Microenvironment and Immunology

Cellular Factors Promoting Resistance to Effective Treatment of Glioma with Oncolytic Myxoma Virus

Franz J. Zemp1,2, Brienne A. McKenzie1,2, Xueqing Lun1,2, Karlyne M. Reilly3, Grant McFadden4, V. Wee Yong5,6, and Peter A. Forsyth1,2,7

Abstract

Oncolytic virus therapy is being evaluated in clinical trials for human glioma. While it is widely assumed that the immune response of the patient to the virus infection limits the utility of the therapy, investigations into the specific cell type(s) involved in this response have been performed using nonspecific pharmacologic inhibitors or allogeneic models with compromised immunity. To identify the immune cells that participate in clearing an oncolytic infection in glioma, we used flow cytometry and immunohistochemistry to immunophenotype an orthotopic glioma model in immunocompetent mice after Myxoma virus (MYXV) administration. These studies revealed a large resident microglia and macrophage population in untreated tumors, and robust monocyte, T-, and NK cell infiltration 3 days after MYXV infection. To determine the role on the clinical utility of MYXV therapy for glioma, we used a combination of knockout mouse strains and specific immunocyte ablation techniques. Collectively, our experiments identify an important role for tumor-resident myeloid cells and overlapping roles for recruited NK and T cells in the clearance and efficacy of oncolytic MYXV from gliomas. Using a cyclophosphamide regimen to achieve lymphoablation prior and during MYXV treatment, we prevented treatment-induced peripheral immunocyte recruitment and, surprisingly, largely ablated the tumor-resident macrophage population. Virotherapy of cyclophosphamide-treated animals resulted in sustained viral infection within the glioma as well as a substantial survival advantage. This study demonstrates that resistance to MYXV virotherapy in syngeneic glioma models involves a multifaceted cellular immune response that can be overcome with cyclophosphamide-mediated lymphoablation. Cancer Res; 74(24); 7260–73. ©2014 AACR.

Introduction

In North America, approximately 25,000 new cases of malignant brain tumors were diagnosed in 2013 (1, 2). Malignant gliomas constitute 80% of these tumors, and have a universally fatal outcome with the most severe having an average survival of only 12 to 15 months despite our best treatment regimens (3). Clearly more effective treatments are needed.

Oncolytic virus therapy shows promise as an experimental therapeutic in malignant gliomas in preclinical models and has proven to be safe in early malignant glioma clinical trials (4, 5). This experimental therapy uses replication-competent viruses that exploit natural or engineered viral trophic restrictions to selectively infect and kill cancer cells. Oncolytic virus therapy is an appealing treatment strategy for several reasons. As a replicating virus, it allows for self-potentiation, while the multiple mechanisms by which an oncolytic virus can kill tumors could potentially overcome challenges that are faced by other targeted therapies. For instance, oncolytic viruses have been demonstrated to direct antitumoral responses through direct cell killing, destroying tumor vasculature, eliciting antitumor cytokine responses, and provoking antitumor immune responses (6).

The ability of oncolytic viruses to elicit antitumoral effects will largely rely on their ability to infect and replicate within the tumor. This has been demonstrated in glioma xenograft models in immunocompromised mice, whereby tremendous efficacy is accompanied by robust viral replication (7–10). However, preclinical studies in immunocompetent glioma models have demonstrated that viral infection is dramatically reduced and rapidly cleared (11–15). We have found similar results in malignant glioma models using Myxoma virus (MYXV), a rabbit-specific member of the Poxviridae family.
family that does not infect humans or other nonlago-
morph species. In immunocompromised models of malig-
nant gliomas, we found lengthy viral replication and durable
responses, with treatment often resulting in "cures" (16–19).
In contrast, we have demonstrated a lack of sustained viral
replication within tumors in situ and no significant survival
benefit in syngeneic glioma models that possess fully func-
tional immune systems (11, 15).

These results in immunocompetent preclinical models
reflect those seen in early clinical trials, which demonstrated
that viral clearance of oncolytic herpes viruses occurs rap-
idly after intratumoral injections and is accompanied by an
effective cell-mediated antiviral-immune response (20, 21).
Thus far, most patients with malignant glioma do not
respond to oncolytic virus therapy, but a small proportion
of long-term survivors have occurred in a recent phase I/II
safety trial treating recurrent glioblastoma with oncolytic
delta24RGD Adenovirus (Frank Tufaro; personal communi-
cation). Hence, understanding the host factors that limit
oncologic virus replication and accelerate its immune clear-
ance would be critical to improving the efficacy of this
approach in the clinic.

Several studies have looked at different immune effectors
that potentially limit oncolytic infection within gliomas. For example, a robust recruitment of monocyctoid cells
following oncolytic virus treatment of intracranial glioma
models has been shown through immunohistochemical
staining [oncologic Vaccina virus (JX-594; ref. 14), herpes
simplex virus (HSV; refs. 22, 23), measles virus (24), and
MYXV (15)]. Similar results have been found in clinical
glioma samples after oncolytic virus therapy with HSV
(20) and adenovirus (23) administration. Targeting the
recruitment of these cells with immunosuppressive com-
ounds has been shown to inhibit the recruitment of these
cells and increase virotherapy effectiveness (14, 15, 22);
however, the pleiotropic nature of these reagents compli-
cates the interpretation that immunosuppressive drugs act
on specific cell types alone. Furthermore, human malignant
gliomas are heavily infiltrated with glioma-infiltrating mon-
ocyes/macrophages (GIM) before treatment, (25–27) and
there have been no studies to date that have attempted to
separate the role of glioma resident versus treatment-
recruited monocytes in gliomas in vivo.

A role for NK cells in mediating oncolytic virus clearance
from gliomas has recently been reported. First identified as
recruited in response to glioma HSV virotherapy in a syngeneic
rat model (22), inhibition of these cells with various pleiotropic
drugs has demonstrated increased oncolytic glioma infection
(22, 28). Specific interrogation of NK cells recruited by HSV
glioma virotherapy suggested that viral distribution through-
out the tumor is limited through the swift destruction of
infected tumor cells by NK cells (29). Thus, NK cells have also
been suggested to be important in mediating viral distribution
and clearance in experimental models of malignant gliomas.

