Autologous T-cell Therapy for Cytomegalovirus as a Consolidative Treatment for Recurrent Glioblastoma

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Abstract

Glioblastoma multiforme (GBM) is one of the most aggressive human brain malignancies. Even with optimal treatment, median survival is less than 6 months for patients with recurrent GBM. Immune-based therapies have the potential to improve patient outcome by supplementing standard treatment. Expression of human cytomegalovirus (CMV) antigens in GBM tissues provides the unique opportunity to target viral antigens for GBM therapy. Here, we report findings of a formal clinical assessment of safety and potential clinical efficacy of autologous CMV-specific T-cell therapy as a consolidative treatment for recurrent GBM. From a total of 19 patients with recurrent GBM, CMV-specific T cells were successfully expanded from 13 patients (68.4%), 11 of whom received up to four T-cell infusions. Combination therapy based on T-cell infusion and chemotherapy was well tolerated, and we detected only minor adverse events. The overall survival of these patients since first recurrence ranged from 133 to 2,428 days, with a median overall survival of 403 days. Most importantly, 4 of 10 patients that completed the treatment remained progression free during the study period. Furthermore, molecular profiling of CMV-specific T-cell therapy from these patients revealed distinct gene expression signatures, which correlated with their clinical response. Our study suggests that a combination therapy with autologous CMV-specific T cells and chemotherapy is a safe novel treatment option and may offer clinical benefit for patients with recurrent GBM. Cancer Res; 74(13); 3466–76. ©2014 AACR.

Introduction

Glioblastoma multiforme (GBM) is one of the most malignant human adult brain tumors. Current treatment regimes include surgical resection, radio-, and chemotherapy, but patient prognosis remains poor with a median survival after initial diagnosis of less than 15 months (1) and a 5-year survival rate of less than 10% (2). GBM is incurable and inevitably recurs after initial therapy. Median survival for recurrent GBM is 3 to 6 months, and most patients do not survive longer than 1 year (2–4). Although chemotherapy, especially the introduction of temozolomide, has been shown to increase survival to some degree (1), dramatic improvements in outcome for patients with GBM have remained elusive. Therefore, much interest has focused on immunotherapeutic approaches. Strategies under investigation include tumor lysate vaccines, tumor antigen vaccines, and targeting of immunomodulatory molecules (reviewed in ref. 5).

Accumulating evidence indicates human cytomegalovirus (CMV) as a contributing factor to glioma progression (6, 7), and CMV has been suggested as a therapeutic target (8). Although not classified as an oncogenic virus, CMV can increase cellular proliferation, angiogenesis, and immune evasion, thus enabling several hallmarks of cancer (9, 10). Recently, an onco-accessory function of CMV has also been described in a mouse model of glioblastoma using murine cytomegalovirus infection (11). More importantly, CMV antigens and nucleic acids have been detected in histologic sections of GBM but not in surrounding healthy tissue (12–18). Although these histologic findings have been disputed (19–21), more recent studies have confirmed the presence of CMV sequences in malignant cells (22, 23). Vaccination of one patient with dendritic cells pulsed with autologous GBM lysate elicited a CMV-specific immune response, further supporting the presence of CMV antigens in GBM tissue (24). This provides an opportunity to target viral antigens with immune-based therapies. Low levels of CMV antigen expression in tumor cells were found to be associated with longer survival of patients with GBM (15, 16), thus indicating that antiviral therapy could improve GBM prognosis. In addition, recent studies supplementing standard GBM therapy with antiviral valganciclovir treatment for more than 6 months demonstrated a survival benefit for patients with GBM (25, 26).
We have explored the feasibility and safety of an autologous T-cell–based GBM immunotherapy targeting CMV antigens. We have recently shown that CMV-specific CD8⁺ T cells in patients with GBM have reduced functional capacity, but that this limitation can be reversed following in vitro stimulation. Adoptive transfer of these cells into a single patient with recurrent GBM in combination with standard chemotherapy was associated with long-term disease-free survival (27). Here, we report the findings from a formal clinical assessment of this initial finding as a phase 1 clinical trial. We demonstrate that autologous CMV-specific T-cell therapy is safe with minimal side effects and may offer clinical benefit for patients with recurrent GBM.

