The HSP90 Inhibitor Ganetespib Synergizes with the MET Kinase Inhibitor Crizotinib in both Crizotinib-Sensitive and -Resistant MET-Driven Tumor Models

Naoto Miyajima, Shinji Tsutsumi, Carole Sourbier, Kristin Beebe, Mehdi Mollapour, Candy Rivas, Soichiro Yoshida, Jane B. Trepel, Ying Huang, Manabu Tatokoro, Nobuo Shinohara, Katsuya Nonomura, and Len Neckers

Abstract

The proto-oncogene MET is aberrantly activated via overexpression or mutation in numerous cancers, making it a prime anticancer molecular target. However, the clinical success of MET-directed tyrosine kinase inhibitors (TKI) has been limited due, in part, to mutations in the MET kinase domain that confer therapeutic resistance. Circumventing this problem remains a key challenge to improving durable responses in patients receiving MET-targeted therapy. MET is an HSP90-dependent kinase, and in this report we show that HSP90 preferentially interacts with and stabilizes activated MET, regardless of whether the activation is ligand-dependent or is a consequence of kinase domain mutation. In contrast, many MET-TKI show a preference for the inactive form of the kinase, and activating mutations in MET can confer resistance. Combining the HSP90 inhibitor ganetespib with the MET-TKI crizotinib achieves synergistic inhibition of MET, its downstream signaling pathways, and tumor growth in both TKI-sensitive and -resistant MET-driven tumor models. These data suggest that inclusion of an HSP90 inhibitor can partially restore TKI sensitivity to previously resistant MET mutants, and they provide the foundation for clinical evaluation of this therapeutic combination in patients with MET-driven cancers.

Cancer Res; 73(23); 7022–33. ©2013 AACR.

Introduction

The proto-oncogene product MET is a receptor tyrosine kinase whose ligand is the hepatocyte growth factor/scatter factor (HGF/SF). HGF binding to MET induces receptor dimerization and trans-phosphorylation, and promotes activation of several signaling networks including phosphoinositide 3-kinase (PI3K)–AKT and mitogen-activated protein/extracellular signal–regulated kinase (MEK) pathways (1). Activating point mutations in the MET kinase domain are implicated in the etiology of hereditary papillary renal carcinoma and have also been detected in sporadic papillary renal carcinoma, lung cancer, and gastric cancer (2–5). Furthermore, amplification of the MET gene locus has been detected in patients with gastric and metastatic colorectal cancers (6, 7). Cell lines engineered to express high levels of wild-type MET or constitutively active mutant MET display a proliferative, motogenic, and invasive phenotype, and form metastatic tumors in nude mice (8–11).

MET is a validated molecular target for cancer therapy, and MET tyrosine kinase inhibitors (TKI) represent a promising treatment modality. Crizotinib, an orally available ATP-competitive and selective small-molecule inhibitor of MET, exhibits marked antitumor activity in several MET-dependent xenograft models (12). A recent phase 2 study of the dual MET/VEGFR2 inhibitor foretinib in patients with papillary renal cell carcinoma reported an overall response rate (using the Response Evaluation Criteria in Solid Tumors version 1.0) of 13.5%, and the presence of a germline MET mutation was highly predictive of a response (13). However, recent studies suggest that primary (de novo) resistance to TKIs is likely to be encountered in tumors harboring certain activating MET mutations. Thus, although crizotinib can inhibit the activity of most MET mutants, some constitutively active mutants, including MET-L1213V and -Y1248H, are resistant to this inhibitor (12, 14). Additional studies have suggested that these mutations not only mediate primary resistance to MET inhibitors but may also play a role in acquired resistance to MET-TKI (15–17).

HSP90 is a molecular chaperone whose association is required for the stability and function of multiple mutated or overexpressed signaling proteins, including many kinases, that promote the growth and survival of cancer cells. Small-molecule HSP90 inhibitors cause HSP90 to dissociate from its
clients, resulting in their destabilization and eventual degradation. HSP90 inhibitors have been validated in numerous preclinical tumor models, and have shown promising activity in several clinical trials (18–20).