In this study, we immunophenotyped the glioma microen-
vironment before and after intratumoral administration of
MYXV therapy in an immunocompetent orthotopic glioma
model. We have previously found in this model that MYXV
is cleared from these tumors between 3 and 7 days after
intratumoral infection (11). Here, we determined the contri-
bution of various immunocytes responsible for the swift viral
clearance of MYXV using a variety of transgenic animals with
specific immunocyte deficiencies. To our knowledge, this is
the first study to utilize transgenic animals to specifically
target immunocyte populations to assess their involvement
in mediating oncolytic virus infection, replication, immune
clearance, and treatment efficacy in immunocompetent malig-
nant glioma models.

Materials and Methods

Cell lines and viruses

The K1492 cell line was derived from C57Bl/6j Trp53+/−/
Nf1−/− mice (30). These lines were cultured in DMEM (#11965,
Invitrogen) containing 10% FBS (Invitrogen) and tested for
mycoplasma at regular intervals. Detailed description of
these lines when orthotopically grafted into C57Bl/6 mice,
as well as a characterization of their in vitro and in vivo
response to MYXV treatment has previously been provided
(11). Generation, propagation, and titration of the viruses
(vMyx-GFP and vMyx-FLuc) has previously been demonstrated
(11, 15–19, 31, 32).

Mouse strains

Female wild-type (C57Bl/6j, #000664), CCR2-null (B6.129S4-
Ccr2tm1Ifc/J, #004999), RAG1-null (B6.129S7-
Rag1tm1Mom/J, #002216), and IL2Rγ-null (B6.129S4-
Il2rgtm1Wjl/J, #003174) mice were purchased from Jackson Laboratories. RAG1/IL2Rγ mice
were created by crossing female RAG1 mice to male IL2Rγ
mice. Six- to 8-week-old mice were used for all experiments.
The animals were housed in a vivarium maintained on a
12-hour light/dark schedule with a temperature of 22 ± 1°C
and a relative humidity of 50% ± 5%. Food and water were
available ad libitum.

Intracranial surgeries

Orthotopic injections of K1492 were performed by injecting
5 × 10⁶ cells in PBS in a 2 μL volume in the right striatum of
mice as described previously (11, 15–17). Animals were fol-
lowed until they lost ≥20% of body weight or had trouble
ambulating, feeding, or grooming. Intracranial injection of
vMyx-FLuc or vMyx-GFP was performed on day 14 for all lines
in survival or bioluminescent studies, and on day 12 for ilow
cytometric studies. Viral treatments were stereotactically
administered through the same burr hole created for the tumor
implantation. All animal work procedures were in accordance
with the Guide to the Care and Use of Experimental Animals
published by the Canadian Council and all protocols were
reviewed and approved by the Animal Care Committee of the
University of Calgary (Protocol #AC12-004).

Monitoring viral infection in vivo

Bioluminescence from the vMyx-FLuc–infected tumors was
imaged with the Xenogen IVIS 200 System by intraperitoneal
injection of d-Luciferin, FireFluor, potassium salt (5 μL/g;
#119222, Caliper Life Sciences) made to 30 mg/mL in PBS and
filter sterilized through a 0.2-μm filter. Luminescence was analyzed by drawing a region of interest around the entire skull and measuring the total emission from that area in units of total FLUX.

Analyses of the number of functional virions were performed by crushing the tumor-bearing hemisphere in 500 μL of cold PBS. Samples were then sonicated (2 × 2 seconds; 10% amplitude; Fisher Scientific Sonic Dismembrator Model 500). This slurry was then spun at 3000 × g for 15 minutes and frozen. Supernatants were freeze thawed three times and then titered on BGMK cells. Titers of MYXV are reported as focus forming units (FFU), measured as fluorescent infection foci on these cells.

Flow cytometry

Mice bearing 12-day K1492 glioma orthografts were treated with vMyx-GFP or PBS, or left untreated, and then sacrificed for flow cytometry 3 days after treatment. Mice were cardiac perfused with 6 mL of PBS and the tumor-bearing hemisphere placed in RPMI1640 (Gibco) with 2% FBS. Homogenized brains were passed through a 70 μm filter and fractionated through a Percoll (GE Healthcare) gradient. Purified immunocytes were then Fc blocked (BD Mouse Fc Block, #553141) and then stained with CD45 [PE rat IgG1, anti-mouse CD45 (BD, #553081)], PE rat IgGk anti-mouse isotype (BD #556925), CD11b [PerCP-Cy5.5 rat IgG1, anti-mouse CD11b (BD #561114), PerCP-Cy5.5 rat IgGk anti-mouse isotype (BD #552991)], CD3ε [APC Ar Ham IgG1, anti-mouse CD3ε (BD #533066), APC Ar Ham IgGk anti-mouse isotype (BD #553874)], Ly6G [PE-Cy7 rat IgG1, anti-mouse Ly6G isotype (BD #566061), PE-Cy7 rat IgGk, anti-mouse CD11b (BD #552784)], and/or NK1.1 [PE-Cy7 mouse IgG2a, anti-mouse CD56 (BD #559027)]. Stained cells were then analyzed on an Attune Acoustic Focusing Cytometer with blue/red lasers using the corresponding software. All cells were initially gated on the "live cell" fraction based on forward and side scatter plots, with the CD45/CD11b plots using a large initial gate to include larger macrophage and neutrophil populations. Flow experiments examining NK and T-cell populations used a small initial gate on just lymphocytes to avoid off-target staining in the macrophage and neutrophil populations.

Immunohistochemistry

K1492 brains were placed in 10% buffered formalin for a minimum of 48 hours. Brains were then cut coronally approximately 0.5 mm behind the injection site and mounted in paraffin blocks, and the first and last slides cut were stained using standard H&E by AIRG Histology Services. Immunohistochemistry was performed using M-T7e (McFadden Laboratory; 1:1,000) and IBa1 (Wako, #019-19741; 1:1,000) primary antibodies and these were detected using a biotinylated goat anti-rabbit (Vector; 1:300) and Vectastain ABC Elite Reagent (Vector Labs, PK-6100). Slides were mounted, counterstained with hematoxylin, and viewed with a Zeiss inverted microscope (Axiovert 200M) and a Carl Zeiss camera (AxioCam MRC).

