NF-κB Activity Regulates Mesenchymal Stem Cell Accumulation at Tumor Sites

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Abstract
Mesenchymal stem cells (MSC) accumulate at tumor sites when injected into tumor-bearing mice, perhaps offering cellular vectors for cancer-targeted gene therapy. However, the molecular mechanisms involved in MSC targeting the tumors are presently little understood. We focused on MSC–endothelial cell (EC) adhesion following TNF-α stimulation in an attempt to elucidate these mechanisms. Interestingly, stimulation of MSCs with TNF-α enhanced the adhesion of MSCs to endothelial cells in vitro. This adhesion was partially inhibited by blocking antibodies against vascular cell adhesion molecule-1 (VCAM-1) and very late antigen-4 (VLA-4). It is well known that TNF-α induces VCAM-1 expression via the NF-κB signaling pathway. Parthenolide has an anti-inflammatory activity and suppressed NF-κB activity by inhibition of IκB phosphorylation after TNF-α stimulation and strongly inhibited TNF-α–induced VCAM-1 expression on MSCs. In vivo imaging using luciferase–expressing MSCs revealed that the bioluminescent signal gradually increased at tumor sites in mice injected with untreated MSCs. In contrast, we observed very weak signals at tumor sites in mice injected with parthenolide-treated MSCs. Our results suggest that NF-κB activity regulates MSC accumulation at tumors, by inducing VCAM-1 and thereby its interaction with tumor vessel endothelial cells. These findings have implications for the ongoing development of efficient MSC-based gene therapies for cancer treatment. Cancer Res 73(1); 364–72. ©2012 AACR.

Introduction
Mesenchymal stem cells (MSC) are nonhematopoietic stem cells with high-proliferative potency and have the ability to differentiate into multiple lineages. They are detected in several adult and fetal tissues, including bone marrow, adipose tissue, and umbilical cord blood. MSCs have generated a great deal of interest in their potential use in regenerative medicine due to their ability to migrate to damaged tissues and to produce cytokines. Furthermore, MSCs can be easily genetically modified with viral vectors to be used as novel cellular vehicles in gene therapy protocols. MSCs are also used to treat severe acute GVHD, because they accumulate at inflammatory lesions and have immunomodulatory activity.

Interestingly, recent studies indicated that MSCs also have the ability to accumulate in tumors. Therefore, they can be used as cellular vehicles for cancer-targeted gene therapy. Intravenous injection of engineered MSCs expressing IFN-β was reported to inhibit the growth of melanoma pulmonary metastasis (1) and breast cancer (2) in mice and also prolonged the survival of mice with glioma xenografts (3). Furthermore, interleukin (IL)-12, which improves immune surveillance against cancer cells (4), and chemokine CX3CL1 (fractalkine), which is able to activate T cells and natural killer (NK) cells (5), were used as therapeutic molecules. We have also shown that retrovirus vector–producing MSCs also effectively inhibit tumor growth (6). In this context, treatment has been developed using retroviral vectors expressing the thymidine kinase of herpes simplex virus combined with the produg ganciclovir.

The ability of MSCs to specifically localize the multiple tumors, makes them extremely attractive for targeted cancer therapy. The most likely cause of preferential migration was considered to be the release of chemotactic gradients from tumor tissues. MSCs have a variety of chemokine and cytokine receptors and respond functionally to ligands in vitro. Tumors are known to produce a large amount of chemokines and cytokines, which could serve as ligands for the receptors on MSCs (7). Therefore, the mechanism of MSC accumulation at the site of tumors seems to be based on their migratory ability. Nevertheless, although various growth factors and chemokines, such as platelet-derived growth factor (PDGF), hepatocyte growth factor (HGF), and stromal cell–derived factor-1α (SDF-1α) may be involved, the detailed molecular mechanisms of MSC accumulation at tumors are poorly understood.

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In the present study, we focused on MSC–endothelial cell (EC) adhesion following TNF-α stimulation in an attempt to elucidate the mechanism of MSC accumulation at tumors.

