Jab1/CSN5 Negatively Regulates p27 and Plays a Role in the Pathogenesis of Nasopharyngeal Carcinoma

Yunbao Pan1,3, Qingxiu Zhang3, Ling Tian3, Xin Wang2, Xiaohang Fan1, Huizhong Zhang2, Francois X. Claret3,4, and Huiling Yang1

Abstract

Nasopharyngeal carcinoma (NPC) is an Epstein-Barr virus–associated malignancy most common in East Asia and Africa. Aberrant expression of Jab1/CSN5, a negative regulator of the cell-cycle inhibitor p27, is correlated with reduced p27 expression and associated with advanced tumor stage and poor prognosis in several human cancers. In this study, we examined the functional relationship between Jab1 and p27 protein expression in NPC. Immunohistochemical analysis showed an inverse association between Jab1 and p27 in NPC tissue samples, and overexpression of Jab1 correlated with poor survival in patients with NPC. Mechanistically, Jab1 and p27 were found to interact directly in NPC cells, with Jab1 mediating p27 degradation in a proteasome-dependent manner. Knockdown of Jab1 resulted in a remarkable increase in p27 levels and inhibition of cell proliferation, indicating that Jab1 targets p27 for degradation, thereby controlling its stability. Jab1 depletion also enhanced the antitumor effects of cisplatin in NPC cells. Together, our findings suggest that Jab1 overexpression plays an important role in the pathogenesis of NPC through Jab1-mediated p27 degradation. Jab1 therefore represents a novel diagnostic marker and therapeutic target in patients with NPC. Cancer Res; 72(7); 1890–900. ©2012 AACR.

Introduction

Nasopharyngeal carcinoma (NPC), originating from the epithelial lining of the nasopharynx, is a squamous cell carcinoma (1). This neoplasm has remarkable ethnic and geographic distribution, with a high prevalence in southern China, Southeast Asia, Northern Africa, and Alaska (2, 3). The annual incidence peaks at 50 cases per 100,000 people in the endemic regions but is rare in the Western world (1 per 100,000; ref. 2). Etiologic studies have indicated that Epstein-Barr virus (EBV) infection, environmental factors, and genetic susceptibility are associated with NPC (1, 4). In China, more than 95% of NPCs are nonkeratinizing carcinoma, whereas less than 5% are keratinizing carcinoma; thus, in our study, we use CNE1 (keratinizing carcinoma) and CNE2 (nonkeratinizing carcinoma) cells to represent the 2 main histologic types (5).

To develop better treatment approaches, it is important to understand the molecular basis of the development and progression of NPC. Genetic alterations have been reported in NPCs including a frequent reduction of p27 expression (9, 10). Furthermore, low p27 expression levels may contribute to the aggressive behavior of NPCs (11). Overexpression of cyclin E mRNA has been shown to predict poor prognosis in NPC and has been correlated with an advanced stage of NPC and a low overall survival rate (12). Recently, it has been reported that PIK3CA gene amplification is frequently observed in advanced-stage NPC, which emphasizes the association between PIK3CA gene amplification and poor prognosis (11). It has also been shown that Akt promotes cell proliferation and survival in NPCs (4, 13). However, additional molecular abnormalities resulting in the deregulation of cell-cycle progression may also occur. Jab1/CSN5 (Jab1 hereafter), as we initially identified as a c-Jun coactivator, is also known as the fifth component of the COP9 signalosome (CSN) complex (CSN5; refs. 14, 15). Jab1 promotes cell proliferation and inactivates p27 by inducing transllocation of p27 from the nucleus to the cytoplasm, which accelerates p27 degradation through the ubiquitin-dependent proteasome pathway and promotes cell-cycle progression (16).
p27 is a universal cyclin-dependent kinase (Cdk) inhibitor that directly inhibits the enzymatic activity of cyclin–Cdk complexes, resulting in cell-cycle arrest at G1 (17). In addition, p27 protein levels are increased in quiescent cells and rapidly decrease after cells are stimulated with mitogens (18). Although transcriptional regulation is possible, the cellular abundance of p27 is primarily regulated at the posttranslational level by the ubiquitin–proteasome pathway (19). Jab1 overexpression is correlated with a loss of p27 and a lower rate of survival in patients with breast cancer, suggesting a role in breast cancer pathogenesis (20). This inverse association between Jab1 and p27 expression has also been observed in anaplastic large cell lymphoma (21), ovarian cancer (22), pancreatic adenocarcinomas (23, 24), and other cancer types (25–27). However, the mechanisms leading to p27 downregulation in NPCs remain undefined. Because Jab1 overexpression is correlated with the loss of p27 in several cancers and low p27 expression is associated with higher tumor grades (28), we hypothesized that Jab1 functions as a negative regulator of p27 and as such may play a role in the pathogenesis of NPCs.