Immunocyte ablation studies and inactivation studies

Clodronate liposomes were obtained from www.clodronates.org. K1492-bearing C57Bl/6 mice were adminis-tered 200 μL of clodronate liposomes or control liposomes (vehicle) via tail vein injection at −1, 0, 1, and 2 days after MYXV treatment. Functional grade low-endotoxin CD161 (NK1.1; PK130 antigen) or isotype control antibody was purchased from AbLab. Antibodies were administered on 12 and 14 dpi, with vMyx-FLuc administered on 14 dpi. Cyclophosphamide (Sigma) was diluted in sterile PBS and administered intraperitoneally at indicated doses.

Statistical analysis

All data were processed and graphed in either MS Excel 2010 or Prism GraphPad v5.0. Statistics were also performed in these programs. All data was assumed to be normally distributed, and F tests were used to determine whether homoscedastic or hetereoscedastic t tests were needed. All t tests were two-sided and values were considered to be statistically significant at P < 0.05. In vivo luciferase monitoring produced highly variable data, thus distributions could not be considered normal, and nonparametric Mann–Whitney (Wilcoxon rank sum) tests were conducted. Survival curves were generated by the Kaplan–Meier method and statistics were determined using the log-rank Mantel–Cox test.

Results

Microglia and macrophages predominantly reside in the glioma with further peripheral recruitment of monocytoid cells following treatment

We have previously characterized the murine K1492 glioma cell line as a suitable malignant glioma model in C57Bl/6 mice to preclinically assess MYXV treatment (11). This line was derived from Trp53<−/−/Nf1<−− mice that spontaneously develop high-grade gliomas that recapitulate many clinical features of the human disease (30). In vitro, K1492 is readily infected by MYXV, supports robust viral replication, and is very susceptible to viral-mediated cell death; (11) however, this same cell line granted intracranial and given intratumoral administration of MYXV was refractory to treatment, with no evidence for viral replication or treatment efficacy.

We analyzed the immunocyte composition of the K1492 tumor microenvironment with or without MYXV treatment. We focused on immunocyte populations that were present 3 days after MYXV treatment as most, but not all, of the virus is cleared from the tumor by this time (11). Using conventional neuroimmunological markers for immunocytes in the brain, we first considered CD45R<−/−CD11b<− cells as microglia, CD45<−/−CD11b<− as GIMs, and CD45<−/−CD11b<− cells as lymphocytes.

Comparing the normal mouse brain with the untreated K1492 brain confirmed that these murine malignant gliomas, similar to human malignant gliomas, contain a significant number of immunocytes (Fig. 1A). A nearly 2-fold increase in CD45R<−/−CD11b<− microglia could be seen, making up 66.4% ± 2.1% of the total CD45R<−/− cells found in the tumor-bearing hemisphere. These microglia appeared to have an activated phenotype, as demonstrated by the large shift in this population displaying increased levels of CD45 and CD11b (53). The
remaining CD45<sup>-</sup> cells were CD45<sup>high</sup> peripheral immunocytes (29.6% ± 1.8%), which were largely dominated by CD45<sup>high</sup> CD11b<sup>+</sup> GIMs (80.9% ± 1.9% of CD45<sup>high</sup>), but also included CD11b<sup>-</sup> lymphocytes (20.7% ± 1.4% of CD45<sup>high</sup>).

Three days after intratumoral MYXV injection, we found a significant loss of CD45<sup>low</sup>CD11b<sup>+</sup> microglia, returning to levels seen in the normal mouse brain. Infection also resulted in the recruitment of additional peripheral CD45<sup>high</sup> cells, reaching levels nearly 2-fold over the untreated K1492 glioma. These CD45<sup>high</sup> cells were largely CD11b<sup>+</sup> (GIMs; 70.3% ± 3.6%), but also contained a large proportion of CD11b<sup>-</sup> cells (lymphocytes; 30.6% ± 3.4%). In addition, a population of cells straddling the CD11b gate in the CD45<sup>high</sup> compartment was observed. These changes were in response to viral inoculation and not a consequence of brain injury, as sterile PBS administration into K1492 tumors did not significantly alter any immunocyte populations (Supplementary Fig. S1A). Finally, as these experiments used a GFP-expressing MYXV, any infected immunocytes in which the virus could initiate replication would be visible in the FITC channel during flow measurements. We found no signal in the FITC channel, suggesting that any viral infection or replication at this time point were in nonimmunocyte populations.

To confirm the extent of microglial and GIM infiltration into K1492 tumors, we stained sections with the conventional myeloid marker Iba1, which stains both microglia and macrophages. We found that microglial and GIM infiltration was extensive, occurring both within and around the tumor (Fig. 1B). Microglia could be seen in a "gradient of activation," whereby ramified/unactivated microglia could be observed farther away from the tumor with the gradual transformation into the activated/amoeboïd phenotype as they approached the tumor (Supplementary Fig. S2A). In response to MYXV infection, 1 day after treatment, we observed dense clusters of polymorphonuclear cells surrounding and within the infected areas (Supplementary Figs. S2 and S3). By 3 days after treatment, these areas of infection were largely cleared of viral protein and these polymorphonuclear cells. Areas of residual infection had intense leukocyte infiltration and the onset of necrosis. This was also apparent at 7 days after treatment, where there was a complete absence of viral protein but large...
areas of focal necrosis were present (Supplementary Fig. S2, black arrows).

While such extensive microglia/GIM infiltration has been known to occur in both human malignant gliomas and immunocompetent rodent models of disease, (25–27) we wanted to confirm that this infiltration was not an artifact of the K1492 orthotopic model. We confirmed a similar pattern of Iba1+ cellular infiltration in the K1861 orthorthax graft in C57 mice (Supplementary Fig. S4A). Furthermore, we found that in spontaneous high-grade gliomas in NPCs (Trp53−/−/Nfi−/−) mice, microglia, and or GIM infiltration was extensive (Supplementary Fig. S4B).

