Intratracheal Administration of a Nanoparticle-Based Therapy with the Angiotensin II Type 2 Receptor Gene Attenuates Lung Cancer Growth

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Abstract
Targeted gene delivery, transfection efficiency, and toxicity concerns remain a challenge for effective gene therapy. In this study, we dimerized the HIV-1 TAT peptide and formulated a nanoparticle vector (dTAT NP) to leverage the efficiency of this cell-penetrating strategy for tumor-targeted gene delivery in the setting of intratracheal administration. Expression efficiency for dTAT NP–encapsulated luciferase or angiotensin II type 2 receptor (AT2R) plasmid DNA (pDNA) was evaluated in Lewis lung carcinoma (LLC) cells cultured in vitro or in vivo in orthotopic tumor grafts in syngeneic mice. In cell culture, dTAT NP was an effective pDNA transfection vector with negligible cytotoxicity. Transfection efficiency was further increased by addition of calcium and glucose to dTAT/pDNA NP. In orthotopic tumor grafts, immunohistochemical analysis confirmed that dTAT NP successfully delivered pDNA to the tumor, where it was expressed primarily in tumor cells along with the bronchial epithelium. Notably, gene expression in tumor tissues persisted at least 14 days after intratracheal administration. Moreover, bolus administration of dTAT NP–encapsulated AT2R or TNF-related apoptosis-inducing ligand (TRAIL) pDNA markedly attenuated tumor growth. Taken together, our findings offer a preclinical proof-of-concept for a novel gene delivery system that offers an effective intratracheal strategy for administering lung cancer gene therapy. Cancer Res; 72(8); 2057–67. © 2012 AACR.
expressed in the mesenchyme of the fetus and to a limited extent in adult tissues (28). AT2R is known to inhibit cell proliferation and stimulate apoptosis in cardiovascular and neuronal tissues in vitro (29). Our previous studies revealed that AT2R deficiency significantly altered chemical carcinogen–induced tumorigenesis in mouse colon (30) and lung (31). A recent study indicates that host AT2R deficiency stimulates the growth of murine pancreatic carcinoma grafts (32). These results suggest that AT2R expression plays an important role in tumor growth.

TRAIL is a naturally occurring cytokine that acts by binding as a homotrimer to death receptor (DR)–4 or -5 and recruiting an adaptor receptor, such as FADD or caspase-8. Because activated caspase-8, in turn, activates a caspase pathway that induces extrinsic apoptotic cell death (33), TRAIL is known to be a strong anticancer gene candidate (34). In fact, TRAIL gene therapy has been tested in multiple mouse cancer models with success (35, 36). Targeted gene delivery to cancer tissue should reduce side effects in healthy tissue and enable TRAIL as a more toxic therapeutic for lung cancer. dTAT NPs are a realistic gene delivery system, and the dTAT solution can be detected primarily in the tumor tissues of the lung (dTAT/pLUC or dTAT/pAT2R) administered via intratracheal spray. For in vitro studies. For in vivo mouse studies, 25 μL of 10% glucose was added to the dTAT/pDNA solution. Finally, 15 μL of 0.3 mol/L CaCl2 was added to the stabilized solution. The final solution was mixed vigorously by pipette. Before use, dTAT/pDNA NPs were allowed to equilibrate for 20 minutes at 4°C.

Preparation of dTAT/pDNA NP

The dTAT/pDNA NPs were prepared by mixing via pipette 10 μL pDNA (0.1 μg/pDNA/μL) and 15 μL dTAT (1 μg/dTAT/μL) solutions. The resultant dTAT/pDNA solution was stabilized by adding either 25 μL of 10% glucose, 0.2 mol/L NaCl, or 0.2 mol/L KCl solution for in vitro studies. For in vivo mouse studies, 25 μL of 10% glucose was added to the dTAT/pDNA solution. Finally, 15 μL of 0.3 mol/L CaCl2 was added to the stabilized solution. The final solution was mixed vigorously by pipette. Before use, dTAT/pDNA NPs were allowed to equilibrate for 20 minutes at 4°C.

In vitro cell transfection studies

LLC cells were trypanosed, counted, and diluted to a concentration of approximately 80,000 cells/mL. Then, 0.1 mL of that dilution was added to each well of a 96-well plate, and the cells were incubated in a humidified atmosphere at 5% CO2 at 37°C for 24 hours. Immediately before transfection, cells were washed once with PBS and 100 μL sample (20% of NPs to 80% of serum-free cell culture medium) was added to each well. Cells were incubated with the NP for 5 hours. The transfection agent was then aspirated and 100 μL of fresh serum-containing medium was added, followed by further incubation. Luciferase expression was determined at 24 hours after transfection by the Luciferase Assay System from Promega according to the manufacturer’s recommended protocol. The light units were normalized against protein concentration in the cell extracts, which was measured using the BCA Protein Assay (Thermo Scientific). Transfection results were expressed as relative light units (RLU) per mg of cellular protein.

