Microenvironment and Immunology

Tumor Microenvironmental Conversion of Natural Killer Cells into Myeloid-Derived Suppressor Cells

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Abstract
How myeloid-derived suppressor cells (MDSC) emerge in the tumor environment remains unclear. Here, we report that GM-CSF can convert natural killer (NK) cells into MDSCs. When transferred into tumor-bearing mice, adoptively transferred NK cells lost their NK phenotype and were converted into Ly6ChighLy6Ghigh MDSC. This conversion was abolished by exposure to IL-2 either in vitro or in vivo. Notably, we found that of the 4 maturation stages based on CD11b/CD27 expression levels, only the CD11bhighCD27low NK cells could be converted into CD11b+Gr1+MDSC ex vivo. Transfer of CD27high NK cells from tumor-bearing mice into tumor-bearing recipients was associated with conversion to MDSC in a manner associated with reduced numbers of CD11bhighCD27high and CD11bhighCD27low NK cell populations in the recipients. Our results identify a pathway of MDSC development from immature NK cells in tumor-bearing hosts, providing new insights into how tumor cells modulate their host immune microenvironment to escape immune surveillance. Cancer Res; 73(18); 5669–81. ©2013 AACR.

Introduction
The tumor environment recruits diverse suppressor cells, such as M2 macrophages, regulatory T cells, and myeloid-derived suppressor cells (MDSC; refs. 1-3), to help cancer cells evade the attack by effector cells, leading to the subversion of the immune surveillance (3). Of the suppressor cells, MDSCs comprise a mixed population of immature myeloid cells that accumulate in various pathologic conditions, particularly in tumors. Recently, the morphologic heterogeneity of MDSCs has been classified by the expression of Ly6C and Ly6G molecules, which has led to the identification of 2 MDSC subsets, i.e., CD11b+Ly6C+Ly6Clow polymorphonuclear (PMN)-MDSCs and CD11b+Ly6C-Ly6Ghigh monocytic (Mo)-MDSCs (4). MDSCs use multiple mechanisms to suppress the adaptive and innate immune systems and increased numbers of MDSCs correlate with a poor prognosis in patients with cancer (5, 6). Recent studies have shown that a small number of transcription factors regulate aberrant myelopoiesis, leading to MDSC expansion (7-9). Despite the advances in the field, the MDSC developmental pathway remains in part elusive.

In contrast with MDSCs, natural killer (NK) cells present barriers to various tumors (10). NK cell depletion leads to enhanced tumor growth in a mouse tumor model, indicating clearly the involvement of NK cells in tumor surveillance (11). In addition, it has been shown that improved disease prognosis is associated with the extent of NK cell infiltration in non-small cell lung carcinomas and colorectal cancers (12). However, in a number of cases, NK cells in the tumor environment exhibit maturation and functional defects, decreasing the absolute number of cells (13-15). Although a number of studies are underway to determine the cause of NK cell abnormalities, further investigations will be required.

In this study, we describe the unprecedented phenomenon that conventional NK cells in a specific maturation state in the tumor environment are converted into MDSCs in tumor-bearing mice. The number of NK cells decreased significantly as the implanted tumor grows, which may be attributed to the conversion of CD11bhighCD27low NK cells into MDSCs rather than being matured into CD11bhighCD27high phenotype. NK cell-activating cytokine, interleukin (IL)-2, inhibited the development of MDSCs from NK cells in vitro and in vivo. Overall, in vivo IL-2 treatment induced NK cell expansion, augmented the activity of the cells, and may result in the regression of tumor growth.

Materials and Methods

Mice and cell lines
All experiments were approved by the Institutional Animal Care and Use Committee of Seoul National University (Seoul, Korea). Six-week-old BALB/c, C57BL/6 mice were purchased

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from Charles River Laboratories and C57BL/6 CD45.1 congenic mice were purchased from JAX. The mice were bred and maintained in the Animal Facility for Pharmaceutical Research at Seoul National University under specific pathogen-free conditions.

CT26 colon adenocarcinoma cell line, EL4 lymphoma cell line, and TC-1 lung carcinoma were obtained from the American Type Culture Collection. Human Her-2/neu-expressing transfectoma Her-2/CT26 cells were developed by transduction of CT26 using a retroviral vector system (16, 17). Cells were maintained in Dulbecco’s Modified Eagle Medium or RPMI-1640 medium supplemented with 10% heat-inactivated FBS, 1% penicillin–streptomycin (G418 was supplemented for transfectoma). Cell lines were periodically authenticated by morphologic inspection and passaged for not more than 3 to 4 weeks from thawing.