Ablation of both tumor-resident and treatment-recruited GIMs

Given that one of the largest populations of cells infiltrating the untreated and treated K1492-tumor-bearing hemisphere was the CD45shbshCD11b− "GIMs," this was the first population assessed for their role in mediating MYXV clearance and treatment failure in our model. It has been shown under neuroinflammatory conditions that CCR2 expression is necessary for monocyte recruitment to the CNS in experimental autoimmune encephalitis, (34) ischemic injury, (35), and viral infection (36), but this has not been tested in a murine malignant gliomas. As expected, we found that CCR2-null mice bearing K1492 malignant gliomas had significantly fewer CD45shbshCD11b− cells than glioma-bearing wild-type mice (2.8-fold decrease; Fig. 2A). This decrease was slightly less prominent after viral infection (1.4-fold), but accompanied by a large population CD45shbshCD11b+Ly6G+ granulocytes (Fig. 2A, yellow). Using this additional marker, we determined that GIMs (CD45shbshCD11b+Ly6G−) were reduced 5.8-fold in the untreated and 2.5-fold in the MYXV-treated CCR2-null animals (Fig. 2B). Furthermore, we found that the granulocyte population was increasingly recruited or failed to be cleared in the CCR2-null mice, resulting in a 5-fold increase in these cells (CD45shbshCD11b+Ly6G−) at 3 days after MYXV treatment compared with wild-type animals.

To confirm and visualize the loss of GIMs in the K1492 tumors, we looked at Iba1 staining of K1492-tumors in wild-type or CCR2-null mice (Fig. 2C). We found that the loss of Iba1 staining was prominently from within the glioma, while the population of Iba1+ cells surrounding the glioma remained largely unchanged. This suggested that GIMs were recruited within the tumor while recruited microglia (whose activation and numbers remained unchanged by flow analysis; Fig. 2A) remained at the tumor border in the CCR2-null mice. This staining pattern for glioma-associated microglia and macrophages has been previously observed in a syngeneic rat glioma model (23).

We have previously shown that monitoring the bioluminescence from a luciferase reporter expressed by MYXV (vMyx-FLuc) is a reliable surrogate marker for viral titers in the tumor-bearing mouse brain (4, 11). Using this virus, we found a significant increase in bioluminescent activity in K1492-bearing CCR2-null mice, resulting in 3.3-, 8.1-, and 5.0-fold increase at 1, 2, and 3 days after treatment, respectively (Fig. 3A). The increase in viral bioluminescence at 3 days after treatment corresponded to a 3×10-fold increase in the amount of virus recovered from the tumor-bearing hemisphere at the same time point (Fig. 3B). The large increase in bioluminescent activity was not a result of off-target infection in the knockouts, as nontumor CCR2-null animals had similar clearance kinetics to the wild-type, albeit there was an increase in bioluminescence at 2 days after treatment (Supplementary Fig. S5A) without mouse mortality (Supplementary Fig. S5B). Further, flow cytometric analysis of the vMyx-GFP–treated CCR2-null mice we did not observe any GFP+ immunocytes, suggesting that this enhanced infection was not due to the immunocyte infection. To confirm the increase in viral infection within the tumor, we stained for viral protein M-T7 in the tumor-bearing CCR2-null animals at 3 days after treatment (Fig. 3C) and found large areas of infected tumor associated with large groups of polymorphonuclear cells. This was reminiscent of what we observed in 1-day posttreatment wild-type animals (Fig. 1B), indeed suggesting that viral clearance from the tumor was delayed. These areas of infection were located adjacent to substantial areas of focal necrosis, suggesting that tumor death accompanied this increased infection. The increase in viral activity in the CCR2-null mice resulted in glioma-bearing mice living 30% longer following treatment (13 days posttreatment vs. 17 days posttreatment, P = 0.0073; Fig. 3D).

Finally, as CCR2 loss could potentially inhibit recruitment of other important immunocytes to the gloma after viral infection, we analyzed the NK and T-cell populations recruited after infection. We found that these cells were not inhibited in their recruitment, but tended to be enhanced, significantly in the case of NK cells (Supplementary Fig. S6). Indeed, here we identified the CD45shbshCD11b+ "straddling population" to be NK cells (NK1.1+ in gray), suggesting that the NK cells that fall within CD11b+ gate are also causing an overrepresentation of GIMs in the wild-type and CCR2-deficient animals.

Taken together, the above results show that GIMs inhibit the initial oncolytic virus infection within the glioma. However, as GIMs may either be residing within the tumor before treatment ("tumor-resident" GIMs) or recruited to the tumor after viral administration ("treatment-recruited"), and both types are reduced in CCR-null mice, we next wanted to determine which population mediated viral clearance. To selectively ablate treatment-recruited GIMs, we administered daily intravenous clodronate liposomes to ablate circulating monocytes (37). As clodronate liposomes are unable to cross the blood–brain barrier, (38) treatment would not affect tumor-resident GIMs or microglia while eliminating recruited monocyteid cells. This was confirmed by flow cytometric analysis of clodronate liposomes and vehicle-treated gliomas (Fig. 4A). Furthermore, we found that depletion of these cells resulted in an increase in granulocytes as well as other important immunocytes to the glioma after viral infection, we analyzed the NK and T-cell populations recruited after infection. We found that these cells were not inhibited in their recruitment, but tended to be enhanced, significantly in the case of NK cells (Supplementary Fig. S6). Indeed, here we identified the CD45shbshCD11b+ "straddling population" to be NK cells (NK1.1+ in gray), suggesting that the NK cells that fall within CD11b+ gate are also causing an overrepresentation of GIMs in the wild-type and CCR2-deficient animals.
Collectively, these data suggest that it is the tumor-resident GIMs, and not the treatment-recruited GIMs, that reduce the intratumoral MYXV infection, viral clearance, and efficacy in this immunocompetent malignant glioma model.

We next attempted to target these tumor-resident GIMs pharmacologically using minocycline tetrachloride. Minocycline is a member of the tetracycline family of antibiotics that can readily cross the blood-brain barrier, has anti-glioma activities, and can inhibit the activity of both microglia and monocytes/macrophages (39–41). We adopted a dosing regimen previously shown to inhibit microglia activation in the mouse brain (41), and found that administration of minocycline mimicked the viral clearance kinetics seen in the CCR2-null mice (Supplementary Fig. S7A), with a significant 10-fold increase over control animals at 2 days after treatment. This increase in initial infection, however, was not accompanied by a combinatory survival advantage, nor did minocycline alone have any response in this model (Supplementary Fig. S7B).

