HIC1 Silencing in Triple-Negative Breast Cancer Drives Progression through Misregulation of LCN2

Guangcun Cheng¹, Xueqing Sun¹, Jinglong Wang¹, Gang Xiao¹, Xiumin Wang¹, Xuemei Fan¹, Lidong Zu¹, Mingang Hao¹, Qing Qu², Yan Mao², Yunjing Xue¹, and Jianhua Wang¹

Abstract
The tumor suppressor gene HIC1 is frequently deleted or epigenetically silenced in human cancer, where its restoration may improve cancer prognosis. Here, we report results illuminating how HIC1 silencing alters effect or signals in triple-negative breast cancer (TNBC), which are crucial for its pathogenesis. HIC1 expression was silenced only in TNBC compared with other molecular subtypes of breast cancer. Restoring HIC1 expression in TNBC cells reduced cell migration, invasion, and metastasis, whereas RNAi-mediated silencing of HIC1 in untransformed human breast cells increased their invasive capabilities. Mechanistic investigations identified the small-secreted protein lipocalin-2 (LCN2), as a critical downstream target of HIC1 in TNBC cells. Elevating LCN2 expression in cells expressing HIC1 partially rescued its suppression of cell invasion and metastasis. Notably, autocrine secretion of LCN2 induced by loss of HIC1 activated the AKT pathway through the neutrophil gelatinase–associated lipocalin receptor, which is associated with TNBC progression. Taken together, our findings revealed that the HIC1–LCN2 axis may serve as a subtype-specific prognostic biomarker, providing an appealing candidate target for TNBC therapy. Cancer Res; 74(3); 862–72. © 2013 AACR.

Introduction
Triple-negative breast cancer (TNBC) consists of a diverse and heterogeneous group of tumors that by definition lack estrogen and progesterone receptors and amplification of the HER2 gene (1). Although systematic therapeutic approaches have reduced cancer-specific mortality, TNBC remains associated with high rates of cancer recurrence and metastasis, which makes the development of effective treatments difficult (2). Therefore, identifying new biomarkers of TNBC progression may be of value for the prevention of metastasis and the design of effective therapeutic strategies for this disease.

Hypermethylated in cancer 1 (HIC1) is a tumor suppressor gene that is frequently epigenetically silenced, including in prostate, liver, colorectal, lung, and breast cancer (3–5). Recent studies showed that HIC1 promoter hypermethylation is not only present in solid tumors but also in normal breast ductal (6), brain (7), and prostate epithelium tissues (3). These findings indicate that other inhibitory mechanisms besides hypermethylation may exist, such as the modification of HIC1 by SUMOylation or acetylation or mutation/inactivation of the positive HIC1 regulator p53 (8, 9).

HIC1 encodes a sequence-specific transcriptional represor with an N-terminal POZ domain and 5 Krippe like C2H2 zinc fingers at the C-terminal that can bind a specific motif consisting of a 5′-(G/C)GGGCA(C/A)CC-3′ sequence centered on GGCA (reverse 5′-TGCCC/A-3′) named HiRE (HIC1 Responsive Element; refs. 10 and 11). The upstream regulation of HIC1 is mediated by p53 through a regulatory feedback loop in which HIC1 directly represses the transcription of SIRT1, which deacetylates and thereby inactivates p53 (11). Recently, downstream target genes of HIC1 responsible for developmental and cell-cycle control have been identified, including the histone deacetylase SIRT1 (11), the transcription factor ATOH1 (12), the G-protein–coupled receptor CXCR7 (13, 14), Cyclin D1, P57KIP2 (CDKN1C; ref. 9), and ephrin-A1, a cell surface ligand for Eph tyrosine kinase receptors (15). However, given the many potential physiologic roles of HIC1, few HIC1 target genes have been characterized.