**Antibodies and flow cytometry**

Anti-CD11b, CD27-APC, anti-Ly6G-PE/Cy7 (BioLegend), anti-F4/80, CD4 (eBioscience), CD11c (BD Biosciences), CD14, CD19, NK1.1, Gr1-PerCP/Cy5.5, anti-CD122, CD45.1, Ly6C-FITC, and anti-CD45.2-PacificBlue (BioLegend) antibodies were used. IL-2–neutralizing Ab and IL-2–receptor β-blocking Ab were obtained from S4B6 and TMB1 hybridsomas, respectively, and were kind gifts from Charles D. Surh (Scripps Institute, La Jolla, California). To analyze the stained cells, FACSCalibur (BD Biosciences) instrument and FlowJo (Treestar) were used.

**In vitro T-cell suppression assay**

The DO11.10 cells (1 × 10⁵/well) were stimulated with OVA protein (grade V; Sigma-Aldrich) and were cocultured with or without cytokine-induced MDSCs for 72 hours. For the final 20 to 24 hours, we added 1 μCi per well [³H]-thymidine. The incorporation of [³H]-thymidine into the divided cells was detected using a liquid scintillation counter (Wallac).

**ROS production, arginase 1 activity, and NO production**

For ROS detection, cells were incubated in the presence of 2.5 μmol/L DCFDA ± 30 ng/mL phorbol 12-myristate 13-acetate (PMA) for 30 minutes. The fluorescence intensity of DCFDA was analyzed by flow cytometry.

Cells were incubated with 2 ng/mL IFN-γ and 100 ng/mL lipopolysaccharide (LPS) for 18 hours. To detect NO production, supernatants were collected and mixed with equal volumes of Greiss reagent (1% sulfanilamide in 5% phosphoric acid and 0.1% N-1-naphthylethylenediamine dihydrochloride in DW), after 10 minutes the absorbance at 540 nm was measured. The concentrations were determined by the standard curve of serial dilution of sodium nitrite. Arginase 1 activity was measured in cell lysates (lysed with 0.1% Triton X-100). Subsequently, 50 μL of 10 mmol/L MnCl₂, 50 mmol/L Tris/HC1 were added and the enzyme was activated by heating for 10 minutes at 55°C. Arginine hydrolysis was conducted by incubating the lysates with 0.5 mol/L arginine (pH 9.7) at 37°C for 80 minutes. The reaction was stopped with 400 μL of acid mixture (H₂SO₄/H₃PO₄/H₂O = 1/3/7). Urea production was measured at 540 nm after addition of 9% α-isonitrosopropiophenone (control absorbance subtracted from specific absorbance).

**In vivo IL-2/αIL-2 Ab complex (IL-2 complex) treatment**

To verify the effect of IL-2 on MDSC accumulation and function in vivo, we used an IL-2 complex. Mice received an intraperitoneal injection of 1.5 μg rmIL-2 plus 50 μg αIL-2 antibody. Before injection, rmIL-2 and the αIL-2 Ab were mixed and incubated at room temperature for 15 minutes.

**Tumor model and isolation of tumor-infiltrating lymphocytes**

C57BL/6 mice were injected subcutaneously with 2 × 10⁵ TC-1 tumor cells. Injection of IL-2 complex began on day 1 after tumor injection. IL-2 complex treatment was administered every other day for a total of 5 or 10 times. The volume of the implanted tumor was evaluated for 19 days following the subcutaneous tumor inoculation. To separate the tumor-infiltrating lymphocytes, the tumors were collected and weighed and single-cell suspensions were prepared. The tumor was cut into small pieces and was incubated at 37°C for 0.5 hours in RPMI-1640 containing 1 mg/mL collagenase (Roche), 500 μg/mL DNase I, and 25 μg/mL hyaluronidase (Sigma).

**Cell sorting**

To sort CD49b⁺ cells, splenocytes were prepared from tumor-bearing mice. To enrich the desired cell population, CD4⁺, CD8⁺, B220⁺, and Ly6G⁺ cells were depleted using microbeads (Miltenyi Biotec). CD11b⁻Ly6Cs⁻/Ly6G⁻ CD49b⁻ cells were sorted by using FACS Aria II. To sort the conventional NK cells, splenocytes were enriched by the depletion of CD4, CD8, CD19, and Ly6G⁺ cells using microbeads. CD3⁺CD19⁻Gr1⁻CD122⁻NK1.1⁻/NKp46⁻ cells were sorted.

