Notch Suppresses Angiogenesis and Progression of Hepatic Metastases

Debarshi Banerjee1, Sonia L. Hernandez1, Alejandro Garcia2, Thaned Kangsamaksin3, Emily Sbiroli2, John Andrews2, Lynn Ann Forrester2, Na Wei1, Angela Kadenhe-Chiweshe2, Carrie J. Shawber2,3, Jan K. Kitajewski3,4, Jessica J. Kandel2, and Darrell J. Yamashiro1,2,4

Abstract

The Notch pathway plays multiple key roles in tumorigenesis, and its signaling components have therefore aroused great interest as targets for emerging therapies. Here, we show that inhibition of Notch, using a soluble receptor Notch1 decoy, unexpectedly caused a remarkable increase in liver metastases from neuroblastoma and breast cancer cells. Increased liver metastases were also seen after treatment with the γ-secretase inhibitor PF-03084014. Transgenic mice with heterozygous loss of Notch1 demonstrated a marked increase in hepatic metastases, indicating that Notch1 signaling acts as metastatic suppressor in the liver microenvironment. Inhibition of DLL1/4 with ligand-specific Notch1 decoys increased sprouting of sinusoidal endothelial cells into micrometastases, thereby supporting early metastatic angiogenic growth. Inhibition of tumor-derived JAG1 signaling activated hepatic stellate cells, increasing their recruitment to vasculature of micro-metastases, thereby supporting progression to macrometastases. These results demonstrate that inhibition of Notch causes pathologic activation of liver stromal cells, promoting angiogenesis and growth of hepatic metastases. Our findings have potentially serious implications for Notch inhibition therapy. Cancer Res; 75(8); 1592–602. ©2015 AACR.

Introduction

The four transmembrane Notch receptors and five membrane-bound ligands, DLL1, DLL3, DLL4, JAG1, and JAG2, classically function in development and differentiation, but also play a critical role in cancer (1–4). Aberrant Notch activation was first discovered in T-cell acute lymphoblastic leukemia (1) and later found in a variety of solid tumors (2–5). Notch functions in tumor angiogenesis are also well documented, with DLL4 highly expressed in tumor vasculature (6, 7).

Consequently, targeting Notch pathway components is currently a focus of preclinical and clinical research (8–14). Yet the widespread functions and highly pleiotropic nature of Notch raises the possibility of unanticipated effects on host tissues. For example, γ-secretase inhibitors (GSI), which prevent cleavage and activation of Notch receptors, cause serious gastrointestinal toxicity due to induction of goblet cell hyperplasia, a direct result of Notch inhibition (15). DLL4 inhibition in animal studies can cause aberrant activation of endothelial cells (EC), leading to formation of vascular tumors (16).

Here, we show that inhibition of Notch signaling causes a remarkable increase in spontaneous liver metastasis from neuroblastoma and breast cancer cells. Similarly, heterozygous loss of Notch1 in host animals leads to a marked increase in liver metastasis. Our data indicate that this effect is due to decreased Notch activation in liver sinusoidal endothelial cells (SEC) and hepatic stellate cells (HSC). Our findings demonstrate that perturbing Notch signaling can induce pathologic activation of hepatic stromal cells, leading to the growth of metastatic deposits.

Materials and Methods

Cell culture

The NGP cell line was obtained from Garrett Brodeur, Children’s Hospital of Philadelphia, Philadelphia, PA, and authenticated by short tandem repeat profiling. SH-SY5Y and MDA-MB-231 cell lines were obtained from the ATCC, BALB/c SEGs from CellBiologics, and human HSCs from ScienCell Research.

Lentiviral production and transfection

NGP was stably transduced with pLKO.1 Notch1 shRNA lentiviral plasmid (Sigma-Aldrich) as described previously (14). For other transfections, lentiviral plasmid pCC1, encoding Fc, N11–36 decoy, N11–24 decoy, N11–13 decoy or N110–24 decoy, were...
cotransfected with other plasmids (pCCL-GFP, pVSVG, pPRE, pHSV-rev) in HEK293T cells by Eugene (Promega).