To test our hypothesis, we assessed Jab1 and p27 expression in a series of 45 NPC and 30 nasopharyngeal inflammation tissue specimens. We found that Jab1 overexpression was associated with no or low expression of p27 in these samples. To further elucidate the role of Jab1 in p27 degradation in NPCs, we infected NPC cell lines with an adenoviral vector overexpressing Jab1 and found that p27 levels were significantly reduced. We also detected a direct physical interaction between Jab1 and p27 in NPC cells. Furthermore, inhibition of endogenous Jab1 expression with specific siRNAs resulted in a substantial increase of p27 levels and inhibition of cell proliferation, indicating that Jab1 controls the stability of p27 by targeting it for degradation in NPCs. Interestingly, siRNA-mediated depletion of Jab1 inhibited cell proliferation and enhanced the antitumor effects of cisplatin in NPC cells. This may suggest that Jab1 is a potential target for treating NPCs.

Materials and Methods

Patients and tissue samples

All patients were from the Cancer Center of Sun Yat-Sen University (Guangzhou, Guangdong, PR China) in 2003. The study group consisted of 36 men and 9 women with NPC who underwent radiotherapy and the control group consisted of 13 men and 17 women with nasopharyngeal inflammation. Patients who had preoperative diagnosis and did not receive preoperative chemoradiation treatment were selected for this study on the basis of the availability of archived paraffin-embedded NPCs and nasopharyngitis tissue blocks for immunohistochemical analysis. Ethical approval was obtained from the cancer center and fully informed consent from all patients before sample collection. Surgical staging of tumors had been done according to the American Joint Committee on Cancer tumor-node-metastasis system and tumor grading was based on currently used histopathologic criteria.

Reagents

Cell culture medium was from Mediatech Inc. and FBS was obtained from Gibco. The antibodies used were Jab1 (Santa Cruz); p27 and PARP (BD Biosciences PharMingen); caspase-3, lamin A/C, and Myc-tag (Cell Signaling Technology; and Flag and β-actin (Sigma-Aldrich). The Lipofectamine Plus and Oligofectamine reagents were from Invitrogen. NE-PER nuclear and cytoplasmic extraction reagents and Western Lightning Chemiluminescence Plus reagent were from Thermo Scientific Pierce. Annexin V/PI kit was from BD Biosciences. Cell Proliferation Kit was from Roche.

Human tissues and immunohistochemical analysis

A total of 75 formalin-fixed, paraffin-embedded human primary NPCs and noncancerous nasopharyngeal specimens from 45 patients with NPC and 30 patients with nasopharyngitis were analyzed. The p27 and Jab1 levels in the formalin-fixed, paraffin-embedded tissue sections were measured by immunohistochemical analysis, as previously described (24). Briefly, heat-induced retrieval of Jab1 and p27 antigens was conducted, and the slides were incubated with the primary antibodies. An immunoreaction was detected with the LSAB+ kit from DakoCytomation. We used 3,3'-diaminobenzidine as the chromogen and hematoxylin as the counterstain. Expression levels of Jab1 and p27 were evaluated by counting at least 500 tumor cells in representative high-power fields. Tumor cells were considered positive for Jab1 or p27 when nuclear or cytoplasmic staining was present. For each tumor, we determined a proportion score and an intensity score. The proportion score represented the estimated fraction of positively stained cells (0, <5%; 1, 5%–35%; 2, 35%–65%; and 3, >65%). The intensity score represented the estimated average staining intensity of the positive cells (0, no staining; 1, weak staining; 2, intermediate staining; and 3, strong staining). The overall amount of protein present in each tumor was then expressed as the average of the proportion and intensity scores for negative and positive cells (ranges, 0 and 0.5–3, respectively). We defined the scores 0.5 to 1.0, 1.5 to 2.0, and 2.5 to 3.0 as negative (−), weakly positive (+), and highly positive [(++)], respectively.