Assessment of cytotoxicity (MTS assay)

Cytotoxicity of polymers was determined by the CellTitre 96 Aqueous Cell Proliferation Assay (Promega). LLC cells were grown as described in the transfection experiments. Cells were treated with the samples for approximately 24 hours. The medium was then removed and replaced with a mixture of 100 μL fresh culture medium and 20 μL MTS reagent solution. The cells were incubated for 3 hours at 37°C in the 5% CO2 incubator. The absorbance of each well was measured at 490 nm using a microtiter plate reader (SpectraMax, M25; Molecular Devices Corp.) to determine cell viability.

Cell viability analysis

The MTT assay was conducted to examine the effect on LLC cell proliferation in vitro of dTAT/pXT2R or of high
concentration glucose, NaCl, or KCl in the dTAT/pDNA solution. In brief, 700 LLC cells were seeded in 96-well plates 24 hours before the addition of dTAT/pAT2R, glucose, or salt solution. The cancer cells were treated with dTAT/pAT2R alone, dTAT/pAT2R with additional glucose or salt solution, or glucose or salt solution alone in serum-free DMEM (0.25 or 0.5 μg of dTAT/pDNA per well) at 37°C for 5 hours, and then the medium was replaced with DMEM containing 10% FBS. After 48-hour incubation at 37°C, the MTT assay was carried out as previously described (37). The same procedure was used to examine the potential adverse effect of dTAT alone (1.87 or 0.5 μg/mL) on the viability of LLC, A549, and BEAS-2B cells. In this study, the MTT assay was conducted after 1, 3, or 5 days of incubation in serum-containing DMEM or BEBM medium.

**Gene expression analysis using real-time PCR**

Transfection of pAT2R into LLC cells was confirmed by real-time PCR. About 5,000 LLC cells were seeded in 24-well plates 24 hours before the addition of dTAT/pAT2R. The cancer cells were treated with dTAT/pAT2R in serum-free DMEM (1 or 2 μg of dTAT/pDNA per well) at 37°C for 5 hours and then were allowed to grow in DMEM containing 10% FBS at 37°C for 48 hours. Then, gene expression was analyzed as previously described (37). AT2R primers were 5'-ACTTCGGGCTTGTGAACATC-3' (forward), and 5'-TAAAT-CAGCCACACCGAGGT-3' (reverse); 18S ribosome RNA primers were 5'-TGGCGTCAGAGCAAGCAGA-3' (forward) and 5'-GAGGTTCCGAAGACAGTCA-3' (reverse).

**In vivo studies**

All animal experiments were done under strict adherence with Kansas State University (Manhattan, KS) Institutional Animal Care and Use Committee protocols. Wild-type female C57BL/6 mice obtained from the Jackson Laboratory were housed in a clean facility and held for 10 days to acclimatize. **Experimental design I:** LLC cells were seeded at 30,000 cells per well in a 24-well plate and incubated 24 hours in 10% FBS containing medium. Medium was then changed to medium containing either the dTAT/pLUC (1 μg of luciferase pDNA/well) or blank dTAT and incubated for 5 hours. After changing the media back to NP-free, 10% FBS containing media, cells were further incubated. Five days after treatment, 100, 1,000, or 10,000 cells were subcutaneously injected into the backs of the mice. At various time points up to 1 week, animals were imaged with a Caliper IVIS Lumina II biophotonic imager. Images were collected using a 6-minute exposure time.

In Experimental design II and III, each mouse was injected via the tail vein with 2 × 10^6 LLC cells suspended in 200 μL of PBS. **Experimental design II:** Seven days after LLC cell injection, these mice were injected intratracheally using an intratracheal sprayer (Pern-Century Inc.) with 50 μL of PBS (n = 12) or 50 μL of dTAT/pLUC (containing 0.7 μg pDNA, n = 12). On days 3, 7, 10, and 14 after intratracheal administration of the dTAT/pLUC, mice were sacrificed and lungs were dissected for histologic analysis of tumor multiplicity and size. In addition, luciferase expression in the lung was analyzed immunohistochemically. **Experimental design III:** On day 7 after LLC injection, these mice were intratracheally treated using the sprayer with 50 μL of PBS (n = 6), dTAT alone (n = 5), dTAT/pAT2R (n = 5), or dTAT/pTRAIL (n = 5). After sacrifice on day 15 after LLC inoculation, lungs were fixed in 10% buffered formalin and used for histologic and immunohistochemical analysis.

