CD44 Proteolysis Increases CREB Phosphorylation and Sustains Proliferation of Thyroid Cancer Cells

Valentina De Falco1, Anna Tamburrino1, Simona Ventre1, Maria Domenica Castellone1, Mouhannad Malek2, Serge N. Mani3, and Massimo Santoro1

Abstract

CD44 is a marker of cancer stem-like cells and epithelial–mesenchymal transition that is overexpressed in many cancer types, including thyroid carcinoma. At extracellular and intramembranous domains, CD44 undergoes sequential metalloprotease- and γ-secretase–mediated proteolytic cleavage, releasing the intracellular protein fragment CD44-ICD, which translocates to the nucleus and activates gene transcription. Here, we show that CD44-ICD binds to the transcription factor CREB, increasing S133 phosphorylation and CREB-mediated gene transcription. CD44-ICD enhanced CREB recruitment to the cyclin D1 promoter, promoting cyclin D1 transcription and cell proliferation. Thyroid carcinoma cells harboring activated RET/PTC, RAS, or BRAF oncogenes exhibited CD44 cleavage and CD44-ICD accumulation. Chemical blockade of RET/PTC, BRAF, metalloprotease, or γ-secretase were each sufficient to blunt CD44 processing. Furthermore, thyroid cancer cell proliferation was obstructed by RNA interference–mediated knockdown of CD44 or inhibition of γ-secretase and adoptive CD44-ICD overexpression rescued cell proliferation. Together, these findings reveal a CD44-CREB signaling pathway that is needed to sustain cancer cell proliferation, potentially offering new molecular targets for therapeutic intervention in thyroid carcinoma. Cancer Res; 72(6); 1449–58. ©2012 AACR.

Introduction

CD44 is a glycosylated transmembrane glycoprotein that is implicated in tumor growth and metastasis (1–3). CD44 undergoes intramembrane proteolysis by metalloprotease that cleaves the extracellular juxtamembrane stem domain, and, then, by γ-secretase that cleaves the transmembrane domain (4–7). The first cleavage results in the shedding of the ectodomain (ecto-CD44) and the release of the membrane-bound CD44 C-terminal fragment (CD44-CTF). The second cleavage causes the release of the CD44 intracellular domain fragment (CD44-ICD; refs. 5, 6). Shedding of ecto-CD44 regulates cell–extracellular matrix interaction (8), whereas CD44-ICD translocates to the nucleus and activates gene transcription (7).

Papillary thyroid carcinoma (PTC) often features RET/PTC, BRAF oncoproteins, both of which signal through the extracellular signal–regulated kinase (ERK) pathway (9–11). RET/PTC result from the fusion of the tyrosine kinase domain of the RET receptor with the N-terminus of heterologous proteins (9). RET tyrosine 1062 (Y1062) plays an important role in RET and RET/PTC signaling, acting as the binding site for several proteins, thus leading to ERK and PI3K (phosphoinositide 3-kinase)/AKT signaling (12–16).

CD44 cleavage produces CD44-ICD in cells transformed by oncogenic RET point mutants (17). CD44 is overexpressed in PTC and in cell lines harboring RET/PTC or BRAF oncogenes (18, 19). Here, we show that CD44-ICD triggers activation of the CREB transcription factor; in thyroid cancer cells, RET/PTC-BRAF signaling cascade uses such a CD44-ICD-CREB axis to sustain cell proliferation.

