useful predictive markers as well as novel therapeutic targets in malignant GISTs. As miRNAs are well preserved in FFPE specimens (41), miR-196a could be a reliable biomarker for risk assessment. We also provide evidence that *HOTAIR* is significantly upregulated in high-risk GISTs, indicating that this lincRNA also could be a useful biomarker, as well as a novel therapeutic target. Further study of the causes and functions of *HOXC* locus activation in GISTs may provide new strategies for the treatment of GIST patients.

#### Disclosure of Potential Conflicts of Interest

T. Nishida has received a research grant from Novartis Pharma K.K. The remaining authors disclose no conflicts of interest.

#### Acknowledgments

The authors thank Dr. William F. Goldman for editing the manuscript and M. Ashida for technical assistance.

#### Grant Support

This study was supported in part by grants-in-aid for Scientific Research (B) from the Japan Society for Promotion of Science (Y. Shinomura), A3 foresight program from the Japan Society for Promotion of Science (H. Suzuki), a grant-in-aid for the Third-term Comprehensive 10-year Strategy for Cancer Control (M. Toyota, H. Suzuki), a grant-in-aid for Cancer Research from the Ministry of Health, Labor, and Welfare, Japan (M. Toyota, H. Suzuki), and the Takeda Science Foundation (H. Suzuki).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received May 31, 2011; revised December 19, 2011; accepted January 6, 2012; published OnlineFirst January 18, 2012.

#### References

- Shinomura Y, Kinoshita K, Tsutsui S, Hirota S. Pathophysiology, diagnosis, and treatment of gastrointestinal stromal tumors. *J Gastroenterol* 2005;40:775–80.
- Rubin BP, Heinrich MC, Corless CL. Gastrointestinal stromal tumour. *Lancet* 2007;369:1731–41.
- Corless CL, Fletcher JA, Heinrich MC. Biology of gastrointestinal stromal tumors. *J Clin Oncol* 2004;22:3813–25.
- Fletcher CD, Berman JJ, Corless C, Gorstein F, Lasota J, Longley BJ, et al. Diagnosis of gastrointestinal stromal tumors: A consensus approach. *Hum Pathol* 2002;33:459–65.
- Yamaguchi U, Nakayama R, Honda K, Ichikawa H, Hasegawa T, Shitashige M, et al. Distinct gene expression-defined classes of gastrointestinal stromal tumor. *J Clin Oncol* 2008;26:4100–8.
- Igarashi S, Suzuki H, Niinuma T, Shimizu H, Nojima M, Iwaki H, et al. A novel correlation between LINE-1 hypomethylation and the malignancy of gastrointestinal stromal tumors. *Clin Cancer Res* 2010;16:5114–23.
- He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet* 2004;5:522–31.
- Esquela-Kerscher A, Slack FJ. Oncomirs—microRNAs with a role in cancer. *Nat Rev Cancer* 2006;6:259–69.
- Croce CM. Causes and consequences of microRNA dysregulation in cancer. *Nat Rev Genet* 2009;10:704–14.
- Calin GA, Croce CM. MicroRNA signatures in human cancers. *Nat Rev Cancer* 2006;6:857–66.
- Choi HJ, Lee H, Kim H, Kwon JE, Kang HJ, You KT, et al. MicroRNA expression profile of gastrointestinal stromal tumors is distinguished by 14q loss and anatomic site. *Int J Cancer* 2010;126:1640–50.
- Haller F, von Heydebreck A, Zhang JD, Gunawan B, Langer C, Ramadori G, et al. Localization- and mutation-dependent microRNA (miRNA) expression signatures in gastrointestinal stromal tumours (GISTs), with a cluster of co-expressed miRNAs located at 14q32.31. *J Pathol* 2010;220:71–86.
- Maruyama R, Choudhury S, Kowalczyk A, Bessarabova M, Beresford-Smith B, Conway T, et al. Epigenetic regulation of cell type-specific expression patterns in the human mammary epithelium. *PLoS Genet* 2011;7:e1001369.
- Suzuki H, Takatsuka S, Akashi H, Yamamoto E, Nojima M, Maruyama R, et al. Genome-wide profiling of chromatin signatures reveals epigenetic regulation of microRNA genes in colorectal cancer. *Cancer Res* 2011;71:5646–58.