HSP90 inhibitors have been reported to destabilize MET (21, 22), as well as several MET-activated downstream signaling proteins, including AKT and RAF (23–25). Based on these findings, HSP90 inhibitors are expected to effectively interdict MET signaling at multiple points to inhibit the proliferation of MET-driven cancer cells. Indeed, the HSP90 inhibitors ganetespib (STA-9090) and SNX-2112 have each shown potent activity in several preclinical MET-driven tumor models, including those resistant to MET-TKI (26, 27). However, a detailed study of the MET-HSP90 interaction and its role in supporting MET activation has not been reported, nor has the possible synergy between HSP90 inhibition and MET-TKI been rigorously examined.

In the current study, we show that HSP90 interacts preferentially with activated MET, regardless of whether activation depends on HGF engaging the receptor or is mediated by kinase domain mutation. Further, we show that the activated MET fraction is most sensitive to HSP90 inhibition. The HSP90 inhibitor ganetespib displays synergy, both in vitro and in vivo, with the MET-TKI crizotinib in cells expressing wild-type MET. Unexpectedly, low-dose ganetespib also partially restores crizotinib sensitivity, both in vitro and in vivo, to cells and tumors expressing TKI-resistant MET mutants. Our findings support the use of HSP90 inhibitors to overcome or delay the initiation of resistance to MET-TKI, and they provide the basis for clinical evaluation of this combination in patients with MET-driven cancers.

Materials and Methods
Cell lines
MKN45, H1993, and HEK293 cell lines (American Type Culture Collection) were maintained under 5% CO2 at 37°C in either RPMI1640 (MKN45, H1993), or Dulbecco’s Modified Eagle Medium (DMEM; HEK293) supplemented with 10% FBS. NIH3T3 cell lines stably expressing either wild-type or mutant MET proteins (V1238I, H1112Y, Y1248H, L1213V, M1268T, and V1110F) were kindly provided by Dr. Laura Schmidt [National Cancer Institute (NCI), Bethesda, Maryland]. These cells were cultured in DMEM with 10% FBS and 0.5 mg/mL G-418.

Antibodies and reagents
Antibodies to MET, phospho-Tyr1234/35 MET, Akt, phospho-Ser473 Akt, Erk1/2, phospho-Thr202/Tyr204 Erk1/2 (Cell Signaling Technology), HSP90 (StressGen), V5 (Invitrogen), ubiquitin (Santa Cruz Biotechnology), and α-tubulin (Calbiochem) were used for immunoprecipitation and/or immunoblotting. Geldanamycin was obtained from the Developmental Therapeutics Program, NCI. Ganetespib and crizotinib were obtained from Synta Pharmaceuticals. Mouse immunoglobulin G (IgG), MG132, and recombinant human HGF were purchased from Millipore, Sigma, and R&D Systems, respectively. FuGene6 (Roche) was used for transient transfection.

Immunoprecipitation and immunoblotting
These experiments were performed as previously described (28). Briefly, cells were lysed in buffer containing 20 mmol/L Hepes, 100 mmol/L NaCl, 1 mmol/L MgCl2, 0.1% Nonidet P-40, 20 mmol/L Na3MoO4, phosphatase, and protease inhibitors. Immunoprecipitates or cell lysates were resolved by 4% to 20% SDS–PAGE, transferred to a nitrocellulose membrane, and probed with respective antibodies.

Plasmid constructs
Human MET cDNA was kindly provided by Dr. Don Bottaro (NCI, Bethesda, Maryland). Chimeric mutant MET/EGFR was generated using PCR and ligated into a pcDNA vector (Invitrogen) in-frame with the C-terminal V5 tag. A point mutation was made by using QuikChange (Stratagene) according to the manufacturer’s instructions.

Cell proliferation assay
Cells were seeded in 96-well plates at a density of 5,000 cells per well and incubated for 24 hours, followed by addition of drugs. After 48 hours, MTT solution (Sigma) was added and plates were incubated at 37°C for 3 hours. The optical density at 570 nm was determined by a spectrophotometer (Bio-TEK). The combination index (CI) was calculated by the median-effect method of Chou and Talalay (29) using CalcuSyn software (Biosoft).

