Epigenetic Targeting of Ovarian Cancer Stem Cells

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Abstract

Emerging results indicate that cancer stem–like cells contribute to chemoresistance and poor clinical outcomes in many cancers, including ovarian cancer. As epigenetic regulators play a major role in the control of normal stem cell differentiation, epigenetics may offer a useful arena to develop strategies to target cancer stem–like cells. Epigenetic aberrations, especially DNA methylation, silence tumor-suppressor and differentiation-associated genes that regulate the survival of ovarian cancer stem–like cells (OCSC). In this study, we tested the hypothesis that DNA-hypomethylating agents may be able to reset OCSC toward a differentiated phenotype by evaluating the effects of the new DNA methyltransferase inhibitor SGI-110 on OCSC phenotype, as defined by expression of the cancer stem–like marker aldehyde dehydrogenase (ALDH). We demonstrated that ALDH\textsuperscript{1+} ovarian cancer cells possess multiple stem cell characteristics, were highly chemoresistant, and were enriched in xenografts residual after platinum therapy. Low-dose SGI-110 reduced the stem-like properties of ALDH\textsuperscript{1+} cells, including their tumor-initiating capacity, resensitized these OCSCs to platinum, and induced reexpression of differentiation-associated genes. Maintenance treatment with SGI-110 after carboplatin inhibited OCSC growth, causing global tumor hypomethylation and decreased tumor progression. Our work offers preclinical evidence that epigenome-targeting strategies have the potential to delay tumor progression by reprogramming residual cancer stem–like cells. Furthermore, the results suggest that SGI-110 might be administered in combination with platinum to prevent the development of recurrent and chemoresistant ovarian cancer. Cancer Res; 74(17); 4922–36. ©2014 AACR.

Introduction

Epithelial ovarian cancer causes more deaths than any other female reproductive tract cancer (1,2). The majority of women diagnosed with advanced-stage epithelial ovarian cancer experience tumor recurrence associated with the development of chemoresistance, and platinum-resistant ovarian cancer is uniformly fatal (3). A new paradigm explaining tumor relapse involves the persistence of “cancer stem cells” that were characterized in several solid tumors, including ovarian cancer (4–6). Although chemotherapy may succeed initially at decreasing the size and number of tumors, it leaves behind residual malignant cells, which we hypothesize are enriched in tumor progenitors or “cancer stem cells.”

Ovarian cancer stem cells (OCSC) have been isolated from established ovarian cancer cell lines, ascites, and primary and metastatic tumors (4,7,8). They share several characteristics with normal stem cells, including the ability to form anchor-age-independent spherical aggregates, express stem cell markers, undergo membrane efflux, form clones in culture and in addition exhibit enhanced tumor-forming ability (9). Although a number of technical approaches have been successfully used to isolate OCSCs (sphere-forming, cell-surface markers, stem cell gene reporter assays), the use of an assay measuring aldehyde dehydrogenase isomorph 1 (ALDH) enzymatic activity has been recently proposed and is used to define CSCs in multiple other tumor types (10,11).

Ovarian CSCs are hypothesized to be largely (or entirely) responsible for emergence of chemoresistant tumors, because they possess many of the phenotypes associated with drug resistance (e.g., enhanced DNA repair, diminished apoptotic responses, increased efflux mechanisms, and quiescent state; refs. 4,12). Moreover, similar to normal embryonic or tissue stem cells, CSCs are believed to harbor a significantly altered epigenome (6,13), and it has been hypothesized that DNA-hypomethylating agents could “reset” these cells toward

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differentiation (14). Indeed, several hypomethylating agents were originally characterized as inducers of cancer cell differentiation (6, 15). However, it has become clear that hypomethylating agents or other epigenetic modulators alone cannot eradicate relapsed tumors. Preclinical studies from our and other groups have established the rationale for combining DNA methylation inhibitors with existing chemotherapeutic agents to overcome acquired drug resistance in ovarian cancer (16–20). On the basis of those studies, we recently completed a phase II trial using a DNA methylation inhibitor as a resensitizer to traditional chemotherapy in patients with recurrent ovarian cancer and showed that this combination has clinical and biologic activity (21), justifying other rationally designed epigenetic treatment strategies in ovarian cancer.

On the basis of the above considerations, we hypothesized that hypomethylating agents, in combination with chemotherapeutics, may target drug-resistant OCSCs, possibly leading to tumor eradication. In the current study, we isolated and characterized ALDH$$^{+}$$ OCSCs from ovarian cancer cell lines and human tumors. ALDH$$^{+}$$ cells were significantly more chemoresistant and tumorigenic compared with ALDH$$^{-}$$ cells in orthotopic tumor initiating assays. Treatment with SGI-110, a second-generation DNA methyltransferase inhibitor (DNMTI), resensitized OCSCs to platinum. A model recapitulating the emergence of recurrent tumors showed an increased percentage of ALDH$$^{+}$$ OCSCs in residual tumors after platinum. Maintenance therapy with SGI-110 during platinum-induced remission inhibited the emergence of platinum-resistant tumors. We suggest that epigenomic targeting using SGI-110 may be useful as a “maintenance” clinical strategy after platinum-based therapy in ovarian cancer.

Materials and Methods

Cell lines, patient samples, culture conditions, and reagents

Ovarian cancer cell lines (A2780, A2780_CR5, and SKOV3) were maintained in RPMI-1640 medium (Invitrogen), with supplements as described previously (see Supplementary Materials and Methods; ref. 22). Cisplatin-resistant variant A2780_CR5 was established from five-round IC$$_{50}$$ survival monocular selection by continuous exposure to increasing concentration of cisplatin (22). A2780 and SKOV3 ovarian cancer cells were authenticated in 2012 by the ATCC. Advanced high-grade serous ovarian tumors were surgically collected (IRB-approved protocol IUCRO-0280), enzymatically dissociated, and cultured, as previously described (4). SGI-110 was provided by Astex Pharmaceuticals Inc. and cisplatin was purchased from Calbiochem.

