Evolutionary Action Score of TP53 Coding Variants Is Predictive of Platinum Response in Head and Neck Cancer Patients

Abdullah A. Osman1, David M. Neskey2, Panagiotis Katsonis3, Ameeta A. Patel1, Alexandra M. Ward1, Teng-Kuei Hsu3, Stephanie C. Hicks4, Thomas O. McDonald5, Thomas J. Ow6, Marcus Ortega Alves7, Curtis R. Pickering1, Heath D. Skinner8, Mei Zhao1, Eric M. Sturgis9, Merrill S. Kies9, Adel El-Naggar10, Federica Perrone11, Lisa Licitra12, Paolo Bossi12, Marek Kimmel5, Mitchell J. Frederick1, Olivier Lichtarge3, and Jeffrey N. Myers1

Abstract

TP53 is the most frequently altered gene in head and neck squamous cell carcinoma (HNSCC), with mutations occurring in over two thirds of cases; however, the predictive response of these mutations to cisplatin-based therapy remains elusive. In the current study, we evaluate the ability of the Evolutionary Action score of TP53-coding variants (EAp53) to predict the impact of TP53 mutations on response to chemotherapy. The EAp53 approach clearly identifies a subset of high-risk TP53 mutations associated with decreased sensitivity to cisplatin both in vitro and in vivo in preclinical models of HNSCC. Furthermore, EAp53 can predict response to treatment and, more importantly, a survival benefit for a subset of head and neck cancer patients treated with platinum-based therapy. Prospective evaluation of this novel scoring system should enable more precise treatment selection for patients with HNSCC. Cancer Res; 75(7); 1205–15. ©2015 AACR.

Introduction

Head and neck squamous cell carcinoma (HNSCC) has an incidence of over 40,000 new cases annually in the United States, and over 500,000 worldwide with an associated disease-specific mortality exceeding 50% (1). The treatment of locally advanced head and neck cancer has evolved over the past three decades and often requires complex, multimodality therapy, including surgical resection, and/or external beam radiation with or without neoadjuvant, concurrent, or adjuvant cisplatin-based chemotherapy (2, 3). Currently, there are no molecular biomarkers to guide selection among these various treatment options. TP53 is the most frequently altered gene in human cancers and recent data from whole-exome sequencing of HNSCC reveals that this gene is mutated in 60%–80% of human papillomavirus negative (HPV−) cases (4, 5). The TP53 gene has been called the "cellular gatekeeper" due to its central role in response to cell stressors such as DNA damage, hypoxia, and oncogenic stress. Cellular DNA damage often leads to stabilization and accumulation of wtTP53, which in turn leads to enhanced transcription of p21 and subsequently cell-cycle arrest, apoptosis, and senescence. The increase in p53 stability depends critically on the phosphorylation of serine/threonine residues (6–9).

Although mutations in TP53 have been shown to have predictive significance for response to platinum-based therapy in several studies, it remains unclear how to stratify patients into response categories based on TP53 status (10–14). Recently, we developed an algorithm termed Evolutionary Action (EAp53) that accurately stratifies patients whose tumors have TP53 mutations associated with especially poor outcomes (high risk), from other mutations with outcomes similar to patients with wild-type TP53 (low risk) and have validated EAp53 as a reliable prognostic marker (D.M. Neskey and colleagues; submitted for publication). We hypothesize that high-risk p53 mutations identified by the EAp53 scoring system are associated with an abnormal functional activity that...
contributes to cisplatin resistance in head and neck cancer. Therefore, to determine whether EAp53 has utility as a predictive biomarker of response to cisplatin in HNSCC, we used both preclinical laboratory-based models and retrospective clinical data to assess the response of tumors expressing no p53, wtp53, or a series of low- and high-risk p53 mutations to cisplatin. The first aspect of our preclinical model found that in clonogenic survival assays, cell lines expressing high-risk p53 mutations are more resistant to cisplatin treatment than cell lines expressing low-risk mutations or wild-type p53.

To further characterize the preclinical response of the TP53 mutations stratified by Evolutionary Action to cisplatin therapy, tumors harboring these mutations were created in an orthotopic mouse model of tongue cancer. Concordant with the differential effect of TP53 mutations on cisplatin response observed in vitro, mice with tumors harboring wild-type p53 or low-risk mutations showed a significant response to cisplatin therapy, whereas the tumors derived from cells either null for p53 expression or with high-risk p53 mutations did not show any growth inhibition with cisplatin therapy. In an effort to correlate the clinical utility of the EAp53 to predict response to cisplatin in patients with HNSCC, the TP53 mutational status of a patient cohort of 68 patients treated for locally advanced HNSCC of the oral cavity with cisplatin-based induction chemotherapy followed by surgical resection was determined. Results from this analysis confirmed our preclinical findings wherein patients’ tumors with high-risk mutations were significantly less responsive to cisplatin-based chemotherapy than tumors with low-risk mutant or wild-type p53. These results indicate that the TP53 mutational status may be a useful biomarker for predicting response to cisplatin-based chemotherapy in HNSCC patients.