![Figure 2. CCR2-deficient mice fail to recruit GIMs to the K1492 tumor before or after Myxoma treatment. Wild-type (WT) or CCR2-deficient (CCR2) C57Bl/6 mice implanted with K1492 cells and analyzed by flow cytometry at 15 days after implantation, 3 days after treatment with Myxoma virus (5 × 10^6 FFU vMyx-GFP) or untreated. A, quantified numbers of each cell type isolated from the tumor-bearing hemisphere (left; n = 6) accompanied by representative CD45/CD11b scatter plot (right). Highlighted in yellow are the Ly6G^+ granulocytes. B, addition of the Ly6G antibody to the flow cytometry analysis demonstrates granulocytic recruitment in MYXV-treated animals and refines the GIM classification of our data (n = 3). Error bars represent SE and asterisks represent significant differences within mouse strain but between treatment groups. Plus signs represent significant differences between mouse strains within treatment groups (P < 0.05); there is no significant difference between WT-K1492 and CCR2-K1492 MYXV for GIMs. C, K1492-bearing CCR2-deficient mice were harvested at 14 days after injection for formalin-fixed paraffin sections (top row, ×25; second row, ×200). Representative staining from three animals per group.](cancerres.aacrjournals.org)
The role of T- and NK cell populations in mediating MYXV efficacy

We were intrigued to discover a role for tumor-resident GIMs in mediating early viral infection kinetics within syngeneic tumors. However, the enhanced viral infection did not persist for more than 3 days, suggesting that other recruited immunocytes may participate in its rapid clearance. Given the recent finding that NK cells mediate the premature clearance of oncolytic HSV from glioma xenografts (28, 29), we next analyzed the extent of NK (CD45highCD3−/−/NK1.1+ and NK1.1+DX5+) and T (CD45highCD3−/−/NK1.1−) cells recruited to K1492 tumors MYXV treatment (Supplementary Fig. S6). We found a resident population of both NK and T cells within the K1492 tumor before MYXV treatment and a significant increase (~2.5-fold) of both cell types after MYXV administration.

To determine the importance of NK, T, and B cells in MYXV clearance, we next used IL2Rg−null transgenic mice, which are devoid of NK cells and have limited numbers of poorly functioning T and B lymphocytes (42). Immunophenotyping of these severely immunocompromised mice bearing K1492 before MYXV infection showed, as expected, that they retained the tumor-resident GIMs and microglia, but were unable to recruit any subsequent CD45high leukocytes 3 days after treatment (Supplementary Fig. S8).

Measuring the rate of viral clearance in the K1492-bearing IL2Rγ-null animals, we found persistent bioluminescent activity that did not dissipate from the mouse brain and remained until the mice were sacrificed because of tumor burden (Fig. 5A). This signal did not significantly change from the original measurement at 1 day after treatment, suggesting that the virus was unable to be cleared but was not effectively amplifying within the tumor. Non–tumor-bearing animals also had a persistent bioluminescent signal, but it was much smaller than that of tumor-bearing animals (Supplementary Fig. S9A). We confirmed persistent infection with the tumor by staining for viral protein M-T7, and found...
Evidence of viral infection within the tumor 7 days after infection (Fig. 5C). The staining pattern of viral protein was interesting, with large areas of viral infection localized to the tumor margins and scattered cells expressing M-T7 within the tumor core. This persistent infection translated into significantly longer survival times, with glioma-bearing IL2Rg-null animals living >50% longer on average after MYXV treatment (9.5 days vs. 14.5 days after MYXV infection; \( P = 0.0002 \)) as compared with K1492-bearing wild-type mice. Nontumor-bearing animals did not succumb to viral treatment (Supplementary Fig. S9B).

We were intrigued by the persistent MYXV infection we observed in these IL2Rg-null animals, and wanted to determine the potential separate roles of the T- and NK cell populations. To test this, we utilized K1492 implanted into T- and B-cell–deficient RAG1 animals (who have intact NK function), and further treated these mice with the NK cell–depleting NK1.1 antibody. We found that a single intraperitoneal injection of this antibody depleted splenic NK cells by >80% in wild-type mice (data not shown). Interestingly, the NK1.1 treatment in wild-type animals had very little effect on viral clearance kinetics (Fig. 6A), and we did not find a survival advantage to NK cell depletion in this model (Fig. 6B). We found that treating glioma-bearing RAG1 animals with MYXV-FLU C resulted in a persistent bioluminescent signal that was statistically increased over wild-type beginning at 5 days after treatment (Fig. 6C); however, this sustained viral infection did not result in a survival advantage (Fig. 6D). Finally, using a combination of both B- and T-cell depletion (RAG1 animals) and NK cell ablation (NK1.1), we found a persistent infection that was significantly increased from day 3 through 10 over the isotype control in RAG1 animals (Fig. 6E). This elevated and persistent infection produced a significant 30% survival increase compared with the isotype-treated controls (18 vs. 24 days after infection, \( P = 0.0342 \); Fig. 6F).

**Figure 4.** Clodronate liposome depletion of peripheral monocytes stops the recruitment of GIMs following Myxoma treatment but has little effect on viral clearance and treatment efficacy. Two-hundred microliters of clodronate liposomes (CL) or vehicle (Veh) was given via tail vein injection at −1, 0, 1 and 2 days after MYXV infection (5 \( \times 10^7 \) FFU, vMyx-GFP). A, quantified numbers of cells analyzed by flow cytometric analysis of GIM and granulocytes in clodronate liposomes or vehicle (Veh) depleted animals (\( n = 4 \)) accompanied by a scatter plot (right) indicating cell populations. Yellow, Ly6G+ cells. B, analysis of viral bioluminescence from vMyx-FLuc (5 \( \times 10^5 \) FFU) in clodronate liposomes (CL)- or vehicle treated animals (\( n = 7/group \)). C, viral recovery assay from vehicle or clodronate liposome–treated mice bearing 14 day K1492 tumors and infected with 1.0 \( \times 10^7 \) FFU vMyx-FLuc 3 days after treatment. Error bars represent SE and asterisk represents significant difference (\( n = 3/group; P < 0.05 \)). D, Kaplan–Meier survival curve of clodronate liposomes or vehicle–treated animals bearing K1492 tumors and treated on day 14 with vMyx-FLuc (5 \( \times 10^7/FFU \)); dpTx, days after treatment.
antibody treatment in either wild-type or RAG1 animals without virotherapy had no change in overall survival (data not shown). These data suggest that ablation of both NK and T cells within the tumor are required to significantly enhance persistence of intratumoral viral replication and to significantly prolong survival in immunocompetent malignant glioma animal model.