**Histologic analysis**

Fixed lung tissues were sectioned at 4 μm and stained with hematoxylin and eosin (H&E) for histologic examination. Quantitative evaluation of tumor nodules in the lungs was conducted as previously described (38).

**Immunohistochemistry for luciferase, Ki-67, and AT2R in the tumor nodules**

After deparaffinization and rehydration of the tissue sections, immunohistochemistry was conducted with antibodies to luciferase (1:1,000; Novus Biologicals), Ki-67 (1:100; Abcam), and AT2R (1:100; Abcam), followed by incubation with biotin-conjugated antibodies against goat IgG or rabbit IgG (1:50; Vector Laboratories), reacted with the avidin–biotin peroxidase complex reagent (Vector Laboratories), visualized with 3,3'-diaminobenzidine tetrahydrochloride (Sigma). To determine the Ki-67 labeling (proliferative) index, 10 nodules were selected randomly by light microscopy, and the number of Ki-67-positive cells in each area was counted. The index was assessed as the percentage of Ki-67-positive cells per tumor cells.

**TUNEL assay**

Terminal deoxynucleotidyl transferase–mediated dUTP nick end labeling (TUNEL) assay was conducted using the DeadEnd Colorimetric TUNEL System (Promega) as previously described (38). The fold change was calculated by dividing the percentage of TUNEL-positive tumor cells in the treated tumors by those in untreated tumors.

**Statistical analysis**

All data are reported as mean ± SE. Statistical significance was assessed by one-way ANOVA. Group comparisons were deemed significant for 2-tailed P values below 0.05.

**Results**

**dTAT/pLUC NP caused efficient gene transfection with low cytotoxicity in vitro**

In studies reported here, dTAT and pLUC complexes were thoroughly mixed by pipetting, and CaCl_2 was added to decrease the NP size through “soft” cross-links of dTAT and pDNA (26). Here, the reduction in the size of dTAT/pLUC NP likely led to some of the noted increase in transfection. A CaCl_2 concentration of 69.2 mmol/L consistently produced small dTAT/pLUC NP caused efficient gene transfection with low cytotoxicity in vitro.
dTAT assay. LLC cells were incubated with up to 10 mg/mL of dTAT or PEI for approximately 24 hours. Cytotoxicity profiles of dTAT peptides showed moderate cytotoxicity (IC$_{50}$/C$_{24}$ 4,075 mg/mL; Fig. 1A), whereas branched PEI was strongly cytotoxic (IC$_{50}$/C$_{24}$ 28 mg/mL). Although the cell viability of LLC, A549, and BEAS-2B was slightly decreased by treatment with dTAT alone solution (1.86 or 3.72 mg/mL) at days 3 and 5, there was no statistically significant difference among the groups (Supplementary Fig. S1).

Luciferase gene expression was measured 48 hours after transfection to study the ability of dTAT NP to transfect LLC cells (Fig. 1B). Different N/P ratios of the dTAT or branched PEI (N/P 10) NP were studied using different concentrations of CaCl$_2$ (0, 17.3, 34.6, and 69.2 mmol/L) as a condensing agent after NP formation. Most dTAT NP showed a high level of gene expression at and above 34.6 mmol/L of added CaCl$_2$ for the various N/P ratios when compared with branched PEI, which had excellent transfection efficiency only in the absence of CaCl$_2$. The study revealed that the highest transfection efficiency by dTAT was achieved at 69.2 mmol/L, CaCl$_2$ and an N/P ratio of 33. It is important to note that gene expression was not detectable for dTAT/pLuc at CaCl$_2$ levels up to 17.3 mmol/L.

dTAT-based AT2R gene transfection attenuated growth of lung cancer cells in vitro