Materials and Methods

Cell lines

TPC-1 and BCPAP cell lines were obtained, respectively, in 1990 from M. Nagao (National Cancer Center Research Institute, Tokyo, Japan) and in 1994 from N. Fabien (CNRS, Oullins, France). The anaplastic thyroid carcinoma cell lines ACT-1 (NRAS Q61K), HTH7 (NRAS Q61R), HTH74 (negative for HRAS, KRAS, and NRAS mutations) and C643 (HRAS G13R) were obtained in 2005 from N. Onoda (Osaka University of Medicine, Japan; ACT-1) and from N.E. Heldin (University Hospital, Uppsala, Sweden; HTH7, HTH74, and C643). These cell lines were DNA profiled by short tandem repeat (STR) analysis in 2009 (20) and shown to be unique and identical to those reported in Schweppe and colleagues (21). In brief, 15 STR loci were tested by using the Applied Biosystems AmpFISTR Identifier kit (ABI no. 432288); DNA profiles were compared manually with the American Type Culture Collection (ATCC) database and to the DNA profiles reported by Schweppe and colleagues (21). Human primary culture of thyroid cells (P5)
were obtained from F. Curcio (University Udine, Italy) in 2003 and passaged in our laboratory, as described (22), for fewer than 2 months after resuscitation. They were tested in 2010 by proliferation rate and expression of thyroid differentiation markers (TG, TPO, and TSHR). Nthy-ori 3-1 (hereafter referred to as NTHY), a human thyroid follicular epithelial cell line immortalized by the SV40 large T gene, was obtained from American Type Culture Collection (ATCC) in 2010. They were tested by ATCC for identity by DNA profiling of STR sequences and were passaged in our laboratory for fewer than 3 months after resuscitation. PC Cl 3 (hereafter referred to as ‘PC’) is a differentiated thyroid follicular cell line derived from 18-month-old Fischer rats in our laboratory in 1987 (23). They were kept in culture for fewer than 3 months after resuscitation and tested in 2010 by proliferation rate, dependence on 6 hormones for growth and expression of thyroid differentiation markers (TG, TPO, and TSHR). HEK293T cells were purchased in 2006 from the ATCC and tested by ATCC for identity by DNA profiling of STR sequences. HEK293T cells were passaged in our laboratory for fewer than 3 months after resuscitation. The PC ICD and BCPAP ICD cell lines were obtained by a stable transfection with the CD44-ICD construct by using the Fugene HD reagent from Roche Diagnostics. A pool of several cell clones was isolated by G418 selection. Transient transfections were carried out with the Fugene HD reagent from Roche Diagnostics. All these cell lines were grown in standard conditions as detailed in Supplementary Methods.

Plasmids
The RET/PTC constructs used in this study were cloned in pBABE or pcDNA3(Myc-His; Invitrogen) and described elsewhere (14). RET/PTC3(K–) is a kinase-dead mutant, carrying the substitution of the catalytic lysine (residue 758 in full-length RET) with a methionine. In RET/PTC3(4YF) mutant the substitution of the catalytic lysine (residue 758 in full-length RET) with a methionine. In RET/PTC3(3YF). These mutants were generated by site-directed mutagenesis using the QuickChange mutagenesis kit (Stratagene). BRAFV600E and HRAS(V12) have been added back in RET/PTC3(3YF). These mutants were generated by site-directed mutagenesis using the QuickChange mutagenesis kit (Stratagene). BRAFV600E and HRAS(V12) plasmids are described elsewhere (14). The full-length rat CD44 was cloned into pDEST47 vector and tagged at the C-ter with GFP (17); CD44-ICD was cloned in pDEST47 (GFP tagged), was cloned into pDEST47 vector and tagged at the C-ter with GST. Recombinant proteins containing the different domains of CREB fused to GST are described elsewhere (24). Pull-down assays were carried out by standard procedures (Supplementary Methods).

Antibodies, compounds, and proteins
The antibody against the cytosolic portion of CD44 (CD44ctyo) and anti-RET are described elsewhere (14, 17). A list of commercial antibodies, compounds, and recombinant proteins used in this study is provided in Supplementary Methods.

Cell growth and staining
For growth curves, cells were seeded in triplicate and counted at the indicated time points. Thyroid cancer cells were maintained in Dulbecco’s modified Eagle’s medium (DMEM) 2.5% FBS and PC cells in medium supplemented with 5% calf serum and without TSH. DNA synthesis was measured by the 5′-bromo-3′-deoxyuridin (BrdUrd) Labeling and Detection Kit from Boehringer Mannheim, as indicated by manufacturer (Supplementary Methods).

ELISA assay
Extracellular shedding of the soluble ectodomain of human standard CD44 (CD44st) was measured using the Instant ELISA (Bender MedSystems) according to the manufacturer’s instructions. Conditioned media from cell cultures were analyzed in triplicate at 450 nmol/L with an microplate reader (Model 550; Bio-Rad).

Protein studies
Immunoblotting and immunoprecipitation experiments were conducted according to standard procedures. For nuclear extraction, cells were lysed by shearing with 15 passages through a 26-gauge needle mounted in a 1-mL syringe. Nuclei were recovered by centrifugation at 3,000 × g for 10 minutes (Supplementary Methods).