Aldefluor assay and flow cytometry

ALDH$$^{+}$$ enzymatic activity was measured using the Aldefluor assay (STEMCELL Technologies; details can be found in Supplementary Materials and Methods; ref. 11).

Cell survival assay

MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide] assay was used to evaluate both the chemosensitivity of ovarian cancer cells (A2780_CR5, ALDH$$^{+/-}$$ derived from A2780_CR5) to cisplatin and the platinum resensitization by SGI-110 by determining the 50% growth inhibitory (IC$$_{50}$$) dose values (see Supplementary Materials and Methods).

Cell-cycle analysis

Cell-cycle analysis was conducted as described in Supplementary Materials and Methods.

Sphere and colony formation assays

Sphere formation assays were conducted as described previously (5) and in Supplementary Materials and Methods. Colony formation assay was performed by sorting 500 untreated or drug-treated ALDH$$^{+/-}$$ cells into 6-well coated high-adhesion plates (Corning). Cells were seeded in 2 mL RPMI-1640 (Invitrogen) medium with 10% FBS (Atlanta Biologicals), 1% L-glutamine (Corning), and 1% penicillin/streptomycin (Corning), cultured for 8 days, plates were washed with 2 mL PBS, fixed with 3 mL 10% formalin (Sigma) for 15 minutes, and stained with crystal violet for 5 minutes (0.025% w/v; Sigma). The number of colonies was counted in each well, excluding small (<50 cells) colonies (23).

Differentiation assay

ALDH$$^{+}$$ cells were FACS sorted from control-treated (100 nmol/L DMSO) or SGI-110 (100 nmol/L per day for 3 days)-treated Aldefluor-stained A2780_CR5 ovarian cancer cells, and 50,000 cells then were plated under adherent conditions with either differentiation medium (DMEM/F12 with 10% FBS) or standard RPMI-1640 medium containing 10% FBS, 1% L-glutamine and 1% penicillin/streptomycin, as described (5). The number of ALDH$$^{+}$$ cells on days 7, 14, 21, 28, and 42 was determined using FACS analysis.

In vivo xenograft experiments

All animal studies were conducted according to a protocol approved by the Institutional Animal Care and Use Committee of Indiana University. Female nude, athymic, BALB/c-nu/nu mice (5–6 weeks old; Harlan) were injected subcutaneously (s.c.) with 20,000 ALDH$$^{+}$$ or ALDH$$^{-}$$ cells sorted from either SGI-110 (100 nmol/L/day for 3 days) or vehicle (DMSO)-treated aldefluor$$^{+}$$ A2780_CR5, and ALDH$$^{+}$$ or ALDH$$^{-}$$ were isolated from three high-grade serous human tumors (1,500 cells per mouse). Before subcutaneous injection, cells were resuspended in 100 µL 1:1 RPMI-1640 mixed with Matrigel (BD Biosciences), as described (5). Tumor length ($l$) and width ($w$) were measured weekly using digital calipers and tumor volume ($v$) was calculated as $v = \frac{1}{2} l \times w^2$. Mice were euthanized when tumors were >2 cm in diameter or at end of study. For the carboplatin-response studies, mice were injected intraperitoneally with 2 $\times$ 10$^6$ A2780 cells and subsequently treated with carboplatin (Hospira) at 50 mg/kg, intraperitoneally or PBS ($n = 6–9$ animals per group) weekly for 3 weeks beginning 3 days after injection of cells. For the maintenance study, mice were injected with A2780 cells and treated with carboplatin for 3 weeks, as described. At the completion of the carboplatin treatment, mice were randomized to receive SGI-110 (2 mg/kg) or vehicle, subcutaneous twice-a-week for 2
weeks (n = 12 per group). Mice were euthanized and peritoneal tumors were counted, weighed, and volumes determined as described. Tumors were transferred to tubes containing the medium RPMI-1640 for immediate isolation of cancer cells, or snap frozen in liquid nitrogen or BHT buffer (Qiagen) and then stored at –80°C until DNA and RNA extraction.

Isolation of tumor cancer cells and growth of spheroids
Xenografts were minced and enzymatically dissociated in Dulbecco’s Modified Eagle Medium/F12 (Invitrogen), collagenase (100 U/ml; Sigma-Aldrich), and hyaluronidase (100 U/ml; Sigma-Aldrich), as previously described (5).

DNA extraction, bisulfite conversion, and DNA methylation profiling
Genomic DNA was extracted from A2780 xenografts from mice treated with SGI-110 or control, by using the QiAamp DNA Mini Kit (Qiagen). Sodium bisulfite conversion was performed using the EZ DNA Methylation-Gold kit (Zymo Research), according to the manufacturer’s instructions. After bisulfite conversion, methylation of CpG sites was determined by Infinum HumanMethylation450 BeadChips (Illumina) following a procedure provided by Illumina, at the University of Chicago Genomics Core, Knapp Center for Biomedical Discovery (Chicago, IL). Data quality verification and levels of methylation of the 485,000 CpG sites included in the array were generated by the Illumina GenomeStudio Data Analysis Software. The Illumina Infinium 450k array was used to analyze DNA methylation in promoter site regions. The method measures the methylation levels over 482 k CpG probes. The average percentage of methylation levels were expressed as β-values and ranged from 0 (completely unmethylated) to 1 (completely methylated). Data have been deposited in NCBI’s Gene Expression Omnibus and are accessible through GEO series accession number GSE55613 (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE55613).