To examine agents that effectively stabilize dTAT/pDNA NP and cause effective transfection, dTAT/pAT2R solution was mixed with glucose, KCl or NaCl solution before condensing complexes with CaCl$_2$, and the efficacy of AT2R expression and cell viability was evaluated. Real-time PCR revealed that all of the agents caused effective DNA transfection at 2 µg dTAT/pAT2R per well (Fig. 2A). As shown in Fig. 2B, dTAT/pAT2R transfection significantly attenuated viability of LLC cells compared with dTAT alone. Among the agents mixed with the dTAT/pAT2R solution, the glucose-stabilized dTAT/pAT2R decreased cell viability most effectively, whereas treatment using dTAT alone slightly inhibited tumor cell growth (Fig. 2B). Incubation of LLC cells with the solution containing glucose, NaCl, or KCl alone without dTAT or pDNA had no adverse effect on cell viability (Supplementary Fig. S2). Taken together, these results suggest that glucose was the most effective agent in stabilizing dTAT/pDNA NP and mediating effective transfection.
effective agent for dTAT/pDNA NP transfection but by itself had no effect on tumor growth in vitro.

**In vivo imaging easily detects dTAT/pLUC NP–transfected lung carcinoma cells in mice**

Because the magnitude of gene expression is one key to successful gene therapy, gene expression was evaluated using an in vivo imaging system after dTAT/pLUC-transfected LLC cells were transplanted into the backs of mice. In this experiment, luciferase expression was detected by in vivo imaging 24 hours after transplantation with a minimum number of 100 luciferase-transfected cells. Luciferase expression by dTAT/pLUC was easily detectable for at least 7 days after transplantation when more than 1,000 luciferase-transfected cells were transplanted (Fig. 3). These results clearly indicate that dTAT NP–based gene transfection would be detectable for an in vivo mouse study.

**Administration of dTAT/pLUC NP via intratracheal spray caused luciferase expression preferentially in lung tumor cells**

The effectiveness of intratracheally administered dTAT/pLUC NPs, the luciferase expression sites, and the effect on tumor growth were quantitatively determined using LLC lung tumor–bearing mice. Immunohistochemical detection of luciferase expression in the lung indicated that the primary expression sites are tumor cells and bronchioloalveolar epithelium (Fig. 4A). Strong luciferase expression was detected at 3 days after the intratracheal spray of dTAT/pLUC; this expression lasted until 14 days after the spray without losing much intensity, indicating that dTAT NP–based gene transfection is effective in vivo. In addition, this dTAT/pLUC transfection via intratracheal spray did not show any effects on tumor growth (Fig. 4B). Histologic examination of tumors in H&E-stained sections showed a large number of LLC tumor nodules in mouse lungs treated with either dTAT/pLUC or PBS. The average number of tumors per view and the size of the tumors in both groups were not significantly different between the 2 groups (Fig. 4B).

**Administration of dTAT/pAT2R or pTRAIL NP via intratracheal spray caused significant growth attenuation of lung tumors**

Because dTAT NP–based in vivo pDNA transfection was very effective and intratracheal spray was shown to deliver the dTAT/pLUC into the lung tumors effectively, the effect of dTAT NP–based in vivo transfection of endogenous apoptosis-inducer genes, AT2R and TRAIL, was examined using orthotopic LLC lung tumor–bearing mice. In this study, LLC cells (2 × 10⁶) were inoculated via the tail vein. One week after cancer cell inoculation, during which time a preliminary study revealed that LLC grafts had started growing as microtumors, dTAT/pAT2R or dTAT/pTRAIL (1 μg DNA/50 μL solution) was sprayed once intratracheally. These treatments significantly decreased lung tumor prevalence as compared with PBS- or dTAT/pLUC-treated mice (Fig. 5A). The dTAT/pAT2R and pTRAIL also significantly decreased the macroscopic lung tumor multiplicity (Fig. 5A). Histologic examination of tumors in H&E-stained sections clearly showed a large number and size of LLC tumor nodules in mouse lungs treated with PBS, whereas only a small number of tumors were detected in dTAT/pAT2R-treated mouse lungs (Fig. 5B–D). As expected, dTAT/pTRAIL attenuated both tumor size and the number of tumors significantly (Fig. 5). Interestingly, administering dTAT alone also attenuated tumor multiplicity and decreased tumor size (Fig. 5C). The AT2R expression site in the lung was determined immunohistochemically. As shown in Fig. 6, an intense immunoreactivity for AT2R was observed in the tumor cells in the dTAT/pAT2R-treated group but not in other groups (Fig. 6). These microscopic observations suggested that an intratracheal dTAT/pAT2R spray significantly attenuated lung tumor growth by expressing AT2R in the tumor cells. Accordingly, these data support the assertion that intratracheal spray of dTAT/pAT2R is an effective modality for targeted lung cancer gene therapy.

