Loss of miR-204 Expression Enhances Glioma Migration and Stem Cell-like Phenotype

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Abstract

Phenotypic similarities have long been recognized between subpopulations of glioma and neural stem cells. Many of these similar properties, including the robust abilities to self-renew, migrate, and invade, are hallmarks of glioma cells that render them extremely aggressive. However, the molecular mechanisms underlying this character, particularly in glioma stem-like cells that drive this disease, remain poorly understood. Here, we report the results of a differential miRNA expression screen that compared glioma and neural stem cells, where we found that miR-204 was markedly downregulated in both types of cells. Mechanistic investigations revealed that miR-204 simultaneously suppressed self-renewal, stem cell-associated phenotype, and migration of glioma cells by targeting the stemness-governing transcriptional factor SOX4 and the migration-promoting receptor EphB2. Restoring miR-204 expression in glioma cells suppressed tumorigenesis and invasiveness in vivo and increased overall host survival. Further evaluation revealed that the miR-204 promoter was hypermethylated and that attenuating promoter methylation was sufficient to upregulate miR-204 in glioma cells. Together, our findings reveal miR-204 as a pivotal regulator of the development of stem cell-like phenotypes and cell motility in malignant glioma cells. Cancer Res; 73(2); 990-9. ©2012 AACR.

Introduction

Gliomas are the most common malignant primary brain tumors in adults and exhibit a spectrum of aberrantly aggressive phenotype. Despite advances in treatments during past decades, the prognosis of the disease remains poor, with the median survival time approximately 12 to 14 months. The extremely poor prognosis of patients with gliomas is largely due to the high tendency of tumor invasiveness, which leads to severe structural and functional damage to the surrounding brain tissue, incomplete surgical resection, and high frequency of tumor recurrence (1). The molecular mechanisms underlying the aggressive malignant phenotype of glioma cells, however, remain largely unknown.

It has been established that glioma cells, or their subpopulations, share similar biological phenotype and gene expression profiles to those of neural stem cells (2, 3). Two common phenotypic features shared by glioma cells and neural stem cells are their robust abilities to migrate through brain parenchyma (4, 5) and self-renewal (6, 7). For neural stem cells, these characters typical of cellular “stemness” are essential for tissue homeostasis, regeneration, and repairing processes in the central nervous system. In malignant gliomas, however, these properties contribute to tumor invasiveness. Understanding whether there are common factors that simultaneously modulate the malignant phenotypes of glioma cells and the stemness of neural stem cells is expected to provide new insights in developing novel and effective therapies for glioma. In this context, identification of genetic and/or epigenetic factors that modulate the “stemness” phenotype of glioma cells is of great importance.

Deregulation of miRNAs has been implicated in the development and progression of nearly all tumor types (8). miRNAs play pivotal roles in development, particularly in modulating stem cell-specific pathways (9). This prompted us to identify and study miRNAs with common or similar expression patterns in neural stem cells and glioma cells. In this report, we describe the miR-204 as a miRNA that is significantly downregulated in both gliomas and neuronal stem cells, through methylation of its promoter. Reintroduction of miR-204 into glioma cells markedly suppresses the motility and invasion of glioma cells as well as their stem cell-like phenotype through targeting multiple regulators. Furthermore, our data show that miR-204 inhibits glioma tumorigenesis and invasion in the brain of mice. These findings, thus, reveal a novel pathway by

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Note: Supplementary data for this article are available at Cancer Research Online (http://cancerres.aacrjournals.org/).

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doi: 10.1158/0008-5472.CAN-12-2895

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which epigenetic modulation of miR-204 renders the abilities of robust self-renewal and aggressive invasion of glioma cells.

Materials and Methods

Cell lines and primary cell culture
Primary normal human astrocytes (NHA) were purchased from Sciencell Research Laboratories (San Diego, CA) and cultured under conditions suggested by the manufacturer. Glioma cell lines, including LN443, LN444, LN464, U118MG, T98G, U251MG, U87MG, D247MG, A172, LN319, LN382T, LN229, LNZ-308, and SNB19, were shared with Dr. Shi-Yuan Cheng’s laboratory at Northwestern University, Chicago, Illinois and cultured in Dulbecco’s Modified Eagle’s Medium (DMEM) supplemented with 10% FBS (HyClone) as previously described (10, 11). All cell lines were authenticated by short tandem repeat fingerprinting at IDEXX RADIL (Columbia, Montana) and Services at the SYSU Forensic Medicine Lab (Guang Zhou China).