Pull-down assay
The GST-CD44-ICD vector codes for a chimeric protein with CD44-ICD fused to the glutathione S-transferase (GST). It was generated by the Invitrogen Gateway technique using as donor the pDEST47-CD44-ICD vector and as recipient the pDEST15 vector. GST-CD44-ICD was purified from pDEST15-CD44-ICD–transformed bacterial lysates using glutathione Sepharose beads. Recombinant proteins containing the different domains of CREB fused to GST are described elsewhere (24). Pull-down assays were carried out by standard procedures (Supplementary Methods).

Reporter assay
All the Firefly luciferase reporters were kindly provided by J. S. Gutkind (NIH, Bethesda, MD; refs. 25–28). Twenty-four hours after seeding, the cells were transiently transfected in triplicate with reporters together with pRL-null, a plasmid expressing the enzyme Renilla luciferase, used as an internal control (Promega Corporation). Luciferase assays were carried out according to standard procedures (Supplementary Methods).

Chromatin immunoprecipitation
Chromatin was extracted from CD44-ICD or empty vector–transfected HEK293T cells, and chromatin immunoprecipitation assay (ChIP) was done with the chromatin immunoprecipitation assay kit (Upstate Biotechnology Inc.) according to manufacturer’s instructions, as described in Supplementary Methods.

RNA silencing
The small inhibitor duplex RNAs (siRNA) were from Dharmaco and were ON-target plus SMARTpool siCD44 human #1-L-009999-00, siCREB human: #L-003619-00-0005, and siCREB rat: #L-092995-00-0010. The siCONTROL Nontargeting Pool (+D-001206-13-05) was used as a negative control. Cells were transfected with 100 nmol/L siRNAs using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics. The Sh29merRNA constructs were transfected with 100 ng siRNA using DharmaFECT reagent. The day before transfection, cells were plated in 35-mm dishes at 40% of confluence in DMEM supplemented with 10% FBS and without antibiotics.
against human CD44 were from OriGene Technologies (TR314080 IDs: TI356313, TI356314, and TI356316). Short hairpin RNA (shRNA) pRS plasmid TR20003 was used as a negative control. Transfection was done in 100-mm dishes using Fugene HD reagent (Roche) with 4 μg shRNA construct. Cells were harvested 48 hours after transfection.

In vitro PP2A dephosphorylation assay

Recombinant HIS-CREB was in vitro phosphorylated by the catalytic subunit of PKA in a buffer containing 1 mmol/L ATP, 10 mmol/L MgCl₂, 50 mmol/L KCl, 10 mmol/L HEPES, and 10% glycerol for 1 hour at 30°C. Phosphorylated CREB was incubated in a phosphatase buffer (40 mmol/L Tris HCl, 34 mmol/L MgCl₂, 4 mmol/L EDTA, 2 mmol/L DTT, and 0.05 mg/mL bovine serum albumin) for 1 hour at 37°C with 10 μU of PP2A catalytic subunit. GST-CD44-ICD, or the GST backbone alone, were added to the samples. The reaction was terminated by adding SDS gel loading dye and samples were run on 10% SDS-polyacrylamide gel. Phospho-CREB signal was detected with anti-phospho(Ser/Thr) PKA substrate antibody and the amount of CREB in each reaction was estimated with anti-HIS antibody.

Statistical analysis

The 2-tailed unpaired Student t test (normal distributions and equal variances) was used for statistical analysis. Differences were significant when P < 0.05. Statistical analysis was done using the Graph Pad InStat software program, version 3.06.3.

Results

CD44-ICD stimulates CRE-mediated transcription

We analyzed the capability of a GFP-tagged CD44-ICD construct to trans-activate a panel of promoter elements, including AP-1 (activating protein-1), SRF (serum response factor), TCF (ternary complex factor), Gli (Glioma-associated oncogene homolog), NF-κB, and CRE (cAMP-responsive element) reporters (25–28). CD44-ICD strongly (about 10-fold, P < 0.01) activated the CRE reporter in HEK293T cells, but not the other promoters (Fig. 1A). This was a specific feature of the