In this study, we identified Lipocalin2 (LCN2, also known as NGAL) as a direct target gene of HIC1 using genome expression profiling. LCN2 is a small-secreted glycoprotein involved in multiple processes, such as immunity, renal development, and apoptosis (16, 17). Pathologically, LCN2 is associated with multiple types of human tumors, including breast cancer (16). Orthotopic studies showed that LCN2-expressing breast tumors display a poorly differentiated phenotype (18, 19) and LCN2 expression is a predictor of poor prognosis in human primary breast cancer (20). However, the mechanisms underlying the role of LCN2 in different subtypes of breast cancer remain controversial (18, 21).
The identification of basal-like tumors by immunohis-
tochemistry remains challenging in clinical practice. Because
TNBCs and basal-like breast cancers share many molecular
and morphologic features, the TNBC phenotype could reliably
be used as a surrogate for basal-like breast cancer (22).

Materials and Methods

Cell cultures

The human breast cancer cell lines MCF-7, MDA-231, MDA-
468, BT474, and SK-BR-3, and the lung cancer cell line A549
were obtained from the American Type Culture Collection
(ATCC) and cultured according to the online instructions of
the manufacturer. The immortalized epithelial cell line HBL-
100 (Cell Bank of Shanghai Institute for Biological Sciences,
Shanghai, China) was maintained in Dulbecco’s Modified Eagle
Medium (DMEM; HyClone) and MCF-10A cells (ATCC) were
maintained in DMEM/F12 (Invitrogen) supplemented with 5%
horse serum, EGF (20 ng/mL), hydrocortisone (0.5 mg/mL),
cholera toxin (100 ng/mL), insulin (10 μg/mL), and 1% pen-
icillin/streptomycin. The cell lines were tested and authen-
ticated by DNA typing in the Shanghai Jiao Tong University
Analysis Core (last test in April 2013).

Western blot analysis

Details are provided in the Supplementary Methods.

Tissue microarray and immunohistochemistry

The immunohistochemical analysis was performed using the
avidin–biotin–peroxidase complex method with an anti-
HIC1 antibody. Details are provided in the Supplementary
Methods.

Construction of lentiviral vectors

To restore the expression of HIC1 in TNBC cells, human full-
length HIC1 cDNA was inserted into the lentivirus vector pHR-
SIN-CIGW. Lentivirus cells were then transduced with the PMD2.
G, PSPA2X, and HIC1 expression vectors using Lipofectamine
2000 (Invitrogen). After 48 hours, culture supernatants were
collected, passed through 0.45 μm filters, and mixed with fresh
media (1:1) and polybrene (8 μg/mL) to infect target cells. Cells
with restored expression of HIC1 were designated as MDA-
231HIC1 and MDA-468HIC1, and the respective controls were
designated as MDA-231GFP and MDA-468GFP. GV248 lentiviral
vectors were used to generate HIC1 knockdown cell lines.
Details are provided in the Supplementary Methods.

Migration and invasion assays

A wound healing assay was used to assess cell migration
ability and cell invasion was examined using a reconstituted
evadacellular matrix membrane (cat. no. 354483; Becton Dick-
inson). The procedures are described in the Supplementary
Methods.

siRNA transfection

siRNA transfection was performed with 50 nmol/L of each
siRNA (GenePharma) using Lipofectamine 2000 according to
the manufacturer’s instructions. The siRNA sequences are
provided in the Supplementary Methods.

Clinical dataset analysis

The classification included in the dataset (Sorlie classifi-
cation method) was used to assign patients to the different
clinical breast cancer subtypes. For survival analyses, overall
survival (OS) and distant metastasis-free survivals (DMFS),
stratified by expression of the gene of interest, were presented
as Kaplan–Meier plots and tested for significance using log-
rank tests. The analysis was performed according to the

cDNA microarray analysis

The Agilent SurePrint G3 Human Gene Expression micro-
array (8 × 60K) was used in this study. Microarray hybrid-
ization, data collection, and analysis were performed at
Oebiotech Biotechnology Corporation according to Agilent
protocols. Detailed procedures are provided in the Supplemen-
tary Methods.

Luciferase reporter assay and chromatin
immunoprecipitation

The procedures are described in the Supplementary
Methods.