**In vivo conversion assay of NK cells**

To determine the conversion of NK cells in vivo, EL4 tumor cells were injected subcutaneously into CD45.2 mice. After 3 weeks, 2.5 × 10⁵ CD27high NK cells (CD11b⁺/CD27high; CD11b⁻/CD27high = 1.2) were isolated from the spleen and were transferred intravenously into CD45.1 naïve mice or mice that were inoculated subcutaneously with 1 × 10⁵ tumor cells 7 days before the adoptive transfer. On day 14 after the transfer, CD45.2⁺CD45.1⁻ cells were analyzed for the expression of NK cell and MDSC markers in the spleen. For conversion assay in intraperitoneal tumor model, EL4 tumor cells were injected subcutaneously into CD45.1 congenic mice. After 3 weeks, NK1.1⁺ cells were isolated from the spleen by FACS ARIA III and transferred intraperitoneally into naïve mice (NK1.1⁻ → naïve host) or mice that had been inoculated with 1 × 10⁵ EL4 tumor cells 5 days before the adoptive transfer (NK1.1⁻ → tumor host), with a daily injection of IL-2 complex until sacrifice (NK1.1⁻ → tumor host + IL-2). On day 9 after the adoptive transfer, cells were collected from the peritoneal cavity and CD45.1⁺CD45.2⁻ cells were analyzed.
Quantitative real-time PCR

Total RNA was extracted using the TRIzol reagent and cDNA was generated with SuperScript reverse transcriptase and oligo (dT) primers (all from Invitrogen Life Technologies). The LightCycler optical system (Roche) and the SYBR Green real-time PCR Kit (Takara) were used for the analysis of gene expression. Target gene values were calculated relative to Hprt expression. The following primer pairs were used:

CD112 antisense (GGGAAGCCGCCAGATGAC); CD116 sense (AAGCAGTCATGCACACCTG), CD116 antisense (TGTTGTCGCTCGTATGGA); CD131 sense (AAGGACCTGCAACCTCATGGCA); CD131 antisense (TGGGCGGTCTGCTCACTCTT); Cebp sense (CCCCAGATGACAGAAGG); Cebp antisense (TGGTGTCGCTGCTCACTCTA); Nfl3 sense (AAGGCCTCCTCATTCTC); Nfl3 antisense (TTCAACCTCGATGCAAAGG); Pa.l sense (GCCCTAGTCACAGGCTTCC); Pa.l antisense (CTCTTACCTATCCTATGCT); Hprt sense (AAAGCTCACCCTTCCCTCATG); and Hprt antisense (ATCCAGAGTCCATGAAAGA).

Statistical analysis

Statistical analyses were conducted using Student’s t test. The results with values of $P < 0.05$ were considered to be statistically significant.

Results

Inverse relationship between the percentage of CD11b+ Ly6Cneg/low cells and MDSCs reveals the conversion of NK-phenotype cells into MDSCs.

To explore the development of tumor-associated MDSCs, we used animal models of Her-2-expressing CT26 colon carcinoma and EL4 thymoma. When these tumor cells were subcutaneously inoculated into mice, we observed that the percentage of Ly6Cneg/low cells (R1) among CD11b+ population gradually declined in the spleen during tumor progression (Supplementary Fig. S1A and S1B). In contrast, Ly6Ghigh PMN-MDSCs (R2) and Ly6Chigh Mo-MDSCs (R3) significantly increased in tumor-bearing mice compared with those in naive mice. This observation led us to hypothesize that the tumor environment promotes the conversion of CD11b+Ly6Cneg/low cells into MDSCs. To address this hypothesis, we cultured CD11b+Ly6Cneg/low cells with cytokines that are known to be relevant to MDSC accumulation (4) and varied depending on the cytokine used (Supplementary Fig. S2A). Considering the number of live converted cells and the proportions of Ly6Chigh/Ly6Ghigh cells after incubation, granulocyte macrophage colony-stimulating factor (GM-CSF) was the most efficient cytokine at converting Ly6Cneg/low cells into MDSCs in vitro (Supplementary Fig. S2B and S2C).

Flow cytometric analysis revealed that the majority of CD11b+Ly6Cneg/low cells expressed CD49b, but not CD14, CD11c, CD4, CD8, and B220 (Supplementary Fig. S2D). Furthermore, fluorescence-activated cell sorting (FACS)-sorted CD49b+ cells (purity > 98%) obtained from tumor-bearing mice were converted into Ly6Ghigh/Ly6Chigh MDSC-like cells upon stimulation with GM-CSF (data not shown). To directly ask whether NK cells can be converted into MDSCs, we purely isolated NK1.1+ and CD49b+ cells from C57BL/6 and BALB/c mice, respectively, and stimulated them with GM-CSF. As depicted in Fig. 1A, GM-CSF downregulated the NK cell markers NK1.1, CD49b, and NKp46, but instead upregulated the expression of MDSC markers Ly6C and Ly6G. Some treatment failed to induce Ly6C and Ly6G on CD4+ or CD8+ T cells and B cells (Supplementary Fig. S3). Similarly, FACS-sorted CD49b+ NKp46+ cells (purity > 95%) from the bone marrow were also converted into Ly6Ghigh and Ly6Chigh cells in the presence of GM-CSF, which was accompanied by a decrease in NK-marker expression (CD49b+, 40.5%; NKp46+, 14.8%). However, CD49b+ NKp46+ cells from the peripheral blood mononuclear cell (PBMC) did not respond to GM-CSF stimulation (data not shown).