Finally, we wanted to investigate whether we could mimic the immunosuppressed state of the IL2RY-null animals in wild-type animals using clinically relevant immunosuppressants. The alkylating agent cyclophosphamide was the logical choice, as it is a clinically utilized antitumor therapeutic, has known lymphoablative properties, and has previously been shown to enhance virotherapy effectiveness in wild-type or IL2RY-K1492-bearing mice treated with Myxoma virus (5 \times 10^6 FFU) in syngeneic C57Bl/6 mice 7 days after treatment with Myxoma virus (5 \times 10^6 FFU) and analyzed by flow cytometry (n = 6). Asterisks represent significant differences between mouse strain but between treatment groups. Plus signs represent significant differences between mouse strains within treatment groups. B, measurement of bioluminescence emitted from wild-type or IL2RY-K1492-bearing mice treated with vMyx-FLUC (5 \times 10^6 FFUs; n = 9 WT, n = 8 IL2RY) 14 days after implantation. C, M-7e staining of K1492-bearing IL2RY mice 7 days after treatment with vMyx-FLUC (5 \times 10^6 FFU). Top panel, \times 4 objective; middle and lower panel, \times 10 objective. D, Kaplan-Meier survival curve of K1492-bearing IL2RY mice treated vMyx-FLUC (5 \times 10^6 FFU) 14 days after tumor implantation. All error bars represent SE. dpTx, days after treatment.

Figure 5. IL2RY-deficient mice have a persistent Myxoma virus infection within the tumor that results in a significant increase in treatment efficacy. Wild-type (WT) or IL2RY-deficient (IL2RY) C57Bl/6 mice implanted with K1492 cells and analyzed by flow cytometry at 15 days after implantation, 3 days after treatment with Myxoma virus (5 \times 10^6 FFU) or untreated. A, quantified numbers of NK cells isolated from the tumor-bearing hemisphere and analyzed by flow cytometry (n = 6). Asterisks represent significant differences within mouse strain but between treatment groups. Plus signs represent significant differences between mouse strains within treatment groups. B, measurement of bioluminescence emitted from wild-type or IL2RY-K1492-bearing mice treated with vMyx-FLUC (5 \times 10^6 FFUs; n = 9 WT, n = 8 IL2RY) 14 days after implantation. C, M-7e staining of K1492-bearing IL2RY mice 7 days after treatment with vMyx-FLUC (5 \times 10^6 FFU). Top panel, \times 4 objective; middle and lower panel, \times 10 objective. D, Kaplan-Meier survival curve of K1492-bearing IL2RY mice treated vMyx-FLUC (5 \times 10^6 FFU) 14 days after tumor implantation. All error bars represent SE. dpTx, days after treatment.

Figure 5. IL2RY-deficient mice have a persistent Myxoma virus infection within the tumor that results in a significant increase in treatment efficacy. Wild-type (WT) or IL2RY-deficient (IL2RY) C57Bl/6 mice implanted with K1492 cells and analyzed by flow cytometry at 15 days after implantation, 3 days after treatment with Myxoma virus (5 \times 10^6 FFU) or untreated. A, quantified numbers of NK cells isolated from the tumor-bearing hemisphere and analyzed by flow cytometry (n = 6). Asterisks represent significant differences within mouse strain but between treatment groups. Plus signs represent significant differences between mouse strains within treatment groups. B, measurement of bioluminescence emitted from wild-type or IL2RY-K1492-bearing mice treated with vMyx-FLUC (5 \times 10^6 FFUs; n = 9 WT, n = 8 IL2RY) 14 days after implantation. C, M-7e staining of K1492-bearing IL2RY mice 7 days after treatment with vMyx-FLUC (5 \times 10^6 FFU). Top panel, \times 4 objective; middle and lower panel, \times 10 objective. D, Kaplan-Meier survival curve of K1492-bearing IL2RY mice treated vMyx-FLUC (5 \times 10^6 FFU) 14 days after tumor implantation. All error bars represent SE. dpTx, days after treatment.

antibody treatment in either wild-type or RAG1 animals without virotherapy had no change in overall survival (data not shown). These data suggest that ablation of both NK and T cells within the tumor are required to significantly enhance persistence of intratumoral viral replication and to significantly prolong survival in immunocompetent malignant glioma animal model.

Finally, we wanted to investigate whether we could mimic the immunosuppressed state of the IL2RY-null animals in wild-type animals using clinically relevant immunosuppressants. The alkylating agent cyclophosphamide was the logical choice, as it is a clinically utilized antitumor therapeutic, has known lymphoablatative properties, and has previously been shown to enhance virotherapy effectiveness in glioma models using a single low-dose (<100 mg/Kg) treatment before virotherapy. Initial studies in our model did not find any significant change in viral infection kinetics or treatment efficacy with single-dose cyclophosphamide at 80 mg/kg (not shown) or 150 mg/kg (Supplementary Fig. S10A). However, repeated doses of 150 mg/kg at −2, 0, and +2 days after treatment resulted in significantly sustained viral infection (Fig. 7A). This dosing schedule itself resulted in significant survival benefit (17 days post-treatment no treatment vs. 25 days posttreatment cyclophosphamide; P = 0.0045); however, the combination of cyclophosphamide and MYXV was significantly more beneficial than either treatment alone (cyclophosphamide vs. MYXV + cyclophosphamide P = 0.0037). Immunohistochemistry for MYXV protein MT7 found considerable staining within the tumors at 7 days after treatment (Fig. 7C), with sizeable necrotic areas surrounding the entire tumor margin.

We analyzed the degree of immunosuppression seen through our dosing regimen by immunophenotyping the K1492 glioma microenvironment after combined MYXV and cyclophosphamide treatment (Fig. 7D; Supplementary Fig. S10C). Cyclophosphamide treatment resulted in a complete failure to recruit any peripheral immunocytes to the tumor, suggesting strong peripheral immunosuppression. Surprisingly, however, we found that...
cyclophosphamide treatment also resulted in the near complete abolition of tumor resident GIMs while having no effect on resident microglial populations. These results demonstrate that lymphoablation in combination with MYXV treatment is a viable therapeutic strategy in high-grade gliomas.