Materials and Methods

**Cell culture**

Bone marrow–derived human MSCs (Lonza Walkersville, Inc.) were cultured in mesenPRO RS medium (Invitrogen). HEK293-derived AD-293 cells (Stratagene), human embryonic fibroblasts WI-38 (RIKEN BRC), human colon adenocarcinoma cell lines SW480 (Cell Resource Center for Biomedical Research Institute of Development, Aging and Cancer, Tohoku University, Miyagi, Japan), and SW480/RFP that were generated by transfection of PacI-digested pAdHM4-CAGFP were grown in RPMI medium (Invitrogen) supplemented with 10% FBS, 100 U/mL penicillin, and 100 μg/mL streptomycin (P/S). Human endothelial progenitor cells (ApproCell Inc.) were cultured in endothelial progenitor cells grown medium (ApproCell Inc.). Human colon adenocarcinoma cell lines Colo205 (Cell Resource Center for Biomedical Research Institute of Development, Aging and Cancer Tohoku University) and Colo205/RFP that were generated by transduction of SW480 with red fluorescent protein-expressing retrovirus vectors (RV-RFP), were grown in Dulbecco’s Modified Eagle’s Medium (DMEM)/F-12 medium (Invitrogen) supplemented with 10% FBS, 100 U/mL penicillin, and 100 μg/mL streptomycin (P/S). Human endothelial progenitor cells (ApproCell Inc.) were cultured in endothelial progenitor cells grown medium (ApproCell Inc.). Human colon adenocarcinoma cell lines Colo205 (Cell Resource Center for Biomedical Research Institute of Development, Aging and Cancer Tohoku University) and Colo205/RFP that was generated by transduction with RV-RFP, were grown in RPMI medium (Invitrogen) supplemented with FBS and P/S. All cultures were kept in an incubator at 37°C and 5% CO₂.

**Adenoviral vectors**

Adenoviral vectors expressing a GFP were constructed by an improved in vitro ligation method (8, 9). The shuttle plasmid pHMCA5-GFP contains a CA promoter (a β-actin promoter/CMV enhancer with a β-actin intron), GFP gene, and a bovine growth hormone (BGH) polyadenylation signal, all of which are flanked by I-CeuI and PI-SceI restriction sites. I-CeuI/PI-SceI-digested pHMCA5-GFP was ligated with I-CeuI/PI-SceI-digested pAdHM4, resulting in pAdHM4-CAGFP. pAdHM41-K7-CAGFP was constructed by ligation of I-CeuI/PI-SceI-digested pHMCA5-GFP with I-CeuI/PI-SceI-digested pAdHM41-K7 (10). Viruses (Ad5-GFP and AdK7-GFP) were generated by transfection of PacI-digested pAdHM4-CAGFP and pAdHM41-K7-CAGFP, respectively, into AD-293 cells with SuperFect (Qiagen) according to the manufacturer’s instructions. Each virus was purified by CsCl₂ step gradient ultracentrifugation followed by CsCl₂ linear gradient ultracentrifugation. Virus particles and biologic titers of each vector preparation were determined as described by Mittereder and colleagues (11). We also created Ad vectors expressing luciferase (Luc) using the shuttle plasmid pHMCA5-Luc, which contains the Luc gene derived from pELuc-test (Toyobo Co. Ltd.). MSCs and fibroblasts were seeded in culture plates or flasks at a density of 1 × 10³ cells/cm², and the next day the cells were treated with each adenovirus vector for 1.5 hours. The medium containing the vectors was removed and replaced with fresh medium.

**Animal models**

All animal experiments were approved by the Jichi Medical University (Tochigi, Japan) ethics committee and carried out in accordance with the NIH Guide for the Care and Use of Laboratory Animals. To create tumor-bearing mice, SW480/RFP cells (3 × 10⁶) were subcutaneously inoculated into 4- to 6-week-old male Balb/c nu/nu mice (Clea Japan Inc.). The mice were used for experiments 7 days after inoculation.