Patients and Methods

Study design, ethics, and patients

This phase 1 clinical study was designed to assess the safety and tolerability of autologous CMV-specific T-cell therapy for recurrent GBM. This clinical trial was conducted according to Declaration of Helsinki principles and was approved by The QIMR Berghofer Medical Research and Uniting Care Health Human Research Ethics Committees. All participants signed a consent form, which was approved by both ethics committees. This study is registered under the Australia New Zealand Clinical Trial Registry (ACTRN12609000338268). Study completion required a minimum of three T-cell infusions, whereas additional infusions could be administered depending on availability of cells. Infusions consisting of 25 to 40 × 10⁶ autologous CMV-specific T cells in sterile saline were administered in 4 (±2) weeks intervals. The infusions were coordinated with periods of chemotherapy to avoid unwanted side effects. Peripheral blood samples collected before each infusion and at regular intervals after infusion were used for haematologic and immunologic monitoring. Each follow-up visit included vital observations and a quality-of-life questionnaire. Magnetic resonance imaging (MRI) was used to assess baseline tumor load before infusion and at regular intervals after first infusion.

In vitro expansion of CMV-specific T cells from patients with GBM

CMV-specific T cells were generated by in vitro stimulation with synthetic peptide epitopes. Peripheral blood mononuclear cells (PBMC) were isolated from peripheral blood by Ficoll gradient, washed, and resuspended in RPMI-1640 supplemented with 10% FBS (growth medium). The cells were cocultured with autologous PBMCs presensitized with CMV peptides (5 μg/mL; Supplementary Table S1) at a responder to stimulator ratio of 2:1 in growth medium. After 3 days and then every 3 to 4 days thereafter, the cultures were supplemented with growth medium containing 120 IU/mL recombinant IL-2 (Komtur Pharmaceuticals). Cells were cryopreserved on day 14, after testing for sterility and CMV-specific T cells using an IFN-γ intracellular cytokine assay. Release criteria were ≥0.1% CMV-specific T cells in total lymphocyte population and >50% cell viability.

Characterization of CMV CTL by intracellular cytokine assay and flow cytometry

PBMCs or cultured T cells were stimulated with peptides corresponding to defined CD8⁺ T-cell epitopes derived from CMV proteins (1 μg/mL) and incubated in the presence of Brefeldin A for 4 hours. For polyfunctional analysis, cells were incubated with a CD107a-antibody, Brefeldin A, and Monensin for 5 hours. After surface staining for CD8, CD4, and CD3, cells were fixed and permeabilized with cytofix/cytoperm and stained for IFN-γ (and IL2 and TNF in polyfunctional assays). Immune monitoring was achieved by surface staining with appropriate peptide-MHC (pMHC) multimers (Immudex) and surface markers, followed by fixation with a transcription buffer set and intracellular staining (all reagents from BD Biosciences, unless otherwise indicated). Stained cells were resuspended in PBS containing 2% paraformaldehyde and acquired using a FACS Canto II or LSR Fortessa with FACSDivaw software (BD Biosciences). Postacquisition analysis was conducted using FlowJo software (TreeStar).

Cell sorting and gene expression analysis by Taqman RT-PCR array

PBMCs were revived from cryopreserved stocks and stained with appropriate CMV dextramers, followed by staining for surface markers to allow for separation of monocytes, B cells, and T cells. After filtering the cell suspension through a nylon mesh for removal of cell clumps, total CD8⁺ T-cell and CMV-specific CD8⁺ T-cell populations were isolated using a FACS Aria (BD Biosciences) cell sorter. Total RNA was purified using the Qiagen RNeasy Micro Kit. An RNA amount equivalent to 3,000 cells was transcribed into cDNA using the high-capacity RNA-to-cDNA-Kit. The cDNA was preamplified with a custom primer pool, loaded into custom-designed Taqman array cards and run on the Viia7 real-time PCR system (all reagents from Life Technologies). The array cards were designed to contain 91 genes that were described to be regulated in CD8 T cells during CMV infection (28). Data were normalized to housekeeping genes (18S, actin, and β2-microglobulin) and analyzed using GeneSpring v12.5 software.