Western blot analysis
Protein extracts from control or treated cells were isolated and subjected to Western blot analysis as described (24). Antibodies for DNMT1, ALDH1A1, and GAPDH were from Cell Signaling Technology. After incubation with horseradish peroxidase–labeled secondary antibodies (Cell Signaling Technology), protein bands were visualized using the ECL reaction (Thermo Scientific).

DNA methylation assay by pyrosequencing
Methylation level for CpG islands of selected genes was determined by pyrosequencing assays as described (21), following a procedure provided by EpigenDx. Average methylation level for each CpG dinucleotide was calculated to indicate the methylation levels of each specific gene.

Statistical analysis
All data are presented as mean values ± SD of triplicate measurements. IC50 dose values were determined by Prism 6 (GraphPad Software), using logarithm-normalized sigmoidal dose curve fitting. The Student t test was used to statistically analyze the significant difference among different groups by using Prism 4.0 (GraphPad Software). P value of 0.05 being considered significant. The genome-wide analysis experiments were conducted using the Partek Genomics Suite (version 6.5). The differences in methylation levels between samples (i.e., the differential methylation levels) were calculated using a mixed-model ANOVA. The resultant P values < 0.05 signified highly significant differential methylation levels at a specific nucleotide site. The analysis built a gene integration network, incorporating physical and predicted interactions, colocalization, shared pathways, and shared protein domains. The visualization of the interactions between the genes in the top functional category was realized using Cytoscape (additional informatics analysis can be found in Supplementary Materials and Methods; ref. 25).

Results
ALDH+ cells are enriched in platinum-resistant ovarian cancer
ALDH activity has been demonstrated to be a global and well-established marker for OCSCs (10, 11). To determine the baseline level of ALDH+ subpopulation in ovarian cancer, the percentages of ALDH+ cells in ovarian cancer cell lines A2780, A2780_CR5, and SKOV3 and biopsies from chemotherapy-naïve high-grade serous ovarian cancer (HGSOC) patients were examined using FACS analysis. The percentage of ALDH+ in A2780 was 0.3% and enrichment (P < 0.01; 3-fold increase) and SKOV3 (0.65%, 2-fold increase; Fig. 1A). In primary tumors, the ALDH+ percentage varied (0.4% to 15%); however, the average percentage of ALDH+ cells was similar to ovarian cancer cell lines (3.53% vs. 1.07%; Fig. 1A and Supplementary Table S2). The presence of ALDH+ cells in ovarian cancer cell lines and...
primary tumors and the relative increase in percentage of ALDH<sup>+</sup> cells in platinum-resistant cell lines supports the potential contribution of OCSCs to platinum resistance and poor clinical outcome.

**Low-dose SGI-110 treatment depletes ALDH<sup>+</sup> cells in ovarian cancer**

As transient exposure to low doses of the DNMTI decitabine has been shown to target CSCs in leukemia and breast cancer (15), we examined the effect of the second-generation DNMTI SGI-110 on OCSCs. Ovarian cancer cell lines and dissociated cells from HGSOCs patients' tumors were treated with SGI-110 (100 nmol/L for 3 days) and the number of ALDH<sup>+</sup> cells was determined by FACS analysis. SGI-110 treatment decreased (P < 0.05) the percentage of ALDH<sup>+</sup> cells in A2780 (0.3% to 0.14%), A2780_CR5 (1.07% to 0.33%), and primary ovarian tumors (2.63% to 0.4%), but not in SKOV3 (Fig. 1A), indicating that low-dose SGI-110 has the potential to target OCSCs in most ovarian cancer cells, but not all.

To investigate whether low-dose SGI-110 resensitizes ovarian cancer cells to platinum therapy, MTT assays were conducted after treatment with cisplatin alone or in combination with SGI-110. As expected, A2780_CR5 cells were more (P < 0.001) resistant to cisplatin than A2780 cells (13.6 vs. 3.4 μmol/L; Fig. 1B), and low-dose SGI-110 treatment decreased (P < 0.05) the IC<sub>50</sub> for cisplatin in both cell lines (A2780, 3.4 to 1.6 μmol/L; A2780_CR5, 13.6 to 6.7 μmol/L; Fig. 1B). Similarly, ALDH<sup>+</sup> cells derived from A2780_CR5 displayed increased (P < 0.05) resistance to cisplatin compared with ALDH<sup>+</sup> cells (38.7 vs. 12.2 μmol/L), and SGI treatment increased (P < 0.05) cisplatin sensitivity of A2780_CR5-derived ALDH<sup>+</sup> cells (38.7 to 6.2 μmol/L), as well as ALDH<sup>+</sup> cells (12.2 to 6.0 μmol/L) but to a lesser extent (Fig. 1B). These results suggest that ALDH<sup>+</sup> cells contribute to chemoresistance and can be resensitized to cisplatin by epigenetic therapy.