**Immunohistochemistry**

Cultured MSCs and fibroblasts were transduced with AdK7-GFP at a concentration of 3,000 virus particles per cell (vp/cell). Two days after transduction, cells were injected into the left ventricular cavities (1 × 10⁶, day 0) of tumor-bearing mice. Mice were sacrificed on day 4, and 7-μm serial cryosections from frozen tissues were processed. Immunohistochemistry was conducted with fluorescein isothiocyanate (FITC)-conjugated anti-GFP antibody (ab6662; Abcam Inc.) on tumor cryosections to detect MSCs or fibroblasts. Nuclei were stained with 4′,6-diamidino-2-phenylindole (DAPI; Vector Laboratories, Inc.). Images were obtained with a fluorescence microscope (BZ-9000; Keyence). SW480/RFP cells (3 × 10⁶) were subcutaneously inoculated into 4- to 6-week-old male Balb/c nu/nu mice. Mice were sacrificed on day 11, serial sections from tumor tissues were processed. Immunohistochemistry was conducted with anti-mouse CD34 monoclonal antibody (MEC14.7; GeneTex Inc.) on tumor section to detect tumor blood vessels. Histofine Simple Stain Mouse MAX PO (Nichirei Biosciences, Inc.) was used as a horseradish peroxidase–conjugated secondary antibody, and 3,3′-diaminobenzidine (DAB) solution was used for brown color development. Sections were then counterstained with Hematoxylin (Wako Pure Chemical Industries, Ltd.). Images were obtained with a fluorescence microscope (BZ-9000).

**In vivo imaging of homing ability to tumors**

Cultured MSCs and fibroblasts were transduced with AdK7-Luc at a concentration of 3,000 and 680 vp/cell, respectively. Two days after transduction, cells were injected into the left ventricular cavities (1 × 10⁶, day 0) of tumor-bearing mice, and then optical bioluminescence imaging was conducted to periodically trace the cells using an in vivo imaging system (IVIS; Xenogen). To detect bioluminescence from MSCs or fibroblasts, the reporter substrate v-luciferin (Ieda Chemical Co., Ltd.) was injected into the mouse peritoneum (75 mg/kg body weight) for scanning. The luminescent intensity at tumor sites was analyzed using Living Image software (Xenogen).

**In vitro migration assays**

Cultured MSCs or fibroblasts were serum-starved for 12 hours. One hundred microliters of tumor conditioned medium (CM), or serum-free medium supplemented with PDGF-BB (10 ng/mL), HGF (30 ng/mL), fibroblast growth factor-β (FGF-β; 20 ng/mL), SDF-1α (150 ng/mL), VEGF-A (25 ng/mL), or monocyte chemoattractant protein-1 (MCP-1; 100 ng/mL) was added to the lower wells of migration chambers (8-μm pore size; Neuro Probe, Inc.). MSCs or fibroblasts (4 × 10³) were added to the upper wells. All recombinant proteins were purchased from R&D systems Inc.. Medium alone (DMEM/F-12) was used as a negative control and treatment with 30% FBS was the positive control. After incubation for 24 hours at...
37°C, cells were labeled with CyQUANT NF dye, and cells attached to the lower surface of the filters were detached with trypsin. Fluorescent intensity was measured using a fluoroscan, and the number of adherent cells was quantified using a standard curve constructed by a known number of cells.

**Flow cytometric analysis of adhesion molecules**

Cultured MSCs, fibroblasts or endothelial cells were stimulated with TNF-α and harvested by trypsinization. Cell aliquots were incubated with FITC-conjugated monoclonal antibodies (BD) against vascular cell adhesion molecule-1 (VCAM-1), CD49d, CD29 (Integrin-B1), and analyzed by flow cytometry (FACScan; BD Biosciences). For each analysis, an aliquot of cells was also stained with isotype control immunoglobulin G (IgG)–conjugated to FITC as a negative control.

**Assay for TNF-α produced in tumor-bearing mice**

SW480/RFP (3 × 10⁶) cells were subcutaneously inoculated into nude mice. Seven days after inoculation, mice were anesthetized with an overdose of isoflurane inhalation. The blood was collected and allowed to coagulate overnight on ice. After centrifugation of the samples (2,000 × g, 30 minutes, 4°C), the serum was removed and stored at −70°C. Tumor, spleen, and liver tissues were homogenized in 1.5 mL of α-minimum essential medium using a tissue homogenizer. The homogenates were then centrifuged (2,000 × g, 30 minutes, 4°C), and the supernatant was removed and recentrifuged (14,000 × g, 30 minutes, 4°C). Serum and supernatants from tissue homogenates were kept at −70°C until use. TNF-α was assayed using a commercially available ELISA kit (mouse TNF-α Instant ELISA; Bender MedSystems) according to the manufacturer’s protocols.