In vivo metastasis assay

Luciferase tagged MDA-231 cells (1 × 10⁶) were injected into
the tail vein of BALB/c nude mice. Tumor growth and metas-
tasis were examined using the Xenogen IVIS Imaging System.
All mouse experiments were performed in an animal center
after obtaining approval from the Shanghai Medical Experi-
mental Animal Care Commission. Details are provided in the
Supplementary Methods.

Statistical analyses

Data are presented as mean ± SD or mean ± SEM. Values of
P < 0.05 were considered statistically significant. The results of
quantitative real-time PCR were analyzed using the Student t
test, 2-tailed. HIC1 staining in tissue microarrays was analyzed
by Pearson χ² test. All analyses were performed with SPSS 11.5.

Results

Expression of HIC1 is silenced in TNBC

Increasing evidence indicates that HIC1 is silenced by
epigenetic modifications in many types of prevalent human
tumors (11–14, 24). We and other groups recently described
the role of HIC1 in tumor progression as partially relying on a
variety of downstream targets (11–14, 24). However, the role
and regulatory mechanism of HIC1 in different molecular
subtypes of breast cancer remain unclear.

We first showed that the expression of HIC1 was weaker in
the TNBC cell line MDA-231 than in luminal-type MCF-7 and BT474
cells, HER2-positive SK-BR-3 cells, and in the normal epithelial
cell line MCF-10A (Fig. 1A). Next, a high-density tissue micro-
array was stained with an anti-human HIC1 antibody. Repre-
sentative images indicated that expression of HIC1 in TNBC
tissues was almost absent as compared with benign, luminal-
type, and HER2-positive tissues (Fig. 1B). Quantitative analysis
confirmed these findings, as shown in Supplementary Table S1.
However, comparison of benign and malignant breast diseases showed no statistically significant differences \( (P = 0.230) \). HIC1 expression was significantly correlated with histologic grade, and estrogen receptor (ER), progesterone receptor (PR), and HER2 levels in breast cancer tissues (Supplementary Table S1).

To further explore the relationship between HIC1 and the clinical prognosis, we evaluated the prognostic value of HIC1 in a large public clinical microarray database of breast cancer that includes data from 1,027 patients (23, 25). The results showed that decreased HIC1 expression was associated with poor prognosis only among patients with basal-like breast cancer (Fig. 1D), but not in the overall breast cancer group (Fig. 1C). Overall, these findings indicate that expression of HIC1 is silenced in TNBC and that it may be associated with the subtype-specific prognoses of breast cancer.

**Effects of restoration of HIC1 expression in TNBC cells**

A critical feature of TNBC is its highly aggressive behavior (26, 27). Based on our findings that HIC1 expression is silenced in TNBC cells, its expression was restored in MDA-231 and MDA-468 cells using a lentivirus vector. Figure 2A and Supplementary Fig. 1A show the restored HIC1 expression in TNBC cells, its expression was restored in MDA-231 and MDA-468 cells using a lentivirus vector. Figure 2A and Supplementary Fig. 1A show the restored HIC1 expression in TNBC cells, its expression was restored in MDA-231 and MDA-468 cells using a lentivirus vector. Supplementary Fig. 1A show the restored HIC1 expression in TNBC cells, its expression was restored in MDA-231 and MDA-468 cells using a lentivirus vector.

To identify potential downstream targets of HIC1, we analyzed the genome-wide transcriptome profile of MDA-231<sup>HIC1</sup> cells and HBL-100<sup>HIC1</sup> cells using Agilent Whole Human Genome Microarrays. The microarray dataset has been deposited in the GEO database (http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE50061).

Fold-change \( (\times 1.5) \) screening between cells with positive HIC1 expression and their respective controls identified 1,364 common genes in MDA-231 and HBL-100 cells and 557 downregulated genes (Fig. 3A). Among the differentially expressed genes, 39 cancer-associated genes were identified that might participate in invasion or metastasis for cluster mapping on the MeV microarray analysis platform (www.tm4.org/mev. html; Fig. 3C).