To further investigate the impact of SGI-110 on OCSCs, we treated ovarian cancer cells with cisplatin (1.67 μmol/L), SGI-110 (100 nmol/L) alone or in combination and examined ALDH<sup>+</sup> cell viability. Cisplatin treatment alone reduced (P < 0.05) the number of viable A2780 (1.80 × 10<sup>6</sup> vs. 4.57 × 10<sup>5</sup>) and SKOV3 (1.50 × 10<sup>5</sup> vs. 8.95 × 10<sup>4</sup>) cells (Fig. 1C), but the number of A2780_CR5-derived ALDH<sup>+</sup> cells increased (1.06 × 10<sup>6</sup> vs. 1.64 × 10<sup>5</sup>) after cisplatin treatment (Fig. 1C). SGI-110 alone inhibited (P < 0.001) the growth of cisplatin-resistant A2780_CR5 (4.58 × 10<sup>6</sup> vs. 2.32 × 10<sup>5</sup>), SKOV3 cells (1.50 × 10<sup>5</sup> vs. 2.92 × 10<sup>4</sup>) as well as A2780_CR5-derived ALDH<sup>+</sup> cells (1.06 × 10<sup>5</sup> vs. 6.09 × 10<sup>4</sup>) and reduced (P < 0.05) the percentage of ALDH<sup>+</sup> cells in A2780 (0.30% vs. 0.14%) and A2780_CR5 (1.5% vs. 0.38%; Fig. 1C and D). As expected, combined treatment with cisplatin–SGI-110 effectively inhibited (P < 0.001) ovarian cancer cell viability (A2780, 1.80 × 10<sup>6</sup> vs. 3.14 × 10<sup>5</sup>; A2780_CR5, 4.58 × 10<sup>6</sup> vs. 1.18 × 10<sup>5</sup>; SKOV3, 1.50 × 10<sup>5</sup> vs. 2.04 × 10<sup>4</sup>) as well as the growth of ALDH<sup>+</sup> cells derived from A2780_CR5 (1.06 × 10<sup>5</sup> vs. 5.43 × 10<sup>4</sup>), and decreased (P < 0.05) the ALDH<sup>+</sup> subpopulation in A2780 (0.30% vs. 0.20%) and A2780_CR5 (1.5% vs. 0.20%; Fig. 1C and D).

To determine whether SGI-110 restored cisplatin sensitivity, effects of treatment with SGI-110 + cisplatin were compared with those induced by cisplatin only. The number of platinum-resistant A2780_CR5, and A2780_CR5-derived ALDH<sup>+</sup> ovarian cancer cells was reduced (P < 0.01) by the combination therapy compared with cisplatin alone (Fig. 1C). The total number of viable A2780_CR5 cells was not reduced by treatment with cisplatin only, and the number of A2780_CR5-derived ALDH<sup>+</sup> cell increased after cisplatin treatment alone (Fig. 1C). The observation that the cisplatin response of A2780_CR5 and A2780_CR5-derived ALDH<sup>+</sup> cells to cisplatin alone was not significant indicates that the effect of SGI-110 + cisplatin on these platinum-resistant cells was not simply additive. In addition, modest activity of single-agent SGI-110 was evident in platinum-resistant cells, inducing more prominent G<sub>0</sub>-G<sub>1</sub> arrest in the platinum-resistant compared with platinum-sensitive ovarian cancer cells (Supplementary Fig. S1), indicating that low-dose SGI-110 exerted a chemosensitization effect. As different cellular backgrounds likely contribute to epigenetic therapy response, SKOV3-derived ALDH<sup>+</sup> population seemed to be more resistant to SGI-110 compared with A2780 and primary tumor–derived cells (Fig. 1C). However, the overall SKOV3 cell population was responsive to the drug, based on increased G<sub>0</sub>-G<sub>1</sub> arrest in SKOV3 cells treated with SGI-110 (Supplementary Fig. S1). Taken together, these data suggest that low-dose SGI-110 exerted antitumor and chemoresensitization effects on OCSC.

**Low-dose SGI-110 reduces ovarian cancer self-renewal and clonogenicity**

We and others previously demonstrated enhanced sphere forming and self-renewal ability of OCSCs when grown in stem cell–selective culture conditions (26–29). To investigate the effect of SGI-110 on sphere formation, A2780 and A2780_CR5 cells were treated with SGI-110/cisplatin alone...
or in combination, and tumor-sphere formation assays were performed. A2780_CR5 cells demonstrated greater \((P < 0.001)\) sphere-forming ability than A2780 \((224 \pm 20 \text{ vs. } 54 \pm 13 \text{ spheres; Fig. 2A and B})\). Moreover, SGI-110 treatment alone markedly inhibited \((P < 0.001)\) the spheroid-forming ability of A2780_CR5 \((224 \pm 20 \text{ vs. } 115 \pm 11)\). The combined SGI-110–cisplatin treatment inhibited \((P < 0.05)\) spheroid formation capability of both the parental line and the resistant subline \((A2780, 54 \pm 13 \text{ vs. } 27 \pm 7; A2780\_CR5, 224 \pm 20 \text{ vs. } 85 \pm 12)\).