Discussion

The human glioma microenvironment is complex, with tumor cell interactions with other glia, extracellular matrix proteins, vasculature, and infiltrating immune cells. Although all of these interactions could, and perhaps do, play some role in determining oncolytic virus therapy efficacy in patients and animal models, interrogating tumor-infiltrating immunocyte populations is a logical approach to investigate potential avenues to improve therapy.

The human glioma-infiltrating immunocyte population is diverse, including an array of immunocytes. However, the most numerous infiltrating immune cells in human glioma are microglia and monocytes/macrophages (25–27), with some reports suggesting they comprise up to one-third of the tumor mass (27). The K1492 syngeneic tumors used in these studies have a similar composition, with substantial infiltration of microglia and recruited peripheral monocytes/macrophages (GIMs), making up >90% of the total immune infiltrate in the K1492 model.

We demonstrate here that GIMs are recruited to the K1492 tumors in a CCR2-dependent manner. Although monocyte recruitment to the CNS through CCR2 has been shown in other CNS pathologies, (34–36) this is the first time that this has been demonstrated in a mouse model of glioma. This could potentially be an important model for testing the role of peripherally derived GIMs in glioma treatment and biology.

In our study, CCR2-deficient mice, but not clodronate liposome–depleted mice, had large increases in tumoral infection over the first 3 days of infection. These data suggest tumor-resident GIMs play the predominate role early in oncolytic infection. Targeting tumor resident as opposed to peripherally recruited monocytes to mediate oncolytic infection rates could pose a challenge clinically. Here we have introduced the idea that minocycline tetrachloride may be able to inhibit these to enhance oncolytic infection. It is important to note that the mechanisms by which minocycline enhances the infection could be diverse, and may not in fact be due to the inhibition of microglia and/or GIMs. Minocycline has been shown to have several anti-inflammatory effects such as the inhibition of...
NFkB and iNOS, as well as antitumor activities such as inhibiting MMPs, angiogenesis, and directly killing of glioma cells (39–41). Credence to the perspective that minocycline could play an important role in glioma treatment is provided by the recently commenced RAMBO trial that has added minocycline to bevacizumab and radiation treatment for recurrent glioma (NCT01580969). Further studies investigating the combinatorial effects of minocycline and oncolytic virus for glioma (NCT01580969). This phenomenon should be considered in future studies examining neuroinflammatory responses by flow cytometry. Furthermore, our studies here fail to address the role that these infiltrating cells may also be playing in altering this inflammatory response by flow cytometry. A surprising result in our study was the loss of microglia in response to MYXV infection. We considered that this could be due to a shift in microglial CD45 expression from CD45<sup>low</sup> to CD45<sup>high</sup>; however, our experiments that ablated much of the infiltrating CD45<sup>high</sup>CD11b<sup>+</sup> populations still resulted in this microglial loss. Iba1 staining after viral infection did not indicate a large loss of these microglia, albeit this stain would not distinguish dead or dying cells. It is our current opinion that these cells were lost during isolation of CNS leukocytes, perhaps as a result of changes in cellular density following viral infection or perhaps as a result of increase in fragility after the inflammatory insult. Figure 7. Repeated low-dose cyclophosphamide treatment results in prolonged tumor infection, resulting in significant combined efficacy. This dosing resulted in lymphoblastation of tumor-resident and treatment-recruited immunocytes. Wild-type animals bearing K1492 gliomas were treated with 150 mg/kg of cyclophosphamide 2 days before, the day of, and 2 days after (−2, 0, and +2 days after treatment; dpTx) intracranial administration of 5 × 10<sup>6</sup> FFU of vMyx-FLuc. A, analysis of viral bioluminescence from vMyx-FLuc (MYXV, n = 12 at 1–7 days after treatment and n = 8 at 10 days after treatment; MYXV + cyclophosphamide, n = 11 at 1–7 days after treatment and n = 7 at 10 days after treatment). B, Kaplan–Meier survival curve of nontreated (NoTx, n = 5), cyclophosphamide-treated (cyclophosphamide, n = 5), vMyx-FLuc-treated (vMyx, n = 9), and cyclophosphamide and vMyx-FLuc-treated (MYXV + cyclophosphamide, n = 8). C, immunohistochemical staining with H&E or viral protein M-T7 in cyclophosphamide-treated, K1492-bearing animals 3 days postinfection with 5 × 10<sup>6</sup> FFUs of MYXV. D, quantified numbers of each cell type isolated from the tumor-bearing hemisphere and analyzed by flow cytometry (n = 3). Asterisks represent significantly different no treatment (NoTx) control and plus sign represents significantly different from MYXV treatment. All error bars represent SE.
bioluminescence kinetics; however, this could also be a function of the return of NK cells after the initial depletion.

The role of NK cells alone in mediating viral clearance in our syngeneic model does not appear to have the same impact as has been reported in the literature for immunocompromised xenograft models (28, 29). However, in RAG1 immunocompromised animals, NK depletion results in a large increase in viral infection in the tumor, increasing viral persistence and resulting in a significant treatment-induced survival response. This suggests that the NK and T-cell populations have overlapping functions in our model. It is interesting to speculate on the nature of the T cells that are recruited so early after infection. Most studies interrogating viral-induced T-cell responses in the CNS interrogate classical T-cell responses, beginning 7 days or later postinfection. However, the prompt recruitment seen here suggests these T cells are playing an innate-like role clearing the infection. Nonclassical T-cell such as natural killer T cells (NKT) and gamma-delta T cells (γδT) would be good candidates to investigate further. Both NKT and γδT cells have known innate antiviral effector functions, are absent in RAG1-null animals, and are known to be swiftly recruited during CNS inflammation (44–46). These innate-like T cells are similar in that they have a limited, predetermined T-cell receptor diversity that is capable of rapid effector function in response to defined foreign- or self-antigens. Furthermore, they both contain NK cytotoxicity receptors, such as the NK2D receptor, that upon ligation of their ligands can instigate cytotoxic responses against stressed or infected cells (47). It was recently shown that NK cell–mediated clearance in orthotopic glioma xenografts was dependent on NK cell natural cytotoxicity receptors Nkp30 and Nkp46 that recognized oncolytic HSV-infected glioma cells (29). It would be very interesting to investigate the roles of NKT and γδT cells in our model and to see whether these cells were functioning in an NK-like fashion, perhaps dependent on virally induced NKG2D ligand expression in MYXV-infected glioma cells.