In summary, our data clearly demonstrate that high-risk TP53 mutations are associated with decreased sensitivity to cisplatin not only in preclinical studies but also in an analysis of a neoadjuvant chemotherapy clinical trial. Prospective clinical studies will be necessary to confirm the utility of TP53 status stratified by EA as a predictive biomarker of response to cisplatin-based therapy for HNSCC patients, which will potentially enable the personalization of therapy for patients that will most likely benefit from this treatment strategy.

**Materials and Methods**

**Cell lines**

Two HNSCC cell lines, UMSCC-1 and PCI-13, were selected for their lack of p53 expression due to a splice-site in UMSGCC1 (hg19:chr17:7578370C>T) and a deletion in PCI13 (hg19: chr17:7579670_7579709del). UMSCC-1 was provided by Dr. Thomas Carey (University of Michigan, Ann Arbor, MI) in February 2010. PCI-13 was acquired from Dr. Jennifer Grandis (University of Pittsburgh, Pittsburgh, PA) in August 2008. The naturally occurring HNSCC cell lines, HN30 (wtp53) and HN31 (mutp53) were obtained in December 2008 from the laboratory of Dr. John Ensele (Wayne State University, Detroit, MI). The cell lines and their isogenic derivatives were tested and authenticated against the parental cell lines by our group using short-tandem repeat analysis (15) within 6 months of use for the current study. Details regarding cell culture, reagents, and generation of stable cell lines were previously described (D.M. Neskey and colleagues; submitted for publication).

**Classification by evolutionary action scoring system**

Missense TP53 mutations were divided into “high-risk” and “low-risk” groups based on the model described previously (D.M. Neskey and colleagues; submitted for publication).

**Immunoblotting**

Cells grown on 10-cm plates were treated with clinically relevant dose of cisplatin (1.5 μmol/L) for 24 hours and washed with cold PBS. Western blotting was performed using standard techniques previously described (16) and primary antibodies to anti-p53 (Santa Cruz Biotechnology, sc-126), anti-phospho-p53 serine 15 (Cell Signaling Technology, 9284), anti-p21 (Calbiochem, OP64), and anti-β-actin (Sigma Aldrich, A1978) were used.

**Transcriptional activity of TP53**

Transcription of p21, a canonical p53 target, was measured via luciferase reporter activity using a vector containing the 2.4 kb p21 promoter and firefly luciferase (pWWP-LUC; Addgene). UMSCC-1 and PCI-13 cells expressing various TP53 constructs, HN30, and HN31 were cotransfected with pWWP-LUC and a constitutively active Renilla luciferase construct using Lipofectamine 2000. After 48 hours, cells were treated with 1.5 μmol/L cisplatin and incubated for 24 hours before collection. Luciferase reporter activity was measured as previously described (17). The results for the p21 reporter assay are relative to the cisplatin-treated wild-type (WT), which was standardized to 100 relative light units.

**Quantitative reverse transcription PCR analyses**

The effect of TP53 mutants on transcription of three downstream target genes (p21, MDM2, and NOXA) were determined by qRT-PCR. HNSCC (UMSCC1, PCI-13) cells stably expressing the TP53 mutant constructs were treated with cisplatin (1.5 μmol/L) for 24 hours before isolation of total RNA using RNAasy Mini Kit (Qiagen). Reverse transcription was performed using the high capacity cDNA Reverse Transcription Kit (Applied Biosystems) according to manufacturer’s instructions and a detailed description is included in the Supplementary Materials and Methods. The GAPDH gene was used as an internal control. Triplicate samples were examined. The expression of each target gene was normalized against GAPDH, which was calculated by the \(\frac{ΔC_t\text{ of target gene} - 100}{ΔC_t\text{ of GAPDH}}\), and results were presented as fold change of expression.

**mRNA expression arrays**

Total RNA was isolated from cell lines by using TRI reagent and hybridized to Affymetrix GeneChip Human Exon 1.0ST Arrays (Affymetrix) according to manufacturer’s instructions and a detailed description is included in the Supplementary Materials and Methods. The expression of TP53 target genes in pBabe and each of other groups was calculated and heatmaps were generated depicting the expression patterns of these genes.

**Clonogenic survival assay**

HNSCC cells stably expressing the TP53 constructs were seeded in 6-well plates at various densities, which allowed for approximately equal number of colonies in the control wells for each construct. The next day, cells were treated with increasing doses of cisplatin (0.01–2 μmol/L) dissolved in dimethyl sulfoxide (DMSO) for 24 hours and cultured for 10 to 14 days.
to allow for colony formation of at least 50 cells. The cells were stained with crystal violet and analyzed as previously described (17). Each experiment was repeated more than three times and treatments were performed in triplicates. An IC50 for each TP53 construct was calculated as the mean IC50 from each clonogenic assay.