Cell-cycle analysis
Cells were plated at a density of 3 × 104 in 10-cm dishes and incubated for 24 hours, followed by the addition of drugs. After 48 hours, cells were fixed with 70% ethanol for 4 hours at −20°C, and then suspended in a solution containing 0.04% digitonin. Cells were incubated at 37°C for 1 hour in a solution containing RNase A (100 μg/mL; Novagen) and propidium iodide (50 μg/mL), and then analyzed with a FACScan flow cytometer (Becton Dickinson).

Colony formation assay
Cells were plated at a density of 1 × 104 in 60-mm dishes containing 0.4% top low-melting agarose and 0.5% bottom low-melting agarose medium, and cultured for 3 weeks. Colonies with a diameter of more than 0.1 mm were counted in five random high-power fields.

Migration and invasion assay
Twelve-well polycarbonate Transwell chambers with 8-μm pores (Corning) coated with Matrigel (BD Biosciences) were used. Cells (1 × 105) were added to the top well in serum-free medium with indicated drugs. In the bottom well, medium with 10% FBS was used as the chemottractant. After 48 hours, cells on the upper surface of the filter were carefully removed with cotton pads. Migrated cells were fixed in 3.7% formaldehyde and stained with 4′,6-diamidino-2-phenylindole (DAPI). The invaded cells were fixed and stained with Diff-Quik Stain kit (Dade Behring). The number of cells in five random high-power fields was determined.
Animal experiments

Animal experiments and procedures were carried out in accordance with the Guide for the Care and Use of Laboratory Animals as adopted and promulgated by the National Institutes of Health. To establish tumor xenografts, cells (1 × 10^6) were injected into the flank of female Nu/Nu mice (6 weeks of age; Taconic). After tumors reached a mean volume of 100 mm^3, mice were administered crizotinib in water by oral gavage or ganetespib in the diluent 10/18 DRD [10% dimethyl sulfoxide (DMSO), 18% Cremophor RH40, 3.6% dextrose in water] by intraperitoneal injection. Tumor volume was calculated as the product of length × width^2 × 0.5. Fractional tumor volume (FTV) relative to untreated controls was determined as described previously (30). On the final day of the study, mice were humanely euthanized, and tumors were resected and pulverized using a homogenizer (Kinematica). Protein lysates were subjected to immunoblotting.

Statistical analysis

To determine the statistical significance of experimental data, we used the unpaired Student t test followed by the Bonferroni post hoc test for multiple comparisons. Data represent the mean ± SD. All P values less than 0.05 were considered statistically significant relative to control, and designated with an asterisk (*). Statistical analysis was done with the JMP software (SAS Institute).

Results

Wild-type MET is an HSP90-dependent kinase

We assessed the interaction of HSP90 and wild-type MET (wtMET) in the gastric cancer cell line MKN45, which over-expresses wtMET. As shown in Fig. 1A, endogenous HSP90 coprecipitated, albeit weakly, with wtMET. Previous reports by us and others have suggested that the activated states of some HSP90-dependent kinases have a greater dependence on HSP90 (31–33). To determine if this were the case for MET, we compared the degree of wtMET–HSP90 interaction in the presence and absence of the MET ligand HGF. Coimmunoprecipitation of HSP90 with MET clearly increased upon HGF stimulation and correlated with an increased population of activated (phosphorylated) MET (Fig. 1B). Next, we compared the sensitivity of total and activated wtMET protein to HSP90 inhibition. Exposure of MKN45 cells to the HSP90 inhibitor geldanamycin (GA) reduced the steady-state expression of both total and activated MET protein in a dose- and time-dependent manner, but the activated MET fraction was most sensitive to HSP90 inhibition (Fig. 1C). Loss of MET protein expression occurred much more rapidly in the presence of geldanamycin than after inhibition of protein synthesis (Supplementary Fig. S1B). Taken together, these data suggest that Hsp90-mediated stabilization of activated MET is required for optimal MET kinase activity. To provide further support for this hypothesis, we examined the ability of constitutively active MET-Y1248H to phosphorylate its substrate Gab1 (35) in NIH3T3/MET-Y1248H cells treated with the Hsp90 inhibitor ganetespib (Supplementary Fig. S2A). Importantly, Gab1 is not an Hsp90 client, and total endogenous Gab1 expression was not affected by ganetespib treatment. Nonetheless, ganetespib caused a dose-dependent reduction of Gab1 phosphorylation in these cells.