**Animals**

Rag2/Il2rg double knockout (Rag2<sup>-/-</sup>, Il2rg<sup>-/-</sup>) in a C57BL/6Jc57Bl10ScSnAi background (Taconic, model 4111), were crossed to a pan-eGFP-expressing mouse [C57BL/6-Tg(CAG-EGFP)1Osb/J, model 003291; The Jackson Laboratory]. Resulting F1s were back-crossed to Rag2/Il2rg double knockout to obtain Rag2<sup>-/-</sup>, Il2rg<sup>-/-</sup>, eGFP<sup>+</sup> mice. These were then back-crossed to the Rag2/Il2rg double knockout for nine generations. Notch1<sup>+/+</sup> mice, were crossed with Rag2<sup>-/-</sup>, Il2rg<sup>-/-</sup>, eGFP<sup>-</sup> mice. Notch1<sup>+/+</sup>, Rag2<sup>-/-</sup>, Il2rg<sup>-/-</sup>, eGFP<sup>+</sup> mice were then back-crossed with Rag2<sup>-/-</sup>, Il2rg<sup>-/-</sup>, eGFP<sup>+</sup> mice to generate Notch1<sup>+/+</sup>, Rag2<sup>-/-</sup>, Il2rg<sup>-/-</sup>, eGFP<sup>+</sup> mice and Notch1<sup>+/+</sup>, Rag2<sup>-/-</sup>, Il2rg<sup>-/-</sup>, eGFP<sup>-</sup> control mice.

**Tumor xenografts**

Procedures were approved by Columbia University Institutional Animal Care and Use Committee. For intrarectal tumors, 4- to 6-week-old female nude mice (Taconic) were anesthetized with ketamine and xylazine, an incision made at the left flank, and 10<sup>6</sup> cells injected into the renal parenchyma. For intracardiac liver metastases studies, 10<sup>5</sup> to 10<sup>6</sup> cells were injected into the left cardiac ventricle, and bioluminescence imaging obtained after 20 minutes to monitor distribution of tumor cells.

**GSI**

Eighteen days after implantation of NGP cells, mice were randomized for GSI treatment, PF-03084014 (provided by Pfizer), formulated in 0.5% methylcellulose (vehicle) was administered orally by gavage at 125 mg/kg twice daily for 10 days.

**Bioluminescence analysis**

Mice were injected i.p. with 75 mg/kg α-Luciferin (PerkinElmer), anesthetized with isoflurane, and imaged with a Xenogen IVIS200. For survival studies, mice were sacrificed when bioluminescence flux reached a threshold value 6 × 10<sup>9</sup> (photons/s). For ex vivo liver imaging, mice were injected with α-Luciferin, sacrificed and the liver dissected, imaged and bioluminescence measured.

**Assessment of liver metastasis**

Livers were fixed in 4% paraformaldehyde, paraffin-embedded, sectioned (5 μm) at 50 μm intervals, and hematoxylin and eosin stained. Diameters of metastatic nodules from three nonconsecutive sections were measured. For quantification by bioluminescence, a liver piece was homogenized in lysis buffer, centrifuged, supernatant mixed with LARII reagent and bioluminescence measured with a luminometer and normalized to the liver piece weight.

**Circulating tumor cells**

Blood was collected by cardiac puncture, lysed by centrifugation, supernatant mixed with LARII reagent and bioluminescence measured with a luminometer.

**Quantification of vasculature**

Vascular parameters were determined as previously described (14, 18). For antibodies see Supplementary Table S1.