**Orthotopic nude mouse model of oral cavity cancer**

All animal experimentation was approved by the Animal Care and Use Committee of the University of Texas MD Anderson Cancer Center. Our orthotopic nude mouse model of oral cavity cancer has been previously validated and described in the literature (18). UMSCC1, PCI 13, and cells expressing either, a high-risk, low-risk TP53 mutation, a null pBabe TP53 vector or wild-type TP53 along with HN30 and HN31 cells were used in the study and a detailed description of the technique is included in the Supplementary Materials and Methods.

**Patient cohort and TP53 sequencing**

A cohort of 68 patients with oral cavity squamous cell carcinoma (SCC) treated with platinum-based induction chemotherapy followed by surgery was collected from two clinical trials to investigate the predictive value of EAp53. Patient demographic, clinical data, and cisplatin-based treatment regimens were previously published (12, 19). Patient data, specimens, and TP53 sequences were collected under Institutional Review Board–approved protocols. DNA was extracted from tissue of patients enrolled in the trials and different techniques were used to determine TP53 sequence (20, 21). Detailed description of DNA isolation and TP53 sequencing is included in the Supplementary Materials and Methods. Patients with either TP53 wild-type or missense mutations were then scored by the EAp53 system into low- or high-risk categories as previously described (D.M. Neskey and colleagues; submitted for publication). The Evolutionary Action classification score was then correlated with clinicopathologic factors and patient outcome to determine associations with treatment response and survival.

**Statistical analysis**

ANOVA analysis with Student t tests were carried out to analyze in vitro data. For mouse studies, the two-tailed t test was used to compare tumor volumes between control and treatment groups. Survival was determined using the Kaplan–Meier method and compared using log-rank tests. Fisher exact test or χ2 test were used to calculate the ORs between treatment and clinical response. P values <0.05 were considered significant.

**Results**

DNA damage-induced functional activity of p53 in response to cisplatin treatment is impaired in HNSCC cells expressing low- and high-risk TP53 mutations

To examine in preclinical models whether the response to cisplatin therapy correlates with TP53 mutational status stratified by the Evolutionary Action method, the p53 function of cell lines that either exogenously express various p53 constructs including wild-type, low-, or high-risk mutant isoforms or endogenously express wild-type p53 (HN30) or a high-risk mutation (HN31) was assessed in these cells following treatment with cisplatin and analyzed by Western blot analysis. As expected, low basal expression levels of p53 and p21 were increased after cisplatin treatment in cells expressing wild-type TP53 (Fig. 1A and B and Supplementary Fig. S1A). In addition to the p53 and p21 induction after cisplatin treatment in the cells stably expressing wtP53, there was a similar level of p53 phosphorylation observed in the cells compared with the HN30 cell line, which endogenously expressed wtP53, indicating that the stably expressed wild-type TP53 was functionally active. In contrast, cells expressing mutant TP53 endogenously or exogenously, had higher basal levels of p53 with minimal induction of p53 or p21 after cisplatin treatment, indicating a lack of functional p53 (Fig. 1A and B and Supplementary Fig. S1A). Regardless of mutational status, phosphorylation of p53 following cisplatin was a ubiquitous finding, but p21 induction was most evident in cells expressing wtP53 and low-risk mutant p53. The data suggest that while p53 phosphorylation occurs in response to DNA damage, many of the high-risk mutations are not functional with respect to p21 induction, and that low risk mutations may retain partial wtP53 function.

Mutations in TP53 alter the DNA binding domain conformation and disrupt the ability of p53 to bind to target gene promoters and consequently to transactivate downstream genes (22) Thus, the ability of TP53 mutants to modulate the expression of classical wtP53 responsive target genes such as p21, MDM2, and Noxa was examined by both p21 promoter luciferase assay and qRT-PCR. After cisplatin treatment, cells with wtP53 have an increase in promoter activity while the low-risk mutations have a stable level of activity and high risk p53 have suppressed levels of p21 promoter activity relative to their basal levels and to the basal levels of the empty vector control (Fig. 1C and Supplementary Fig. S1B). The mRNA levels of the target genes, p21, MDM2, and Noxa were significantly elevated in response to cisplatin treatment in HNSCC cells harboring either an exogenously expressed or endogenous wild-type p53. Cells with low-risk mutations in the PCI13 cell line showed a trend toward increased target gene expression following cisplatin treatment relative to cells lacking p53, specifically the p21 level in A161S, or MDM2 and Noxa levels in Y236C (Fig. 1D). This is in contrast to cells harboring high-risk mutations where the mRNA levels after cisplatin treatment were similar to cells lacking p53 (Fig. 1D). Similar observations were seen in the low-risk UMSCC1 cell lines, specifically MDM2 level in A161S or MDM2 in Y236C. UMSCC1 cells harboring high-risk mutations were more variable in the target gene expression after cisplatin treatment, which may represent unique properties of these constructs or the cell background (Supplementary Fig. S1C). The difference between p21 mRNA and protein levels observed in PCI13 and UMSCC1 cells expressing the high-risk mutant (R175H) is possibly due to posttranslational modification event that resulted from less degradation of the mutant protein and therefore enhanced its stability upon cisplatin treatment. It could also be related to p63 and p73 isoforms being differentially expressed in these cells upon cisplatin addition. The p63 and p73 are well known p33-related proteins that act as transcriptional activators of p21 and apoptotic inducers upon DNA damage in tumor cells (23). Taken together, these results reveal that cells with a mutated p53 have increased basal levels of protein but following cytotoxic stress, there is decreased promoter activity of the canonical target, p21, and low mRNA levels of downstream target genes compared with cells expressing wild-type p53.
HNSCC cells bearing high-risk TP53 mutations are highly resistant to cisplatin treatment in vitro