To better understand where HSP90 interacts with MET and how it might affect its kinase activity, we examined the αC-β4 loop located in the MET kinase domain. The amino acid composition of this short loop was shown to be important for HSP90 binding to a number of kinases including ErbB2 (36, 37). We reported previously that although the kinase domains of HSP90-dependent ErbB2 and HSP90-independent EGFR (ErbB1) are highly homologous, they vary in sequence in the αC-β4 loop. Replacement of the αC-β4 loop in ErbB2 with that of the EGFR abrogated HSP90 binding, whereas replacing the EGFR αC-β4 loop with that of ErbB2 conferred Hsp90 association and dependence (37). When we compared the sequence of the αC-β4 loops in EGFR and MET, we found a divergence in eight of 10 residues between amino acids 1,132 and 1,141 (numbering for MET kinase domain residues; highlighted in gray, Fig. 2C). To investigate whether these residues are an important determinant of MET/HSP90 interaction and dependence, we used the unpaired Student t test followed by the Bonferroni post hoc test for multiple comparisons. Data represent the mean ± SD. All P values less than 0.05 were considered statistically significant relative to control, and designated with an asterisk (*). Statistical analysis was done with the JMP software (SAS Institute).
interaction, we replaced the eight divergent residues in V5-tagged MET-Y1248H with those found in the aC-β loop of the EGFR (V5-MET-Y1248H/EGFR, highlighted in red, Fig. 2C). After transient transfection into HEK293 cells, we immunoprecipitated the tagged MET proteins with antibody to V5 and we assessed the relative association of both MET-Y1248H and MET-Y1248H/EGFR with HSP90. Although V5 immunoprecipitation affinity-purified equivalent amounts of tagged MET proteins from cell lysates, coimmunoprecipitated HSP90 was present in only trace amounts in MET-Y1248H/EGFR immune pellets. This was in distinct contrast to the amount of HSP90 found associating with MET-Y1248H protein (Fig. 2D), confirming that the amino acid composition of the aC-β loop in the MET-kinase domain determines HSP90 association.

We noticed that, concurrent with loss of HSP90 association, the activated fraction of immunoprecipitated MET-Y1248H/EGFR was dramatically reduced compared with MET-Y1248H. To confirm this result, we blotted equivalent amounts of protein lysate from the transfected cells with antibody to phospho-MET. Substitution of the aC-β loop in MET-Y1248H nearly completely abrogated its constitutive phosphorylation (Fig. 2E). To ascertain whether reduced constitutive activation also reflected an inability to respond to ligand, we compared the ability of HGF to activate MET-Y1248H/EGFR and wtMET in transiently transfected serum-starved HEK293 cells. The data show that, although constitutive phosphorylation of MET-Y1248H/EGFR is even less than that of wtMET (Fig. 2F, "HGF−" lanes), both proteins are comparably activated by HGF (Fig. 2F, "HGF +" lanes). Further, we found...
in vitro MET-TKI contrast, the inclusion of geldanamycin restores sensitivity to

Miyajima et al.