Figure 1. A, luciferase assays were carried out in HEK293T cells to measure effects of a GFP-tagged CD44-ICD (ICD) on the indicated reporters. B, CRE-LUC reporter assay in HEK293T cells transiently transfected with GFP-tagged full-length CD44 or CD44-ICD (ICD). C, HEK293T cells were transiently transfected with CycD1(CCND1)-LUC and CREB siRNA or control scrambled siRNA (siCTR). CREB silencing and CD44-ICD (ICD) expression was verified by immunoblot. In A–C, cotransfected Renilla luciferase was used for normalization. Results are reported as fold change with respect to the empty vector (–). Triplicates ± SD are shown. P values were determined by the 2-tailed unpaired Student t test. CD44 or CD44-ICD (ICD) expression was verified by immunoblot with anti-GFP antibody (inset). D, immunoblot stained with cyclin D1 antibody of HEK293T cells cotransfected with CD44-ICD-GFP together with p300 or p300LYRR-dominant negative mutant. CD44-ICD expression was verified with anti-GFP. Anti-tubulin was used for normalization. E, HEK293T cells were transiently transfected with CD44-ICD. Mock or anti-CREB immunoprecipitated chromatin was analyzed by semiquantitative PCR with primers spanning the CRE site on the CCND1 promoter. Input DNA levels are shown for normalization. Data in D and E are representative of 3 independent experiments.
cleaved CD44-ICD, because full-length GFP-tagged CD44 did not significantly ($P > 0.05$) stimulate the CRE reporter (Fig. 1B).

**CD44-ICD sustains CREB/p300-mediated CCND1 expression**

CRE elements bind the CRE (cAMP-responsive element binding protein) family of transcription factors (29, 30). Cyclin D1 (CCND1) is a prototypic CREB transcriptional target, containing a CRE element upstream of the mRNA start site (at −58 bp; refs. 25, 31). We studied the effect of CD44-ICD on a CCND1-luciferase reporter. HEK293T cells were kept without serum for 24 hours after the transfection to reduce the growth factor–dependent CCND1 promoter activity. CD44-ICD stimulated (approximately 12-fold, $P < 0.01$) the CCND1 promoter (Fig. 1C, left).

We silenced endogenous CREB by transiently transfecting CREB siRNA (siCREB) in CD44-ICD–transfected HEK293T cells; CREB knockdown (about 70% of reduction) was verified by immunoblot (Fig. 1C, right). CREB silencing, but not a scrambled control, obstructed CD44-ICD–mediated activation of CCND1-luciferase (Fig. 1C, left, $P < 0.01$).

The coactivator CBP/p300 binds to active S133-phosphorylated CREB and triggers CREB-mediated gene transcription (32). Previous data showed that CD44-ICD stimulates p300-mediated transactivation (33). Thus, we cotransfected CD44-ICD with p300 or a p300-dominant negative construct (LYRR; ref. 34). CD44-ICD upregulated (about 3-fold) endogenous CCND1 expression; the p300-dominant negative mutant blocked CD44-ICD–induced CCND1 expression, whereas wt p300 slightly increased it (Fig. 1D).

Phosphorylated CREB is recruited to CRE sites in DNA (35). We carried out a ChIP to measure CRE binding to the CRE element of the CCND1 promoter. CD44-ICD expression increased the binding of CREB to the CCND1 promoter by about 5-fold (Fig. 1E).

Taken together, these results showed that CD44-ICD stimulates CREB recruitment to CCND1 promoter and CREB/p300-mediated transcription of CCND1.

**CD44-ICD increases CREB phosphorylation on serine 133**

Nuclear extracts were prepared from HEK293T cells transfected with CD44-ICD and phosphoS133 (CREB principal activation site) levels were measured by immunoblot. As a positive control, we treated cells with the adenylate cyclase activator forskolin (FSK). Besides CREB, the phospho-CREB antibody recognizes phosphorylated CREB family members CREM (30 kDa) and ATF-1 (38 kDa). Expression of CD44-ICD induced a robust increase of pS133 CREB as well as of phospho-CREM (30 kDa) and ATF-1 (38 kDa). Expression of CD44-ICD induced the robust increase of pS133 CREB as well as of phosphorylated ATF-1 (Fig. 2A, left).