Cell cultures

Human NPC cell lines CNE1 (well-differentiated, from Cancer Center, Sun Yat-Sen University), CNE2 (poorly differentiated, from Cancer Center, Sun Yat-Sen University), HONE1 (poorly differentiated, the generous gift of Prof. Ronald Glaser, The Ohio State University Medical Center, Columbus, OH), and C666.1 (undifferentiated, the generous gift of Prof. Kwok-Wai Lo, Prince of Wales Hospital, The Chinese University of Hong Kong, Shatin, N.T., Hong Kong) were cultured in RPMI medium containing 10% FBS and penicillin–streptomycin sulfate. 293T human embryonic kidney cells were cultured in Dulbecco’s Modified Eagle’s Medium (Invitrogen) with 10% FBS and penicillin–streptomycin. HOK16B (normal keratinocyte cells, the generous gift of Prof. Jeffrey N. Myers, MD Anderson Cancer Center, Houston, TX) was cultured in keratinocyte serum-free media containing 30 μg/ml bovine pituitary extract, 0.2 ng/ml EGF, 5% FBS, and...
penicillin–streptomycin sulfate. All cell lines were incubated at 37°C in an atmosphere of 5% CO₂.

**Cell extracts and immunoblotting**

Cells in the log phase of growth were collected, washed twice in cold PBS, and lysed as described previously (29). Nuclear fractions of the indicated cell lysates were separated with NE-PER nuclear and cytoplasmic extraction reagents according to the manufacturer’s directions. Proteins were separated by 10% SDS-PAGE, transferred to nitrocellulose membranes, and probed with anti-Jab1, anti-p27, anti-Myc, anti-PARP, and anti-caspase-3. β-Actin was used as the internal positive control for all immunoblots. Immunoreactive bands were detected with horseradish peroxidase–conjugated secondary antibodies with the Western Lightning Chemiluminescence Plus Reagent. Activity of PARP and caspase-3 was measured as percentage of cleavage band intensity to the total bands and calculated as follows: % PARP = (c/Tc)/(100% × Tt/Tc), where Tc is the intensity value of the cleavage bands and Tt is the intensity value of the total bands.

**Adenoviral vectors and gene transduction**

A recombinant adenoviral vector expressing a doxycycline-regulated (Tet-Off) form of human Jab1 (Ad-Myc-Jab1) was constructed, as previously described (24). NPC cells were transduced for 48 hours with a regulatory virus (adeno-X Tet-Off, Clontech) and Ad-Myc-Jab1 at a multiplicity of infection of 50 in a growth medium containing 10% FBS in the presence or absence of 1 μg/ml doxycycline, a tetracycline analogue.

**Proteasome inhibition assays**

NPC cells were treated with the proteasome inhibitors LLnL (35 μmol/L) or MG132 (30 μmol/L), or LLM (25 μmol/L), and dimethyl sulfoxide (DMSO) as a control. Activity of PARP and caspase-3 was measured as percentage of cleavage band intensity to the total bands and calculated as follows: % PARP or caspase-3 = 100% × Tc/Tt, where Tc is the intensity value of the cleavage bands and Tt is the intensity value of the total bands.

**siRNA, DNA transfection, and coimmunoprecipitation assays**

For the siRNA analysis, Jab1 siRNA and control siRNA oligonucleotides were cloned into an RNAi vector (BD Biosciences PharMingen) according to the manufacturer’s instructions and as described by Kouvaraki and colleagues (24). The Myc-Jab1 and Flag-p27, GFP-Jab1, and Cherry-p27 plasmids were transfected into either 293T or CNE1 cells with the Lipofectamine Plus reagent, and cells were either lysed for coimmunoprecipitation assays or examined with a fluorescence microscope.