Tissue specimens and patient information
Twofly freshly dissected glioma tumor specimens were clinically and histopathologically diagnosed at the First Affiliated Hospital of the Sun Yat-sen University. Six normal brain tissues were obtained by donation from individuals who died in traffic accidents and confirmed to be free of any pathologically detectable lesions. Prior consents of donors and approvals from the Institutional Research Ethics Committee were obtained.

Plasmid, retroviral infection, and transfection
A DNA fragment containing the hsa-miR-204 precursor flanked by a 500-bp genomic sequence at either end was inserted into the retroviral vector pMSCV-puro (Clontech). The open reading frames (ORF) of SOX4 and EphB2 genes generated by PCR were cloned into the retroviral vector pMSCV-neo (Clontech). Retroviral production and infection were carried out as previously described (10). 3’-Untranslated regions (UTR) of SOX4 and EphB2 were amplified and cloned into the downstream region of a luciferase gene in a modified pGL3 control vector (Promega). The miR-204 mimics and miR-204 inhibitor oligonucleotides and their corresponding control oligonucleotides were synthesized at Ribol Biotech (Guangzhou, China). Transfection of plasmids or oligonucleotides was carried out using the Lipofectamine 2000 reagent (Invitrogen) according to the manufacturer’s instruction.

Single cell tracks assay
Cells (5 × 10^4/well) were seeded in 6-well plates. Single cell tracks were obtained by the Axio Observer-Z1 time-lapse microscopy system (Carl Zeiss). Experiments were conducted according to the manufacturer’s instruction and repeated independently at least 3 times.

Flow cytometric analysis
Cells were dissociated into single-cell populations and labeled with a phycoerythrin-conjugated CD133/2 (293C3) antibody (Miltenyi Biotec). The expression level was analyzed using the EPICS XL flow cytometer with EXPO32 ADC software (Beckman Coulter).

Tumor xenograft live imaging
Glioma xenografts stably expressing firefly luciferase together with miR-204 or the corresponding control vector were orthotopically implanted in the brains of nude mice as previously described (11) and monitored using the IVIS Spectrum Live Imaging System (Caliper Life Sciences). Image calibration and visualization were carried out using the Life Imagine 4.2 software (Caliper Life Sciences).

Bisulfite genomic sequencing
Genomic DNA from primary NHA, glioma cell lines, and clinical specimens was bisulfite-modified with the Epitect Bisulfite Kit (Qiagen). Bisulfite-treated DNA was amplified with bisulfite-sequencing PCR (BSP) primers located in the TRPM3 promoter. PCR products were cloned using the pGEM-T Easy Vector System (Promega). Plasmids from single colonies were purified and sequenced.

Microarray data visualization
TCGA dataset were obtained from (12). GEO expression profile was obtained from (13). All microarray data were processed on the MeV 4.6 platform (14).

Statistical analysis
Statistical analyses, with the exception of the microarray data, were carried out using the SPSS 11.0 statistical software package. P values of less than 0.05 in all cases were considered statistically significant.

Results
miR-204 was downregulated in gliomas
To identify miRNAs with similar expression patterns in glioma and neural stem cells, we retrieved miRNA expression profiles from 2 datasets, The Cancer Genome Atlas (TCGA) and the GEO dataset GSE29759. As shown in Fig. 1A, miR-204 emerged as one of the most prominent downregulated miRNAs in the TCGA glioblastoma (GBM) dataset, accompanied by several other downregulated miRNAs reported previously, namely, miR-124,106a,17-5p and members of the miR-181 family (15, 16). As documented previously, miRNAs upregulated in gliomas, including miR-21 (17), miR-9 (20, 21), and miR-10b, were also screened out. (22, 23). Comparing the miRNA profiles in the TCGA dataset with those in neural stem cells identified that miR-204 was downregulated in both glioma and neural stem cells (Fig. 1B). To validate the expression pattern of miR-204 in gliomas, quantitative reverse transcription PCR was conducted with 6 normal brain tissue samples and 20 freshly dissected glioma samples. Consistent with the data presented in the TCGA profile, expression of miR-204 was markedly downregulated in high-grade glioma samples (WHO tumor grades III and IV) and to a lesser degree, decreased in WHO tumor grades I and II glioma samples, compared with normal brain tissues (Fig. 1C). Notably, 10 out of 14 glioma cell lines displayed significantly downregulated expression of miR-204 relative to that in 2 primary cultured normal human astrocytes (NHA; Fig. 1D).
Restoration of miR-204 in glioma cells suppressed cellular migration and invasion