To further investigate the observed conversion of NK cells into MDSC-like cells, we examined whether IL-2 affects this process, as this cytokine activates NK cells and potentiates NK cell-mediated antitumor activity. Notably, addition of IL-2 not only inhibited the expression of Ly6C and Ly6G triggered by GM-CSF, but also maintained the expression of NK cell markers NK1.1, CD49b, and NKp46 (Fig. 1A).

To further investigate the role of IL-2 in vivo, NK1.1+ cells were isolated (purity > 97%; Fig. 1B) from tumor-bearing CD45.1 mice and transferred into CD45.2 mice inoculated intraperitoneally with tumor cells 5 days before the adoptive transfer. Donor cells from the peritoneal cavity were analyzed. Up to 66% of the transferred Ly6Cneg/lowNK1.1+ cells were converted to Ly6C+ and/or Ly6Ghigh MDSCs, whereas only 20% of the cells retained NK1.1+ phenotype (Fig. 1C). When the recipients were additionally given CD122-biased IL-2/IL-2–neutralizing-Ab complex (IL-2 complex; refs. 18, 19), most of the transferred cells maintained NK cell phenotype (Ly6C and/or Ly6Ghigh, > 3.6%; NK1.1+, 99%). In addition, there was minimal conversion of NK1.1+ cells in naive recipients (Ly6C and/or Ly6Ghigh, 7%; NK1.1+, 87.4%). Taken together, these results showed that NK cells can be converted into MDSC-like cells in tumor-bearing host in vivo, or by tumor-associated cytokines including GM-CSF ex vivo, and IL-2 can inhibit this conversion.

IL-2 reverses the suppressive activity of MDSCs converted from NK-phenotype cells and subverts tumor environment.

To examine whether the MDSC-like cells converted from NK cells possess immunosuppressive activity, we obtained the Ly6Cneg/lowLy6Ghigh cells converted from CD49b+ cells after stimulation with GM-CSF and cocultured them with DO11.10 splenocytes whose T cell receptor recognizes OVA presented by MHC II. As shown in Fig. 2B, addition of the MDSC-like cells significantly inhibited OVA-induced proliferation of DO11.10 T cells, which resembled the suppressive activity of purified PMN- and Mo-MDSCs (Fig. 2A). However, IL-2 significantly reversed the suppressive activity of the converted CD49b+ cells (Fig. 2B). The converted cells also displayed ROS/NO production (Fig. 2C and D) and arginase 1 activity (Fig. 2E), by which Ly6Cneg/lowLy6Ghigh cells from C57BL/6 and BALB/c mice, respectively, and stimulated them with GM-CSF. As depicted in Fig. 1A, GM-CSF downregulated the NK cell markers NK1.1, CD49b, and NKp46, but instead upregulated the expression of MDSC markers Ly6C and Ly6G. Some treatment failed to induce Ly6C and Ly6G on CD4+ or CD8+ T cells and B cells (Supplementary Fig. S3). Similarly, FACS-sorted CD49b+ NKp46+ cells (purity > 95%) from the bone marrow were also converted into Ly6Ghigh and Ly6Chigh cells in the presence of GM-CSF, which was accompanied by a decrease in NK-marker expression (CD49b+, 40.5%; NKp46+, 14.8%). However, CD49b+ NKp46+ cells from the peripheral blood mononuclear cell (PBMC) did not respond to GM-CSF stimulation (data not shown).

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NK cells after GM-CSF stimulation were likely bona fide MDSCs cells with suppressive activity as well as with MDSC-signature secretion profiles.

To analyze the effect of IL-2 on MDSC populations and tumor growth, mice were treated with IL-2 complex every other day after TC-1 tumor inoculation. The tumor growth was suppressed by IL-2 treatment (Fig. 3A and B), as the proportion of MDSCs per tumor weight was reduced, although NK1.1+ cells were increased among tumor-infiltrating leukocytes (Fig. 3C). However, IL-2 cessation reaccelerated the tumor growth, Ly6C/high MDSCs were also replenished and, importantly, NK1.1+ cells were decreased in the tumor bed (Fig. 3C). In the spleen, the IL-2 counteracting effect on the frequency of MDSCs and NK cells was also prominent, although it did not have a significant effect on the cell numbers (Fig. 3D and E). This can be explained by previous studies showing that IL-2 directly or indirectly enhances the survival of granulocytes and monocytes (20-22).