Statistical analysis

Mortality and progression-free survival (PFS) were characterized using Kaplan–Meier curves, and the corresponding median survival times were obtained for the intention-to-treat (ITT) and per-protocol (PP) patient populations of 11 and 10 patients, respectively. The ITT patient population included patients who received at least one T-cell infusion and the PP patient population included patients who received at least three infusions. Time to death or censoring was defined as the time between the last follow-up date and the date of the first recurrence. Time to progression or censoring was defined as the time between the date of the subsequent recurrence after the treatment or the last follow-up date and the date of the first recurrence. Left truncations occurred since patients did not receive the treatment at the date of the first recurrence and have been accounted for.
Table 1. Characteristics of patients with GBM and treatment history before T-cell therapy

<table>
<thead>
<tr>
<th>GBM ID</th>
<th>Age at diagnosis</th>
<th>Gender</th>
<th>Final histology</th>
<th>Time to recurrence (d)</th>
<th>Recurrences before T-cell therapy</th>
<th>Time to T-cell therapy (d)</th>
<th>Number of operations</th>
<th>Gliadel use</th>
<th>XRT/TMZ before T-cell therapy</th>
<th>Additional treatment before T-cell therapy</th>
</tr>
</thead>
<tbody>
<tr>
<td>01</td>
<td>60</td>
<td>M</td>
<td>GBM</td>
<td>300</td>
<td>2</td>
<td>706</td>
<td>2</td>
<td>2</td>
<td>Yes</td>
<td>None</td>
</tr>
<tr>
<td>02</td>
<td>49</td>
<td>M</td>
<td>Gliosarcoma</td>
<td>404</td>
<td>1</td>
<td>473</td>
<td>2</td>
<td>0</td>
<td>Yes</td>
<td>Avastin</td>
</tr>
<tr>
<td>03</td>
<td>50</td>
<td>M</td>
<td>GBM</td>
<td>208</td>
<td>3</td>
<td>1,572</td>
<td>4</td>
<td>2</td>
<td>Yes</td>
<td>Etoposide, thalidomide</td>
</tr>
<tr>
<td>04</td>
<td>72</td>
<td>F</td>
<td>GBM</td>
<td>439</td>
<td>1</td>
<td>519</td>
<td>2</td>
<td>2</td>
<td>Yes</td>
<td>None</td>
</tr>
<tr>
<td>05</td>
<td>52</td>
<td>M</td>
<td>GBM</td>
<td>378</td>
<td>1</td>
<td>—</td>
<td>2</td>
<td>0</td>
<td>Yes</td>
<td>None</td>
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<tr>
<td>06</td>
<td>32</td>
<td>F</td>
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<td>1,058</td>
<td>1</td>
<td>2,117</td>
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<td>Yes</td>
<td>Avastin</td>
</tr>
<tr>
<td>07</td>
<td>54</td>
<td>M</td>
<td>GBM</td>
<td>143</td>
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<td>241</td>
<td>2</td>
<td>0</td>
<td>Yes</td>
<td>Carboplatin</td>
</tr>
<tr>
<td>08</td>
<td>66</td>
<td>M</td>
<td>GBM</td>
<td>499</td>
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<td>—</td>
<td>2</td>
<td>0</td>
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</tr>
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<td>09</td>
<td>74</td>
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<td>GBM</td>
<td>235</td>
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<td>None</td>
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<td>10</td>
<td>37</td>
<td>M</td>
<td>AA</td>
<td>1,630</td>
<td>3</td>
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<td>3</td>
<td>0</td>
<td>No</td>
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</tr>
<tr>
<td>11</td>
<td>21</td>
<td>M</td>
<td>GBM</td>
<td>639</td>
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<td>—</td>
<td>2</td>
<td>0</td>
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</tr>
<tr>
<td>12</td>
<td>47</td>
<td>F</td>
<td>GBM</td>
<td>4,552</td>
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<td>—</td>
<td>2</td>
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<tr>
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<td>57</td>
<td>M</td>
<td>GBM</td>
<td>244</td>
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<td>318</td>
<td>2</td>
<td>0</td>
<td>Yes</td>
<td>None</td>
</tr>
<tr>
<td>14</td>
<td>47</td>
<td>F</td>
<td>GBM</td>
<td>1,105</td>
<td>1</td>
<td>—</td>
<td>2</td>
<td>0</td>
<td>Yes</td>
<td>Avastin</td>
</tr>
<tr>
<td>15</td>
<td>60</td>
<td>M</td>
<td>GBM</td>
<td>197</td>
<td>1</td>
<td>283</td>
<td>1</td>
<td>0</td>
<td>Yes</td>
<td>None</td>
</tr>
<tr>
<td>16</td>
<td>50</td>
<td>M</td>
<td>GBM</td>
<td>989</td>
<td>1</td>
<td>1,092</td>
<td>1</td>
<td>0</td>
<td>Yes</td>
<td>None</td>
</tr>
<tr>
<td>17</td>
<td>23</td>
<td>M</td>
<td>GBM</td>
<td>436</td>
<td>1</td>
<td>—</td>
<td>1</td>
<td>0</td>
<td>Yes</td>
<td>Avastin</td>
</tr>
<tr>
<td>18</td>
<td>61</td>
<td>F</td>
<td>GBM</td>
<td>323</td>
<td>N/A</td>
<td>—</td>
<td>2</td>
<td>0</td>
<td>Yes</td>
<td>None</td>
</tr>
<tr>
<td>19</td>
<td>41</td>
<td>F</td>
<td>GBM</td>
<td>413</td>
<td>2</td>
<td>704</td>
<td>3</td>
<td>0</td>
<td>Yes</td>
<td>Carboplatin, lomustine, avastin</td>
</tr>
</tbody>
</table>