Among the genes markedly regulated in MDA-231 and HBL-100 cells, we focused on LCN2 based on our previous microarray
data in PC3 and C4-2B cells (Fig. 3B; ref. 14). Because the regulatory region of the LCN2 promoter might contain several functional HIC1 consensus binding sites (HiRE, HIC1 responsive element; refs. 10 and 11), we next explored the mechanisms by which LCN2 expression is modulated by HIC1 in these cell lines. Real-time PCR and ELISA confirmed that HIC1 downregulated the expression of LCN2 at the mRNA and protein levels, consistent with the above microarray assays (Fig. 3D). Similar results were obtained in the lung cancer cell line A549 (Fig. 3D). By contrast, shRNAs knockdown of HIC1 in HBL-100 cells significantly enhanced LCN2 expression at the mRNA and protein levels (Fig. 3E). These findings suggest that the expression of LCN2 is modulated by HIC1.

**LCN2 is a direct target gene of HIC1**

To determine whether LCN2 is a direct target gene of HIC1, we firstly constructed a series of LCN2-truncated promoter/reporter fusion plasmids containing progressive 5’ deletions from −2955 to +18, based on data showing that many putative HIC1 binding elements centered on TGCCC/A are observed on its promoter (10, 11), which allowed us to perform luciferase promoter–reporter assays (Fig. 4A). These constructs were then transfected alone or with the pcDNA3-FLAG-HIC1 expression vector into MDA-231, MCF-7, and SK-BR-3 cells to measure promoter activities in the absence or presence of HIC1. The data showed that the construct containing the LCN2 promoter showed more than 3-fold higher activity than the basic group (Fig. 4B). However, transient transfection of MDA-231 cells with HIC1 markedly inhibited LCN2 promoter activity in a dose-dependent manner (Fig. 4B and C). A similar inhibitory effect was observed in MCF-7 and SK-BR-3 cells (Fig. 4B and C). The effect of the HIC1 expression vector on the inhibition of LCN2 promoter activity in MDA-231 and MCF-7 cells was maintained in the −1427/+18, −774/+18, and...
–252/+18 truncated constructs, but lost in the smaller construct (–62/+18; Fig. 4D). These results suggest that the regulatory region primarily involved in the HIC1-mediated repression of LCN2 is located in the –252/+18 upstream region of the promoter, which contains 3 putative HIC1 binding sites, namely M1, M2, and M3 (Fig. 4E).

To verify this possibility, these 3 putative binding sites were deleted to abolish HIC1 binding. As shown in Fig. 4F, the D_M2 mutated construct significantly decreased the inhibitory effect of HIC1 as compared with the control, whereas the D_M1 and D_M3 constructs did not affect its function. These results suggest that the putative M2 site in the LCN2 promoter is essential for HIC1-mediated repression, whereas the M1 and M3 sites may have little effect. To confirm this, LCN2 expression was downregulated by restoring HIC1 at the mRNA and protein levels in MDA-231 cells (top), and similar results were obtained in the lung cancer A549 cell line (bottom). Three independent experiments were performed. E, inactivation of endogenous HIC1 resulted in LCN2 upregulation at the mRNA (left) and protein levels (right).

Taken together, these results demonstrate that endogenous HIC1 proteins can be directly recruited onto the LCN2 promoter to repress its expression, whereas the silenced HIC1 expression in TNBC cells may facilitate the escape of LCN2 from the suppressive modulation.

**LCN2 partially rescues HIC1-induced phenotypes in TNBC cells**

To further demonstrate that LCN2 is a critical target gene involved in the HIC1-induced phenotypes of TNBC cells, we firstly examined the effect of LCN2 on cell invasion in vitro. Exogenous LCN2 significantly promoted invasion in MDA-231 cells compared with the control (Fig. 5A); however, in MDA-231HIC1 cells, LCN2 partially rescued the reduced invasion caused by HIC1 re-expression.