Figure 2. Activated MET depends on HSP90 association for its stability. A, constitutively active mutant MET strongly interacts with HSP90. NIH3T3 cells stably expressing wild-type (wt) MET or constitutively active mutant MET (Y1248H) were lysed and subjected to IP with anti-MET antibody followed by immunoblotting with anti-HSP90, anti-MET, and anti-p-MET antibodies. B, constitutively active mutant MET is more sensitive to HSP90 inhibition than is wtMET. NIH3T3 cells stably expressing wtMET or constitutively active mutant MET (Y1248H) were incubated in medium supplemented with serum including HGF and increasing concentrations of GA for 24 hours. Cells were collected and subjected to immunoblotting with indicated antibodies. C, alignment of the amino acid sequence of MET and EGFR kinase domain αC-β4 loops. Amino acid sequences (in one-letter code) of MET and EGFR in the αC-β4 loops were aligned by using MacVector software. Star denotes identity, and dot denotes similarity. Residues that differ between MET and EGFR are shaded; MET/EGFR denotes MET with highlighted residues from EGFR. Amino acid residues in MET and EGFR are numbered from the starting methionine. D, MET-Y1248H/EGFR loses association with HSP90. Expression vectors encoding V5-tagged MET-Y1248H, MET-Y1248H/EGFR or an empty vector (pcDNA, used as a control) were transfected into HEK293 cells. After 48 hours, cells were lysed and subjected to IP with anti-V5 antibody followed by immunoblotting with anti-HSP90, anti-V5, and anti-p-MET antibodies. E, steady-state phosphorylation of MET-Y1248H/EGFR is reduced compared with MET-Y1248H. Expression vectors encoding V5-tagged MET-Y1248H, MET-Y1248H/EGFR or an empty vector (pcDNA) were transfected into HEK293 cells. After 48 hours, cells were lysed and subjected to immunoblotting as shown. F, MET-Y1248H/EGFR retains HGF sensitivity. HEK293 cells were transfected with V5-tagged empty plasmid, wtMET, or MET-Y1248H/EGFR (2 μg each). After 48 hours, cells were serum-starved overnight and then treated for 20 minutes with 1 μmol/L HGF. Samples were then lysed in buffer (20 mmol/L Hepes, 100 mmol/L NaCl, 1 mmol/L MgCl₂, 0.1% NP-40, 20 mmol/L Na₃MoO₄, phosphatase, and protease inhibitors). Equal amounts of total cell lysate were resolved by Western blotting and probed with indicated antibodies. After normalization to total MET protein, HGF increased the activated fractions of wtMET and MET-Y1248H/EGFR 4.8- and 8.9-fold (by densitometric analysis), respectively. G, HEK293 cells were transfected as in (F), treated with HGF and exposed to increasing concentrations of crizotinib (16 hours). MET-Y1248H/EGFR, which displays reduced interaction with Hsp90, is markedly more sensitive to the TKI compared with MET-Y1248H.

MET-Y1248H/EGFR to be more sensitive to crizotinib compared with MET-Y1248H (Fig. 2G), supporting an inverse correlation between Hsp90 interaction and crizotinib sensitivity. This hypothesis is consistent with in vitro data showing that the TKI sensitivity of constitutively active, bacterially expressed MET-Y1248H protein is antagonized by preincubation with purified Hsp90 before in vitro kinase assay. In contrast, the inclusion of geldanamycin restores sensitivity to MET-TKI in vitro (Supplementary Fig. S2B).

The MET inhibitor crizotinib synergizes in vitro with ganetespib in wtMET-driven cells

Most MET-TKIs are ATP-competitive inhibitors and preferentially target the inactive form of MET (38). In contrast, the data presented here show that HSP90 inhibitors appear to preferentially target the activated MET and, at least in vitro, appear to enhance the sensitivity of activated MET to TKIs. To determine whether a similar phenomenon occurs in cells, we first assessed the effect of the MET-TKI crizotinib and of the
second-generation HSP90 inhibitor ganetespib, alone or combined, on the activation state of wtMET, and on downstream signaling to AKT and ERK1/2 in the wtMET-overexpressing gastric carcinoma cell line MKN45 and the non–small cell lung carcinoma cell line H1993. In both cell lines, the addition of low-dose ganetespib dramatically enhanced the impact of crizotinib on the activation of all three kinases (Fig. 3A). Although AKT itself is an HSP90 client, the lowest dose of ganetespib (10 nmol/L) had minimal activity when used alone in either cell line.

To investigate whether enhanced inhibition of these signaling pathways correlated with enhanced cellular activity, we...
performed MTT assays using MKN45 cells. The addition of increasing concentrations of ganetespib to a fixed set of crizotinib concentrations shifted the dose–response curves to the left in a (ganetespib) concentration-dependent manner (Fig. 3B), suggesting that the combination of these two drugs might be synergistic. We confirmed this to be the case by calculating the CI using the median-effect method of Chou and Talalay (29). CI values of less than 1 indicate synergy, whereas CI values of more than 1 reflect antagonism. Based on these data, ganetespib and crizotinib clearly synergize in MKN45 cells (CI = 0.6; Fig. 3C). Cell-cycle analysis revealed an increase in G1 phase in ganetespib/crizotinib-treated cells (as well as a small increase in sub-G1), and this was not observed in cells treated with either drug alone (Fig. 3D).