Taken together, the results of these experiments demonstrate that resistance to Myxoma virus infection and replication in syngeneic animals is, not surprisingly, a multifaceted mechanism. The diverse nature of the mechanisms that are limiting this therapy in our model suggests that pan-inhibitors of the immune system could be the most efficacious in enhancing oncolytic virus therapeutic responses, as we and others have shown with cyclophosphamide (14–16, 22). Interestingly, in addition to peripheral lymphocyte ablation, our study here demonstrated that cyclophosphomimide treatment was able to ablate GIMs in our model. This was an unexpected observation, as the cytotoxic activity of bioactive cyclophosphamide (phosphoramid mustard) is through DNA alkylation, thus targeting actively dividing cells. We do not anticipate that these GIMs are turning over so quickly from the circulating population, as we did not see ablation of these GIMs with clodronate liposome–mediated monocye depletion. It will be interesting to interrogate the loss of these GIMs with this cyclophosphamide dosing regimen, and to determine whether this is a viable strategy for depleting GIMs in other models. Furthermore, based on our transgenic animal experiments, we could predict that this is what the immune cell composition would look like in a MYXV-treated CCR2/IL2Rγ double knockout mouse. It is curious then why we did not achieve a initial increase in viral infection following cyclophosphamide treatment as observed in the CCR2-null mice; however, it is possible that the dose 2 days before treatment was insufficient to deplete the GIM population, and that subsequent doses were necessary. If this were the case, it could postulated that starting cyclophosphamide even earlier before oncolytic virus administration could result in even greater tumoral infection and combinatorial efficacy.

It is important to note that maximizing oncolytic virus clinical utility exclusively through the enhancement of viral infection and virus-mediated cell death ignores the potential of this therapy to stimulate antitumor immune responses. Indeed, numerous oncolytic viruses have been genetically modified to enhance antitumor immune responses, making oncolytic viruses akin to an immunotherapy (48). We demonstrate here that for MYXV to develop an oncolytic treatment response, a threshold of infection must be reached through the inhibition of innate immune responses. Achieving this through genetic lymphoablative precludes any cellular immunotherapeutic responses (and is clinically irrelevant); however, transient lymphoablative or immune-inhibiting therapeutics may allow sufficient tumoral infection and cell death to induce a robust multifaceted antitumor immune response. It is important to consider that most cancers are tremendously heterogeneous, including gliomas (49), and maximizing the viral production of tumor antigen by maximizing initial oncolytic virus infection rates could help initiate a broader immunotherapeutic response. This, however, is a double-edged sword, as initial innate immune responses are what orchestrate subsequent adaptive responses. Thus, a balance must be achieved to allow sufficient viral infection and antigen generation, while not inhibiting the generation of appropriate adaptive responses. It is not yet clear whether MYXV is capable of generating an adaptive immunotherapeutic response in gliomas, as it has been shown in other cancer models (31). Additional investigation probing the nature of the glioma microenvironment before and after virotherapy, as well as the specific mechanisms by which the innate immune response limits MYXV infection rates, could lead to a more targeted approach to enhance viral infection while initiating antitumor immunity.

Disclosure of Potential Conflicts of Interest

G. McFadden has ownership interest (including patents) in Viron Therapeutics. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions

Conception and design: F.J. Zemp, X. Lun, V.W. Yong, P.A. Forsyth Development of methodology: F.J. Zemp, X. Lun, P.A. Forsyth Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): F.J. Zemp, B.A. McKenzie, K.M. Reilly, P.A. Forsyth Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): F.J. Zemp, G. McFadden, V.W. Yong, P.A. Forsyth Writing, review, and/or revision of the manuscript: F.J. Zemp, B.A. McKenzie, X. Lun, V.W. Yong, P.A. Forsyth Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): V.W. Yong, P.A. Forsyth Study supervision: X. Lun, P.A. Forsyth
Acknowledgments

The authors thank Dr. Douglas Mahoney and Dr. Jennifer Chan for critical reading of the manuscript and/or guidance throughout these experiments. The authors also thank Molly Chisholm and staff in the Mouse Double Barrier Unit at the University of Calgary (Calgary, Alberta, Canada).

Grant Support

This work was funded by the Canadian Cancer Society Research Institute (CCSRI) with funds raised by the Canadian Cancer Society (P.A. Forsyth), a Program Project Grant from the Terry Fox Foundation (P.A. Forsyth), the V Foundation (P.A. Forsyth), and the Canadian Institutes of Health Research (CIHR; W.V. Yong). During this work, F.J. Zemp was funded by Alberta Innovates Health Solutions, Vanier Scholarship from CIHR, and the Isaac Walton Killam scholarship. P.A. McKenzie was funded by CIHR. G. McFadden’s laboratory is funded by NIH grants R01 AI086607 and R01 CA136541. K.M. Reilly’s laboratory is funded by the Intramural Research Program of NIH, NCI (ZIA BC 010541).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received March 25, 2014; revised September 17, 2014; accepted October 2, 2014; published OnlineFirst October 21, 2014.

References

34. Huang DR, Wang J, Kivisakk P, Rollins BJ, Ransohoff RM. Absence of monocyte chemotactic protein 1 in mice leads to decreased local


Cellular Factors Promoting Resistance to Effective Treatment of Glioma with Oncolytic Myxoma Virus

Franz J. Zemp, Brienne A. McKenzie, Xueqing Lun, et al.


Updated version
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-14-0876

Supplementary Material
Access the most recent supplemental material at:
http://cancerres.aacrjournals.org/content/suppl/2014/10/21/0008-5472.CAN-14-0876.DC1

Cited articles
This article cites 46 articles, 17 of which you can access for free at:
http://cancerres.aacrjournals.org/content/74/24/7260.full#ref-list-1

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, use this link http://cancerres.aacrjournals.org/content/74/24/7260. Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.