Activated pS133 CREB binds p300 and this results in a transcriptionally active complex. Thus, we analyzed CREB–p300 interaction by immunoprecipitating CREB from nuclear extracts of HEK293T cells cotransfected with CD44-ICD and p300, and staining the immunoblot with p300 antibody. CD44-ICD, as well as FSK treatment, increased (about 5-fold) the CREB-p300 interaction (Fig. 2A, right).

These results showed that CD44-ICD expression increases levels of S133-phosphorylated CREB and its binding to p300.

**CD44-ICD forms a protein complex with CREB**

We transfected HEK293T cells with V5-tagged CD44-ICD and stained anti-CREB immunoprecipitates with V5 antibody. CD44-ICD formed a protein complex with CREB (Fig. 2B, To...
verify whether the interaction between CREB and CD44-ICD was direct or mediated by other proteins, we carried out a pull-down assay using recombinant CREB and GST-CD44-ICD proteins. Figure 2C shows that CREB and CD44-ICD readily interacted in vitro.

We used different domains of CREB expressed as recombinant proteins fused to GST (24) to pull-down myc-tagged CD44-ICD expressed in HEK293T cells. As shown in Fig. 2D, CD44-ICD bound 2 contiguous domains of CREB: Q2 (aa 160–283), the constitutive glutamine-rich activation domain, and bZIP (aa 284–341), the DNA binding/dimerization domain. CD44-ICD did not bind the isolated DbZIP construct (aa 1–283) of CREB that contains Q2 but not bZIP. It is possible that, in the absence of bZIP, the presence of the amino-terminal part of the protein interferes with the interaction between Q2 and CD44-ICD. The fact that CD44-ICD binds to a CREB domain different to that binding to p300 (KID domain; ref. 29) is consistent with the possibility that a complex of 3 proteins, CREB, p300, and CD44-ICD is formed.

**CD44-ICD reduces the rate of CREB dephosphorylation**

Various serine/threonine kinases, that is, PKA, RSK, and MSK, are able to phosphorylate CREB on S133 and to stimulate CREB binding to DNA (29, 35, 36). We evaluated the effects of the blockage of these kinases. To inhibit PKA, we either treated cells for 2 hours with the H89 compound, a potent selective inhibitor of PKA, or transfected cells with a plasmid encoding the PKA-specific peptide inhibitor (PKi; ref. 37). RSK and MSK are activated by MEK (mitogen-activated protein/ERK kinase; ref. 36); thus, to inhibit RSK and MSK, we treated cells for 2 hours with the MEK inhibitor U0126. Nuclear extracts were analyzed by Western blot with anti-pS133 CREB and normalized with anti-CREB. The inhibition of both PKA and MEK efficiently blocked CD44-ICD–induced CREB phosphorylation (Supplementary Fig. S1). These results prompted us to hypothesize that, in the presence of CD44-ICD, the increase of CREB phosphorylation levels occurs because of a reduced rate of CREB dephosphorylation, rather than to activation of a specific CREB kinase. To evaluate this possibility, we stimulated serum-starved HEK293T cells for 40 minutes with the cAMP-analog N6-benzoyl-cAMP to induce S133 CREB phosphorylation, and then measured CREB binding to CD44-ICD. GST-CD44-ICD pulled down a larger amount of CREB upon induction of CREB phosphorylation (Fig. 3A), indicating that CD44-ICD, although able to bind dephosphorylated CREB, binds preferentially to pS133 CREB.

Then, we induced CREB phosphorylation with N6-benzoyl-cAMP in the presence or not of GFP-tagged CD44-ICD and chased pS133 CREB dephosphorylation by immunoblot. Figure 3B shows that the pS133 CREB half-life was increased by CD44-ICD; CREB pS133 levels did not decrease up to 12 hours in cells

![Figure 3](https://www.aacrjournals.org/doi/fig/10.1158/0008-5472.CAN-11-3320/fig3.png)
expressing CD44-ICD, whereas the half-life of pS133 CREB was about 8 hours in mock-transfected cells (Fig. 3B).

Finally, we carried out an in vitro CREB dephosphorylation assay. Recombinant HIS-tagged CREB was phosphorylated in vitro by PKA and then incubated with PP2A phosphatase in the presence or the absence of GST-CD44-ICD. As shown in Fig. 3C, GST-CD44-ICD, but not the GST backbone, protected phosphoCREB from PP2A-mediated dephosphorylation. These findings showed that CD44-ICD reduces the rate of CREB dephosphorylation on S133.