Next, we determined the effect of combination therapy on the transformed phenotype of MKN45 cells by assessing anchorage-independent colony formation. Low-dose crizotinib (10 nmol/L) and ganetespib (10 nmol/L) alone had marginal effects on colony formation. However, a combination of both drugs inhibited MKN45 colony formation by more than 90% (Fig. 3E). In addition, combination treatment more effectively inhibited MKN45 cell migration, compared with single-agent treatment at equivalent concentrations (Supplementary Fig. S3).

**Crizotinib-resistant MET mutants retain sensitivity to HSP90 inhibition**

*De novo* or acquired resistance to MET-TKI as a consequence of MET mutation remains a key therapeutic challenge to the clinical utility of these drugs. Several germline and somatic mutations in the tyrosine kinase domain of MET have been identified in hereditary papillary renal cancers, and these tumors are MET-driven (5, 13). A majority of these MET mutants are constitutively active, and some demonstrate *de novo* resistance to MET-TKI (9, 39). Because MET activity depends on association with HSP90, we speculated that HSP90 inhibitors might also retain activity toward a range of MET mutants, including those that are TKI resistant. To examine this possibility, we compared the inhibitory activity of single-agent crizotinib with that of single-agent ganetespib in six MET mutants—four that retain TKI sensitivity (MET-V1238I, MET-H1112Y, MET-M1268T, and MET-V1110I) and two that are resistant to MET inhibitors (MET-Y1248H and MET-L1213V). For TKI-sensitive mutants, both MET phosphorylation and the activity of downstream signaling pathways (pAKT and pERK) were equally or more potently inhibited by ganetespib than by crizotinib (Fig. 4A and Supplementary Fig. S4). However, in contrast to crizotinib, ganetespib retained equivalent or greater inhibitory activity in TKI-resistant mutants compared with wtMET (Fig. 4B).

To evaluate whether combination therapy was beneficial in the MKN45 model, we chose crizotinib (12.5 mg/kg) and ganetespib (12.5 mg/kg) dosing that provided approximately 50% growth inhibition when used singly (Supplementary Fig. S5). Treatment with this drug combination proved to be effective in both xenograft models, although at the maximum dose used in this study (50 mg/kg administered 3 times per week), somewhat greater activity was seen in wtMET xenografts (95% inhibition of tumor growth) compared with MET-Y1248H xenografts (82% growth inhibition; Supplementary Fig. S5).

To determine whether there may be therapeutic benefit in combining ganetespib with crizotinib *in vivo*, we employed two distinct xenograft models. MKN45 xenografts (expressing wtMET) were sensitive to MET-TKI (91% inhibition of tumor growth 36 days after tumor inoculation in mice treated daily with 50 mg/kg crizotinib) in agreement with previous reports (12, 27), whereas MET-Y1248H NIH3T3 xenografts were predictably MET-TKI resistant (59% inhibition of tumor growth at 21 days after daily treatment with 150 mg/kg crizotinib; Supplementary Fig. S5). In contrast, ganetespib was effective in both xenograft models, although at the maximum dose used in this study (50 mg/kg administered 3 times per week), somewhat greater activity was seen in wtMET xenografts (95% inhibition of tumor growth) compared with MET-Y1248H xenografts (82% growth inhibition; Supplementary Fig. S5).

To evaluate whether combination therapy was beneficial in the MKN45 model, we chose crizotinib (12.5 mg/kg) and ganetespib (12.5 mg/kg) dosing that provided approximately 50% growth inhibition when used singly (Supplementary Fig. S5). Treatment with this drug combination proved to be significantly more potent (95% growth inhibition) compared with single-agent treatment (56% growth inhibition for crizotinib alone and 64% growth inhibition for ganetespib alone; Fig. 6A).

In mice bearing TKI-resistant MET-Y1248H xenografts, we similarly observed that combination therapy with ganetespib and crizotinib (37.5 and 125 mg/kg, respectively) inhibited tumor growth by 90%, which was significantly greater than the inhibition achieved by treatment with either drug alone (65% growth inhibition for ganetespib and 31% growth inhibition for crizotinib; Fig. 6B).