**Cell proliferation assay**

The MTT assay was used to evaluate cell viability, as described previously (29). Briefly, 48 hours after transfection, NPC cells were seeded in 96-well plates (500 cells per well for growth curves or 2,000 cells per well for cisplatin treatment) in 100 μL of RPMI-1640 medium with 8% FBS. After the indicated incubation period, the MTT labeling reagent (final concentration, 0.5 mg/mL) was added, and the spectrophotometric absorbance of the samples was read with a microplate (ELISA) reader at 570 nm. The ratios (%) of cell survival or colony formation were calculated as the ratio of the indicated treatment group to the control group as follows: % survival ratio = 100% × Nc/Nt, and % inhibition ratio = 100% × (1−Nc/Nt), where Nt is the optical density (OD) value (colony number) of the treatment group and Nc is the OD value (colony number) of control group.

**Colony formation assay**

Forty-eight hours after transfection, NPC cells (200 cells per well) were plated in 6-well plates for growth analysis in RPMI-1640 medium with 8% FBS. The following day, the cells were treated with 0 or 0.5 μmol/L cisplatin for 48 hours. After 10 days, the cells were fixed with methanol, stained with 0.1% crystal violet, and scored by counting with an inverted fluorescence microscope.

| Table 1. Characteristics of 45 patients with NPC included in the study |
|-----------------------------|----------------|
| Characteristics             | No. of patients (N = 45) |
| Gender (male/female)        | 36/9 |
| Median age, y               | 41 |
| Tumor histologic type       | |
| Squamous cell carcinomas    | 32 (71%) |
| Nonkeratinizing carcinomas  | 13 (29%) |
| Differentiation             | |
| Poor                        | 32 (71%) |
| No differentiation          | 13 (29%) |
| Lymph node metastasis       | |
| Positive                    | 40 (89%) |
| Negative                    | 5 (11%) |
| Stage                       | |
| I                           | 0 |
| II                          | 12 (27%) |
| III                         | 20 (44%) |
| IV                          | 13 (29%) |
microscope, with the standard definition that a colony consists of 50 or more cells.

**Flow cytometric analysis of the cell cycle and apoptosis**

Forty-eight hours after transfection, the cells were collected and fixed overnight in 75% cold ethanol at −20°C. Cells were washed twice in cold PBS and labeled with propidium iodide (PI) and analyzed immediately after staining with a FACScan flow cytometer (BD Biosciences) and FlowJo software.

Flow cytometric analysis was used to differentiate between living, early apoptotic, late apoptotic/necrotic, and necrotic cells by staining with Annexin V-FITC and PI. Briefly, after treating cells with 5 μmol/L cisplatin for 48 hours, the transfected cells were labeled with Annexin V-FITC and PI according to the manufacturer’s recommendations. Quantification of Annexin V-FITC and PI binding was conducted with a FACScan flow cytometer.

**Statistical analysis**

Fisher’s exact test was used to compare the expression and localization of Jab1 and p27 with various clinicopathologic variables. The Spearman test was used to analyze the association between cytoplasmic Jab1 and p27. Overall survival was defined as the time from diagnosis to death. Kaplan–Meier analysis was used to examine the association of Jab1 and p27 expression and survival. Statistical analysis for the results was done by Student t test when only 2 groups or one-way ANOVA when more than 2 groups were examined. Differences between
groups were stated to be statistically significant when \( P < 0.05 \).

All computations were carried out with SPSS 16.0 (SPSS).

**Results**

**Patient characteristics and demographics**

To analyze the Jab1 and p27 expression patterns, we used 45 NPC tissue samples (median age, 41; range, 20–73 years) and 30 nasopharyngeal inflammation tissue samples (median age, 36; range, 14–65 years). The clinicopathologic characteristics of the patients with NPC are summarized in Table 1.