Expression of CD44-ICD-CREB complex in human thyroid carcinoma cell lines

We selected thyroid cancer cells as a model whereby to study effects of CD44-ICD. Initially, we looked for CD44-ICD, in 2 PTC cell lines, TPC-1 and BCPAP, that feature the RET/PTC1 rearrangement and the BRAFV600E mutation, respectively; as a control, we used P5, a normal thyroid primary cell culture. Both PTC cell lines, but not nontransformed thyrocytes, expressed full-length CD44 as well as CD44-ICD (Supplementary Fig. S2A). Moreover, pS133 CREB as well as phosphoATF-1 levels were detected in BCPAP and TPC-1, whereas they were only barely detectable in P5 cells (Supplementary Fig. S2A).

GST-CD44-ICD, but not the GST backbone, was able to pull down pS133CREB and phosphoATF-1 in both BCPAP and TPC-1 cell lysates (Supplementary Fig. S2B, left). Finally, in thyroid cancer cells, the pCREB-CD44-ICD interaction was shown by immunoprecipitating pS133-phosphorylated CREB and staining the blot with anti-CD44cyto antibody (Supplementary Fig. S2B, right).

These findings showed that thyroid cancer cells express CD44-ICD and that, in these cells, CD44-ICD exists in a complex with CREB.

RET/PTC and BRAFV600E oncogenes induce CD44 cleavage

We treated TPC-1 and BCPAP cells for 48 hours with BB94, a broad-spectrum metalloprotease inhibitor; moreover, we treated the RET/PTC1-positive TPC-1 cell line with ZD6474, a RET kinase inhibitor, and the BRAFV600E-positive BCPAP cell line with U0126, a MEK inhibitor. EctoCD44 (sCD44st, soluble standard CD44) shedding into the cell culture media was measured by ELISA. EctoCD44 release was detected in TPC-1 and BCPAP conditioned media (Supplementary Fig. S3A, left); treatment with BB94, ZD6474, or U0126 blocked ectoCD44 shedding (P < 0.01), indicating that RET and BRAF signaling stimulates CD44 cleavage (Supplementary Fig. S3A, left). In TPC-1, treatments with BB94, ZD6474, and UO126 downregulated CD44-ICD (Supplementary Fig. S3A, middle). CD44 downregulation by siRNA proved that the protein band with a relative molecular weight of about 10 kDa was indeed CD44-ICD (Supplementary Fig. S3A, right). Finally, γ-secretase blockade by COMp X or DAPT decreased the amount of CD44-ICD, whereas it increased the amount of a polypeptide of approximately 31 kDa, which corresponds to the molecular weight of CD44-CTF (Supplementary Fig. S3A, middle). This effect was consistent with a block of transmembrane conversion of CD44-CTF to CD44-ICD. In contrast, CD44-CTF did not accumulate upon RET/PTC1 or MEK inhibition, which suggests that the RET/PTC1 and MEK block acts upstream CD44-CTF generation, possibly at the level of metalloprotease-mediated CD44 cleavage (Supplementary Fig. S3A, middle).

We coexpressed a GFP-tagged full-length CD44 with the 2 most prevalent RET/PTC oncogenes, RET/PTC1 and RET/PTC3. RET/PTC oncogenes stimulated the generation of CD44-ICD and CD44-CTF (Supplementary Fig. S3B). That the band migrating above CD44-ICD corresponded to CD44-CTF was suggested by the fact that when we used the metalloprotease inhibitor BB94 in cells coexpressing CD44-GFP and RET/PTC1, this band was strongly reduced (Supplementary Fig. S4B). RET/PTC-mediated CD44 cleavage depended on RET/PTC kinase activity and on the integrity of tyrosine 1062, because CD44-ICD did not accumulate when kinase-dead (K–) and (Y1062F) RET/PTC1 mutants were expressed (Supplementary Fig. S3C, left). CD44-ICD did not accumulate when the RET/PTC3 4YF mutant (in which the 4 major RET signaling
tyrosines are mutated to phenylalanine) was expressed (Supplementary Fig. S3C, right), but CD44 cleavage was rescued when Y1062 was added back (3YF; Supplementary Fig. S3C, right). We treated RET/PTC1-transfected HEK293T cells with U0126. Treatment with U0126 blocked CD44-ICD generation (Supplementary Fig. S3D). Transient expression of the myctagged constitutively active forms of RAS (RASV12) and BRAF (BRAFV600E) induced CD44-ICD formation (Supplementary Fig. S3D), which further supports the concept that the ERK pathway plays an important role in CD44 cleavage. Accordingly, also human thyroid carcinoma cell lines featuring mutant RAS alleles upregulated both CD44 and CD44-ICD with respect to nontransformed thyrocytes (Supplementary Fig. S5).