To examine the long-term impact of SGI-110 on OCSCs, spheroid formation and colony formation assays were performed using ALDH\(^+\) cells derived from A2780_CR5 treated with cisplatin and (or) SGI-110 and allowed to recover for 4 days. As shown in Fig. 2C and Supplementary Fig. S3, low-dose SGI-110, either alone or in combination with cisplatin, inhibited \((P < 0.05)\) spheroids \((30 \pm 16 \text{ vs. } 13 \pm 2 \text{ SGI-110 or } 30 \pm 16 \text{ vs. } 10 \pm 5 \text{ SGI-110 + cisplatin})\) and colony \((72 \pm 17 \text{ vs. } 58 \pm 5 \text{ SGI-110 or } 72 \pm 17 \text{ vs. } 42 \pm 10 \text{ SGI-110 + cisplatin})\) formation capability of ALDH\(^+\) cells. To further examine the short-term effect of low-dose SGI-110 treatment on self-renewal, the same assay was performed on the A2780_CR5 ALDH\(^+\)/ALDH\(^-\) cells. Treatment with SGI-110 alone had no effect on ALDH\(^+\) ovarian cancer cell growth rate (Supplementary Fig. S2), eliminating growth rate as a major contributing factor to the sphere or colony formation capability of ALDH\(^+\) cells. ALDH\(^+\) formed a greater \((P < 0.05)\) number of spheroids compared with ALDH\(^-\) cells \((87 \pm 52 \text{ vs. } 212 \pm 97)\) (Fig. 2D, Supplementary Fig. S4), but no difference in colony formation was observed for ALDH\(^+\) vs. ALDH\(^-\) (Fig. 2E). In addition, low-dose SGI-110 treatment inhibited \((P < 0.05)\) spheroid-forming capability of ALDH\(^+\) cells \((212 \pm 97 \text{ vs. } 112 \pm 50; \text{Fig. 2D})\) and reduced \((P < 0.001)\) clonogenicity of ALDH\(^+\) \((113 \pm 25 \text{ vs. } 23 \pm 10)\) and ALDH\(^-\) \((90 \pm 17 \text{ vs. } 18 \pm 5)\) cells (Fig. 2E).

Moreover, serial passaging indicated that ALDH\(^+\) cells maintained their sphere-forming ability over multiple passages, whereas the number of cell aggregates formed by ALDH\(^-\) cells was reduced from passage 2 to 3 \((7.7 \pm 1.4 \text{ vs. } 40.0 \pm 1.4; \text{Supplementary Fig. S5})\). Although ALDH\(^+\) ovarian cancer cells were incapable of long-term survival in stem cell culture conditions, the cells demonstrated limited survival in anchorage-independent conditions by grouping together to formed loosely adherent cell clusters \((30)\). Those clusters were unable to undergo serial passage and, therefore, were not considered true “spheroids.” Thus, the initial 3-day low-dose SGI-110 treatment inhibited ALDH\(^+\) sphere-forming ability over three passages \((P < 0.05; \text{Supplementary Fig. S5})\).

As A2780_CR5 ALDH\(^+\) cells generated a greater \((P < 0.05)\) number of ALDH\(^+\) cells in DMEM than in the RPMI-1640 medium, and SGI-110 treatment decreased \((P < 0.01)\) the ALDH\(^+\) subpopulation under either culture condition (Fig. 2F). Although A2780_CR5 ALDH\(^+\) cells persisted and maintained stemness properties during the initial culture period, the proportion of ALDH\(^+\) cells declined \((P < 0.05)\) by 42 days in culture (Fig. 2F), indicating that, A2780_CR5 ALDH\(^+\) cells were able to repopulate ALDH\(^+\) and ALDH\(^-\) and low-dose SGI-110 therapy–induced ALDH\(^+\) differentiation (Supplementary Fig. S6).

Low-dose SGI-110 blunts tumorigenicity of ovarian CSCs by targeting ALDH\(^+\) cells

To examine the effect of SGI-110 on OCSC tumor formation \textit{in vivo}, untreated and SGI-110–treated ALDH\(^+\)/ALDH\(^-\) cells derived from the A2780_CR5 ovarian cancer cell line and primary patient tumors were evaluated in a xenograft model (Fig. 3A). ALDH\(^+\) cells from human tumors \((1,500 \text{ cells per mouse; Fig. 3B})\) or A2780_CR5-derived ALDH\(^+\) \((20,000 \text{ cells per mouse; Fig. 3C})\) displayed robust \((P < 0.05)\) tumor-initiating capacity compared with ALDH\(^-\) cells. Importantly, untreated or SGI-110 pretreated primary tumor-derived ALDH\(^+\) cells were nontumorigenic under these conditions (Fig. 3B), whereas 20,000 A2780_CR5-derived ALDH\(^+\) cells exhibited reduced \((P < 0.05)\) tumor formation ability compared with A2780_CR5-derived ALDH\(^+\) cells \([\text{area under the curve (AUC), } 83 \pm 124 \text{ vs. } 1259 \pm 44; \text{Fig. 3C}]\). Furthermore, increasing the number of ALDH\(^+\)/ALDH\(^-\) cells used in xenograft assays accelerated tumor growth and shortened tumor-initiation time for ALDH\(^+\) and ALDH\(^-\) cells (Supplementary Table S3). Treatment with low-dose SGI-110 for 5 days \textit{in vitro} (before injection into mice) prolonged \((P < 0.05)\) the time to tumor initiation and reduced tumor volume in ALDH\(^+\) xenografts (Supplementary Table S3). To examine the effects of SGI-110 on OCSCs, the percentage of ALDH\(^+\) cells in untreated versus SGI-110–treated A2780_CR5-derived ALDH\(^+\) xenograft tumors was assessed by FACS. ALDH\(^+\) cells were highly enriched \((P < 0.05)\) in the A2780_CR5-derived ALDH\(^+\) xenografts compared with ALDH\(^+\) tumors \((71.2\% \pm 8.7\% \text{ vs. } 13.3\%\), suggesting that ALDH\(^+\) cells were unable to dedifferentiate into ALDH\(^-\) cells. SGI-110 reduced \((P < 0.05)\) ALDH\(^+\) subpopulation \((71.2\% \pm 8.6\% \text{ to } 55.8\% \pm 0.9\%)\) in xenografts (Fig. 3D).