To investigate whether miR-204 could modulate the migration and invasiveness of glioma cells, the SNB19 and LN382T glioma cell lines, which possessed the lowest levels of miR-204 expression among all tested glioma cell lines, were selected for further studies. By retrovirally transducing glioma cells with miR-204 at expression levels comparable with that of NHAs (Supplementary Fig. S1), we first examined the effect of miR-204 on cell migration using the wound-healing assay. As shown in Fig. 2A, compared with the vector-control cells, which spread to the center line within 16 hours, miR-204-transduced cells exhibited considerably slower migration and decreased cell spreading. Furthermore, we tracked the movement of glioma cells, using a time-lapse imaging system, and examined whether miR-204 altered the migrating behavior of individual glioma cells. As shown in Fig. 2B, miR-204-transduced-SNB19 and -LN382T cells generally retained their original positions, whereas vector-control cells moved away to a higher extent. Specifically, we measured the migration tracks of 10 individual cells within 5 hours and observed that miR-204-transduced cells moved within a very limited area. In contrast, the control cells spread out in a markedly larger area, with approximately 1.5- to 3-fold of increase in migration distance (Fig. 2C). To further determine the effect of miR-204 expression on motility of glioma cells, a transwell penetration assay was conducted. As shown in Fig. 2D, less number of miR-204 cells invaded across the membrane precoated with Matrigels when compared with control cells. Importantly, the 3-dimensional (3D) spheroid invasion assay showed that vector-control cells displayed a highly aggressive penetrating growth after 5 days, whereas the miR-204 transduced-cells grew in spherical colonies (Fig. 2E). Taken together, these observations suggested that miR-204 strongly inhibited the migration and invasion of glioma cells.

miR-204 Attenuated stem cell-like phenotype of glioma cells

Malignant gliomas tend to relapse after surgical resection, and this characteristic is believed to be largely attributable to the stem cell-like properties of a fraction of cells within a glioma tumor (2). As miR-204 is expressed at very low levels in both glioma and neural stem cells but at high levels in differentiated normal brain tissues, we determined the potential role of mir-R-204 in the development and maintenance of the stem cell-like property of glioma cells. Sphere-forming assay showed (Fig. 3A) that miR-204 overexpressing-cells formed much smaller neural spheres after 7 days of culture compared with control cells (~2-fold smaller in diameter), indicating markedly decreased self-renewal ability by miR-204.
To further investigate the potential role of miR-204 in suppressing the stemness property of glioma cells, we examined the expression of pluripotency factors in neuronal spheres formed by miR-204-transduced cells and control cells. As shown in Fig. 3B, the expression levels and patterns of pluripotency factors, including SOX2, NANOG, KLF4, and OCT4, were similar between spheres formed by vector control- and miR-204-transduced cells. However, at 48 hours after stimulation with 1% serum, miR-204-expressing cells rapidly lost their expression of pluripotency factors, retaining only 2% of expression in the neuronal spheres after induction for differentiation, while the vector-control cells retained expression of pluripotency factors at approximately 30% of the initial levels (Fig. 3B). Consistent with these results, at both time points, CD133-positive proportions were significantly lower in miR-204-transduced cells than those in control cells, and a faster declining rate of CD133 positivity was also displayed in miR-204-transduced cells (Fig. 3C). In further experiments, we co-stained for a glioma stem cell marker, Nestin, and an astrocyte differentiation marker, GFAP, during the process of differentiation. We observed a significantly faster decrease of Nestin and simultaneous increase of GFAP expression in cells transduced with miR-204 as compared with those in control cells (Fig. 3D). Taken together, these data showed that reintroduction of miR-204 in glioma cells reduced the stem cell-like population in glioma and greatly
attenuated the ability of stem cell-like glioma cells to retain stemness.