**Migration assay**

HSCs expressing GFP were seeded (1.5 × 10<sup>4</sup> cells/well) in the upper chamber of CytoSelect 24-well cell migration plate (8-μm pore size; CellBiotics). NGF cells expressing ligand decoys or Jag1-siRNA transfected were seeded to the upper chamber (1.5 × 10<sup>4</sup> cells/well). RPMI-1640 + 10% FBS was added to the lower chamber. After 48 hours, migrating HSCs were counted by fluorescence microscopy from eight random fields from three inserts. To verify inhibition of Notch signaling, HSC–GFP cells were FACs sorted and cell lysates immunoblotted for Hey1 and Hes1.

**Statistical analysis**

All statistical analysis was performed using Prism5 software (GraphPad). Survival was determined by log-rank (Mantel–Cox). Data were analyzed for normality by the D’Agostino-Pearson omnibus K2 test. Normal data were analyzed by unpaired t test or ANOVA with post hoc analysis by Tukey multiple comparison test. Nonnormal data were analyzed nonparametrically with the Mann–Whitney or Kruskal–Wallis test with post hoc analysis by Dunn multiple comparison test. Bioluminescence data were transformed Y = log(Y), then analyzed by ANOVA, with post hoc analysis by the Tukey multiple comparison test.

**Results**

Combined blockade of Notch and VEGF prolongs survival but increases liver metastasis

We previously demonstrated that combining Notch and VEGF blockade disrupted angiogenesis (14), and therefore hypothesized that this would translate into prolonged survival in comparison with VEGF blockade alone. The Notch1 decoy (N1D) is composed of EGF repeats 1 to 36, and blocks Notch activation by both DLL and JAGGED ligands (13). The neuroblastoma cell line NGP-expressing N1D (NGP-N1D) or LacZ (NGP-LacZ) was implanted into the left kidney of nude mice, and treated with the anti-VEGF antibody bevacizumab (BV). We monitored tumor size in vivo using bioluminescence and sacrificed animals at a predetermined photon flux. Combined Notch and VEGF blockade (NGP-N1D-BV) prolonged median survival as compared with bevacizumab alone (NGP-LacZ-BV; Fig. 1A, 43 vs. 36 days, P = 0.048). There was no difference in the tumor weight between NGP-N1D-BV compared with NGP-LacZ-BV (1.88 ± 0.95 vs. 1.46 ± 0.91 gm, mean ± SD).

Unexpectedly, at day 35, we noted bioluminescent signal contralateral to the primary tumors in NGP-N1D-BV mice, with increasing signal at day 50 (Fig. 1B). Necropsy of NGP-N1D-BV mice demonstrated multiple liver nodules (Fig. 1C). In comparison, few NGP-LacZ-BV mice had gross liver nodules (17/28 vs. 4/25, respectively, P = 0.0017, Fisher exact test). Metastatic burden as measured by bioluminescence of liver homogenates was 10-fold higher in NGP-N1D-BV (Fig. 1D, P < 0.001). The number and size of hepatic metastases was increased (Fig. 1E), with metastatic foci increased in NGP-N1D-BV (Fig. 1F, mean 11.2 vs. 2.8 metastatic foci/mm<sup>2</sup>, P < 0.01), with parallel increases in metastases’ mean diameter (Fig. 1G; 744 vs. 195 μm, P < 0.0001). Thus, combined Notch and VEGF inhibition while prolonging survival, paradoxically led to increased liver metastasis.
Combined Notch and VEGF blockade does not increase circulating tumor cells

We have previously demonstrated that combined blockade of Notch and VEGF decreased the vasculature of neuroblastoma xenografts and disrupted the interaction of ECs with pericytes (14). Similar results were obtained when the vasculature of the primary tumor of the survival experiment was examined. There was a 23% decrease in the EC marker endomucin in NGP-N1D+BV tumors (Supplementary Fig. S1A, P < 0.05). Although almost all endomucin(+) vessels had adjacent α-smooth muscle actin (α-SMA; +) pericytes (96% for NGP-LacZ+BV, 94% NGP-N1D+BV), the pericytes in NGP-N1D+BV often appeared dissociated from the ECs (Supplementary Fig. S1B). This apparent disruption of tumor vasculature suggested that Notch blockade might facilitate the entry of tumor cells into the circulation, thus promoting liver metastasis. We therefore quantified circulating tumor cells (CTC) by bioluminescence from blood obtained at time of sacrifice (Fig. 1H). There was no difference in CTCs, indicating that concurrent Notch and VEGF inhibition does not increase tumor cell intravasation.