NOTE: Time to recurrence is calculated from date of first surgery to date of tumor recurrence based on MRI findings. Number of operations refers to tumor debulking surgeries only. Time to T-cell therapy is calculated from date of first diagnosis. Shaded areas indicate patients that did not complete the trial.

Abbreviations: AA, anaplastic astrocytoma; d, days; N/A, no information available; TMZ, temozolomide chemotherapy; XRT, radiotherapy.

*Patients that were initially diagnosed with low-grade malignancies.
Table 2. Clinical follow-up of adoptive T-cell therapy of patients with recurrent GBM

<table>
<thead>
<tr>
<th>GBM ID</th>
<th>Total cells expanded</th>
<th>Number of cells per infusion</th>
<th>Time to progressive disease after first infusion (d)</th>
<th>Treatment in addition to T cells</th>
<th>Current status (December 31, 2013)</th>
</tr>
</thead>
<tbody>
<tr>
<td>01</td>
<td>2.6 x 10^7</td>
<td>3 x 10^7</td>
<td>4</td>
<td>TMZ</td>
<td>Alive</td>
</tr>
<tr>
<td>02</td>
<td>1.8 x 10^7</td>
<td>2.8 x 10^7</td>
<td>4</td>
<td>Avastin, TMZ</td>
<td>Died 1,447</td>
</tr>
<tr>
<td>03</td>
<td>1.4 x 10^7</td>
<td>2 x 10^7</td>
<td>3</td>
<td>Avastin, carboplatin</td>
<td>Died 1,304</td>
</tr>
<tr>
<td>04</td>
<td>Failed</td>
<td>N/A</td>
<td>0</td>
<td>TMZ</td>
<td>Died 1,010</td>
</tr>
<tr>
<td>05</td>
<td>Failed</td>
<td>N/A</td>
<td>0</td>
<td>TMZ</td>
<td>Died 1,750</td>
</tr>
<tr>
<td>06</td>
<td>2.9 x 10^7</td>
<td>4 x 10^7</td>
<td>4</td>
<td>Avastin, TMZ</td>
<td>Died 1,134</td>
</tr>
<tr>
<td>07</td>
<td>1.6 x 10^7</td>
<td>3.5 x 10^7</td>
<td>2</td>
<td>Avastin, TMZ</td>
<td>Died 353</td>
</tr>
<tr>
<td>08</td>
<td>1.3 x 10^7</td>
<td>2.5 x 10^7</td>
<td>4</td>
<td>Avastin, TMZ</td>
<td>Died 462</td>
</tr>
<tr>
<td>09</td>
<td>Failed</td>
<td>N/A</td>
<td>0</td>
<td>TMZ</td>
<td>Died 100</td>
</tr>
<tr>
<td>10</td>
<td>Failed</td>
<td>N/A</td>
<td>0</td>
<td>Avastin, CCNU</td>
<td>Died 100</td>
</tr>
<tr>
<td>11</td>
<td>1.5 x 10^7</td>
<td>3.2 x 10^7</td>
<td>4</td>
<td>Avastin, TMZ</td>
<td>Died 34</td>
</tr>
<tr>
<td>12</td>
<td>10.6 x 10^7</td>
<td>2.5 x 10^7</td>
<td>4</td>
<td>Avastin, TMZ</td>
<td>Died 287</td>
</tr>
<tr>
<td>13</td>
<td>9.2 x 10^7</td>
<td>4 x 10^7</td>
<td>3</td>
<td>Avastin, TMZ</td>
<td>Died 143</td>
</tr>
<tr>
<td>14</td>
<td>1.9 x 10^7</td>
<td>3 x 10^7</td>
<td>3</td>
<td>Avastin, TMZ</td>
<td>Died 317</td>
</tr>
<tr>
<td>15</td>
<td>1.3 x 10^7</td>
<td>3 x 10^7</td>
<td>3</td>
<td>Avastin, TMZ</td>
<td>Died 392</td>
</tr>
</tbody>
</table>