ALDH\(^+\) cells overexpress stemness-associated genes

To confirm the stem-like properties of the ALDH\(^+\) ovarian cancer cells, we examined the mRNA levels of several known stem cell–associated genes \((4, 11, 30)\). Compared with nOSE, HGSOCs \((n = 5)\) displayed increased \((P < 0.05)\) expression of stem cell–associated genes, \textit{NOTCH3}, \textit{OCT4}, and \textit{ALDH1A1} (Fig. 4A). Importantly, increased \((P < 0.05)\) expression levels of \textit{ALDH1A1} \((162.4 \pm 28.8\text{-fold}), \textit{BMI1} (8.8 \pm 0.2\text{-fold}), \textit{NANOG} (9.5 \pm 0.8\text{-fold}), \textit{NOTCH3} (1.9 \pm 0.6\text{-fold}), and \textit{OCT4} (7.1 \pm 1.7\text{-fold}) and decreased \((P < 0.05)\) expression levels of the differentiation-related gene \textit{HOXAI0} \((4.1 \pm 0.7\text{-fold})\) and \textit{HOXA11} \((2.4 \pm 0.8\text{-fold})\) were observed in A2780_CR5-derived ALDH\(^+\) compared with ALDH\(^-\) cells (Fig. 4B and Supplementary Fig. S7).

Low-dose SGI-110 induces differentiation of ALDH\(^+\)

Methyltransferases DNMT1, 3A and 3B are the main effectors of DNA methylation. Deregulated levels of DNMTs have been reported in cancer \((31)\) and in ovarian cancer \((32, 33)\), and in association with platinum resistance. Therefore, \textit{DNMT1}, 3A, and 3B expression levels were measured in platinum-sensitive \((A2780)\) and –resistant \((A2780\_CR5)\) sublines and in A2780_CR5-derived ALDH\(^+\) and ALDH\(^-\) cells. \textit{DNMT1} was \((P < 0.05)\) significantly upregulated in cisplatin-resistant A2780_CR5 and ALDH\(^+\) cells, and \textit{DNMT3A} and 3B were overexpressed in ALDH\(^-\) cells, suggesting that aberrant
Figure 3. SGI-110 decreases tumorigenesis by targeting ALDH\(^+\) cells. A, schematic diagram of the approach used to study in vivo tumorigenesis of low-dose SGI-110–untreated and -treated ALDH\(^+\)/– cells. B, primary xenograft tumor growth curve of 1,500 patient-derived ALDH\(^+\) or ALDH\(^–\) cells pretreated with SGI-110 (100 nmol/L) for 72 hours or DMSO in mice (\(n = 3\) for each group) during 8 weeks. Average of AUC was calculated and shown in the histogram. ALDH\(^+\)/– cells isolated from three high-grade serous human tumors (1,500 cells per mouse). C, xenograft tumor growth curve of 20,000 A2780_CR5–derived ALDH\(^+\) or ALDH\(^–\) cells pretreated with SGI-110 (100 nmol/L) for 72 hours or DMSO in mice in 7 weeks (\(n = 3\) for each group). AUC was calculated and shown in the histogram. D, average number of ALDH\(^+\) population present in untreated or SGI-110 (100 nmol/L)-pretreated A2780_CR5-derived ALDH\(^+\) or ALDH\(^–\) xenograft tumors. Mean values \(\pm\) SD of three independent experiments are shown (\(*\), \(P < 0.05\); **, \(P < 0.01\); ***, \(P < 0.001\)).
methyltransferase (DNMT) activity in OCSCs (Fig. 4B). We then assessed stem-cell, differentiation-related genes, and DNMT isoforms (DNMT1, DNMT3A, and DNMT3B) and differentiation-associated genes (HOXA10 and NOTCH3) were measured by qRT-PCR in A2780_CR5 (platinum-resistant), ALDH− and ALDH+ cells derived from A2780_CR5 compared with parental A2780 cells. C, scheme of low-dose SGI-110 time-course treatment. D, average expression of stemness genes (BMI1, NANOG, NOTCH3, and OCT4) were measured by qRT-PCR in A2780_CR5 compared with untreated ALDH+ cells. E, average expression of the differentiation-associated gene HOXA10 mRNA measured by qRT-PCR in A2780_CR5-derived ALDH+ cells over SGI-110 (100 nmol/L) treatment compared with untreated ALDH+ cells. Average of DNA methylation level of HOXA10 was measured using pyrosequencing. Average expression of mRNA DNMT1 (F, top) and ALDH1A1 (F, bottom) was measured by qRT-PCR in A2780_CR5-derived ALDH+ cells at indicated time points over SGI-110 (100 nmol/L) treatment compared with untreated ALDH+ cells. Three independent experiments were performed and mean values ± SD are calculated (**, P < 0.01; ***, P < 0.001).
Figure 5. *In vivo* effects of carboplatin on xenograft growth and ovarian CSCs. A, effects of carboplatin on weights, volumes, and metastases sites of xenograft tumors derived from A2780 cells. Bars represent average measurements ±SD; ***, *P* < 0.001 (n = 6 per group). B, percentage of ALDH*+ cells in control or carboplatin-treated xenografts. Cells were isolated by mechanical and enzymatic digestion and ALDH*+ cells were detected by FACS. Bars represent average of four measurements ±SD; ***, *P* < 0.001 (left). Representative FACS histograms are shown on the right. C, spheroid formation by cells dissociated from control or carboplatin-treated xenografts; bars represent average of three measurements ±SD; **, *P* < 0.01 (left). Phase microscopy shows morphology of spheres formed by cells dissociated from control or carboplatin-treated xenografts (×100 magnification, right).
Injection of A2780 cells

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<th>Day</th>
<th>Carboplatin</th>
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Tumor collection