**CD49b+ cells are prone to conversion into MDSCs in the tumor environment**

To investigate whether the "MDSC-philic" cytokine-induced conversion of NK-phenotype cell into MDSCs is a property acquired in response to environmental cues or is an inherited feature, CD49b+ cells were isolated from naive, 3- and 5-week tumor-bearing mice. The longer NK-phenotype cells remained in the tumor environment, the more likely they were to become Ly6C/Ly6G/high MDSCs following cytokine treatment. CD49b+ cells from the naive and 3-week tumor-bearing mice responded to GM-CSF; however, 50% to 60% of the cells remained CD49b+ cells and the amount of CD49b+ cells from the 5-week tumor-bearing mice was reduced to 19% (Fig. 4A). In addition, IL-2-mediated inhibition was less potent in CD49b+ cells from the 5-week tumor-bearing mice than the 3-week tumor-bearing mice. This observation was confirmed by the increased induction of CD49b- population, which was 17% in the 5-week
NK Cell Conversion into MDSCs

Figure 2. IL-2 reverses the suppressive activity of MDSCs converted from CD49b+ cells. Purified PMN-, Mo-MDSCs, and CD49b+ cells (A) or cytokine-treated CD49b+ cells (B) were cocultured with DO11.10 Tg splenocytes in the presence of 250 μg/mL OVA proteins for 3 days. For the final 20 to 24 hours, we added 1 μCi per well [3H]-thymidine into the divided cells was detected using a liquid scintillation counter. (–) control, DO11.10 splenocytes + OVA proteins; (–) control, DO11.10 splenocytes alone. The data represent the mean ± SEM. *, P < 0.05; **, P < 0.01; ***P < 0.001. C, the level of ROS in purified CD49b+, PMN- and Mo-MDSC, or cytokine-treated CD49b+ cells (for 4 days, 1 day rest in media before stimulation) was measured by fluorescence intensity of DCFDA labeling after PMA stimulation for 30 minutes. Filled, no PMA and no DCFDA; dotted, no PMA and DCFDA; solid, PMA and DCFDA. D and E, purified CD49b+, PMN- and Mo-MDSC, or cytokine-treated CD49b+ cells (for 4 days, 1 day rest in media before stimulation) were stimulated with 2 ng/mL IFN-γ and 100 ng/mL LPS for 18 hours, supernatants were collected, and nitrite concentration was measured (E). Arginase 1 activity (urea production) was measured in cell lysates (F).

tumor-bearing mice and 7% in the 3-week tumor-bearing mice in the presence of GM-CSF and IL-2 (Fig. 4A). These data indicate that NK cells may enter a conversion state that is biased toward MDSCs in the tumor environment.

The change in responsiveness to IL-2 was further shown by analyzing the related-cytokine receptors. CD49b+ cells from the 5-week tumor-bearing mice preferentially expressed GM-CSF–receptor β-chain (CD131), whereas the expression of IL-2–receptor β-chain (CD122) was decreased compared with the expression in the naive or the 3-week tumor-bearing mice. The expression of GM-CSF–receptor α-chain (CD116) did not increase significantly in the 5-week tumor-bearing mice (Fig. 4B). Although the expression of Nfil3, an essential regulator of NK cell development (23, 24), was decreased, the expression levels of Cebpα and Pu.1, essential transcription factors for granulocyte and monocyte development (25, 26), were significantly increased in CD49b+ cells from the 5-week tumor-bearing mice compared with the naive mice (Fig. 4B). To delineate whether the conversion was driven by these transcription factors, we analyzed the kinetics of gene expression during the cytokine stimulation. The expression of both genes was significantly downregulated, as early as 6 hours after IL-2/GM-CSF stimulation, compared with the stimulation with GM-CSF alone. The gap in the gene expression level was sustained during the cytokine stimulation. The expression of both genes was significantly downregulated, as early as 6 hours after IL-2/GM-CSF stimulation, compared with the stimulation with GM-CSF alone. The gap in the gene expression level was sustained during the cytokine stimulation.

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drive the conversion of NK-phenotype cells. However, further investigation will be needed to provide direct evidence for this hypothesis.

**CD11b**<sup>high</sup>**CD27**<sup>high</sup> subset of NK cells is converted into MDSCs in tumor-bearing mice

Despite prominent difference in the responsiveness to GM-CSF, we observed no significant difference in the expression of surface/transcription markers related to the lineage specification, NK cell receptors, the secretion of effector/suppressive cytokines between NK cells from naive and tumor-bearing mice (data not shown). To provide more definitive evidence for which NK cells in the tumor environment were converted into MDSCs, we further analyzed the difference in the extent of NK cell maturation based on CD11b/CD27 expression (27) between the naive and tumor-bearing mice. The numbers of total NK cells were significantly decreased in the spleen and bone marrow of the mice inoculated with tumor cells for 3 weeks compared with naive mice (Fig. 5A and B). Importantly, the numbers of CD11b<sup>high</sup>CD27<sup>high</sup> and CD11b<sup>high</sup>CD27<sup>low</sup> subsets among NK cells were significantly reduced in tumor-bearing mice compared with naive mice, whereas those of the other subsets were relatively similar (Fig. 5C and D).