Reintroduction of miR-204 induced loss of invasion and tumorigenecity in vivo

Next, we test whether miR-204 is involved in tumorigenesis and invasiveness of glioma cells in vivo. An orthotopic glioma xenograft model was employed by xenotransplanting luciferase-expressing glioma cells transduced with miR-204 or corresponding control vectors in the brains of nude mice. Tumor growth was monitored using a live animal bioluminescence imaging system during the experiment. As shown in Fig. 4A, both miR-204-transduced cell lines exhibited significantly slower growth after implantation. For the highly proliferative SNB19 cells, results from luminescence images revealed a high degree of suppression on the growth of miR-204-transduced cells in contrast to the larger intracranial tumor formed by the control cells at day 10 after implantation. All mice bearing vector-control cells died 18 days after implantation, while only 1 mouse implanted with miR-204-transduced SNB19 cells died at day 21 (Fig. 4B). Similar inhibition of tumorigenicity of miR-204 transduction was also observed in LN382T cells, which are known to grow more slowly. Specifically, 1 mouse with miR-204 cells died after 35 days of tumor cells implantation, and all vector-cell-implanted mice were dead at day 29 (Fig. 4B). These data suggested that miR-204 was a strong inhibitor of glioma tumorigenecity in vivo.

Furthermore, immunohistochemical (IHC) analysis showed that tumors formed by vector-control cells exhibited an extensive branch-like growing pattern that spread into the surrounding tissue. By contrast, miR-204-overexpressing cells formed an oval-shaped tumor with smooth margins and...
noninvasive front (Fig. 4C), further validating that the invasive behavior of glioma could be suppressed by miR-204 in vivo.

**Core regulators of migration and stemness were direct targets of miR-204**

As miRNAs exert their functions by targeting multiple transcripts that are coordinately orchestrated in a biological process, we screened for targets of miR-204 using the TargetScan Program and identified 2 conserved binding sites for miR-204 in the 3'-UTR region of the SOX4 gene, a known core regulator governing the stemness of both glioma and neural stem cells (24, 25). Meanwhile, EphB2, which is known to be required for the migration of both glioma and neural stem cells (26–29), was also found as a potential target gene of miR-204 (Fig. 5A). To validate whether SOX4 and EphB2 were targets of miR-204, we transiently expressed miR-204 in SNB19 and LN382T cells, both of which possess low endogenous miR-204 expression. Significantly decreased protein expression of both SOX4 and EphB2 was observed after transfection of a miR-204 mimic oligonucleotide. In contrast, inhibition of miR-204 by transfecting a miR-204 inhibitor oligonucleotide in LN443 and U251MG cells, which express high levels of endogenous miR-204, led to elevated expression of both proteins (Fig. 5B). Furthermore, in the in vivo glioma model described earlier, lower levels of SOX4 and EphB2 staining were detected in tumors formed by miR-204-transduced cells, as compared with those in the control tumors (Supplementary Fig. S2).

To determine whether inhibition of miR-204 on SOX4 and EphB2 expression is mediated by posttranscriptional effects on the 3'-UTRs of both genes, we constructed a luciferase reporter plasmid containing 3'-UTRs for SOX4 and EphB2, respectively. As shown in Fig. 5C, ectopic expression of miR-204 markedly decreased, while inhibition of miR-204 was significantly elevated with the reporter luciferase activities (Fig. 5C), suggesting that miR-204 posttranscriptionally inhibited SOX4 and EphB2 expression. Furthermore, mutagenesis in miR-204 target sites of the SOX4 or EphB2 3'-UTR linked to the luciferase reporter confirmed the site-specific effect of miR-204 (Fig. 5C). To examine the interaction between miR-204-containing miRNP and target mRNAs, we pulled down Ago1 and its associated RNA in cells transfected by control miRNA or the miR-204 mimic. Quantitative PCR (qPCR) assessment of the SOX4 and EphB2 3'-UTR regions revealed high levels of SOX4 and EphB2 transcripts enrichment in the miRNP of miR-204 mimic-transfected cells, strongly suggesting that miR-204 could guide miRNP to target SOX4 and EphB2 (Fig. 5D). Finally, we assessed the expression of SOX4 and EphB2 in 20 glioma specimens shown in Fig. 1C, and found significant inverse correlation between miR-204 and its 2 targets (Supplementary Fig. S3). This observation indicates miR-204 may act as a regulator for SOX4 and EphB2 expression in clinical glioma specimens.

To determine the functional significance of SOX4 and EphB2 genes in miR-204-induced phenotype, we further stably expressed SOX4 ORF, or EphB2 ORF, that did not contain the 3'-UTR in miR-204-transduced cells. As shown in Fig. 5E, restored SOX4 expression could significantly abrogate the inhibitory effect of miR-204-induced sphere formation. Similarly, reexpression of EphB2 ORF in miR-204-transduced cells restored the invasiveness of glioma cells induced by miR-204.
expression (Fig. 5F). Taken together, these data revealed that by coordinately suppressing core regulators of glioma stemness and migration, miR-204 conferred the abilities of self-renewal and invasion in glioma cells.