**Injection of A2780 cells**

**Carboplatin**

**Carbo-SGI**

**Vehicle**

**Relative numbers of spheroids**

**Sphere formation assay**

**Effects of SGI-110 on DNA methylation in xenografts**

**Effects of SGI-110 on DNA methylation profiles**

**Gene networks affected by SGI-110**

- Signaling
- Transcription regulation
- Transport
- Cell adhesion
- Morphogenesis
- Apoptosis
- Proteolysis
- Cell development
- GTPase regulation
- Metabolism
- Protein synthesis
the SGI-110–hypomethylating effect. SGI-110 also reduced (P < 0.05) the mRNA expression levels of DNMT1 (Fig. 4F), DNMT3A, and 3B (Supplementary Fig. S8A and S8B) and ALDH1A1 (Fig. 4F) in ALDH+ cells up to 14 days (P < 0.05). Three-day low-dose SGI-110 treatment also resulted in decreased DNMT1 and ALDH1A1 protein levels in A2780, A2780_CR5, and SKOV3 cells (Supplementary Fig. S8C–S8E). These data support that low doses of SGI-110 promote differentiation of ALDH+ ovarian cancer cells and suppress their stem-like properties.

**Enrichment in ALDH+ cells after platinum in ovarian cancer xenografts**

To test the hypothesis that CSCs persist in ovarian tumors after platinum-based chemotherapy, we used an intraperitoneal xenograft model derived from parental (platinum-sensitive) A2780 ovarian cancer cells treated with carboplatin or vehicle (control). Tumor volume, weight, and number of metastases were significantly decreased (P < 0.001) by weekly treatment with carboplatin at 50 mg/kg (Fig. 5A). Vehicle and carboplatin-treated tumors were dissociated to single-cell suspension at the end of treatment and cells were analyzed for Aldefluor positivity, and for ability to form spheres in anchorage-independent conditions. The percentage of ALDH+ cells was increased (P < 0.001) approximately 20-fold in tumors residual after carboplatin compared with vehicle-treated tumors (Fig. 5B). Cells dissociated from carboplatin-treated tumors formed increased (P < 0.001) numbers and size of spheres compared with cells dissociated from control tumors (Fig. 5C), consistent with an OCSC phenotype. In all, carboplatin significantly decreased tumor growth in vivo but also contributed to enriching the OCSC population in residual tumors.

**DNA demethylation induced by SGI-110 delayed recurrence of ovarian cancer xenografts**

To determine whether DNA hypomethylation induced by SGI-110 prevents tumor recurrence after maximal response to platinum therapy, mice bearing intraperitoneal A2780-derived xenografts and treated with carboplatin were randomized to a 2-week treatment with SGI-110 or vehicle (Fig. 6A, n = 12 mice per group). Treatment with SGI-110 decreased (P < 0.05) the total tumor weight and volume compared with control-treated mice (Fig. 6B). Cells dissociated from SGI-110–treated tumors significantly reduced spheroid formation capability in vitro (Fig. 6C, P < 0.05), consistent with inhibition of stem cell properties. To demonstrate that SGI-110 induced global DNA hypomethylation consistent with its DNMT inhibitory properties, Illumina Infinium HumanMethylation450 arrays were used to quantify DNA methylation in control and SGI-110–treated ovarian xenografts. More than 62,000 methylation sites and 10,000 CpG islands were found to be significantly hypomethylated in SGI-110–treated tumors compared with controls (Table 1). The substantial global DNA hypomethylation induced by SGI-110 in vivo was also demonstrated by a decrease of 6% in β-values across all CpG islands (P < 0.0001, Fig. 6D) and through unsupervised hierarchical clustering analysis of methylation sites in control and SGI-110–treated xenografts (Fig. 6E). To understand biologic processes represented by the genes whose promoter CpG islands were significantly hypomethylated in response to SGI-110, we grouped those genes into well-defined functional gene ontology (GO) categories, using DAVID (Database for Annotation, Visualization, and Integrated Discovery; ref. 34). Of the 84 genes meeting the criteria (described in Materials and Methods, Supplementary Table S4), 65 had well-defined GO categories, and 40 could be grouped into ten functional categories containing at least three assigned genes. These categories represent important biologic processes, including metabolism, apoptosis, proteolysis, cell development, morphogenesis, cell adhesion, transport, signaling, transcriptional regulation, and GTPase regulation (Fig. 6F), suggesting that hypomethylation induced by SGI-110 alters critical pathways in cancer. Genes included in these networks include PCDH10, a gene known to be downregulated in cancer through DNA methylation (35), miR-203 that is epigenetically silenced in myeloma and involved in apoptosis control (36), PTKe, involved in epithelial–mesenchymal transition (37), and others.

**Discussion**

Our results demonstrate that ALDH+ ovarian cancer cells possessing stem cell characteristics are enriched in platinum-resistant ovarian cancer cell lines, human tumors, and xenografts residual after platinum therapy. The novel DNMT1 SGI-110 inhibits ALDH+ cell viability, sphere formation, and tumor-initiating capacity, represses stem cell–associated gene transcription, and resensitizes platinum-resistant ovarian cancer cells to platinum. In vivo, maintenance treatment with SGI-110 after carboplatin induces profound global hypomethylation and delays tumor progression. Collectively, our data suggest that a strategy targeting DNA methylation in ovarian cancer exerts potent antitumor activity by allowing elimination of ALDH+ cells enriched in residual, platinum-resistant tumors. Our data have several implications. First, we assert that ALDH1A1 expression and activity characterizes ovarian cancer cells with stem cell properties,
in agreement with reports from other groups (6, 30, 38). Aldefluor positivity detectable by FACS identifies the enzymatic activity of ALDH1, a member of the ALDH family that metabolizes reactive aldehydes (39). Although ALDH positivity has been recognized as a stem cell marker in various tissues, the role of the enzyme in the functions of CSC remains elusive. A potential function relates to its regulatory role in the synthesis of retinoids, which play a critical role in cellular differentiation. Whether the enzyme has other functions important to the maintenance of CSCs remains not known. Here, we show that ALDH+ cells derived from ovarian cancer cell lines and from primary ovarian tumors are more resistant to platinum, express stem cell restricted transcription factors, and are able to generate spheres and tumors in vivo.