To elucidate whether the reduction of those NK cell populations was attributed to the observed conversion, we separated the lineage<sup>+</sup> (CD3<sup>+</sup>CD19<sup>+</sup>Gr1<sup>+</sup>) CD122<sup>+</sup>NKp46 or NK1.1<sup>+</sup> cells from tumor-bearing mice into 4 stages based on CD11b/CD27 expression level (Supplementary Fig. S4A and

Figure 3. IL-2 subverts immunosuppressive tumor environment. A–E, C57BL/6 mice were injected subcutaneously with TC-1 tumor cells. Injection of IL-2 complex began on day 1 after tumor injection. IL-2 complex treatment was administered every other day for a total of 5 or 10 times. A, the volume of the implanted tumor was evaluated for 19 days following the subcutaneous tumor inoculation. At day 21 after tumor inoculation, tumor weight was measured (B) and the percentages and numbers of MDSCs and NK cells were analyzed in the tumor bed (C; %/tumor weight) and spleen (D and E). The data represent the mean ± SEM.

*P < 0.05; **P < 0.01; ***P < 0.001.
S4B) and cultured them in the presence of GM-CSF. Notably, CD11b<sup>high</sup>CD27<sup>high</sup> NK cells obtained from tumor-bearing mice were converted into CD11b<sup>+</sup>Gr1<sup>+</sup>MDSC phenotype (Fig. 6A and B), whereas their naive counterpart and CD11b<sup>low</sup>CD27<sup>low</sup>, CD11b<sup>low</sup>CD27<sup>high</sup>, CD11b<sup>high</sup>CD27<sup>low</sup> NK cells from tumor-bearing mice hardly did (data not shown). Comparable purity of CD122<sup>+</sup>NKp46<sup>+</sup> cells for sorting showed that the unwanted cells were present equally (0.8%–1.3%) in 4 sorted populations (Supplementary Fig. S4A), indicating that this phenomenon was not due to myeloid precursors in the tumor environment. Moreover, the GM-CSF–treated CD11b<sup>high</sup>CD27<sup>high</sup>NKp46<sup>+</sup> cells yielded a 3.6-fold greater number of converted cells compared with those from NKp46<sup>+</sup> cells (purity > 98.5% and 98.1%, respectively, Supplementary Fig. S5A and S5B). Furthermore, we evaluated the conversion efficiency based on the starting cell numbers. While the efficiency was only 2% in NKp46<sup>+</sup> cells, this increased to 10% in CD11b<sup>high</sup>CD27<sup>high</sup>NKp46<sup>+</sup> cells (Supplementary Fig. S5C), of which
tendency was consistent with the proportion of CD11b<sup>high</sup> CD27<sup>high</sup> population included in the purified NKp46<sup>+</sup> cells (Supplementary Fig. S5A, bottom). This discrepancy in which “NK marker/C0” cells equally included between NK populations indicates that the conversion arose from “NK marker/+” cells.

Annexin V staining revealed that 54% of the GM-CSF–treated CD11b<sup>high</sup>CD27<sup>high</sup>NKp46<sup>+</sup> cells were apoptotic during the course of conversion (Supplementary Fig. S5D).

The expression of CD122, NKp46, and NK1.1 was downregulated in the presence of GM-CSF alone, whereas retained in GM-CSF/IL-2 (Fig. 6A and B). The morphology of the CD11b<sup>high</sup>CD27<sup>high</sup> population was similar to the other populations of NK cells and the naïve CD11b<sup>high</sup>Gr1<sup>+</sup>NK cells, which exhibited a shape consistent with lymphocytes (Supplementary Fig. S4C). In contrast, ring-, segmented band-, and monocyte-shaped cells were observed after in vitro stimulation with GM-CSF. In the presence of GM-CSF/IL-2, the cells exhibited the morphology of activated NK cells (Fig. 6C).

Another NK-activating cytokine, IL-15, also exerted an inhibitory effect on conversion upon addition of low (20 ng/mL) and high (50 ng/mL) concentrations, although low levels of CD11b<sup>+</sup>Gr1<sup>+</sup> cells (1.2%) were present after the addition of low-concentration of IL-15 (Fig. 6D).

To show this phenomenon in vivo, we purified CD45.2<sup>+</sup>CD27<sup>high</sup> NK cells from tumor-bearing mice and transferred the cells into congenic CD45.1 mice. In tumor-bearing recipients, 12% of the transferred cells were converted into CD11b<sup>+</sup>Gr1<sup>+</sup> MDSC phenotype (Fig. 7A and B). In addition, the expression of CD122 and NK1.1 was decreased. In contrast, the phenotype of CD27<sup>high</sup> NK cells was retained in naïve...
recipients. Moreover, CD11b<sup>high</sup>CD27<sup>low</sup> NK cells and CD27<sup>high</sup>NK cells from tumor-bearing and naive mice, respectively, were rarely converted into MDSCs. Furthermore, the number of CD11b<sup>high</sup>CD27<sup>low</sup> and CD11b<sup>high</sup>CD27<sup>high</sup> NK cells that arose from transferred CD27<sup>high</sup>NK cells was reduced significantly in tumor-bearing recipients compared with naive recipients (Fig. 7C–E).