NOTE: Shaded areas indicate patients that underwent venesection but did not complete the trial. Values in parentheses indicate days. Abbreviations: TMZ, temozolomide chemotherapy; N/A, not applicable; C2, CMV-specific T-cell therapy.

Patient death was not related to GBM.
appropriately. The analyses were performed using SAS Enterprise Guide 4.3 and R 3.0.1.

Results

Patient characteristics

Nineteen patients with recurrent GBM were recruited for this study. Eligibility criteria for the study included (i) age 18 years or above; (ii) geographically accessible for follow-up; (iii) ability to provide informed consent; (iv) Eastern Cooperative Oncology Group performance status 0, 1, 2, or 3; (v) life expectancy of at least 3 months; (vi) positive CMV serology; and (vii) previous histologic diagnosis of GBM (WHO grade IV) and radiologic and/or clinical evidence of tumor progression or recurrence. Four patients had to be withdrawn before venesection due to progressive disease, while insufficient CMV-specific T cells were expanded from two patients due to low precursor frequency or poor cell viability. All patients received standard treatment with maximal safe surgical debulking at primary diagnosis, external beam radiotherapy and chemotherapy (Table 1). CMV-specific T cells were successfully expanded from 13 patients, but two patients had to be withdrawn due to progressive illness and one patient discontinued the intervention after two infusions due to progressive disease. In total, 10 patients completed a minimum of three infusions as required per protocol (Table 2).

In vitro expansion and functional characterization of CMV-specific CD8+ T cells

Autologous CMV-specific T cells were successfully expanded from 13 of 19 patients (Table 2). Ex vivo analysis of CMV-specific T cells from patients with GBM using pMHC

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Cancer Res; 74(13) July 1, 2014

Cancer Research

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multimers showed antigen-specific T-cell frequencies ranging from 0.026% to 18.4% of total CD8\(^+\) T cells (median, 4.2%). Following in vitro stimulation with HLA-matched CMV peptide epitopes, a significant increase in antigen-specific T cells was observed (range, 4.2%–92.7% of total CD8\(^+\) T cells; median, 49.85%; \(P < 0.0001\), Fig. 1A and B, and Supplementary Table S2). Phenotypic characterization showed that these T cells were predominantly CD3\(^+\) with a combination of both CD8\(^+\) and CD4\(^+\) T cells (Fig. 1C). The majority of CMV-specific T cells were CD45RA\(^-\) CD27\(^+\) and/or CD57\(^+\) a phenotype characteristic of effector cells (Fig. 1D). Intracellular cytokine analysis revealed that high proportions of these in vitro expanded CD8\(^+\) T cells expressed IFN\(\gamma\) in response to stimulation with CMV epitopes (Fig. 1E and F). Furthermore, in-depth functional analysis of these T cells from some patients showed that these cells displayed polyfunctional profile and also expressed CD107a (data not shown).

Safety and clinical evaluation of CMV-specific T-cell–based immunotherapy for recurrent GBM