Notch blockade is sufficient to promote liver metastasis

To determine whether Notch blockade alone could promote liver metastasis, we implanted NGP-LacZ or NGP-N1D intravenously. NGP-N1D tumors had increased liver homogenate bioluminescence (40-fold, P < 0.05), metastatic foci (9.2-fold, P < 0.05), and diameters (0.75 vs. 0.17 mm, P < 0.01; Supplementary Fig. S2A–S2C).

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To determine whether increased metastasis was due to an effect on the primary tumor or the liver, we injected NGP-LacZ or NGP-N1D cells into the left cardiac ventricle (19). We also treated with bevacizumab or placebo to determine whether VEGF blockade contributed to liver metastases. Intracardiac injection of NGP-N1D resulted in a markedly higher liver metastatic burden by ex vivo liver bioluminescence (Fig. 2A), and by total liver flux compared with NGP-LacZ (Fig. 2B, 113-fold, P < 0.05). Treatment with bevacizumab did not alter the metastatic pattern, with NGP-N1D+BV having increased liver metastases compared with NGP-LacZ+BV (Fig. 2B 93-fold, P < 0.05). Quantification of liver homogenate bioluminescence yielded similar results with increased metastases for N1D and N1D+BV (Supplementary Fig. S3A). Notch blockade resulted in significantly larger metastases (Fig. 2C and D; 3.00 and 2.91 mm, NGP-N1D and NGP-N1D+BV, vs. 1.55 and 1.51 mm, NGP-LacZ, and NGP-LacZ+BV). These results demonstrate that Notch inhibition is sufficient to promote liver metastases, and does not require concurrent VEGF blockade.

To determine whether the effect of Notch blockade on liver metastasis was cell line specific, we expressed the N1D in the MYCN-nonamplified neuroblastoma cell line SH-SYSY (SH-SYSY-N1D). Intracardiac injection of SH-SYSY-N1D resulted in markedly increased liver metastatic burden (Fig. 2E and F; 1.065-fold, P < 0.001), and larger metastases (Fig. 2G, 2.03 vs. 0.92 mm, P < 0.0001), compared with SH-SYSY-GFP.

To determine whether this effect was tumor-type specific, we engineered the breast cancer cell line MDA-MB-231 to express the N1D (MDA-MB-231-N1D). Intracardiac injection of MDA-MB-231-N1D resulted in a 23% decrease in the EC marker endomucin in the primary tumor of the survival experiment was examined. There was a 23% decrease in the EC marker endomucin in NGP-N1D+BV tumors (Supplementary Fig. S1A, P < 0.05). Although almost all endomucin(+) vessels had adjacent α-smooth muscle actin (α-SMA; +) pericytes (96% for NGP-LacZ+BV, 94% NGP-N1D+BV), the pericytes in NGP-N1D+BV often appeared dissociated from the ECs (Supplementary Fig. S1B). This apparent disruption of tumor vasculature suggested that Notch blockade might facilitate the entry of tumor cells into the circulation, thus promoting liver metastasis. We therefore quantified circulating tumor cells (CTC) by bioluminescence from blood obtained at time of sacrifice (Fig. 1H). There was no difference in CTCs, indicating that concurrent Notch and VEGF inhibition does not increase tumor cell intravasation.

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231-N1D resulted in increased liver metastasis as compared with controls (Fig. 2H), with a 2.1-fold increase in liver flux (Fig. 2I, *P < 0.05), and a 11.7-fold increase in liver homogenate bioluminescence (Supplementary Fig. S3B, *P < 0.05).