Based on a comparison of expected and obtained FTVs in single agent- and combination-treated mice (50), crizotinib...
Figure 4. MET kinase domain mutants retain sensitivity to HSP90 inhibition. A, ganetespib inhibits MET phosphorylation and downstream signaling in TKI-sensitive MET mutants. NIH3T3 cells stably expressing MET mutants (V1238I and H1112Y) were treated for 16 hours with increasing concentrations of either crizotinib or ganetespib, lysed, and subjected to immunoblotting. B, ganetespib inhibits MET phosphorylation and downstream signaling in TKI-resistant MET mutants. NIH3T3 cells stably expressing MET mutants (Y1248H and L1213V) were treated and analyzed as described earlier.

Discussion

Although small-molecule kinase inhibitors have demonstrated clinical efficacy in cancer, patients who initially respond to such targeted therapy frequently will develop resistance. Mechanisms of resistance include the growth advantage provided by the appearance of drug-resistant kinase mutations (40), as well as a process termed ‘oncogene switching,’ whereby TKI-treated cells utilize an alternative kinase to drive shared downstream signaling pathways (41). Recent studies have revealed that HSP90 inhibition can overcome both forms of kinase inhibitor resistance. For example, HSP90 inhibition suppresses EGFR activity and downstream signaling in erlotinib-resistant EGFR/T790M-expressing cells (42), and suppresses ALK activity and signaling in cells expressing crizotinib-resistant EML4-ALK/L196M (43). Similarly, targeting HSP90 prevents escape of ErbB2-driven breast cancer cells from chronic ErbB inhibition and escape of MET-amplified tumor cells from MET-TKI (44, 45).

TKI-resistance–conferring MET kinase domain mutations, including Y1248H and L1213V, have been identified in hereditary papillary renal cell carcinomas (14), and long-term exposure to MET TKI in vitro leads to acquisition of these mutations in MET-expressing gastric cancer cells (15, 16). Such kinase-activating mutations are thought to interfere with ATP-competitive TKI binding, and they likely contribute to both primary and acquired drug resistance in MET-dependent cancers. We have shown that a panel of MET proteins (both TKI-sensitive and TKI–resistant) retains dependence on HSP90 and remains sensitive to HSP90 inhibition. Importantly, we show that the active state of MET displays the strongest dependence on HSP90, whether activation is induced by HGF or is a consequence of kinase domain mutation (ligand-independent).

Because of the preference of HSP90 for activated MET, these data suggest the possibility that HSP90 inhibition might synergize with MET-TKI. Recent preclinical evaluation of the effects obtained upon simultaneously exposing acute myelogenous leukemia cells expressing activated FLT3 tyrosine kinase to both a FLT3-directed TKI and a HSP90 inhibitor lends support to this possibility, as does a recent
study reporting the combinatorial benefit of ganetespib, crizotinib, and other TKIs in the context of EML4-ALK–driven non–small cell lung cancer (46, 47). Our data show that simultaneous treatment with MET-TKI and HSP90 inhibitor causes a true synergistic inhibition of cell growth in wild-type MET-expressing MKN45 gastric cancer cells. Synergy of this drug combination in these cells was reflected by enhanced inhibition of downstream signaling pathways, significantly greater inhibition of colony growth and cell motility, and by significantly greater growth inhibition of MKN45 xenografts in vivo.

Unexpectedly, we found that HSP90 inhibition also partially restored crizotinib sensitivity to two TKI-resistant MET mutants. Combination treatment dramatically inhibited MET signaling, colony growth, cell invasion, and xenograft growth in vivo. Although some of these effects may be due to the inhibition of additional HSP90 clients functioning within or parallel to MET-driven signaling pathways, the data we have presented here indicate that, at least for MET, HSP90 inhibition directly affects MET sensitivity to a TKI. Our model is in general agreement with a recent study proposing that TKI binding stabilizes HSP90-dependent kinases and obviates the need for HSP90 interaction (48), because we have shown that it is the activated state of MET that interacts most strongly with and is most dependent on HSP90. Another report has suggested that TKI binding promotes kinase degradation by denying access to HSP90 (49), although in the case of MET it appears that this would be the case only if the TKI bound to the active conformation of the kinase, promoted dissociation of HSP90, and limited a return to the inactive conformation. The unexpected combinatorial benefit we have shown for TKI-resistant MET highlights the complex influence of HSP90 on kinase conformation, especially in the context of activating mutations, and our model provides a rationale for the increased dependence of other constitutively active kinase mutations on HSP90. For example, others have shown that HSP90 inhibitor synergizes with a Bcr-Abl TKI in TKI-resistant chronic myelogenous leukemia cells (50). Although the benefit of simultaneous treatment with TKI and HSP90 inhibitor should be evaluated on a case-by-case basis, a rationale for such a strategy clearly exists in certain settings. Specifically, our data suggest that a MET/HSP90 inhibitor combination regimen is a viable strategy to be explored in patients