However, endogenous MDSCs may cause cell death of transferred CD27<sup>high</sup>NK cells, by which the matured NK cells were reduced. To investigate this, we evaluated the cell death of donor cells. The frequency of Annexin V<sup>+</sup> donor cells tended to increase in the tumor-bearing host compared with the naive host, although the difference was not statistically significant (Supplementary Fig. S6A). To examine whether MDSCs rendered donor NK cells apoptotic, we transferred MDSCs (4 × 10<sup>6</sup>/injection) into naive recipients 1 day before and after NK cell transfer. There was no difference in the frequency of apoptotic donor cells in the MDSC-treated naive hosts compared with the naive hosts (Supplementary Fig. S6A). In the same experimental setting, the donor cells were significantly more converted into MDSCs in the tumor-bearing hosts than in the naive or MDSC-treated naive hosts (Supplementary Fig. S6B). To more directly clarify the effect of MDSCs on NK cells during maturation, we stimulated CD45.1<sup>+</sup>CD27<sup>high</sup>NK cells with IL-12, -15, and -18 to mature them in vitro (Supplementary Fig. S6C; ref. 28). These cytokines preferentially upregulated the expression of KLRG1 and CD11b, maturation markers for NK cells, compared with the untreated NK cells. However, no differences were observed in the maturation and apoptosis of NK cells even when CD45.1<sup>+</sup>MDSCs were added 3 or 6 times.
compared with other conditions. These results suggested that the reduction of NK cells could be attributed to their conversion into MDSCs, and possibly cell death, in the tumor environment. However, the viability and maturation of NK cells did not seem to be influenced by endogenous MDSCs, at least in the EL4 tumor model.

**Discussion**

The importance of NK cells for eradicating cancer cannot be overemphasized. Several studies have reported that the tumor environment impairs the development and function of NK cells (13, 15) and even diminishes the number of NK cells in patients with chronic myelogenous leukemia (14). In this study, we determined that NK cells at a specific maturation stage were converted into immunosuppressive MDSCs, a process that was impeded by IL-2 in vitro and in vivo. The inverse relationship between the percentage of Ly6c<sup>high</sup> cells and MDSCs provided us a clue as to which NK-phenotype cells might be converted into MDSCs. This observation led us to investigate the conversion of NKp46<sup>+</sup>/NK1.1<sup>+</sup> cells and the Lin<sup>−</sup>CD122<sup>+</sup> conventional NK cells. It has been reported that the expression of NKp46 is restricted to NK cells and is induced in a number of minor T-cell populations, whereas myeloid cells, such as DCs, neutrophils, and macrophages, do not express NKp46 (29), indicating that the conversion showed in this...
Apart from the question of contribution, the observation that CD11b<sup>low</sup>CD27<sup>neg/low</sup> NK cells were not converted, indicating that this phenomenon was not because of the myeloid-biased differentiation by the cytokines released by tumor. We are supposed to clarify this by tumor cell knockdown experiments using shRNA or neutralization of cytokines in future studies.

Because the exact roles of CD27 in NK cell biology have not been fully investigated, it aroused our curiosity whether CD27<sup>-</sup>CD27<sup>ligand signaling influenced NK cells or MDSCs. However, upon CD27-mediated signaling by the addition of recombinant ligands in vitro, no remarkable effect was found other than increased apoptosis during the conversion of NK cells into MDSCs (data not shown), although the ligands may play other roles in vivo.

We showed that the conversion of Lin<sup>-</sup>CD122<sup>+</sup> conventional NK cells was prominent in only CD11b<sup>high</sup>CD27<sup>high</sup> populations (Fig. 6). Four sorted populations of NK cells showed comparable purity for CD11b<sup>high</sup>CD27<sup>low</sup> NK cells (Supplementary Fig. S4A), which indicates that the unwanted cells are present equally. Moreover, our morphologic analysis showed that all 4 populations of NK cells isolated from tumor-bearing mice had no difference (Supplementary Fig. S4C). Nevertheless, NK cells in the other maturation states were not converted, indicating that this phenomenon was not because of the myeloid-biased differentiation by the tumor environment, rather NK cells in a specific maturation state were converted into MDSCs.

Several studies have endeavored to explain the development, function, migration, and fate of MDSCs (31–33). A recently published study has determined that the myeloid precursors of MDSCs that reside in the spleen are relocated to tumors (34, 35). Moreover, novel transdifferentiation pathway from monocytic-to-granulocytic MDSCs driven by epigenetic regulation in the tumor environment, which is beside the classical divergent development of monocytes and granulocytes, has been revealed (33). From a different perspective, we suggest that NK-phenotype cells could be additional precursors of MDSCs. Accordingly, the precise contribution of NK cells to MDSC expansion must be examined; however, the depletion of NK cells by antibodies did not significantly influence the accumulation of MDSCs in the tumor-bearing mice (data not shown). Apart from the question of contribution, the observation that the effector NK cells were converted into suppressor cells indicated the aggravation of the immunosuppressive environment in the tumor-bearing mice, which may also apply to patients with cancer (14).