To determine whether EAp53 has utility as a marker that can predict HNSCC response to cisplatin therapy, we assessed the response of HNSCC cells expressing no p53 (pBabe empty vector), wtp53, or a series of low- and high-risk p53 mutations to cisplatin in clonogenic survival assays. Figure 2A is representative images of clonogenic survival assay in HNSCC cell lines. As shown in Fig. 2B and C, in both genetic backgrounds, the high-risk mutant p53 clones were highly resistant to cisplatin, with 4 of 5 clones having an average IC50 of 0.95 μmol/L > 0.8 μmol/L, when exposed to cisplatin for a 24-hour period. We have determined this in vitro exposure of 0.8 μmol/L to be equivalent to the high dose of cisplatin (i.e., 100 μmol/L) given to patients in humans. Low-risk mutant p53 clones were less resistant to cisplatin with an average IC50 of 0.72 μmol/L that is statistically significant when compared with the high-risk mutant p53 clones (Fig. 2B and C). In addition, the clones expressing wild-type p53 had lower IC50 values (0.15 μmol/L) compared with clones with null pBabe empty vector (0.44 μmol/L) in both PCI-13 (P < 0.001) and UMSSC1 (P < 0.003). These data suggest that introduction of low- and high-risk TP53 mutations into HNSCC cell lines resulted in a gain-of-function (GOF) phenotype for resistance to cisplatin therapy. Furthermore, our laboratory has shown that the endogenous mutp53 of HN31 also confers a relative cisplatin resistance with an IC50 of 0.60 μmol/L compared with its isogenic wtp53 counterpart, HN30, which has an IC50 of 0.14 μmol/L (24). To further address loss-of-function (LOF) versus GOF, TP53 was knocked down in the isogenic pair of cell lines HN30 (wtp53) and HN31 (HRmutp53) and cells were then examined for cisplatin sensitivity (Supplementary Fig. S2A and S2B). The shRNAp53 HN30 cells become more resistant to cisplatin (IC50: 0.32 μmol/L), arguing that loss of wtTP53 function can make tumors less sensitive to cisplatin. Interestingly, knock-down of mutant HR p53 in HN31 made cells more sensitive to cisplatin with an IC50 value very close to the HN30 p53 knockdown (IC50 of 0.30 μmol/L vs. 0.32 μmol/L), indicating a GOF associated with the HR mutation. Collectively, the data argue that in vitro both loss of wtTP53 and a HR-associated GOF contribute to increased cisplatin resistance.

Expression profile of high-risk TP53 mutations shows a lack of p53 transcriptional activity while low-risk mutations retain some residual function

Given the apparent GOF phenotype seen in our in vitro studies of the p53 mutants, we performed mRNA expression profiling in an effort to identify genes and pathways specific to the high-risk mutp53 that could explain their relative resistance to cisplatin. The principal component analysis of the gene expression profiles for cisplatin-treated UMSSC1 cell lines, wtp53, pBabe, low-risk mutation (A161S), or high-risk mutation (C238F), revealed that the high-risk mutation expression profiles were more similar to the pBabe cell line, which lacks p53 expression. This is apparent from component 1 (x-axis), which accounted for 40% of variance in expression. Furthermore, the low-risk mutation profile had smaller variances (20%) in expression from the pBabe and high-risk mutations as seen by the large component 2 (y-axis) contribution. In contrast, the wtTP53 had the largest variances from the high-risk mutation and pBabe and was also distinct from the low-risk mutation profiles (Fig. 3A and B).

Ordinal logistic regression models were performed to identify genes that contribute to either a GOF or LOF phenotype where the magnitude of expression from highest to lowest is either high-risk, low-risk, wtp53, and pBabe or wtTP53, low-risk, high-risk, and pBabe respectively. Surprisingly, based on our in vitro data, the number of significant genes with the false discovery rate set at <10%, that contribute to a LOF phenotype was dramatically higher than the number of genes associated with a GOF phenotype, 1190 and 0, respectively (Fig. 3B and C). Furthermore, the
beta-uniform mixture plot analyses for the two potential phenotypes reveal an enrichment of genes with low P values leading to nonuniform distribution of genes in LOF analysis compared with the more uniform distribution in the GOF analysis (Fig. 3D and E). These results validate the higher P value cutoff used in the ordinal logistic regression analyses for LOF compared with GOF (Fig. 3B and C). As expected, the primary pathways that were driving these expression patterns were regulated by p53. Overall, the results from the mRNA expression profile reveal that the introduction of a high-risk p53 mutation leads to a reduction in the wild-type p53 function to levels similar to pBabe cells that lack p53 expression. In addition, introduction of a low-risk mutation results in an expression pattern suggestive of residual wild-type p53 function as seen by the intermediate level of expression of TP53 target genes (Fig. 3F and Supplementary Table S4).