Next, the invasive ability of MDA-231 cells was examined after restoring LCN2 expression. Effective restoration of HIC1 and LCN2 expression in luciferase-tagged MDA-231 cells was confirmed by Western blotting and ELISA (1#, 2#, 3#, and 4# in Fig. 5B, left). LCN2 overexpression in MDA-231HIC1 cells markedly increased invasion compared with the control (Fig. 5C, 2# and 1#). However, coexpression of LCN2 in MDA-231HIC1 cells
partially rescued the HIC1-induced suppression of invasion (Fig. 5C, 4# and 3#). To further explore whether LCN2 is involved in modulating tumor metastasis in vivo, luciferase-tagged MDA-231 cells (1#, 3#, and 4#) were transplanted into nude BALB/c mice by tail vein injection. The results showed that lung metastasis was significantly reduced in MDA-231HIC1 cells (3#) compared with the control group (1#; Fig. 5D), consistent with the in vivo results shown in Fig. 2F–H. However, coexpression of LCN2 in MDA-231 HIC1 cells (4#) partially rescued the HIC1-induced reduction of lung metastasis compared with the MDA-231HIC1 cells alone (3#). Moreover, quantification of micrometastasis in lung tissue further confirmed the above observations (Fig. 5E). As expected, LCN2 overexpression in MDA-231 cells markedly increased lung metastasis (Supplementary Fig. S5A). Assessment of the number of micrometastasis also showed that lung metastasis was markedly increased in MDA-231LCN2 mice compared with the control (Supplementary Fig. S5B). Representative images using H&E staining further confirmed the macroscopic findings (Supplementary Fig. S5C).

The AKT pathway is involved in HIC1-induced LCN2 expression in TNBC cells

Based on our in vitro and in vivo findings, we next explored the pathways involved in HIC1-induced LCN2 expression associated with TNBC progression. As shown in Fig. 6A, the AKT pathway was inactivated in MDA-231HIC1 cells (3#) compared with the control group (1#; Fig. 5D), consistent with the in vivo results shown in Fig. 2F–H. However, coexpression of LCN2 in MDA-231HIC1 cells (4#) partially rescued the HIC1-induced reduction of lung metastasis compared with the MDA-231HIC1 cells alone (3#). Moreover, quantification of micrometastasis in lung tissue further confirmed the above observations (Fig. 5E). As expected, LCN2 overexpression in MDA-231 cells markedly increased lung metastasis (Supplementary Fig. S5A). Assessment of the number of micrometastasis also showed that lung metastasis was markedly increased in MDA-231LCN2 mice compared with the control (Supplementary Fig. S5B). Representative images using H&E staining further confirmed the macroscopic findings (Supplementary Fig. S5C).
expression may contribute to the activation of the AKT pathway.

Indeed, Fig. 6D shows that knockdown of endogenous LCN2 by siRNAs decreased AKT activation in MDA-231 cells compared with the scrambled control. Emerging evidence has indicated that LCN2, as a secreted inflammatory factor, functions through neutrophil gelatinase-associated lipocalin receptor (NGALR), a multipass membrane protein (28). To test whether NGALR is involved in the AKT signaling pathway, NGALR expression was reduced by siRNAs in MDA-231 and MDA-468 cells. The results showed that the activation of the AKT pathway was inhibited in both cell lines compared with the respective scrambled controls (Fig. 6E and Supplementary Fig. S6). Further experiments are in progress to elucidate the underlying mechanisms and to determine whether other receptors are involved in the process.

To determine whether the LCN2-activated AKT pathway is involved in cell invasion, several small molecule inhibitors of the pathway were used, namely LY29004 [phosphoinositide 3-kinase (PI3K) inhibitor], BEZ235 (PI3K and mTOR inhibitor), BKM120 (selective PI3K inhibitor), and RAD001 (mTOR inhibitor). As shown in Fig. 6F–G, LCN2-induced cell invasion in MDA-468 cells was significantly inhibited by LY29004, BEZ235, and BKM120, whereas RAD001 had no effect, which could be partly attributed to pAkt473 feedback-loop activation. Taken together, these results demonstrate that HIC1-induced LCN2 expression activates the AKT pathway, which is responsible for promoting TNBC cell invasion.