These findings showed that RET/PTC and BRAF signaling to ERK stimulates CD44 processing.

CD44-ICD sustains CREB phosphorylation and CCND1 expression in thyroid cancer cells

In BCPAP and TPC1 thyroid cancer cells, CD44 silencing by shCD44, but not the empty vector, attenuated CREB phosphorylation on S133 (Fig. 4A). In shCD44-transfected cells, CREB phosphorylation was rescued by the transfection of a V5-tagged rat CD44-ICD mRNA that is shCD44 resistant because it has several mismatches with human shCD44 (Fig. 4A). BB94 and DAPT, able to block CD44 cleavage, reduced CREB pS133 (Fig. 4B). Downregulation of either CREB or CD44 by siRNA, but not a scrambled control, reduced (about 2-fold) the expression of CCND1 in BCPAP and TPC-1 cell lines (Fig. 4C).

These findings showed that CD44-ICD sustains CREB-dependent CCND1 expression in thyroid cancer cells.

CD44-ICD sustains proliferation of thyroid cells

We silenced CD44 by transiently transflecting BCPAP and TPC-1 cells with a human shCD44 plasmid. To exclude off-target effects, cells were cotransfected with the shCD44-resistant rat CD44-ICD construct. CD44 silencing reduced BrdUrd incorporation (P < 0.05), and this effect was rescued by adoptive expression of rat CD44-ICD (P < 0.05; Fig. 5A). CD44-ICD was not able to rescue the reduction of BrdUrd incorporation mediated by CREB silencing (P > 0.05) that is consistent with CREB acting downstream CD44-ICD (Fig. 5A). We treated BCPAP cells with the γ-secretase inhibitor COMP X and, to rescue the effect, we transfected CD44-ICD or the empty vector. As shown in Fig. 5B, COMP X inhibited DNA synthesis in BCPAP, and CD44-ICD was able to revert this effect. We stably expressed CD44-ICD in BCPAP and selected one mass population by G418 treatment. Although treatment with COMP X reduced the growth rate of parental cells (P < 0.01; Fig. 5C, left), it did not change significantly (P > 0.05) the proliferation of BCPAP-ICD cells (Fig. 5C, right).

Nontransformed thyroid PC cells require a mixture of 6 hormones (6H), including TSH, for proliferation (23). In hormone-starved PC cells, transiently transfected CD44-ICD stimulated CRE-mediated transcription (about 9-fold; Fig. 6A; P < 0.01) and activated the CCND1 promoter (almost 4-fold; Fig. 6B; P < 0.01). CREB silencing, but not a scrambled control, obstructed CD44-ICD–mediated activation of CCND1 luciferase (Fig. 6B; P < 0.01). A mass population of CD44-ICD–transfected PC cells (PC ICD pool) showed increased (about 8-fold) CRE-reporter luciferase activity with respect to empty vector–transfected cells (Fig. 6C; P < 0.01). Finally, we measured the rate of DNA synthesis in the absence of TSH. BrdUrd incorporation in PC
ICD cells was higher (about 10-fold) than in empty vector–transfected cells (Fig. 6D; \( P < 0.01 \)). Thus, CD44-ICD is sufficient to trigger proliferation of nontransformed thyrocytes.

**Discussion**

Here we report a novel functional link between CD44 and the CREB transcription factor. CREB is involved in neoplastic transformation and, being activated via PKA by cAMP, is also involved in the growth of normal thyroid follicular cells (38–41). Our data show that CD44-ICD binds CREB and increases pS133 CREB levels. CD44-ICD stimulated CREB-mediated gene transcription and recruitment of CREB to CCND1 gene promoter. We noted that CD44-ICD preferentially binds to phosphorylated CREB and that CD44-ICD expression attenuates the rate of CREB dephosphorylation, thus suggesting that interaction with CD44-ICD impairs CREB dephosphorylation on S133. Accordingly, CD44-ICD protected pCREB from PP2A-mediated dephosphorylation in vitro.