High-risk TP53 mutations are associated with decreased response to cisplatin therapy and overall survival in an orthotopic mouse model of tongue cancer

To further characterize the ability of the EAp53 to predict the mutations to cisplatin therapy, tumors harboring these mutations were created in an orthotopic nude mouse model of tongue cancer. Animals underwent a 4-week course of cisplatin therapy, during which the tumor volumes and overall survival were monitored. Consistent with the differential effect of TP53 status on cisplatin response observed in vitro, tumors in mice injected with tumor cells expressing wild-type p53 showed a significant response to cisplatin therapy, while the tumors derived from cells null for p53 expression harboring the pBabe vector control or high-risk p53 mutations did not show any growth inhibition with cisplatin therapy (Fig. 4A–C). Interestingly, the response of tumors with low-risk mutations was more similar to the response of wtp53-bearing cells, which in agreement with the mRNA levels and expression array data in that the low-risk mutations appear to retain some wild-type p53 function. To compare relative tumor response between tumors to cisplatin, the area under the tumor growth curve was calculated for each animal and the mean AUC was plotted for each treated tumor and their corresponding control (Supplementary Fig. S3). Mice with tumors that harbor endogenous or exogenous wild-type p53 or low-risk mutations have a significant response (P < 0.0001, low- and high-risk mutation versus wild-type TP53).
therapy. This gradient of response corroborates the expression array data and once again implies a partial wild-type p53 function for the low-risk mutations and lack of p53 function for the high-risk mutations. The resistance to cisplatin seen in the high-risk mutations was also associated with decreased survival in animals bearing tumors with high-risk TP53 mutations (Fig. 4D–F). These results demonstrate that the Evolutionary Action method can predict the p53 mutations that are least likely to respond to platinum-based therapy in vivo in an orthotopic murine model of oral cancer.

EAp53 classification predicts response to platinum-based chemotherapy in patients with locally advanced oral cavity cancer

To determine the reliability of EAp53 to predict response to treatment in patients with oral cavity cancer, we identified a cohort of patients with locally advanced oral cavity cancer who received platinum-based induction chemotherapy in the context of prospective clinical trials. This cohort consisted of 68 patients, of which 26 of the tumor samples (38%) had missense mutations of TP53, while 42 tumor samples (62%) had wild-type p53 (Table 1). We have shown that the EAp53 system identified three groups independently, low EAp53 score, high EAp53 score, and wild-type p53 (D.M. Neskey and colleagues; submitted for publication). Univariate analysis in the training set revealed that the low EAp53 score mutations, that is, low risk, and wild-type were not statistically different, whereas the high EAp53 score mutations, termed high-risk mutations, appeared to be distinct from the other two groups (D.M. Neskey and colleagues; submitted for publication). Given the similar outcomes, patients with tumors having low-risk mutations were combined with wild-type p53 (wtP53). Therefore, the TP53 status was further classified by EAp53 into low-risk group (42 wild-type TP53 and 12 low-risk TP53 mutations), and a high-risk group consists of 14 high-risk TP53 mutations (Table 1...
EAp53 Predicts Response to Cisplatin in Head and Neck Cancer

Figure 4.
High-risk TP53 mutations are resistant to cisplatin in an orthotopic nude mouse model. A–C, HNSCC cell lines exogenously expressing PCI13 and UMSCC1 (A and B), and endogenously HN30 and HN31 (C) expressing p53 were injected into the tongue of nude mice. D–F, Kaplan–Meier curves of the orthotopic tongue model created with either cell lines HN30 or HN31 (D) or TP53 isogenic cell lines PCI-13 or UMSCC1 (E and F).

and Supplementary Table S1). Review of the pathologic findings revealed that 13 of the patients (93%) with high-risk TP53 mutations had residual disease, whereas only one patient showed complete response to cisplatin-based therapy. In contrast, 24 of the 54 patients (44%) with wild-type p53 and low-risk TP53 mutations achieved complete pathologic response while 30 patients (56%) had residual disease. These data demonstrate that relative to low-risk TP53, high-risk mutations are greater than 10-fold more likely to have residual disease following cisplatin-based chemotherapy (Table 1; \( P = 0.029 \)). EAp53 status was also found to be better predictor of cisplatin response than the previous classification system developed by Poeta and colleagues (11), which showed no statistically significant association with neoadjuvant response (Table 1; \( P = 0.248 \)) in our cohort. The other clinicopathologic data analyzed were not associated with a response to cisplatin therapy (Table 1). In addition, patients with tumors having high-risk TP53 mutations appear to have decreased overall survival relative to patients with low-risk TP53 status (Fig. 5A; \( P = 0.04 \)) in a Kaplan–Meier analysis. There was also a trend toward decreased disease-free survival in patients with high-risk TP53 mutations but it did not reach statistical significance (Fig. 5B; \( P = 0.08 \)). On univariate and multivariate analyses, the survival benefit of low-risk p53 status in the log rank tests was not observed in Cox Proportional Hazard Ratio Model (Supplementary Table S2 and S3). Overall, these data provide evidence that EAp53 can predict a subset of patient with high-risk TP53 mutations that have a decreased response to platinum-based chemotherapy and a poorer overall survival.