**In vitro adhesion assays**

For adhesion assays, endothelial cells (at 4 passages) were cultured to confluence on fibronectin-coated 96-well plates (20 ng/mL; Sigma-Aldrich, Inc.) and treated with TNF-α (10 ng/mL) for 12 hours before assaying. MSCs and fibroblasts were treated with TNF-α (10 ng/mL) 12 hours before the adhesion assays and incubated with isotype control IgG or anti-VCAM-1 or very late antigen-4 (VLA-4; 10 μg/mL) monoclonal antibodies (mAb) for 1 hour. Cells were labeled with CyQUANT NF dye, and 1 × 10⁴ cells were seeded onto endothelial cells. After 30 minutes of incubation at 37°C, wells were washed thoroughly 3 times with PBS to remove nonadherent cells. Fluorescent intensity was measured using a fluoroscan, and the number of adherent cells was quantified using a standard curve constructed by a known number of cells. In some experiments, MSCs and fibroblasts were pretreated for adhesion studies with one of the following substances: TNF-α (10 ng/mL), anti-VCAM-1 antibody (mouse monoclonal anti-rat, clone 5F10, 10 μg/mL, Eurogentec), or anti-VLA-4 antibody (mouse monoclonal anti-rat, clone 1A29, 10 μg/mL, Research Diagnostics).

**Parthenolide treatment of MSCs**

Parthenolide (Biomol) was reconstituted in dimethyl sulfoxide (DMSO; Sigma-Aldrich, Inc.) to a stock concentration of 0.4 μM/L and subsequently diluted in PBS. MSCs were treated with parthenolide (5 μmol/L) for 6 hours before experiments. To assess the effect of parthenolide treatment of transgene expression, cells were reseeded into 96-well plates, and luciferase assays were conducted using luciferase-expressing MSCs. Cell viability after parthenolide treatment was also examined with Cell Proliferation Kit II [2,3-bis[2-methoxy-4-nitro-S-sulphonyl]H-tetrazolium-5-carboxanilide inner salt (XTT); Roche Diagnostics GmbH] according to the manufacturer’s instructions.

**Western blotting**

Western blot analysis was conducted to measure the NF-κB pathways. Next, MSCs were pretreated with parthenolide or vehicle (DMSO) for 6 hours, and then cultured with TNF-α (10 ng/mL) for 3 hours. Cells were lysed in radioimmunoprecipitation assay (RIPA) buffer containing protease inhibitor (Pierce Biotechnology). Protein extracts were electrophoresed on a 4% to 12% Bis–Tris gel (Invitrogen), and transferred to polyvinylidene difluoride (PVDF) membranes. Membranes were incubated in PVDF blocking reagent (TOYOBO), and then incubated with primary antibodies against the following proteins: IkBa, phospho-IkBα (Ser32), NF-κB p65, phospho-NF-κB p65 Ser536, and α-tubulin (Cell Signaling Technology), followed by incubation with horseradish peroxidase-conjugated goat anti-rabbit IgG or -mouse IgG1 secondary antibody, and detected using a Western blotting detection system (GE Healthcare).

**Immunocytochemistry**

To visualize p65 nuclear translocation, MSCs were pretreated with parthenolide or vehicle (DMSO) for 6 hours and then cultured with TNF-α (10 ng/mL) for 20 minutes. Cells were fixed with 4% formaldehyde and permeabilized with Triton-X 100. After washing with PBS, slides were incubated with rabbit anti-p65 antibody (Cell Signaling Technology), followed by incubation with Alexa Fluor 488–conjugated goat anti-rabbit IgG secondary antibody. The actin cytoskeleton was stained with Alexa Fluor 546–conjugated phalloidin (Invitrogen); nuclei were stained with 1,5-bis[2-(di-methylamino)ethyl]amino-4,8-dihydroxynaphthacene-9,10-dione (DRAQ)-5 dye (Invitrogen). Cells were examined using Keyence BZ-9000.