Second, we demonstrate that ovarian xenografts residual after treatment with platinum are enriched in ALDH+ cells, suggesting that cells with stem cell characteristics escape traditional cytotoxic treatments. Our model is consistent with the proposed concept that stem cells elude the effects of traditional anticancer strategies and can reconstitute recurrent tumors that become recalcitrant to chemotherapy (12). We use A2780, a tumorigenic and one of the few available platinum-sensitive ovarian cancer cell lines, to recapitulate the clinical evolution of ovarian cancer, with massive initial response to chemotherapy, followed by inevitable resurgence of resistant tumors. Although it has been suggested that the genomic signature of A2780 does not fully match that of HGSOC (40, 41), previous studies demonstrated aggressive in vivo growth of A2780 cells (26, 42) and response to platinum, resembling the human disease. Our model supports that tumor recurrence could be attributed to the persistence of platinum refractory stem cells at the end of initial treatment. We propose a novel strategy to target these resistant cells through epigenomic reprogramming by using a novel DNMTI. We show that SGI-110 suppressed the viability of ALDH+ cells, their ability to form spheres in vitro, and their tumorigenic potential in vivo. Importantly, treatment with SGI-110 reseminated platinum-resistant ALDH+ cells to platinum, providing proof-of-concept for further investigating hypomethylating strategies as means to resensitize tumors to chemotherapy.

Third, it has been recognized that embryonic and CSCs harbor distinct DNA methylation profiles (43, 44) that enable tight control of cell differentiation and self-renewal capacity. Therefore, treatment of a stem cell–enriched cell population with DNA-hypomethylating agents would remove the repressive epigenetic brakes, allowing stem cells to undergo differentiation and leave the pluripotent undifferentiated state. Although this concept has been tested in leukemia models (45), it remains unexplored in solid tumors. Here, we show for the first time that the expression levels of all three DNMT isoforms is significantly increased in ALDH+ cells derived from platinum-resistant ovarian cancer cells and that SGI-110 is able to re-set these cells toward differentiation. As histone modifications have been associated with DNA methylation and regulation of stemness-associated genes, and an association between EZH2 and ALDH1A1 expression has been reported (46), it seems plausible that additional epigenetic regulatory mechanisms contribute to maintaining stem cell characteristics. Thus, our studies provide the first proof-of-principle that epigenomic strategies efficiently target ovarian cancer stem cells.

Our results also demonstrate that SGI-110 induces profound hypomethylation in vivo, with tens of thousands of CpG sites becoming demethylated in response to treatment. A distinctly hypomethylated DNA profile emerges, providing reassurance that the novel DNMTI hits its biologic targets in solid tumors in vivo. Future studies will strive to identify the critical genes or pathways responsible for tumor growth inhibition and chemotherapy resensitization in response to this DNMT inhibitor. It is likely that not a single gene, or pathway, but a complex program is reengaged by targeting the epigenome, as we show here by pathway analyses.

Finally, we demonstrate that treatment with SGI-110 after platinum decreases recurrent tumor burden in a platinum-sensitive ovarian cancer intraperitoneal xenograft model that recapitulates first the response to therapy, and second the recurrence of disease after chemical debulking using carboplatin. These data support exploring maintenance treatment with a hypomethylating agent after maximal response induced by traditional treatment. Maintenance strategies after chemotherapy have been investigated with variable level of success in ovarian cancer (47–49) and remain an area of active

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<th>Control vs. SGI-110</th>
<th>Decreased methylation</th>
<th>Increased methylation</th>
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<td>Δβ &gt; 0.2 a</td>
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<td>CpG sites</td>
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<td>CpG regions b</td>
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NOTE: Methylation of CpG sites was determined using Infinium HumanMethylation450 arrays and was expressed as β-values ranging from 0 (unmethylated) to 1 (totally methylated). Significant changes: P < 0.01 (ANOVA) and FDR < 0.05.

aΔβ, Difference in β-values between control and SGI-110 groups.
bCpG islands plus shores and shelves.
Epigenetic Targeting of Ovarian Cancer Stem Cells

exploration. Our study provides the first evidence that epigenome targeting strategies decrease tumor progression by targeting and reprogramming residual CSCs, supporting further refinement of this intervention and translating these findings to the clinic.

Disclosure of Potential Conflicts of Interest
K.P. Nephew received a commercial research grant from and is a consultant/advisory board member for Astex Pharmaceuticals, Inc. D. Matei is a consultant/advisory board member for Astex Pharmaceuticals, Inc. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions
Conception and design: Y. Wang, H. Cardenas, K.P. Nephew, D. Matei
Development of methodology: Y. Wang, F. Fang, S. Condello, Y. Liu, K.P. Nephew, D. Matei
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): Y. Wang, H. Cardenas, S. Condello, M. Segar, Y. Liu, K.P. Nephew, D. Matei
Analysis and interpretation of data (e.g., statistical analysis, bios-statistics, computational analysis): Y. Wang, H. Cardenas, F. Fang, S. Condello, M. Segar, Y. Liu, K.P. Nephew, D. Matei

References

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