Of the 13 patients for whom CMV-specific T cells were generated, 10 patients received three to four T-cell infusions (2–4 \(\times\) 10\(^6\) cells/infusion, Table 2). One patient only received two infusions, whereas two patients died before the availability of T cells. Infusions were generally well tolerated and mostly minor adverse events were recorded (Table 3). The toxicity grading was assigned according to the NCI Common Terminology Criteria for Adverse Events. A single serious adverse event (SAE) possibly related to T-cell therapy was recorded (Patient GBM:19). This patient had a generalized seizure within 12 hours of the first T-cell infusion and was hospitalized for 3 days. The patient had a history of seizures before entering into the current trial. The investigators discussed this SAE with the Data Safety Monitoring Committee, and deemed the SAE was unlikely to be associated with T-cell therapy. No further seizures developed in this patient after subsequent T-cell infusions. These analyses indicate that autologous CMV-specific T-cell infusions are a safe treatment for GBM. The median overall survival (OS) of the 11 patients that received at least one infusion was 403 days (range, 133 to 2,428 days; Fig. 2A). The time to progression for all patients after infusion ranged from 108 to more than 1,783 days, with a median of 246 days (Fig. 2B). Of the 11 patients treated with T-cell therapy, four patients remained progression-free (Table 2). Patient GBM:01 showed the longest stabilization of disease with almost 4 years of PFS after the T-cell infusion. This patient had no other treatment subsequent to CMV-specific T-cell therapy and remains disease-free to date.

Table 3. Safety assessment following adoptive T-cell therapy of patients with recurrent GBM\(^a\)

<table>
<thead>
<tr>
<th>Adverse event</th>
<th>Number of patients affected (attribution score(^b))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grade 1: mild</td>
<td></td>
</tr>
<tr>
<td>Headache</td>
<td>3 (2)</td>
</tr>
<tr>
<td>Fatigue</td>
<td>3 (2)</td>
</tr>
<tr>
<td>Visual hallucination</td>
<td>1 (2)</td>
</tr>
<tr>
<td>Pyrexia of unknown origin</td>
<td>1 (2)</td>
</tr>
<tr>
<td>High blood pressure</td>
<td>1 (2)</td>
</tr>
<tr>
<td>Grade 2: moderate</td>
<td></td>
</tr>
<tr>
<td>Abnormal liver function tests</td>
<td>1 (2)</td>
</tr>
<tr>
<td>Lymphopenia</td>
<td>1 (2)</td>
</tr>
<tr>
<td>Seizure</td>
<td>1 (2)</td>
</tr>
<tr>
<td>Anxiety</td>
<td>1 (2)</td>
</tr>
<tr>
<td>Grade 3: severe</td>
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</tr>
<tr>
<td>Seizure</td>
<td>1 (2)</td>
</tr>
</tbody>
</table>

\(^a\)Severity grade and attribution scores assessed according to the NCI Common Terminology Criteria for Adverse Events. 
\(^b\)Attribution score: 1, unrelated; 2, unlikely; 3, possible; 4, probable; 5, definite. Adverse event logs were compiled from clinical observation (including vital signs), patient interview, and blood samples taken during infusions and at follow-up visits. Events scored as unrelated were excluded from this table.

Figure 2. A, OS of patients with GBM treated with autologous CMV-specific T cells since first recurrence. B, PFS following CMV-specific T-cell adoptive immunotherapy (determined from the date of the first T-cell infusion).
Immunologic and molecular analysis of CMV-specific T-cell therapy

To assess the efficiency of T-cell therapy, we first determined the effect of antigen-specific T-cell frequencies on survival. However, there was no correlation between the number of CMV-specific T cells transferred and the OS and PFS (Fig 3A and B). Ex vivo longitudinal analysis of CMV-specific T cells showed that although in some patients there was a small increase in the precursor frequency of CMV-specific T cells, the number of antigen-specific T cells returned to baseline after the completion of adoptive immunotherapy (Supplementary Fig. S1A and S1B). The functional profile of these T cells also remained unchanged during and after the completion of T-cell therapy (Supplementary Fig. S1C).

We next explored the possibility that gene expression profiling of T-cell therapy may help to distinguish patients that might benefit from adoptive immunotherapy. To test this hypothesis, we used a custom array that allowed quantitative expression analysis of different categories of genes relevant for T-cell function (Supplementary Table S3). This expression profiling revealed that 47 of 91 genes were significantly changed in CMV-specific CD8+ T cells following in vitro expansion (Fig. 3C and Supplementary Fig. S2A and S2B). This gene expression profile was consistent with a signature of activated T cells and was further confirmed by specific antibody staining and flow cytometry (Supplementary Fig. S2C).