15. Taguchi T, Sonobe H, Toyonaga S, Yamasaki I, Shuin T, Takano A, et al. Conventional and molecular cytogenetic characterization of a new human cell line, GIST-T1, established from gastrointestinal stromal tumor. *Lab Invest* 2002;82:663–5.
16. Calin GA, Sevignani C, Dumitru CD, Hyslop T, Noch E, Yendamuri S, et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc Natl Acad Sci U S A* 2004;101:2999–3004.
17. Gupta RA, Shah N, Wang KC, Kim J, Horlings HM, Wong DJ, et al. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. *Nature* 2010;464:1071–6.
18. Luthra R, Singh RR, Luthra MG, Li YX, Hannah C, Romans AM, et al. MicroRNA-196a targets annexin A1: a microRNA-mediated mechanism of annexin A1 downregulation in cancers. *Oncogene* 2008;27:6667–78.
19. Yekta S, Shih IH, Bartel DP. MicroRNA-directed cleavage of HOXB8 mRNA. *Science* 2004;304:594–6.
20. Rinn JL, Kertesz M, Wang JK, Squazzo SL, Xu X, Bruggmann SA, et al. Functional demarcation of active and silent chromatin domains in human HOX loci by noncoding RNAs. *Cell* 2007;129:1311–23.
21. Rink L, Skorobogatko Y, Kossenkova AV, Belinsky MG, Pajak T, Heinrich MC, et al. Gene expression signatures and response to imatinib mesylate in gastrointestinal stromal tumor. *Mol Cancer Ther* 2009;8:2172–82.
22. Maru DM, Singh RR, Hannah C, Albarracín CT, Li YX, Abraham R, et al. MicroRNA-196a is a potential marker of progression during Barrett's metaplasia-dysplasia-invasive adenocarcinoma sequence in esophagus. *Am J Pathol* 2009;174:1940–8.
23. Bloomston M, Frankel WL, Petrocca F, Volinia S, Alder H, Hagan JP, et al. MicroRNA expression patterns to differentiate pancreatic adenocarcinoma from normal pancreas and chronic pancreatitis. *JAMA* 2007;297:1901–8.
24. Guan Y, Mizoguchi M, Yoshimoto K, Hata N, Shono T, Suzuki SO, et al. MiRNA-196 is upregulated in glioblastoma but not in anaplastic astrocytoma and has prognostic significance. *Clin Cancer Res* 2010;16:4289–97.
25. Mueller DW, Bosserhoff AK. MicroRNA miR-196a controls melanoma-associated genes by regulating HOX-C8 expression. *Int J Cancer* 2011;129:1064–74.
26. Li Y, Zhang M, Chen H, Dong Z, Ganapathy V, Thangaraju M, et al. Ratio of miR-196s to HOXC8 messenger RNA correlates with breast cancer cell migration and metastasis. *Cancer Res* 2010;70:7894–904.
27. Shah N, Sukumar S. The Hox genes and their roles in oncogenesis. *Nat Rev Cancer* 2010;10:361–71.
28. Waltregny D, Alami Y, Clausse N, de Leval J, Castronovo V. Overexpression of the homeobox gene HOXC8 in human prostate cancer correlates with loss of tumor differentiation. *Prostate* 2002;50:162–9.
29. Miller GJ, Miller HL, van Bokhoven A, Lambert JR, Werahera PN, Schirripa O, et al. Aberrant HOXC expression accompanies the malignant phenotype in human prostate. *Cancer Res* 2003;63:5879–88.
30. Kikugawa T, Kinugasa Y, Shiraishi K, Nanba D, Nakashiro K, Tanji N, et al. PLZF regulates Pbx1 transcription and Pbx1-HoxC8 complex leads to androgen-independent prostate cancer proliferation. *Prostate* 2006;66:1092–9.
31. Alami Y, Castronovo V, Belotti D, Flagiello D, Clausse N. HOXC5 and HOXC8 expression are selectively turned on in human cervical cancer cells compared to normal keratinocytes. *Biochem Biophys Res Commun* 1999;257:738–45.
32. Zhai Y, Kuick R, Nan B, Ota I, Weiss SJ, Trimble CL, et al. Gene expression analysis of preinvasive and invasive cervical squamous cell carcinomas identifies HOXC10 as a key mediator of invasion. *Cancer Res* 2007;67:10163–72.
33. Braig Z, Mueller DW, Rothhammer T, Bosserhoff AK. MicroRNA miR-196a is a central regulator of HOX-B7 and BMP4 expression in malignant melanoma. *Cell Mol Life Sci* 2010;67:3535–48.
34. Mueller DW, Bosserhoff AK. MicroRNA miR-196a controls melanoma-associated genes by regulating HOX-C8 expression. *Int J Cancer* 2011;129:1064–74.
35. Mansfield JH, Harfe BD, Nissen R, Obenaus J, Srineel J, Chaudhuri A, et al. MicroRNA-responsive 'sensor' transgenes uncover Hox-like and other developmentally regulated patterns of vertebrate microRNA expression. *Nat Genet* 2004;36:1079–83.
36. Tsai MC, Manor O, Wan Y, Mosammaparast N, Wang JK, Lan F, et al. Long noncoding RNA as modular scaffold of histone modification complexes. *Science* 2010;329:689–93.
37. Yang Z, Zhou L, Wu LM, Lai MC, Xie HY, Zhang F, et al. Overexpression of long non-coding RNA HOTAIR predicts tumor recurrence in hepatocellular carcinoma patients following liver transplantation. *Ann Surg Oncol* 2011;18:1243–50.
38. Okada Y, Feng Q, Lin Y, Jiang Q, Li Y, Coffield VM, et al. hDOT1L links histone methylation to leukemogenesis. *Cell* 2005;121:167–78.
39. Krivtsov AV, Feng Z, Lemieux ME, Faber J, Vempati S, Sinha AU, et al. H3K79 methylation profiles define murine and human MLL-AF4 leukemias. *Cancer Cell* 2008;14:355–68.
40. Popovic R, Riesbeck LE, Velu CS, Chaubey A, Zhang J, Achille NJ, et al. Regulation of mir-196b by MLL and its overexpression by MLL fusions contributes to immortalization. *Blood* 2009;113:3314–22.
41. Hui AB, Shi W, Boutros PC, Miller N, Pintilie M, Fyles T, et al. Robust global micro-RNA profiling with formalin-fixed paraffin-embedded breast cancer tissues. *Lab Invest* 2009;89:597–606.