Hypermethylation in the promoter region of the miR-204 gene

We next sought to investigate the molecular mechanism that mediates the downregulation of miR-204 in glioma cells. Interestingly, miR-204 was an intronic miRNA located between exons 7 and 8 of the TRPM3 gene (30). Consistent with a previous report (30), expression of mature miR-204 and pri-miR-204 linearly correlated with that of TRPM3 in NHA and several glioma cell lines (Fig. 6A), indicating that miR-204 and TRPM3 might share common regulatory mechanism(s). To examine the epigenetic modification of this shared promoter region, we retrieved the CpG-rich loci of TRPM3 promoter from the UCSC genome browser (Fig. 6B) and assessed for their methylation status via bisulfite genomic-sequencing PCR (BSP). The BSP result showed that in NHA, as well as in glioma cells with relatively higher levels of miR-204 expression, the CpG islands were mostly not methylated (Fig. 6C). By contrast, in genomic DNA obtained from glioma cells with downregulated miR-204 and from clinical glioma tumor samples, the promoter CpG islands of miR-204 were hypermethylated in most glioma specimens, strongly suggesting an essential role of promoter methylation in miR-204 downregulation (Fig. 6C and Supplementary Fig. S4). To further validate this finding, we treated the cells with DNA methyltransferase inhibitor 5-aza-dC for 72 hours and found that, in cells with low levels of miR-204 expression, inhibition of DNA methylation significantly promoted the expression of miR-204 whereas, in NHA and glioma cells without miR-204 promoter hypermethylation, miR-204 expression was not affected (Fig. 6D). Correspondingly, 5-aza-dC treatment could induce downregulation of SOX4 and EphB2 expression in glioma cell lines examined with low basal
levels of miR-204 expression but could not suppress the expression of SOX4 and EphB2 in cells with high levels of endogenous miR-204 (Supplementary Fig. S5). Taken together, our data suggested that, in glioma cells, miR-204 downregulation was attributable to promoter methylation.

Discussion

Development of stem cell-like properties in glioma cells has been recognized as a hallmark contributing to disease progression and generation of cellular population(s) robustly empowered to migrate and invade the surrounding brain tissue, to be resistant to chemo- or radiotherapies, resulting in frequent tumor recurrence, and to proliferate and colonize from a small number of initiating cells, in addition to other aspects of tumor malignancies (31). Accumulated evidence supports the notion that such a process is regulated by signaling pathways similar to those governing the stemness of neural stem cells. Notably, activation of these pathways requires coordinated expression and cooperative action of several core regulators. For example, SOX4 has recently been identified as a determinant factor of glioma stemness, which forms a complex with OCT4 and subsequently activates the transcription of SOX2, 1 of the core pluripotency reprogramming factors (24, 32). This transcriptional axis is essential for the maintenance of glioma stem cell population and, more importantly, blocking this pathway greatly improves animal survival in a glioma xenograft model (24). As a short half-life (<1 hour) protein (33), the expression level of SOX4 is tightly controlled transcriptionally and posttranscriptionally. In endometrial cancer, downregulation of miR-129-2 mediates the overexpression of SOX4, suggesting a significance of miRNA-mediated regulation in sustaining the expression of this stemness-associated oncoprotein (34). In glioma cells, however, the level of miR-129 has not been reported or screened out in high-throughput assays to have significant change. In this study, we identified miR-204 as a miRNA commonly downregulated in both types of neural cells. We then further determined the targets of miR-204 and the consequent modulation of stem cell-like properties of glioma cells. Our data showed that miR-204 acts to inhibit the expression of SOX4 and subsequently suppress the stem cell-like phenotype of glioma cells, and the miR-204-downregulated status in glioma cells leads to generation of a glioma cell population in which marked stemness is developed. In miR-204-transduced cells, ectopic expression of SOX4 is able to rescue neural sphere-formation ability attenuated by miR-204, confirming that SOX4 is a major target of miR-204 and responsible for maintaining the stem cell population in human gliomas. Notably, miR-204 not only suppresses self-renewal but also migration of glioma cells, which is another hallmark of neural stem cells (5). This study has also identified EphB2 as a major migration regulator directly targeted by miR-204. Previously, EphB2 has been reported to promote stem cell migration in both physiological and injury-repairing conditions (5, 26). Elevated expression and enhanced phosphorylation of EphB2 has also been found in glioma cells (29, 35). More importantly, EphB2 has identified to be a critical promoter of tumor progression.
migration in glioma through interacting with the FAK and activating FAK-mediated signal (28). The mechanism regulating EphB2 expression in glioma cells, however, is still unclear. Our data thus shows a novel posttranscriptional regulator of EphB2, namely, miR-204, epigenetic silencing of which enables elevation of EphB2 in glioma cells and enhances EphB2-mediated migration and invasion.