**Results**

**In vivo imaging of MSC accumulation in tumors**

We used bone marrow–derived human MSCs, which expressed characteristic phenotypic markers for MSCs and differentiated into adipocyte, osteocyte, and chondrocyte under specific culture conditions (Supplementary Fig. S1). Then, fiber-modified adenovirus vectors (AdK7) were used for efficient transduction of MSCs and fibroblasts in this study. When the cells were transduced with GFP-expressing AdK7 vectors at a density of 3,000 vp/cell, transduction efficiency was almost 100% (Supplementary Fig. S2A and S2B). The bioluminescent intensity of MSCs transduced with luciferase-expressing Ad vectors at 3,000 vp/cell was equal to that of fibroblasts transduced at 680 vp/cell (Supplementary Fig. S2C). Mice injected with GFP-expressing MSCs or fibroblasts were sacrificed 4 days after injection for immunohistochemical analysis.
MSCs identified with anti-GFP antibody were detected in the boundaries of tumors and tumor stroma. However, we found no GFP-positive fibroblasts in the tumor tissues (Fig. 1A). We also used bioluminescence imaging to quantitatively investigate the tumor tropism of MSCs. We injected luciferase-expressing MSCs or fibroblasts into mice through the left ventricular cavity, and then conducted optical bioluminescence imaging to periodically trace the cells using IVIS. In mice injected with luciferase-expressing MSCs or fibroblasts, the factors that attracted MSCs also induced migration of fibroblasts. Rather, it seems that chemokines attracted MSCs also induced migration of fibroblasts, the factors that attracted MSCs also induced migration of fibroblasts. Therefore, we propose a hypothesis as follows: factors, as indicated in Fig. 2, attract both MSCs and fibroblasts to the tumor microenvironment, but importantly, MSCs also induced migration of fibroblasts. Rather, it seems that fibroblasts were more strongly attracted to these factors than MSCs.

**In vitro migration assays**

We analyzed the effects of several growth factors (specifically PDGF-BB, HGF, and VEGF), chemokines (specifically MCP-1 and SDF-1α), and SW480 culture-conditioned medium on MSC and fibroblast migration. These factors are commonly expressed in tumor tissues, and are thought to be potential mediators of MSC tropism. We also used serum-free medium as a negative control and medium containing 30% FBS as a positive control. Migration was quantified by direct labeling and counting of cells by a fluorometer (Fluoroskan Ascent FL; Thermo Labsystems). Exposure to PDGF, HGF, or conditioned medium from SW480 cells stimulated significantly greater MSC migration, whereas VEGF and SDF-1α had no significant effect as compared with serum-free medium (Fig. 2). We compared the migration capacity of MSCs and fibroblasts, the factors that attracted MSCs also induced migration of fibroblasts. Rather, it seems that fibroblasts were more strongly attracted to these factors than MSCs.

**In vitro adhesion assays**

The tumors generated in mice in this study strongly induced tumor stroma with defined blood vessels, and MSCs specifically accumulated in this stroma (Fig. 3A). Therefore, we propose a hypothesis as follows: factors, as indicated in Fig. 2, attract both MSCs and fibroblasts to the tumor microenvironment, but importantly, MSCs significantly adhere to endothelial cells as
activated cell sorting analysis. After TNF-α-stimulated culture, we speculated that in endothelial cells, MSCs, and fibroblasts accumulate at tumor sites via blood vessels in tumor stroma. We measured TNF-α-required for induction of adhesion molecule expression. First, we compared with endothelial cells. MSCs effectively adhered to endothelial cells as indicated earlier (Fig. 1B). In contrast, we could not observe definite accumulation at the tumor sites in mice injected with parthenolide-treated MSCs (Fig. 5D and E). Similar results were also obtained by experiments using Colo205 tumor-bearing mice (Supplementary Fig. S4).