We have attempted to identify the conversion of NK cells into MDSCs in a human system. Several studies have shown that MDSCs can be produced from whole PBMCs and monocytes (36–39). However, CD56<sup>-</sup> cells purified from the PBMCs of patients with cancer were not converted into MDSCs (data not shown). These observations are consistent with the results from PBMC data in the mouse system, although the reason for this remains unclear. This indicates that the conversion of NK cells may depend on their anatomical location. Because in mice, NK cells from the spleen and bone marrow and not the PBMCs were converted into MDSCs, the corresponding organs in humans should be investigated. Furthermore, the types and stages of cancer confer complexity on the environments in which NK cells are influenced by various factors, which may cause confusion when predicting the conversion of NK cells into MDSCs in the human system. Therefore, more detailed and extensive investigations of human specimens classified by the type/ stage of cancer and the organs from which NK cells are isolated will be required for future studies.

NK cells are thought to differentiate from common lymphoid progenitors (CLP; refs. 10, 40). However, a number of studies have debated the origin of NK cells and have shown that the cells can be derived from myeloid progenitors (41–43). It has also been reported that CLPs under certain conditions and the earliest progenitors in the thymus have the potential for myeloid development (44, 45). Whether myeloid-derived NK cells exist in the periphery and whether these cells are essential for the conversion shown in this study, are intriguing issues. Even if this were the case, myeloid-derived NK cells are only a fraction of the entire NK cell population because not all of NK.1<sup>+</sup> and CD49b<sup>+</sup> NK cells were converted into MDSCs, between those only CD11b<sup>high</sup>CD27<sup>high</sup> NK cells were converted.

The levels of Cebp<sub>a</sub> and Pu.1, essential transcription factors for the development of granulocytes and monocytes (25, 26), were increased in CD49b<sup>+</sup> cells from the tumor-bearing mice and may constitute a marker for NK cells that are converting into MDSCs. This idea was supported by the results of the kinetic study of the transcription factors; the relative expression levels of Cebp<sub>a</sub> and Pu.1 were higher in GM-CSF–treated CD49b<sup>+</sup> cells compared with GM-CSF/IL-2–treated cells. Several studies have reported that the ectopic expression of Cebp<sub>a</sub> and Pu.1 activates transdifferentiation (46, 47) and that these transcription factors synergistically program distinct responses to NF-κB activation (47), also can be induced by GM-CSF. Conversely, in the presence of IL-2, it is conceivable that STAT5 is phosphorylated and binds to target genes. We also identified the activation of STAT5 in NK cells in the presence of IL-2 by flow cytometric analysis (data not shown). It is assumed that IL-2–induced STAT5 then binds to the promoter of Cebp<sub>a</sub>/Pu.1 in competition with STAT3, thereby inhibiting transcriptional and/or epigenetical regulation by STAT3 as shown by the opposing STAT3–STAT5
regulation of IL-17 expression in $T_p$17 cells (48). The exact molecular mechanisms that regulate the conversion of these cells into MDSCs need to be investigated further.

IL-2 has been used as an immunotherapy for patients with metastatic melanoma and metastatic-renal cell carcinoma because of its strong stimulatory effect on $CD^+$ T cells and NK cells (49, 50). We showed that IL-2 prevented NK cells from converting into MDSCs in vitro and in vivo. However, no significant inhibitory effect on the number of MDSCs was observed in the spleen of IL-2 complex-treated tumor-bearing mice. A previous study reported that the cytokines produced from IL-2–activated NK cells prolong the survival of granulocytes and monocytes (22) and IL-2 complex strongly activates NK cells (18). In this case, the effect on MDSC numbers in IL-2 complex-treated mice could be underestimated because of the increase in granulocytes/monocytes, which share surface markers, such as Ly6C, Ly6G, CD11b, with PMN- and Mo-MDSCs.

Collectively, the data in this study suggest that NK cell reduction resulted from the conversion of NK cells into MDSCs in the middle of the maturation stage, before CD11b$^{hi}$ CD27$^{lo}$ NK cells give rise to the mature CD11b$^{hi}$CD27$^{lo}$ phenotype in the tumor environment. It could be a missing part of mechanisms by which results in NK cell reduction in tumor immunosuppressive environment, suggesting a novel tumor evasion mechanism. It is also noteworthy that our data suggested the possibility of conversion between lymphoid (NK cells) and myeloid cells (MDSCs), which are derived from distinct progenitor cells.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

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