Discussion
Currently, there are not any established molecular biomarkers to predict response to chemotherapeutic agents in HNSCC. Recent whole exome analysis has confirmed that TP53 is the most frequently mutated gene in HNSCC occurring in 60%–80% of cases but the challenge remains to identify the mutations associated with resistance to current cytotoxic therapies and therefore decreased survival outcomes (4, 5). In the current study, we implemented a novel classification system, EAp53, which has the ability to predict response to cisplatin-based therapy not only in preclinical models of HNSCC but also in patients with locally advanced oral cavity squamous cell
carcinoma. We utilized a previously described collection of isogenic head and neck squamous cell carcinoma cell lines harboring a series of TP53 mutations or wild-type TP53, which allowed us to specifically examine the impact of TP53 alterations on response to cisplatin, as the genetic backgrounds of these cell lines are otherwise identical (D.M. Neskey and colleagues; submitted for publication). To confirm that the exogenous expression of p53 in these cells lines accurately represents the function of both wtp53 and mutant p53 (mutp53), cells endogenously expressing either wild-type or mutant p53 were also used for comparison.

The results from this study reveal that relative to wild-type TP53, both high- and low-risk mutations show a decrease in cisplatin-mediated p21 induction, a known transcriptional target of p53 in response to cisplatin treatment. In low-risk mutant TP53 cells, this diminished p21 induction appears to be associated with a reduction in the p21 promoter activity and an intermediate level of mRNA expression relative to wtp53-expressing cells. In contrast, the cell lines with either exogenous high-risk TP53 mutations

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Total</th>
<th>Yes</th>
<th>No</th>
<th>OR</th>
<th>95% CI</th>
<th>P*</th>
<th>Global P*</th>
</tr>
</thead>
<tbody>
<tr>
<td>All patients</td>
<td>68</td>
<td>25</td>
<td>43</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;59</td>
<td>46</td>
<td>19</td>
<td>27</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&gt;59</td>
<td>22</td>
<td>6</td>
<td>16</td>
<td>1.877</td>
<td>(0.620–5.675)</td>
<td>0.265</td>
<td></td>
</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>50</td>
<td>19</td>
<td>31</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Female</td>
<td>18</td>
<td>6</td>
<td>12</td>
<td>0.816</td>
<td>(0.262–2.536)</td>
<td>0.725</td>
<td></td>
</tr>
<tr>
<td>T stage</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>21</td>
<td>12</td>
<td>9</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>32</td>
<td>9</td>
<td>23</td>
<td>3.407</td>
<td>(1.070–10.847)</td>
<td>0.038</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>15</td>
<td>4</td>
<td>11</td>
<td>3.667</td>
<td>(0.874–15.384)</td>
<td>0.076</td>
<td>0.073</td>
</tr>
<tr>
<td>N stage</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>39</td>
<td>14</td>
<td>25</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>12</td>
<td>5</td>
<td>7</td>
<td>0.784</td>
<td>(0.209–2.938)</td>
<td>0.718</td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>8</td>
<td>4</td>
<td>4</td>
<td>0.56</td>
<td>(0.121–5.939)</td>
<td>0.458</td>
<td></td>
</tr>
<tr>
<td>2c</td>
<td>4</td>
<td>2</td>
<td>2</td>
<td>0.56</td>
<td>(0.071–4.421)</td>
<td>0.582</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>5</td>
<td>0</td>
<td>5</td>
<td>6.255</td>
<td>(0.322–121.42)</td>
<td>0.226</td>
<td>0.404</td>
</tr>
<tr>
<td>N stage</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;2b</td>
<td>51</td>
<td>19</td>
<td>32</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2b</td>
<td>17</td>
<td>6</td>
<td>11</td>
<td>1.089</td>
<td>(0.346–3.422)</td>
<td>0.885</td>
<td></td>
</tr>
<tr>
<td>TP53 status</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Wild-type</td>
<td>42</td>
<td>20</td>
<td>22</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mutant</td>
<td>26</td>
<td>5</td>
<td>21</td>
<td>3.818</td>
<td>(1.211–12.03)</td>
<td>0.022</td>
<td></td>
</tr>
<tr>
<td>EA status</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Low risk</td>
<td>54</td>
<td>24</td>
<td>30</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>High risk</td>
<td>14</td>
<td>1</td>
<td>13</td>
<td>10.4</td>
<td>(1.269–85.24)</td>
<td>0.029</td>
<td></td>
</tr>
<tr>
<td>Poeta classification</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-disruptive</td>
<td>61</td>
<td>24</td>
<td>37</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Disruptive</td>
<td>7</td>
<td>1</td>
<td>6</td>
<td>3.9</td>
<td>(0.440–34.387)</td>
<td>0.248</td>
<td></td>
</tr>
</tbody>
</table>

Abbreviations: CI, confidence interval; EA, evolutionary action score.