**Determination of cell proliferation and apoptotic index in the tumors**

To evaluate the effect of the treatments on the proliferative and apoptotic activities of tumor cells, numbers of Ki-67- and TUNEL-positive cells in tumor tissues were determined. Immunohistochemical analysis revealed that while the number of Ki-67-positive cells was slightly decreased in the
dTAT/pAT2R- or pTRAIL-treated tumors, this difference was not significant (Fig. 7A and B). In contrast, both dTAT/pAT2R and dTAT/pTRAIL increased apoptosis. The TUNEL-positive cells were significantly increased in tumors of mice treated with dTAT/pAT2R and pTRAIL relative to those treated with PBS or dTAT (Fig. 7A and C). Treatment with dTAT alone did not significantly alter either cell proliferation or apoptosis. These results indicate that treatment with dTAT/pAT2R increased apoptosis of tumor cells and thus decreased tumor multiplicity and the tumor size.

Discussion

The primary objectives of this study were to examine the efficacy of dTAT as a vector, to determine whether pDNA can be distributed to lung tumor cells and cause robust expression, and to evaluate the effectiveness of dTAT NP–based delivery of AT2R or TRAIL pDNA, as AT2R or TRAIL overexpression is known to attenuate tumor cell growth (34, 36, 37). In addition, efficiency of intratracheal spray of dTAT/pDNA was evaluated using luciferase, AT2R, and TRAIL pDNA. Results indicated that a bolus intratracheal spray of dTAT/pDNA caused robust gene expression, primarily in lung tumor cells. Expression of AT2R and TRAIL significantly attenuated tumor growth. Therefore, the present study introduces an effective in vivo gene delivery system using a cationic peptide, dTAT, for lung cancer therapy.

The first study indicated that dTAT NP–based transfection was comparable with PEI polyplexes (Fig. 1A). Our data indicate that under the conditions tested, dTAT did not show
any practical acute cytotoxicity in vitro until nearly 5 mg/mL concentration for 24 hours, whereas PEI showed strong cytotoxicity at much lower concentrations (Fig. 1A). Evaluation of the cytotoxicity of dTAT alone using other cell types, such as human lung bronchial epithelial cells and human lung adenocarcinoma cells, also revealed similar low cytotoxicity (Supplementary Fig. S1). Glucose or salt solution addition also did not affect viability in these other cell types (Supplementary Fig. S2). The low cytotoxicity was also shown in vivo after intratracheal application, in which all mice receiving dTAT alone or dTAT/pDNA survived during the experimental period and did not show any histologically detectable abnormality or acute inflammatory reaction (data not shown). The low cytotoxicity of the dTAT peptide is in agreement with previous reports of TAT and other similar cell-penetrating peptides (26, 39). Furthermore, our recent dTAT dose-escalation toxicity study in mice with intravenous administration (39) suggests that this dTAT NP–based delivery system is minimally toxic. Accordingly, it is concluded that dTAT NP potentially represents an efficient and safe gene transfection vector, worthy of further in vivo studies.

The second study clarified that addition of glucose, KCl or NaCl to the dTAT/pDNA mixture caused equally effective DNA transfection, but addition of glucose caused the most significant attenuation of cell growth (Fig. 2). Although the first experiment clearly indicated that dTAT alone treatment...
was significantly less cytotoxic than PEI alone, treating cells with dTAT for 2 days attenuated cell viability (Fig. 2B). Growth inhibition by treatment with dTAT alone is consistent with the report that the TAT peptide itself can induce cell death (40). The present study clearly indicated that the cell growth attenuation effect of dTAT is amplified when the dTAT NP is prepared with plasmids encoding an apoptosis-inducer gene such as AT2R, suggesting that dTAT NP can effectively transfect genes in tumor cells and induce tumor cell death.

Gene therapy–dependent tumor growth inhibition requires sustained and robust transgene expression to be effective (26). Accordingly, the intensity and duration of gene expression by dTAT NP transfection were determined by in vivo imaging after transplanting tumor cells transfected in vitro with dTAT/pLUC. Luciferase expression was detectable at days 1 and 3 after transplantation of only 100 dTAT/pLUC-transfected cells. Luciferase expression was detectable for a week after the injection of 1,000 cells or more (Fig. 3). These results indicated that dTAT/pDNA caused efficient transfection both in vitro and in vivo, and the duration of the strong expression is significantly long to prompt further study.