Figure 5. MET TKI and HSP90 inhibitor synergize in TKI-resistant MET mutant-expressing cells. A, NIH3T3 cells stably expressing TKI-resistant MET-Y1248H (left) or MET-L1213V (right) were exposed to increasing concentrations of crizotinib in the presence or absence of defined concentrations of ganetespib, lysed, and subjected to immunoblotting. B, soft agar colony growth of NIH3T3 cells stably expressing TKI-resistant MET-Y1248H was assessed and analyzed as described in Figure 3E. C, combination treatment dramatically inhibits invasion of TKI-resistant mutant MET-expressing cells. NIH3T3 cells stably expressing TKI-resistant mutant MET-Y1248H in serum-free medium containing indicated drugs were added to upper wells of Transwell chambers. The membrane separating top and bottom wells was coated with Matrigel. The bottom wells contained medium with 10% FBS as the chemoattractant. After 48 hours, invaded cells (e.g., cells appearing on the lower surface of the separating membrane) were stained and counted microscopically. Data are shown as mean ± SD of triplicate experiments. *, P < 0.05 vs. control (unpaired Student t test followed by Bonferroni test).
naive MET-dependent cancers, as well in those patients whose cancers have developed resistance to MET-TKI. When delaying or reversing MET-TKI resistance, an effective MET/HSP90 inhibitor combination strategy may also require lower (and, thus, potentially less toxic) concentrations of both drugs.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Authors’ Contributions

Conception and design: N. Miyajima, S. Tsutsumi, M. Mollapour, J.B. Trepel, K. Nonomura, L. Neckers

Development of methodology: N. Miyajima, M. Mollapour

Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): N. Miyajima, C. Sourbier, K. Beebe, C. Rivas, S. Yoshida, J.B. Trepel, M. Tatokoro, N. Shinohara

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): N. Miyajima, C. Rivas, S. Yoshida, J.B. Trepel, K. Nonomura, L. Neckers

Writing, review, and/or revision of the manuscript: N. Miyajima, J.B. Trepel, K. Nonomura, L. Neckers

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): N. Miyajima, Y. Huang

Study supervision: N. Miyajima, K. Nonomura, L. Neckers

Acknowledgments

The authors thank Y.H. Lee and D. Bottaro for the V5-tagged plasmids, human MET cDNA, and for helpful discussions. The authors also thank L. Schmidt for the MET mutants, mutant MET-expressing NIH3T3 cells, careful reading of the article, and for helpful discussions. Ganetespib and crizotinib were provided by W. Ying (Synta Pharmaceuticals, Inc).

Grant Support

This work was financially supported by funds from the Intramural Research Program, National Cancer Institute, Center for Cancer Research.

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received April 25, 2013; revised August 13, 2013; accepted September 3, 2013; published OnlineFirst October 11, 2013.
References


20. Whitesell L, Mimaugh EG, De Costa B, Myers CE, Nechayev L. Inhibition of heat shock protein HSP90-p60sv-src heteroreceptor com-


The HSP90 Inhibitor Ganetespiib Synergizes with the MET Kinase Inhibitor Crizotinib in both Crizotinib-Sensitive and -Resistant MET-Driven Tumor Models

Naoto Miyajima, Shinji Tsutsumi, Carole Sourbier, et al.


Updated version
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-13-1156

Supplementary Material
Access the most recent supplemental material at:
http://cancerres.aacrjournals.org/content/suppl/2013/10/11/0008-5472.CAN-13-1156.DC1

Cited articles
This article cites 49 articles, 30 of which you can access for free at:
http://cancerres.aacrjournals.org/content/73/23/7022.full#ref-list-1

Citing articles
This article has been cited by 3 HighWire-hosted articles. Access the articles at:
http://cancerres.aacrjournals.org/content/73/23/7022.full#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.