Discussion

In this study, we showed that MSC accumulation at tumor sites would be related not only to migratory capacity toward growth factors and chemokines, but also to MSC–EC adhesion following activation by TNF-α. We further showed that NF-κB activity regulates MSC accumulation at tumor sites through the induction of VCAM-1 expression and the resultant interaction with tumor blood vessel endothelial cells. It is thought that MSCs are mobilized into action following tissue damage, such as injury or inflammation typically accompanied by the release of inflammatory cytokines from the damaged tissues, leading to the recruitment of MSCs to the target. Tumors have a microenvironment consisting of large numbers of inflammatory cells (12). This microenvironment promotes the recruitment of MSCs via various soluble factors secreted by the tumor and inflammatory cells, including EGF, VEGF-A, FGF, PDGF, SDF-1α, IL-8, IL-6, granulocyte colony-stimulating factor (G-CSF), granulocyte-macrophage colony-stimulating factor (GM-CSF), MCP-1, HGF, TGF-β1, and urokinase-type plasminogen activator (uPA; ref. 13). However, in our experimental settings, although systemically injected MSCs accumulated at the tumors, subcutaneously injected MSCs did not (data not shown). We also compared the migration capacity of MSCs and fibroblasts toward growth factors and chemokines in vitro. Rather, it seems that fibroblasts were more strongly attracted to these factors than MSCs. Our results suggest that the mechanism of MSC accumulation cannot be explained solely by cytokine-mediated migration. Therefore, we need different viewpoints to clarify the mechanism. The tumors generated in this study strongly induced tumor stroma with large numbers of blood vessels, and MSCs in particular accumulated in the boundaries between the tumors and tumor stroma. Furthermore, MSC accumulation at the site of the tumors was observed only when cells were injected via the left ventricular cavity. Therefore, we focused on MSC–EC adhesion to elucidate the mechanisms involved.

It has previously been reported that the interaction of MSCs with the vascular endothelium resembles leukocyte chemotaxis (14). To analyze these interactions, we referred to a model

**Effects of parthenolide on MSC migration and adhesion**

We propose a hypothesis that if TNF-α–induced VCAM-1 expression is inhibited, MSC accumulation at tumors is also attenuated. It is well known that TNF-α induces VCAM-1 expression through the NF-κB signaling pathway. We used parthenolide, a sesquiterpene lactone that occurs naturally in the Feverfew plant. Although parthenolide has several biologic activities, we focused on its suppressive effect on NF-κB activity. At first, there were no differences in migratory capacity toward growth factors or chemokines with or without parthenolide treatment (Fig. 4A). Next, we assessed the inhibitory effect of parthenolide on NF-κB activity: MSCs were pretreated for 6 hours, and then were stimulated with TNF-α for 3 minutes. Parthenolide suppressed p65 nuclear translocation through the inhibition of IkBα phosphorylation (Fig. 4B and C) and strongly inhibited the TNF-α–induced VCAM-1 expression on MSCs (Fig. 4D). Consequently, and MSC–EC adhesion was strongly inhibited by parthenolide treatment similarly to anti-VCAM-1 blocking antibody (Fig. 4E).

**In vivo imaging of parthenolide-treated MSCs**

First, we examined the effect of parthenolide treatment on transgene expression and cell viability. There were no significant effects on transgene expression and cell viability after parthenolide treatment (Fig. 5A and B). Next, we conducted in vivo imaging using IVIS. We observed definite bioluminescence at tumor sites in the mice injected with untreated MSCs (Fig. 5C), and bioluminescent intensity was gradually increased (Fig. 5E), as indicated earlier (Fig. 1B). In contrast, we could not observe definite accumulation at the tumor sites in mice injected with parthenolide-treated MSCs (Fig. 5D and E).
that has been proposed for endothelial cell regulation of leukocyte infiltration in inflammatory tissues. Leukocyte-endothelial adhesion involves dynamic interactions between leukocytes and endothelial cells, and involves multiple steps. These steps must be precisely orchestrated to ensure a rapid response with minimal damage to healthy tissue (15). Interactions between leukocytes and the endothelium are mediated by several families of adhesion molecules, each of which participates in a different phase of the process. The surface expression and activation of these molecules during an inflammatory response is tightly controlled under normal conditions. Inflammatory cytokines including IL-1 and TNF-α involve induction of adhesion molecules. In our experimental settings, although other inflammatory cytokine levels including IL-1 and IL-6 were low (data not shown), significant production of TNF-α was observed. We do not clearly know the source of TNF-α in the tumor at this time, and that our in vitro data only suggest that the stroma is the primary source.