Thyroid carcinoma overexpresses CD44 and such overexpression is associated to the oncogenic conversion of the ERK signaling pathway (18, 19). Moreover, as in other cell types (42), CD44 was expressed in prospectively identified thyroid cancer stem cells that induce tumors when injected orthotopically into mouse thyroid (43). Thus, it is feasible that CD44 plays a role in thyroid cancer as well as in thyroid cancer stem cells.

Oncogenic RET point mutants induce CD44 cleavage (17). Here, we show that CD44-ICD is expressed in thyroid cancer cell lines harboring RET/PTC or BRAFV600E oncogenes and that RET/PTC and BRAF trigger CD44 cleavage. CD44-ICD is necessary for the proliferation of thyroid cancer cells and sufficient to trigger proliferation of nontransformed thyrocytes. These effects are mediated by CREB and by increased rate of CCND1 transcription. The fact that extracellular shedding of ectoCD44 accompanies the generation of CD44-ICD and that RET and BRAF blockade impairs CD44-CTF accumulation indicates that metalloprotease-mediated cleavage is one level at which CD44-ICD generation is stimulated in cells expressing RET/PTC and BRAF. Accordingly, RET/PTC and BRAF upregulate the transcription of several metalloproteases (44). CD44 cleavage can be triggered by binding to different extracellular ligands, including low molecular weight hyaluronan acid (45). RET, RAS, and BRAF oncogenes stimulate the expression of osteopontin, one extracellular CD44 ligand; whether this facilitates CD44 cleavage remains to be determined.

Our findings support a model whereby CD44 cleavage acts as an amplifier of oncogenes signaling to CREB (Fig. 7). According to this model, RET/PTC and BRAF promote CREB phosphorylation via ERK-mediated activation of CREB kinases and stabilize pS133-CREB through CD44-ICD. In turn, the CD44-ICD-CREB axis stimulates CCND1 transcription and thyroid cell proliferation. Should these results be validated in vivo, they
may prompt the possibility of pharmacologic manipulation of the pathway.

**Disclosure of Potential Conflicts of Interest**
No potential conflicts of interest were disclosed.

**Acknowledgments**
The authors thank AstraZeneca for ZD6474, A. Felciello for PKA inhibitors, J. S. Gutkind for reporter vectors, M. Montminy for CREB recombinant proteins, G. Blandino for p300 vector, F. Curcio for P5 cells, M. Nagao for TPC1, N. Fabien for BCPAP, N. Onoda for ACT-1, and N.E. Heldin for HTH7, HTH74 and C643 cells and also thank J.A. Gilder for text editing and A.M. Cira for technical assistance.

**References**

**Grant Support**
This study was supported by the grant no. 5880 of Associazione Italiana per la Ricerca sul Cancro (AIRC), grant no. PI0-2 of Italian Ministero della Sanita, grants no. E6(1)1100000001 and E6(1)1000201001 of Ministero dell’Univerità e della Ricerca, grant no. FMR-36495 of the European Union Contract (GEN-BISK-T), and grant no. 4915 of ARC and LNCL-comité du Rhone.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received October 4, 2011; revised December 29, 2011; accepted January 13, 2012; published OnlineFirst January 23, 2012.


CD44 Proteolysis Increases CREB Phosphorylation and Sustains Proliferation of Thyroid Cancer Cells

Valentina De Falco, Anna Tamburrino, Simona Ventre, et al.


Updated version  Access the most recent version of this article at: doi:10.1158/0008-5472.CAN-11-3320

Supplementary Material  Access the most recent supplemental material at: http://cancerres.aacrjournals.org/content/suppl/2012/01/23/0008-5472.CAN-11-3320.DC1

Cited articles  This article cites 45 articles, 16 of which you can access for free at: http://cancerres.aacrjournals.org/content/72/6/1449.full.html#ref-list-1

Citing articles  This article has been cited by 3 HighWire-hosted articles. Access the articles at: /content/72/6/1449.full.html#related-urls

E-mail alerts  Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions  To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions  To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.