To identify gene expression patterns that might be of prognostic value, we divided our patient cohort into two groups: (i) patients with GBM who had short PFS (<100 days) and (ii) patients with GBM who either remained progression-free within the study period or developed progressive disease after more than 100 days. In-depth analysis of gene expression data revealed that these two groups of patients showed significant differences in the expression of seven genes, including T-cell transcription factors (EOMES, BCL6, and FOXP3), cytokine/chemokines (IFNG and CCL5), and checkpoint markers (CTLA4 and XAF1; Fig. 3D and E). These analyses suggest that expression profiling of T-cell therapy may provide clues on the potential therapeutic benefit of adoptive immunotherapy.

CMV-specific T cells are present in GBM tumor tissue and show a distinct phenotype compared with peripheral blood

Ex vivo analysis of tumor-infiltrating antigen-specific T cells can provide some critical insights on the immune control of malignant cells. In our study, GBM16 patient who received four infusions of CMV-specific T cells developed progressive disease 4 months after the completion of T-cell infusions and then underwent surgical tumor resection. We isolated T cells from the resected tumor tissue and were able to detect CMV-specific CD8+ T cells (Fig. 4A, top). However, the majority of these antigen-specific T cells failed to express multiple cytokines, including IFNγ, TNF, IL2, and showed poor cytotoxic activity as assessed by CD107a mobilization following stimulation with CMV peptides (Fig. 4A, bottom). Furthermore, staining of CD103 as a marker for tissue resident T cells (Trm) revealed that although approximately one third of CD8+ T cells in the tumor tissue were Trm, none of the CMV-specific T cells expressed CD103 (Fig. 4B). The frequency of CMV-specific T cells in the tumor tissue was approximately 4-fold lower when compared with T cells circulating in peripheral blood at different time points before (d0), during (d35), and after T-cell therapy (d78 and d121, Fig. 4C). Tumor-infiltrating CMV-specific T cells expressed higher levels of PD-1, TIM-3, and CTLA-4 and lower levels of transcription factors T-bet, Eomes, and LEF-1 (Fig. 4C). These observations suggest that tumor-infiltrating antigen-specific T cells in this patient with GBM displayed poor functional capacity and increased expression of inhibitory receptors when compared with T cells from peripheral blood. Similar expression patterns of PD-1, CTLA-4, TIM-3, and transcription factors were detected in the global CD8+ T-cell population (Supplementary Fig. S3). We further detected almost 5-fold higher levels of regulatory T cells (CD4+CD25+FoxP3+) than in peripheral blood, which is consistent with an immunosuppressive environment in tumor tissue (Fig. 4C).

Discussion

The survival of patients with recurrent GBM remains poor despite use of all currently available cytotoxic therapeutics (1, 29–32). Over the last decade, immune-based therapies have emerged as possible tools for the treatment of recurrent GBM (33–35), and exploratory studies have shown improved PFS and OS (36–38). In 2002, Cobbs and colleagues demonstrated expression of the CMV proteins IE-1 and late antigen in GBM tumor biopsies, which were later confirmed by other groups (10, 12–14, 21). Further studies have suggested that CMV-encoded proteins such as viral IL10 and US28, a G-protein–coupled receptor-like protein, may act as tumor promoters in GBM (10, 39, 40). The presence of CMV in GBM has generated considerable interest, especially the potential targeting of the viral proteins using immune-based therapies (18). We have shown previously that CMV-specific T cells from the majority of patients with GBM display reduced multifunctional potentiality and that in vitro stimulation of these T cells can improve their functional profile.
Adoptive transfer of these T cells into one patient with recurrent GBM was shown to be safe with possible clinical benefit.

In the present study, we report the outcome of the first clinical trial for adoptive immunotherapy using CMV-specific T cells in patients with recurrent GBM. We recruited 19 patients with recurrent GBM and of these 11 patients received multiple infusions of autologous in vitro expanded CMV-specific T cells. A number of important conclusions can be drawn from this study. First, adoptive transfer of