Sustained activation of regulatory programs shared by glioma and neural stem cells requires orchestrated transcription and posttranscriptional regulation of gene expression. Because of the multitalent property and the network effect, miRNAs are of great importance in the suppression of pluripotency of embryonic stem cells by targeting reprogramming factors and initiating differentiation (9). During the process of differentiation, an optimal miRNA expression pattern enables proper development of the organism (36). In tumor cells, however, many of the stem cell-specific miRNA expression patterns are reacquired, such as the loss of neural-specific miR-124 expression that promotes the stemness of glioma (15, 37). Consistently, we find that miR-204, which is downregulated, is thereby expected to play significant roles in both glioma cells and neural stem cells. miR-204 downregulation not only promotes cellular stemness but also the biological properties essential to the lethality of the disease, namely, migration and invasion abilities of glioma cells, both in vitro and in vivo. Most importantly, restoration of miR-204 in glioma cells greatly abrogates the aggressiveness of glioma cells.

Lastly, we showed that reduction of miR-204 is attributed to downregulated transcription due to methylation of the promoter of its host gene TRPM3. TRPM3 is a member of the transient receptor potential melastatin (TRPM) family that has moter of it host gene downregulated transcription due to methylation of the promoter of it host gene. Sustained activation of regulatory programs shared by glioma and neural stem cells requires orchestrated transcription and posttranscriptional regulation of gene expression. Because of the multitalent property and the network effect, miRNAs are of great importance in the suppression of pluripotency of embryonic stem cells by targeting reprogramming factors and initiating differentiation (9). During the process of differentiation, an optimal miRNA expression pattern enables proper development of the organism (36). In tumor cells, however, many of the stem cell-specific miRNA expression patterns are reacquired, such as the loss of neural-specific miR-124 expression that promotes the stemness of glioma (15, 37). Consistently, we find that miR-204, which is downregulated, is thereby expected to play significant roles in both glioma cells and neural stem cells. miR-204 downregulation not only promotes cellular stemness but also the biological properties essential to the lethality of the disease, namely, migration and invasion abilities of glioma cells, both in vitro and in vivo. Most importantly, restoration of miR-204 in glioma cells greatly abrogates the aggressiveness of glioma cells.

Column 2...

our current finding that the promoter of TRPM3 is hypermethylated and TRPM3 mRNA is significantly downregulated in gliomas warrants further studies on the potential tumor-suppressive function of the TRPM3 gene in glioma. Moreover, it also remains highly interesting whether TRPM3 and miR-204 might cooperate with each other in the pathogenesis of glioma. Taken together, our present study provides new insights in understanding the stemness phenotype developed by glioma cells and, therefore, might contribute to future development of novel antglioma strategies.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Conception and design: Z. Ying, M-F Li
Development of methodology: Z. Ying, Y. Li, X. Zhu, Y. Yang, H. Tian
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): Z. Ying, Y. Li, J. Wu, H. Tian, W. Li
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): Z. Ying, Y. Li, J. Wu, X. Zhu, Y. Yang, H. Tian, W. Li
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Grant Support
This work was supported by the Ministry of Science and Technology of China (grant no. 973-2011CB11305), the National Science Foundation of China (Grant nos. 81071647, 81071762, 30900415, 81272339, and 81272417), the Guangdong Recruitment Program of Creative Research Groups (grant nos. 2009010058 and 201003060003), the National Science and Technique Major Project (grant nos. 201005022-2, 2012ZX10004213, and 311030), the Key Science and Technique Research Project of Guangzhou Province (grant no. 12292060029). United States National Institutes of Health (grant nos. CA130966 and CA138911), a scholar award from the Zeller Family Foundation, and a research award for Brain Cancer from the James S. McDonnell Foundation.

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Received July 4, 2012; revised November 15, 2012; accepted November 17, 2012; published OnlineFirst November 29, 2012.

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doi:10.1158/0008-5472.CAN-12-2895

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