**LCN2 predicts poor clinical outcome in patients with TNBC**

Based on the above findings, we assessed whether LCN2 expression is associated with TNBC progression. Supplementary
Table S2 shows that serum LCN2 levels were inversely correlated with the ER ($P < 0.001$) and PR status ($P = 0.012$) of patients with breast cancer, consistent with LCN2 expression in breast cancer tissues (19). Compared with the benign group, LCN2 level was markedly higher in patients with TNBC ($P = 0.007$), although the difference did not reach statistical significance ($P = 0.080$) in the HER2-positive subtype (Supplementary Table S2). However, this could be related to the discrete values and sample size limitations. Moreover, the subtype-specific prognostic power of LCN2 was shown in the independent clinical microarray database of breast cancer (23), in which a high level of LCN2 was associated with poor DMFS and OS in patients with basal-like breast cancer (Fig. 7A and B).

Figure 7C shows a schematic model of the function of the HIC1–LCN2 axis in TNBC progression. HIC1 expression is silenced in TNBC by epigenetic modifications, which increases the secretion of LCN2 because of the loss of the direct inhibitory effect of HIC1. LCN2 can activate the AKT pathway via an autocrine loop potentially mediated by the NGALR receptor, therefore promoting TNBC invasion and metastasis.

**Discussion**

HIC1 is a suppressor gene that is frequently hypermethylated in human cancers, including breast carcinomas. Emerging evidence suggests that re-expression of HIC1 is responsible for an improved prognosis in human cancers (29, 30). Breast cancer is a heterogeneous disease and several distinct subtypes exist, of which the triple-negative subtype has the worst clinical prognosis (29, 30). However, the role...
Figure 7. LCN2 predicts poor clinical outcome in TNBC. A, Kaplan–Meier plots of OS in whole datasets and patients with basal-like breast cancer, stratified by LCN2 expression. Data were obtained from the Kaplan–Meier plotter database (23). The P value was calculated by a log-rank test. B, Kaplan–Meier plots of DMFS in patients with breast cancer and patients with basal-like breast cancer, stratified by LCN2 expression. High levels of LCN2 were associated with significantly poorer OS and DMFS in patients with basal-like breast cancer. C, schematic model of the role of the HIC1–LCN2 axis in TNBC progression.

and regulatory mechanism of HIC1 in each subtype remain unclear.

Here, we showed that HIC1 expression was not significantly different between benign and malignant breast cancer tissues. This result could be related to the finding that HIC1 hypermethylation is a late event in some neoplasms, which suggests that other inhibitory mechanism may exist that modulate HIC1 expression (31–33). Our data showed that HIC1 was only silenced in the triple-negative subtype as compared with the benign, luminal, and HER2 subtypes. These findings indicate that HIC1 expression may be linked to a more aggressive type of TNBC and raise the question of whether lower HIC1 expression may serve as a marker of poor prognosis. Indeed, analysis of a public clinical microarray database (23) showed that HIC1 expression in patients with basal-like breast cancer was significantly associated with OS.

Our in vitro and in vivo experiments showed that restoring HIC1 expression in TNBC cells had a marked effect on reducing cell migration, invasion, and metastasis. However, reduction of HIC1 expression by shRNAs in MCF-10A and HBL-100 cells greatly increased their invasive ability. Microarray analysis and in vitro experiments identified LCN2 as an immediately downstream target of HIC1. LCN2 is a small-secreted glycoprotein involved in multiple processes, such as immunity, renal development, and apoptosis (16, 17). It binds iron by associating with 2,5-dihydroxybenzoic acid, a siderophore that shares structural similarities with bacterial enterobactin, and delivers or removes iron from the cell, dependent on the context (34). Indeed, LCN2 is expressed in a variety of tumors, including breast cancer (18). Leng and colleagues reported that inhibition of LCN2 impairs breast tumorigenesis and metastasis in MMTV-ErbB2 (V664E) mice (35). Several recent studies have suggested that LCN2 expression plays a role in breast cancer initiation and progression by promoting epithelial–mesenchymal transition (EMT), modulating MMP9 activity and regulating iron homeostasis in cancer cells (18, 36, 37). However, in this study, LCN2 had no effect on the regulation of EMT or MMP9 activity (Supplementary Figs. S3 and S4), which suggests that the function and potential mechanisms of LCN2 in breast cancer need to be further elucidated (21, 38).