Figure 4. Ex vivo analysis of tumor-infiltrating and peripheral blood circulating CMV-specific CD8+ T cells from a patient with GBM. A, ex vivo HLA-peptide multimer staining and polyfunctional analysis of tumor-infiltrating CMV-specific T cells from patient GBM:16. B, expression of CD103 (a marker of tissue-resident T cells) on total CD8+ and CMV-specific T cells in tumor-infiltrating lymphocytes. C, longitudinal comparative phenotypic analysis of peripheral blood circulating and tumor-infiltrating CMV-specific CD8+ T cells and CD4+ Tregs. Tumor-infiltrating lymphoid cells and PBMC were incubated with HLA-peptide multimers, antibodies specific for CD3, CD4, CD8 and specific markers (as indicated on the y-axis of each box), and then analyzed using a LSR Fortessa with FACSDiva software. Postacquisition analysis was conducted using the FlowJo software.
CMV-specific T cells was completely safe with minimal toxicities. Although the CMV-specific T-cell therapy was provided in combination with standard therapies (Table 2), we did not observe any deleterious impact of these therapies on the adoptively transferred CMV-specific T cells. More importantly, patients did not experience any severe side effects, and the only recorded SAE was deemed unrelated to the treatment.

Second, clinical follow-up analyses showed that CMV-specific immunotherapy was coincident with disease stabilization and prolonged PFS in some patients. The median OS in our study was 57 weeks (range, 19–346 weeks) with a median PFS of >35 weeks (range, 15.4–254 weeks). Most importantly, 4 of the 10 patients who completed T-cell therapy remained progression-free. Although promising, these observations will require confirmation in a formal phase II randomized clinical trial. Interestingly, the positive effects of antiviral therapy in the GBM setting have also recently emerged from a randomized, placebo-controlled study investigating the use of the antiviral drug valganciclovir for the treatment of primary gliomas (25).

Third, we were unable to see any correlation of antigen-specific T-cell frequencies following adoptive immunotherapy and clinical outcome. Although some patients showed a small increase in virus-specific T cells in the peripheral blood following the first few infusions, this effect was transient. Furthermore, phenotypic and functional analysis showed no link between the clinical response and antigen-specific T cells in the peripheral blood. Although the reason for this lack of correlation is unknown, it is probable that the tumor microenvironment and disease burden may affect T-cell function and influence the clinical response to adoptive immunotherapy (41, 42). Indeed, preliminary data from a single patient (GBM:16) who developed progressive disease soon after the completion of adoptive immunotherapy showed that although antigen-specific T cells were detected in the tumor, the majority of these cells lacked multifunctional potentiality and had undetectable expression of CD103, which is a crucial marker for tissue residence. In addition, these T cells showed increased expression of checkpoint inhibitory receptors when compared with the circulating effector cells from peripheral blood, potentially reflecting the impact of the tumor microenvironment. We further detected higher amounts of regulatory T cells, which might contribute to immunosuppression in GBM tissues (43). These observations were supported by gene expression analysis of antigen-specific T cells used for adoptive immunotherapy. Indeed, increased prolonged PFS was coincident with lower expression of checkpoint inhibitory receptors and increased expression of T-cell transcription factors crucial for T-cell function. It is important to emphasize that these observations will require more in-depth analyses in a larger cohort of patients within a randomized phase II clinical study.

Taken together, adoptive immunotherapy of patients with recurrent GBM with CMV-specific T cells is safe and may provide long-term clinical benefits. These studies provide an important platform for a formal assessment of adoptive T-cell immunotherapy in both therapeutic and prophylactic settings. CMV-specific T-cell–based immunotherapy should be considered as a consolidative treatment following primary diagnosis of GBM for the prevention of recurrent disease.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

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References


8. Johnsen JL, Baryawmo N, Soderberg-Naucler C. Is human cytomegalo-
9. Dzurzynski K, Chang SM, Heimberger AB, Kalejta RF, McGregor
Dallas SR, Smit M, et al. Consensus on the role of human cytomeg-
10. Soroceanu L, Cobbss CS. Is HCMV a tumor promoter? Virus Res
2011;157:193-203.
et al. Phase II study of metronomic chemotherapy for recurrent malig-
Phase II trial of continuous dose-intensive temozolomide for recurrent malig-
Induction of CD8+ T-cell responses against novel glioma-associated antigen peptides by vaccination with [alpha]-type t polarized dendritic cells and polysinosinic-polycytidylic acid stabili-
15. Ranganathan P, Clark PA, Kuo JS, Salamat MS, Kalejta RF. Signi
16. Bhatacharjee B, Renzette N, Kowalk TF. Genetic analysis of cyto-
19. Crough T, Beagley L, Smith C, Jones L, Walker DG, Khanna R. Ex vivo functional analysis, expansion and adoptive transfer of cyto-
megalovirus-specific T-cells in patients with glioblastoma multiforme. Immuno
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