To determine whether Notch blockade promotes metastasis to other organs, we examined the spleen, kidney, and bone marrow after intracardiac injection of N1D alone and the lung after intracardiac injection of MDA-MB-231-N1D.
Liver metastasis is promoted by GSI but not by Notch1 knockdown. A, intrarenal NGP tumors were treated 18 days after implantation with PF-003084014 ($n = 7$) or vehicle ($n = 6$) for 10 days and sacrificed 7 days later; $P =$ nonsignificant. B, ex vivo imaging of livers. C, quantification of total flux (photon/s) by ex vivo liver bioluminescence; **, $P < 0.01$. D, ex vivo imaging of livers after intracardiac injection of NGP-Con ($n = 7$) and NGP-N1KD ($n = 8$). E, quantification of total flux (photon/s) by ex vivo liver bioluminescence; $P =$ nonsignificant. F, NGP-Con ($n = 7$), NGP-N1KD ($n = 7$), NGP-LacZ ($n = 6$), and NGP-N1D ($n = 6$) were implanted into the left lobe of the liver, and mice were sacrificed when flux reached $6 \times 10^6$ photons/s; log-rank (Mantel-Cox); $P = 0.0144$.

GSI PF-03084014 promotes liver metastasis

Our results demonstrate that cell lines engineered to express N1D have increased liver metastasis. We also used a pharmacologic approach to inhibit Notch signaling, with the GSI PF-03084014 (20), which has completed phase 1 testing in adults – medicinal approach to inhibit Notch signaling, with the GSI PF-03084014 (20), which has completed phase 1 testing in adults. Treatment of NGP intrarenal tumors began 18 days after implantation, with PF-003084014 administered for 10 days, and sacrificed 7 days later. PF-003084014 did not inhibit the growth of the primary tumors (Fig. 3A). Liver metastases, however, were significantly increased in mice treated with PF-003084014, with a 38-fold higher total liver flux compared with vehicle treated mice (Fig. 3B and C, $P < 0.01$). These results demonstrate that GSI inhibition of Notch signaling can promote liver metastases.

Notch inhibition does not affect prometastatic characteristics of tumor cells

The increased liver metastases could be due to a direct effect on tumor cells or indirectly on the liver. We asked whether N1D inhibition of autocrine or paracrine Notch signaling in tumor cells promoted prometastatic characteristics. NGS expresses Notch1, whereas SH-SY5Y expresses Notch1, 2, and 3, and both cell lines express DLL4, JAG1, and JAG2 (Supplementary Fig. S5A). Yet, despite these expression patterns, N1D did not affect in vitro proliferation, invasion, or migration of either NGS or SH-SY5Y (Supplementary Fig. S5B and S5C).

Knockdown of Notch1 in tumor cells does not promote liver metastases

To determine whether loss of Notch signaling in tumor cells was responsible for liver metastasis, we used shRNA to knockdown Notch1 (N1KD) in NGS (Supplementary Fig. S6A). N1KD, similar to N1D, decreased expression of Notch-responsive genes Hes1 and Hey1, but did not affect proliferation in vitro (Supplementary Fig. S6B). In vivo, intracardiac injection of NGP-N1KD did not increase liver metastasis (Fig. 3D and E), indicating that inhibition of Notch signaling in tumor cells is not responsible for promoting liver metastasis. NGS-N1D and NGS-N1KD cells were also directly implanted in the left lobe of the liver, and growth monitored by bioluminescence. NGS-N1D intrahepatic tumors progressed significantly faster (Fig. 3D; median survival 31 days, $P = 0.0144$) than NGP-N1KD (42 days), or control NGP-LacZ and NGP-Con tumors (37 and 38 days). This suggests that inhibition of Notch in the liver, but not in tumor cells, promotes the growth of metastatic lesions.