**Jab1 and p27 expression patterns in tissue samples from patients with noncancerous inflamed and NPC**

Immunohistochemical analysis of nasopharyngitis and NPC tissue samples showed that Jab1 was localized to the cytoplasm and nucleus of NPC cells. About 74% had positive cytoplasmic staining and 56% had positive nuclear staining for Jab1 in NPCs. The staining in these cases was higher than that in the noncancerous inflamed nasopharyngeal tissue, which was 10% in cytoplasmic and 16% in nuclear, respectively, \( P < 0.01 \) (Fig. 1A). In contrast, nuclear p27 level was higher in noncancerous inflamed nasopharyngeal tissues (67%) than that in the NPC tissues (58%). However, 64% cytoplasmic p27 staining was observed in NPC cases, which was higher than that in noncancerous tissue (37%), \( P < 0.05 \) (Fig. 1B). Furthermore, cytoplasmic Jab1 expression in NPC tissues was inversely associated with nuclear p27 patterns (\( R = -0.389, p = 0.008 \)) and correlated with cytoplasm p27 (\( R = 0.328, P = 0.028 \); Fig. 1C).

**Correlation of Jab1 and p27 expression with clinical outcome**

Survival analysis by the Kaplan–Meier method showed that high expression of Jab1 tended to correlate with a poor prognosis, \( P < 0.01 \) (Fig. 1D). Similar results were obtained from the subsets of patients with nonkeratinizing carcinoma (Supplementary Fig. S1A) and keratinizing squamous cell carcinoma (Supplementary Table S1B).
Jab1 expression is associated with poor prognosis in NPC
carcinoma (Supplementary Fig. S1B). The median for survival time in patients with negative and weakly positive Jab1 tumors was 34 months, compared with 14 months in patients with highly positive Jab1 tumors ($P < 0.01$). Meanwhile, cytoplasmic p27-positive expression correlated with a poor prognosis, the median for survival time was 19 months in positive group whereas 36 months in negative group ($P < 0.05$).

**Jab1 promotes p27 degradation through proteolysis and is sensitive to proteasome inhibitors**

Immunoblotting also showed strong Jab1 expression in NPC cells but not normal keratinocyte cells, where weak Jab1 expression was detected (Fig. 2A). Treatment of NPC cells with specific proteasome inhibitors resulted in a significant increase in total p27 levels (Fig. 2B). To investigate whether Jab1 overexpression in NPC cells downregulates p27 levels, we transduced those 4 cell lines with Jab1 adeno virus in the absence or presence of doxycycline and measured Myc-Jab and p27 levels 48 hours after infection. Using this inducible (Tet-Off) system, Jab1 overexpression led to a significant decrease in p27 levels in all cell lines tested (Fig. 2C).

Downregulation of p27 mediated by Jab1 overexpression was inhibited in cells that had been treated with proteasome inhibitors (LLnL, MG132, and LLM) but not in those treated with DMSO (Fig. 2D), indicating that Jab1 promotes p27 degradation through proteolysis and is sensitive to proteasome inhibitors. These data provide direct evidence of a functional relationship between Jab1 and p27 in NPC cells.

**Jab1 specifically interacts with p27**

To determine whether Jab1 and p27 directly interact, we conducted coimmunoprecipitation analyses. Lysates were immunoprecipitated with anti-Jab1 antibodies and immunoblotted with anti-p27 antibodies (Fig. 3A). We found that endogenous Jab1 indeed interacts with p27 (Fig. 3A). This interaction was specific because the Jab1-containing lysates were not immunoprecipitated with nonimmune rabbit serum. To confirm these results, we examined the ectopic Jab1 and p27 interactions by transfecting Myc-Jab1 and Flag-p27 plasmids into 293T cells. As expected, Myc-Jab1 binds to Flag-p27 (Fig. 3B). Moreover, data showed most p27 in a predominantly nuclear location when Cherry-p27 was transfected into CNE1 cells, but both nuclear and cytoplasmic staining were seen when GFP-Jab1 was induced (Fig. 3C and Supplementary Fig. S3). These data strongly indicate that Jab1 associates with p27 in NPC cells.