# Cancer Research

The Journal of Cancer Research (1916–1930) | The American Journal of Cancer (1931–1940)

## Upregulation of miR-196a and *HOTAIR* Drive Malignant Character in Gastrointestinal Stromal Tumors

Takeshi Niinuma, Hiromu Suzuki, Masanori Nojima, et al.

*Cancer Res* 2012;72:1126-1136. Published OnlineFirst January 18, 2012.

<b>Updated version</b>	Access the most recent version of this article at: doi: <a href="https://doi.org/10.1158/0008-5472.CAN-11-1803">10.1158/0008-5472.CAN-11-1803</a>
<b>Supplementary Material</b>	Access the most recent supplemental material at: <a href="http://cancerres.aacrjournals.org/content/suppl/2012/01/18/0008-5472.CAN-11-1803.DC1">http://cancerres.aacrjournals.org/content/suppl/2012/01/18/0008-5472.CAN-11-1803.DC1</a>

<b>Cited articles</b>	This article cites 41 articles, 13 of which you can access for free at: <a href="http://cancerres.aacrjournals.org/content/72/5/1126.full#ref-list-1">http://cancerres.aacrjournals.org/content/72/5/1126.full#ref-list-1</a>
<b>Citing articles</b>	This article has been cited by 13 HighWire-hosted articles. Access the articles at: <a href="http://cancerres.aacrjournals.org/content/72/5/1126.full#related-urls">http://cancerres.aacrjournals.org/content/72/5/1126.full#related-urls</a>

<b>E-mail alerts</b>	<a href="#">Sign up to receive free email-alerts</a> related to this article or journal.
<b>Reprints and Subscriptions</b>	To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at <a href="mailto:pubs@aacr.org">pubs@aacr.org</a> .
<b>Permissions</b>	To request permission to re-use all or part of this article, use this link <a href="http://cancerres.aacrjournals.org/content/72/5/1126">http://cancerres.aacrjournals.org/content/72/5/1126</a> . Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.