Notch inhibition increases vascularity of liver metastases

One potential target of Notch blockade is the host vasculature. We segregated liver metastases into small ($< 300 \mu m$) versus large ($> 300 \mu m$) lesions, reasoning that progression could be enhanced by alterations in vasculature at either early or late stages. In small metastases ($< 300 \mu m$) of NGS-LacZ+BV, there was a paucity of intrametastatic vessels as demonstrated by a lack of endomucin (Fig. 4A) and ICAM-1 (not shown). In contrast, invading vascular sprouts were common seen in NGS-N1D+BV metastases (Fig. 4A). Quantifying the incidence of vascularized metastases, 74% of NGS-N1D+BV metastases
were endomucin(+) as compared with 21% in NGP-LacZ+BV (P<0.0001). These small metastases lacked αSMA staining (not shown), suggesting that in this early angiogenic phase vessels were not yet invested by pericytes. In examining tumor cell proliferation within metastases, we noted a 3.8-fold increase (P<0.0001) in NGP-N1D+BV compared with NGP-LacZ+BV (Fig. 4A). Examination of large metastases (>300 μm) demonstrated increased vasculature in NGP-N1D+BV as seen with endomucin (Fig. 4B, 3-fold, P<0.05), collagen IV (Supplementary Fig. S7A), and the pericyte markers NG2 (Fig. 4B, P<0.01), and αSMA (not shown).

Liver metastases formed after NGP intracardiac injections were predominantly large (>300 μm, Fig. 2D). NGP-N1D liver metastases (NGP-N1D or NGP-N1D+BV) displayed enhanced angiogenic vasculature by collagen IV (Supplementary Fig. S7B), endomucin (Fig. 4C) and ICAM-1 (not shown), and αSMA (Fig. 4C), compared with NGP-LacZ or NGP-LacZ+BV. Similar results were obtained after intracardiac injection of SH-SYSY-N1D, with Notch blockade markedly increasing vascularity by ICAM-1 (Supplementary Fig. S7C, 13-fold, P<0.0001) and αSMA (Supplementary Fig. S7C, 12-fold, P<0.0001). Thus, Notch blockade increases liver metastasis angiogenesis both at the initial step of SEC sprouting and the later step of pericyte recruitment.

Notch inhibition activates HSCs

HSCs are often considered liver pericytes, and can contribute to tumor angiogenesis (22). Increased recruitment of NG2/αSMA (+) cells to the metastatic vasculature suggests the possibility that Notch blockade affects HSCs. Desmin has been widely used as a marker for quiescent HSCs (23, 24), with αSMA and NG2 commonly regarded as typifying activated HSCs (10). We therefore performed double-label IHC for desmin and αSMA. In normal mouse liver, desmin(+) HSCs are seen in peri-sinusoidal locations and are αSMA(−), indicating quiescence (Supplementary Fig. S8A). After intracardiac injection of NGP cells, quiescent desmin(+)/αSMA(−) HSCs are seen within adjacent liver in NGP-LacZ and NGP-N1D (Supplementary Fig. S8B). Desmin and αSMA signals were scant within NGP-LacZ metastases. In contrast, markedly increased desmin(+)/αSMA(+) signal was detected within NGP-N1D metastases. These results suggest that Notch inhibition activates HSCs, leading to their recruitment into metastases.

Decrease in host Notch1 promotes liver metastasis

To evaluate the role of host Notch1 signaling in liver metastasis, mice with heterozygous deletion of Notch1 (17) were crossed to immunodeficient Rag2−/−, Il2rg−/−, eGFP+/+ mice (Fig. 5A). SECs of Notch1+/−/ mice had a 3-fold higher proliferation rate