**Depletion of Jab1 by siRNA increases accumulation of p27, induces cell-cycle arrest, and inhibits cell proliferation in NPC cell lines**

To assess the effect of silencing Jab1 in human NPC cells, we transfected NPC cells with Jab1 siRNA oligonucleotides or control siRNA oligonucleotides cloned into the RNAi-vector system. Forty-eight hours after transfection, Jab1 protein levels were substantially decreased (>$97\%$) in all Jab1 siRNA–transfected cell lines compared with cells transfected with the control siRNA oligonucleotides (Fig. 4A). The decrease in endogenous Jab1 protein expression was associated with a significant increase in p27 levels and nuclear accumulation of p27 in nucleus (Fig. 4A), suggesting the biologic importance of Jab1 in regulating p27 proteolysis in NPCs.

Next, to determine whether knockdown of Jab1 would lead to an increase in p27 activity, as indicated by inhibition of the G1-S phase progression, we used PI staining to analyze the
proportion of cells that were in the G1 and S-phases of the cell cycle. In CNE1 cells, 24% of the control siRNA-transfected cells were in the S-phase compared with 37% of cells in the G1 phase, and 14% of the Jab1 siRNA–transfected cells were in the S-phase compared with 47% of cells in the G1 phase (Fig. 4B). Similar results were obtained from CNE2 cells (25% in S-phase and 37% in G1 phase for control cells vs. 18% in S-phase and 42% in G1 phase for Jab1 siRNA–transfected cells) and HONE1 cells (33% in S-phase and 28% in G1 phase for control cells vs. 22% in S-phase and 42% in G1 phase for Jab1 siRNA–transfected cells). To confirm the above conclusion, we next conducted a coimmunoprecipitation experiment to measure the amount of p27–cyclin E–Cdk2 inhibitory complex formation. As shown in Fig. 4C, p27 levels in Jab1 knockdown cells were higher when immunoprecipitated with Cdk2, indicating more complex formation. These data suggest that downregulation of Jab1 induces stabilization of p27, therefore increasing p27 levels and activity.

To investigate whether downregulation of Jab1 was involved in growth suppression, we first analyzed whether the siRNA-mediated Jab1 inhibition could recapitulate the tumor suppressor effect in NPC cell lines. The expression levels of Jab1 protein in Jab1 siRNA–transfected NPC cells were significantly less than those in the control siRNA-transfected cells (Fig. 4A). Moreover, the siRNA-mediated Jab1 knockdown also significantly inhibited the in vitro growth and colony formation of NPC cells (Fig. 5A and B). Notably, the growth-inhibitory effects of the Jab1 siRNA knockdown suggest that Jab1 targeting is a mechanism of tumor suppression in NPC cells. In addition, we tested the effect of knocking down Jab1 on apoptosis. Forty-eight hours after the siRNA was transfected, we used Annexin V and PI staining (Fig. 5C) or analysis of cleaved PARP and caspase-3 (Fig. 5D). However, there was no significant difference between the control and Jab1 siRNA–treated groups; thus, we cannot conclude that Jab1 knockdown initiates apoptosis under these conditions.
Jab1 depletion enhances the antitumor effects of cisplatin in NPC

Because cisplatin is the main treatment of NPC, we proposed to investigate that Jab1 may be involved in the antitumor effects of cisplatin. We first tested whether the inhibition of Jab1 enhances the antitumor effects of cisplatin by MTT assay. Data showed that 21%, 18%, and 7% of CNE1, CNE2, and HONE1 cells treated with control siRNA and cisplatin were inhibited, whereas, 38%, 38%, and 26% cells were inhibited by Jab1 siRNA and cisplatin, suggesting that Jab1-knockdown NPC cells were more sensitive to cisplatin than cells treated with control siRNA (Fig. 6A). To test whether Jab1 plays a role in anchorage-independent growth in response to cisplatin, which may reflect in vivo tumorigenicity, we conducted a colony formation assay in NPC cells treated with Jab1 siRNA and cisplatin. We found that the Jab1-knockdown cells had significant inhibition of colony formation (73% in CNE1, 90% in CNE2, and 66% in HONE1) compared with the control siRNA-treated cells (22% in CNE1, 60% in CNE2, and 14% in HONE1) in response to cisplatin (Fig. 6B).