Here, we showed that LCN2 activated the AKT pathway and promoted cell invasion in TNBC cells. The PI3K inhibitors LY29004, BEZ235, and BKM120 partly inhibited LCN2-induced breast cancer cell invasion, whereas the mTOR inhibitor RAD001 had no effect, possibly because of pAkt473 feedback-loop activation. These results suggest that in addition to the involvement of the AKT pathway in LCN2-induced cell invasion, other pathways may be activated via crosstalk. A physiologic receptor for LCN2 has not been defined (39). One of its potential receptors, NGALR, has been reported to internalize iron-laden LCN2. LCN2 is also known to undergo endocytosis through the megalin receptor (40, 41), a single-pass type I membrane protein that acts together with cubilin to mediate high-density lipoprotein endocytosis and may participate in the regulation of parathyroid hormone (42). Few studies have explored the mechanism underlying the role of LCN2 and LCN2 receptors in breast cancer progression. Here, we found for the first time that NGALR knockdown could inhibit the activation...
of the AKT pathway. Moreover, our data showed that PTEN activity is not greatly induced in the presence of LCN2 (Supplementary Fig. S7). These results suggest that the LCN2-induced AKT activation may be associated with the NGALR receptor, and PTEN may not be involved. However, the mechanisms underlying the role of LCN2 in TNBC progression require further investigation.

We showed that the LCN2 level in serum was higher in patients with TNBC than in those with benign and other subtypes of breast cancer, and it was negatively correlated with the ER and PR status. A similar finding was reported in patients with ovarian cancer, where a correlation between LCN2 expression and tumor differentiation was identified (43). Analysis of the clinical microarray database of breast cancer (23) showed that high levels of LCN2 are associated with poor DMFS and OS in patients with basal-like breast cancer, suggesting that serum LCN2 level may serve as a subtype-specific prognostic factor in breast cancer.

In summary, the findings suggest that the HIC1–LCN2 axis may serve as a subtype-specific prognostic biomarker and provide an attractive therapeutic target for the treatment of patients with TNBC.

References


Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Conception and design: G. Cheng, X. Sun, L. Zu, J. Wang
Development of methodology: X. Sun, J. Wang
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): X. Sun, Y. Mao
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): X. Sun, J. Wang, G. Xiao
Writing, review, and/or revision of the manuscript: X. Sun, X. Fan, L. Zu
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): X. Sun, J. Wang, M. Hao, Q. Qu, Y. Xue
Study supervision: X. Sun, J. Wang

Grant Support
Research in the authors’ laboratory is supported by the National Natural Foundation of China (81071747 and 81272404), the National Key Program (973) for Basic Research of China (2011CB510106, 2011CB504300), the Program for Professor of Special Appointment (Eastern Scholar to J. Wang) at the Shanghai Institute of Higher Learning, and Program of the Shanghai Municipal Health Bureau Subject Chief Scientist (XBR20110052).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received August 22, 2013; revised November 18, 2013; accepted November 18, 2013; published OnlineFirst December 2, 2013.
HIC1 Silencing in Triple-Negative Breast Cancer Drives Progression through Misregulation of LCN2

Guangcun Cheng, Xueqing Sun, Jinglong Wang, et al.


Updated version
Access the most recent version of this article at: doi:10.1158/0008-5472.CAN-13-2420

Supplementary Material
Access the most recent supplemental material at: http://cancerres.aacrjournals.org/content/suppl/2013/12/02/0008-5472.CAN-13-2420.DC1

Cited articles
This article cites 43 articles, 15 of which you can access for free at: http://cancerres.aacrjournals.org/content/74/3/862.full#ref-list-1

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.