As we expected, TNF-α enabled MSCs to adhere to endothelial cells through induction of the expression of adhesion molecules, including VCAM-1 and VLA-4. It is generally considered that VCAM-1 on activated endothelium interacts with the VLA-4 on the leukocyte in the model of leukocyte-endothelial cell adhesion. At first, we speculated that VLA-4 on MSCs plays the same important role as leukocytes. Although both VCAM-1 and VLA-4 on endothelium were efficiently induced by TNF-α stimulation, TNF-α-induced expression of VCAM-1 on MSCs is much stronger than that of VLA-4. Furthermore, MSC–EC adhesion was more effectively inhibited by anti-VCAM-1 antibody as compared with the anti-VLA-4 antibody. On the basis of these results, although VLA-4 on MSC have also related to the MSC-EC adhesion, we thought that VCAM-1 on MSC has more important implications for this adhesion. Once MSCs circulate in the bloodstream, adhesion to endothelial cells is the first step in accumulation in tumors. TNF-α exerts its biologic functions through activating the NF-κB signaling pathway. NF-κB is a major cell survival signal that is antiapoptotic. MSC accumulation was significantly decreased through parthenolide inhibition of NF-κB activity. Although several studies have shown that mitogen-activated protein kinase (MAPK) phosphorylation by growth factors are involved in MSC migration (16, 17), parthenolide did not inhibit MAPK phosphorylation (data not shown). Therefore, at least parthenolide treatment did not affect in migration ability of...
MSCs toward growth factors from tumors in this experimental settings. Nevertheless, MSC accumulation was significantly decreased through parthenolide inhibition of NF-κB activity. We did not show histologic evidence in the experiments using parthenolide. However, we show that parthenolide does not inhibit luciferase activity in vitro (and thus does not seem to be toxic), and that therefore the effect observed in vivo should be an effect on recruitment. Although we focused on the function of TNF-α in this study, other inflammatory cytokines including IL-1β and IFN-γ also have ability to induce VCAM-1 expression in target cells (18), and may be involved in MSC accumulation.

TNF-α is a major inflammatory cytokine that plays important roles in diverse cellular events, such as cell survival, proliferation, differentiation, and death. Numerous reports have shown that TNF-α levels in serum are increased in patients with cancer (19, 20), and TNF-α is also related closely to the tumor progression including metastasis. For example, TNF-α intensely induces IL-6 and MCP-1 from cancer-associated fibroblasts and normal fibroblastic cells and has indirect influences on generation of prometastatic microenvironment (21). Furthermore, TNF-α is also released in cardiac infarction, during acute coronary syndromes, and in chronic heart failure; MSCs also accumulate at the site of cardiac infarction (22, 23). These results indicated that proinflammatory cytokines promote homing of stem cells in the heart and that these cytokines have a positive effect on cardiac regeneration. Therefore, activation with TNF-α is one of the critically important steps for MSC accumulation. Moreover, MSC-based tissue-targeted strategies may be adapted for various inflammatory diseases.

In MSC-based cancer-targeted gene therapies, it is thought that therapeutic efficacy is directly linked with accumulation efficiency of MSCs at tumor sites. Our results suggested that combination use of NF-κB inhibitors, including bortezomib, or TNF-α blocking agents, such as infliximab, reduces the
therapeutic efficacy of gene-modified MSCs due to inhibition of the accumulation steps. In contrast, tumor-specific TNF-α–inducing agents would be useful in enhancing therapeutic efficacy, thus further research is required in identifying such agents to more effective therapeutic strategies.

In conclusion, the present study shows that NF-κB activation through TNF-α stimulation and VCAM-1/VLA-4–mediated MSC-EC adhesion may be an important element in MSC accumulation. Although MSCs are useful as cellular vehicles for cancer-targeted gene therapy, past studies have shown that increased MSC accumulation is needed to enhance therapeutic efficacy. Thus, methodology for the enhancement of MSC accumulation should be developed and our findings suggest a solution.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

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