We further examined whether Jab1 knockdown could enhance cisplatin-induced apoptosis in NPC cells. We found that cisplatin induced more apoptosis in Jab1-knockdown cells than that in control siRNA-treated cells, which is 1.4-fold in CNE1 and 4.7-fold in CNE2, respectively (Fig. 6C), when using Annexin V and PI staining, and 1.6-fold in CNE1 and 3.2-fold in

![Figure 5. Knockdown of Jab1 inhibits cell proliferation. A, NPC cells were transfected with Jab1 siRNA for 48 hours, and then cell growth was determined by an MTT assay. B, representative results of colony formation assays with NPC cells treated with control siRNA or 1 nmol/L (+) or 5 nmol/L (++) of Jab1 siRNA. Bottom, quantification of the relative number of colonies is shown. Data represent 3 independent experiments, mean ± SD. *P < 0.05; **P < 0.01. C, cells were treated with control siRNA or Jab1 siRNA for 48 hours and then stained with Annexin V and PI and measured by flow cytometry. Bottom, quantification of the percentage of living cells. Data represent 3 independent experiments, mean ± SD. D, cells were transfected with siRNA for 48 hours, and then cell lysates were immunoblotted with the indicated antibodies for apoptotic analysis. C, cleaved; Cont-si, control siRNA; Jab1-si, Jab1 siRNA.](image-url)
CNE2, when using Hoechst 33342 staining (Supplementary Fig. S2). However, there were no significant differences in apoptosis induced by control and Jab1 siRNA. Proteolytic cleavage of PARP and cleaved caspase-3 is the hallmark of apoptosis. Thus, we also examined the effect of Jab1 siRNA on the proteolytic cleavage of PARP and cleaved caspase-3 in response to cisplatin. Compared with the control siRNA-treated cells, cisplatin consistently induced more proteolytic cleavage of PARP (12% change in CNE1, 30% change in CNE2, and 6% change in HONE1) and cleaved caspase-3 (11% change in CNE1, 50% change in CNE2, and 6% change in HONE1).
change in CNE2, and 31% change in HONE1) in Jab1-knockdown cells (Fig. 6D).

Discussion
The molecular basis of NPC pathogenesis remains poorly defined, and this has hindered the development of new treatments. Therefore, identification of novel mechanisms involved in NPC oncogenesis is needed.

In this study, we identified the role of Jab1-mediated p27 degradation in NPC oncogenesis. First, we found that Jab1 was aberrantly expressed in either cytoplasm or nucleus NPC tissues compared with noncancerous infiltrated nasopharyngeal tissue. All (14 of 14) strong Jab1-positive patients died within 41 months whereas 11 of 31 weak or Jab1-negative patients were alive after 60 months. We also showed that Jab1 protein is overexpressed in NPC cell lines but not in paired normal keratinocyte cells. Our findings on p27 expression also confirm those of previous reports. For instance, Hwang and colleagues (11) reported that in patients with NPC, 47 of 69 cases expressed low levels of p27. In our study, 29 of 45 NPC cases (64%) expressed low or no nuclear p27, and 28 cases (62%) expressed low or no cytoplasmic p27. In addition, the nuclear cytoplasmic translocation of p27 has been observed in human tumors and is associated with poor survival (30–33). It was in agreement with our studies that patients with NPC with cytoplasmic-positive p27 had a poor survival.

Furthermore, cytoplasmic Jab1 expression in NPC tissues was inversely associated with nuclear p27 patterns ($R = -0.389, p = 0.008$) and correlated with cytoplasm p27 ($R = 0.328, P = 0.028$). Our finding of an inverse correlation between Jab1 and p27 expression levels in NPC is in agreement with our previously published results on breast cancer and pancreatic carcinomas (20, 24) and similar associations reported in other epithelial or lymphoid malignancies (21, 22). This suggests that Jab1 has a physiologic role in controlling p27 levels in NPCs. To our knowledge, this is the first study on the expression and function of Jab1 in NPCs to be reported.