To examine the in vivo gene transfection efficiency of dTAT NPs, the expression of luciferase in the lung was monitored immunohistochemically for 14 days after administering dTAT/pLUC via intratracheal spray in LLC tumor–bearing mice. A single spray of dTAT/pLUC caused robust luciferase expression, primarily in the tumor cells and bronchial epithelium, for at least 14 days (Fig. 4A). These studies proved that dTAT NPs cause long-lasting, robust gene expression in vivo. Hence, the current study suggested that gene transfection using dTAT NPs was an effective strategy for in vivo gene therapy and is potentially selective for rapidly dividing lung cancer cells.

In the next study, delivery of the endogenous apoptosis-inducer gene AT2R was examined using LLC tumor–bearing mice. pTRAIL was used as a positive control. As expected, dTAT/pTRAIL attenuated tumor growth macroscopically and microscopically by inducing apoptosis (Figs. 5 and 7), indicating that dTAT NP gene transfection was effective. In the dTAT/pAT2R treatment group, expression of AT2R was detected primarily in the tumor cells, which apparently led to the attenuation of the tumor growth. The degree of cell proliferation and apoptosis in the tumors suggested that bolus intratracheal spray of dTAT/pAT2R probably lowered tumor burden by inducing apoptosis of tumor cells rather than by attenuating cell proliferation (Fig. 7). These results are consistent with previous reports that AT2R is a strong apoptosis inducer, attenuating growth of various cell types (41–43) including human lung cancer cells (37). Accordingly, induction of AT2R overexpression is a potential treatment scheme for lung cancer.

A single intratracheal spray of dTAT alone also attenuated tumor growth significantly (Fig. 5) as compared with the PBS controls. In support of this result, it has been shown that HIV-TAT peptide can directly attenuate growth of polyamine-deprived cancer cells (40). Therefore, tumor growth attenuation in the lungs of LLC graft–bearing mice by dTAT alone may partly be caused by the direct tumoricidal effect of dTAT on the tumor cells. However, all mice receiving dTAT alone survived until sacrificed and showed no abnormal clinical signs or histologic abnormalities in the normal areas of the lung. These results suggest that the dTAT-dependent cell growth attenuation appears to be limited to the tumor cells. As an alternative explanation, the dTAT peptide–dependent tumor attenuation may also be caused by a secondary effect of the dTAT peptide on the tumor microenvironment. This speculation may be supported by the immunohistochemical observations that dTAT NP–dependent luciferase or AT2R expression was recognized in the alveolar epithelium (Figs. 4 and 6) and alveolar macrophages (data not shown). This observation suggests that dTAT peptides are taken up by various types of cells in the lung, although gene expression levels are weaker than those in tumor cells. Because the tumor microenvironment is an important factor for tumor growth regulation (44–46), it is possible that dTAT modulates the tumor microenvironment toward conditions less favorable to tumor growth. Although further studies are required to better understand the effect of dTAT on tumor growth, this dTAT-dependent tumor suppression may be a beneficial adjuvant property of these therapeutic nanoparticles.

In conclusion, although further studies are required to substantiate the in vivo safety of dTAT NPs by formal
multispecies toxicity and pharmacokinetic studies, our data indicate that dTAT NP could be a safe and effective in vivo gene transfection tool. The present study provides clear evidence that intratracheal administration of dTAT NP–based therapeutic gene delivery causes strong gene expression preferentially in tumor cells. A bolus intratracheal administration of dTAT/pAT2R or pTRAIL NP significantly attenuated the growth of fast-growing LLC tumors, suggesting that dTAT NP–based gene therapy is effective and useful for lung cancer treatment. AT2R is a potentially useful gene for lung cancer therapy.

Disclosure of Potential Conflicts of Interest
C. Berkland has Commercial Research Grant and Ownership Interest (including patents) from Savara Pharmaceuticals. No potential conflicts of interest were disclosed by the other authors.

Acknowledgments
The authors thank Marla Pyle and Garret Seiler (Department of Anatomy and Physiology, Kansas State University) for critical reading and constructive comments during the preparation of the manuscript.

Grant Support
This work was supported in part by the Kansas State University (KSU) Terry C. Johnson Center for Basic Cancer Research, KSU College of Veterinary Medicine.
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Received November 2, 2011; revised January 14, 2012; accepted February 13, 2012. Published OnlineFirst March 2, 2012.

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Dean’s fund, Kansas State Legislative Appropriation, The Institute for Advancing Medical Innovation at The University of Kansas and NIH grants P20 RR016786, P20 RR016745, 5P20RR015563 and Kansas Bioscience Authority collaborative cancer research grant. In addition, Savara Pharmaceuticals is gratefully acknowledged for providing financial support for portions of this work.

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