*aUsed Fisher exact test to calculate P values.
*bIf contingency table was larger than 2 × 2, then global P value was calculated using either χ² test or Fisher exact test and P value was calculated for each 2 × 2 subtable. For comparison with the EA p53 system, patient tumors were also classified as disruptive and non-disruptive according to Poeta and colleagues (11). Patients with wild-type TP53 or silent mutations were classified as nondisruptive; however, the association was still not significant even when patients with wild-type TP53 or silent mutations were removed.

carcinoma. We utilized a previously described collection of isogenic head and neck squamous cell carcinoma cell lines harboring a series of TP53 mutations or wild-type TP53, which allowed us to specifically examine the impact of TP53 alterations on response to cisplatin, as the genetic backgrounds of these cell lines are otherwise identical (D.M. Neskey and colleagues; submitted for publication). To confirm that the exogenous expression of p53 in these cells lines accurately represents the function of both wtp53 and mutant p53 (mutp53), cells endogenously expressing either wild-type or mutant p53 were also used for comparison.

The results from this study reveal that relative to wild-type TP53, both high- and low-risk mutations show a decrease in cisplatin-mediated p21 induction, a known transcriptional target of p53 in response to cisplatin treatment. In low-risk mutant TP53 cells, this diminished p21 induction appears to be associated with a reduction in the p21 promoter activity and an intermediate level of mRNA expression relative to wtp53-expressing cells. In contrast, the cell lines with either exogenous high-risk TP53 mutations

![Figure 5.](image_url)
or endogenous mutp53 had decreased p21 promoter activity following cisplatin treatment as previously described (25, 26). The functional activity of high TP53 mutations was partially corroborated by the qRT-PCR results with the high-risk mutations having a decreased level of target gene expression following cisplatin treatment. It has been suggested that loss of upregulation of key p53 target genes may contribute to the GOF phenotype of some mutp53 (25–28). Although mutated p53 is unable to bind to sequence specific DNA of target gene promoters secondary to alterations in the DNA binding domain, it has been suggested that p53 either recognizes target promoters independent of this region or binds the promoters at regions unique from p53-binding sites (26, 29). Furthermore, the loss of upregulation of p53 target genes may be enhanced by the constitutive overexpression of mutp53 due to their inability to effectively activate MDM2, a negative regulator of p53 abundance (30, 31). This latter notion could be supported by our finding that cisplatin treatment reduced the mRNA level of MDM2 in cells expressing low- and high-risk TP53 mutants.

To further assess the effect of suppression of p21 on response to chemotherapy in the EAp53 high-risk mutations, we show in a clonogenic survival assay that HNSCC cells expressing low-risk mutations have an intermediate level of resistance while high-risk TP53 mutations have a high level of resistance to platinum-based therapy relative to the pBabe vector control, confirming a GOF phenotype with regard to cisplatin sensitivity. The differences in cisplatin sensitivity between the wtp53 and mutp53 may be due in part to the inhibition of accelerated cellular senescence by mutat-ed p53 (24).

Although our initial in vitro experiments suggested a GOF phenotype for high-risk TP53 mutations, our attempt to identify pathways driving this characteristic through expression array analysis revealed that the introduction of high-risk mutations actually produces a largely nonfunctional p53 (i.e., suppressed levels of expression of TP53 target genes) that is most similar to a complete absence of the protein. Furthermore, the low-risk mutations have distinct expression variances from cells either lacking p53 or containing a high-risk mutation, which implies this mutation may retain some wildtype functions. The discrepancy between the expression array data and the in vitro experiments could be due to different posttranslational modifications of the low- and high-risk mutations that would not be detected on an mRNA expression array or an altered protein–protein interactome through which p53 gains function through functionally significant interactions with important cellular protein targets (32, 33).

In support of this latter mechanism, we have recently reported that GOF p53 mutations can bind to and inactivate, the master metabolic regulatory protein, AMPK, which leads to gain of oncogenic functions (34). Furthermore, evidence is now accumulating to indicate that different p53 mutations possess different functions in different tissues, potentially reflecting differences in the expression of their cellular targets (35). Therefore, understanding the consequences of each p53 mutation in relationship to disease progression and response to therapy promises to be an extremely complex undertaking and thus highlights the importance of our current study.