Jab1 promotes cell proliferation by interacting directly with p27 and induces nuclear export and subsequent p27 degradation (34). p27 is a critical component of the cell-cycle machinery (16). As an inhibitor of cyclin E–Cdk2, p27 plays a pivotal role in controlling cell proliferation and therefore the cell’s entry into S-phase and exit from G1 phase during development and tumorigenesis (18, 35). We infected NPC cell lines with an inducible Jab1-expressing adenoviral vector and found that p27 levels were significantly reduced. These results are in agreement with those we previously obtained in breast cancer and pancreatic cell lines (20, 24). Furthermore, knockdown of endogenous Jab1 expression with siRNA in NPC cell lines led to a significant increase in total p27 levels and nuclear accumulation of p27 in the nucleus and p27–cyclin E–Cdk2 complex, substantial decreases in the number of S-phase cells, and increases in the number of G1 phase cells. In addition, a direct p27–Jab1 interaction was confirmed by coimmunoprecipitation assays and fluorescent microscopy. These results provide additional evidence that Jab1 controls p27 levels posttranslationally by accelerating p27 degradation in NPCs.

We also showed that loss of Jab1 resulted in growth inhibition in NPC cell lines. Similarly, a study by Supriatno and colleagues (36) showed that suppression of Jab1 inhibited the growth of human head and neck cancer cells (HIN and HSY cells) and delayed tumor growth in murine xenografts. It suggests that strong expression of Jab1 not only represents a prognostic marker for malignant transformation but also contributes to cancer cell proliferation and survival. Furthermore, we found that knockdown of Jab1 sensitized NPC cells to cisplatin chemotherapy. Jab1 siRNA–treated cells were more susceptible to apoptotic stimuli than the control siRNA-treated cells. Taken together, the data implicate Jab1 as a potential biomarker and further suggest exploration of whether decreasing Jab1 levels or function would overcome resistance to cisplatin chemotherapy in NPCs. The significance of these functions of Jab1 in NPCs needs to be explored in future mechanistic studies.

Overall, we found Jab1 to be overexpressed and inversely correlated with p27 expression levels in NPC. Jab1 overexpression associated with low p27 levels or other factors may contribute to uncontrolled cell-cycle progression and tumorigenesis. Depletion of Jab1 by siRNA, which leads to inhibition of proliferation, enhanced cisplatin activity in NPC. Therefore, modulation of the Jab1 gene product is a novel target for therapies in NPCs.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Acknowledgments
The authors thank Kate J. Newberry for editing the manuscript.

Grant Support
This work was supported by a fellowship from China Scholarship Council (2010638087 to Y. Pan), a grant from National Cancer Institute (R01-CA98053 to F.X. Claret); National Natural Science Foundation of China (81071837; 30670627); Natural Science Foundation of Guangdong Province, China (9251009901000005; 06021210); and Scientific and Technological Project of Guangdong, China (2008A030201009, 2010B050700016 to H. Yang).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received October 22, 2011; revised January 25, 2012; accepted January 31, 2012; published OnlineFirst February 20, 2012.

References
4. Yip WK, Leong VC, Abdullah MA, Yusoff S, Seow HF. Overexpression of phospho-Akt correlates with phosphorylation of EGF receptor,
Jab1/CSN5 Negatively Regulates p27 and Plays a Role in the Pathogenesis of Nasopharyngeal Carcinoma

Yunbao Pan, Qingxiu Zhang, Ling Tian, et al.


Updated version
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-11-3472

Supplementary Material
Access the most recent supplemental material at:
http://cancerres.aacrjournals.org/content/suppl/2012/02/20/0008-5472.CAN-11-3472.DC1

Cited articles
This article cites 36 articles, 11 of which you can access for free at:
http://cancerres.aacrjournals.org/content/72/7/1890.full#ref-list-1

Citing articles
This article has been cited by 1 HighWire-hosted articles. Access the articles at:
http://cancerres.aacrjournals.org/content/72/7/1890.full#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.