We further characterized not only the functional spectrum of p53 mutations but also the ability of EAp53 to predict response to cisplatin in vivo with an orthotopic nude mouse model of tongue cancer. These results confirmed that tumors bearing high-risk mutations are more resistant to cisplatin compared with tumors expressing either wtp53 or low-risk mutant p53. The lack of tumor response to treatment in mice with high-risk p53-bearing tumors was similar to tumors lacking p53 expression, which corroborates the expression array data. In addition to having an improved tumor response, animals with tumors harboring wt53 and low-risk mutations treated with cisplatin had an improved survival compared with both their untreated controls and the animals with high-risk p53 tumors.

Finally, in an effort to evaluate the predictive ability of TP53 mutational status stratified by the EA method, we analyzed a cohort of patients with locally advanced oral cavity squamous cell carcinoma who had been treated with platinum-based neoadjuvant chemotherapy followed by surgery (12, 19). This analysis revealed that EAp53 can identify a population of patients with high-risk p53 mutations that do not respond to platinum-based treatment and had decreased overall survival. Although these results are encouraging, there are limitations to this study. The percentage of patients with wt53, 62%, is higher than expected, which could be attributed to either the sensitivity of sequencing TP53 from paraffin tissues or sequencing exons 5–8, as only approximately 80% of mutations occur within the DNA-binding domain (36). Therefore, it is possible that the portion of the wild-type p53 patients’ tumors that did not have a clinical response actually harbored TP53 mutations. Taken together, these explanations may partly account for not only the high percentage of wt53 but also the large number of wt53 nonresponders. Nonetheless, EAp53 was predictive of lack of response to platinum-based treatment in patients with high-risk mutant p53, and identified patients with oral cavity squamous cell carcinoma that have decreased survival. To further validate these clinical findings, additional studies of p53 mutational status in larger cohorts of HNSCC patients that have received neoadjuvant platinum-based induction therapy are ongoing.

In summary, the EAp53 model clearly identifies a subset of high-risk TP53 mutations associated with decreased sensitivity to cisplatin in preclinical models. In addition, EAp53 can predict response to treatment and more importantly a survival benefit for a subset of patients treated with platinum-based therapy. To fully evaluate the role of EAp53 as a predictive biomarker of platinum-based treatment, prospective clinical trials in which patients’ tumors are stratified on the basis of their p53 mutational status and correlated with the response to treatment and survival are necessary.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): A.A. Osman, D.M. Neskey, A.A. Patel, A.M. Ward, T.J. Ow, M.O. Alves, C.R. Pickering, M.S. Kies, A. El-Naggar, L. Licitra, P. Bossi, J.N. Myers
Osman et al.

Writing, review, and/or revision of the manuscript: A.A. Osman, D.M. Neskey, P. Katsonis, S.C. Hicks, T.J. Ow, H.D. Skinner, E.M. Sturgis, M.S. Kies, L. Licina, P. Bossi, M. Kimmel, M.J. Fredericke, O. Lichtarge, J.N. Myers

Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): A.A. Osman, D.M. Neskey, A.A. Patel, A.M. Ward, J.N. Myers

Study supervision: A.A. Osman, A. El-Naggar, M. Kimmel, J.N. Myers

Other (performed experiments): A.A. Osman

Other (provided TP53 mutational results found in an Italian series of HNSCC): F. Perrone

Grant Support

This work was supported by the University of Texas MD Anderson Cancer Center PANTHEON program (philanthropic support to J.N. Myers), the NIH Specialized Program of Research Excellence Grant P50CA097007 (J.N. Myers), the NIH/NIDCR R01 DE14613 and R01 DE024601 (J.N. Myers), Cancer Prevention and Research Institute of Texas [CPRT] RPI20258 (J.N. Myers), National Research Service Award Institutional Research Training Grant T32CA06374 (J.N. Myers), the NIH Program Project Grant C168485 (J.N. Myers), and the Cancer Center Support Grant CA016672 (J.N. Myers). This work was also supported by NIH R01 CA075656 (O. Lichtarge), R01 CA066099 (O. Lichtarge), and NSF CCF 0905536 (O. Lichtarge) and DBI 0851393 (O. Lichtarge), and Pharmacoinformatics Training Program of the Keck Center of the Gulf Coast Consortia NIH grant no. 5 R90 DK071505 (P. Katsonis).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received September 19, 2014; revised December 16, 2014; accepted January 12, 2015; published OnlineFirst February 17, 2015.

References

Evolutionary Action Score of TP53 Coding Variants Is Predictive of Platinum Response in Head and Neck Cancer Patients


Cancer Res 2015;75:1205-1215. Published OnlineFirst February 17, 2015.

Updated version
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-14-2729

Supplementary Material
Access the most recent supplemental material at:
http://cancerres.aacrjournals.org/content/suppl/2015/02/17/0008-5472.CAN-14-2729.DC1

Cited articles
This article cites 36 articles, 15 of which you can access for free at:
http://cancerres.aacrjournals.org/content/75/7/1205.full.html#ref-list-1

Citing articles
This article has been cited by 5 HighWire-hosted articles. Access the articles at:
/content/75/7/1205.full.html#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.