Figure 4. Notch inhibition increases vascularity of liver metastases. A, top, small liver metastases (<300 μm) immunostained for endomucin. Arrows, vascular sprouts. Liver metastases with endomucin(+) sprouts were counted; P<0.0001, Fisher exact test; bar, 50 μm. Bottom, proliferation determined by phosphorylated histone H3 (pH3); mean ± SD; **, P<0.0001. B, large metastases (>300 μm) immunostained for endomucin (top) and NG2 (bottom). Green autofluorescence distinguishes RBC within metastases; mean ± SD; *, P<0.05; **, P<0.01; bar, 100 μm. C, liver metastases from the NGP intracardiac experiment were immunostained for endomucin (top), and αSMA (bottom); mean ± SD; *, P<0.05; **, P<0.01; ***, P<0.001; bar, 200 μm. Nuclei shown by DAPI. Dashed line, metastases.

www.aacrjournals.org Cancer Res; 75(8) April 15, 2015 1597
Published OnlineFirst March 5, 2015; DOI: 10.1158/0008-5472.CAN-14-1493
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as seen with bromodeoxyuridine (BrdUrd; Fig. 5B, P < 0.0001), and a 14-fold increase in ICAM-1 expression as compared with littermate Notch1+/+ (Fig. 5B, P < 0.0001).

To determine whether deficient Notch1 signaling would promote liver metastasis, Notch1+/+ and Notch1+/− mice had intracardiac injection of NGP (106 cells) and were sacrificed at 8 weeks. Notch1+/− mice had markedly increased liver metastatic burden compared with littermate Notch1+/+ mice (Fig. 5C), with a 711-fold increase in liver flux (Fig. 5D, P < 0.01). Histologic examination demonstrated large metastases in Notch1+/− mice (2.79 ± 1.82 mm, mean ± SD), but a paucity of metastases in Notch1+/+ mice (solitary metastasis 0.17 mm), which precluded further analysis of the vasculature.

To increase the number of metastatic liver lesions in Notch1+/+ controls, intracardiac injection of 10-fold more NGP cells (107) was performed and mice sacrificed after 3 weeks. ICAM-1+ vessels in liver metastases from Notch1+/− mice were increased 16-fold compared with Notch1+/+ controls (Fig. 5E, P < 0.0001). Thus, deficient host Notch1 signaling is permissive for liver metastasis and increases early angiogenesis.

Notch regulates the ICAM-1 expression in SECs

Our results demonstrate increased ICAM-1+ vasculature in the livers of Notch1+/− mice and in the hepatic metastases from Notch1+/− mice and N1D-expressing tumors. To determine whether expression of ICAM-1 in SECs is regulated by Notch1, SECs were isolated from mouse liver. mSECs express Notch1, 2, 4, and DLL1, DLL4, JAG1, and JAG2 (Supplementary Fig. S9A). Notch1 knockdown increased the expression of ICAM-1 in mSECs, but not in human umbilical vein endothelial cells (HUVEC; Supplementary Fig. S9B). Conversely, expression of Notch1-intracellular domain (N1IC; ref. 25), decreased the expression of ICAM-1 (Supplementary Fig. S9C). These results demonstrate that ICAM-1 is regulated in SEC by Notch1.

Notch inhibition increases ICAM-1 but not the retention of tumor cells

ICAM-1 has previously been shown to promote the arrest of tumor cells in liver sinusoidal endothelium (26). We therefore speculated that attenuation of Notch signaling, by upregulating ICAM-1, might increase tumor cell retention within the liver. To address this possibility, we used adenovirus expressing either N1D (AdN1D) or control Fc (AdFc). Injection of AdN1D into nude mice, leads to hepatocyte infection, increased liver ICAM-1 expression (Supplementary Fig. S10A and S10B), and secretion of N1D into the circulation (Supplementary Fig. S10C). Three days after AdFc or AdN1D infection, 105 NGP cells were injected intracardially, and 24 hours later mice were sacrificed and liver metastasis quantified (Supplementary Fig. S10C). There was no difference in bioluminescence between AdFc and AdN1D, indicating that Notch inhibition, despite increasing ICAM-1, does not increase the retention of tumor cells within the liver. If, however, mice are sacrificed 6 weeks later, increased liver metastasis is observed (34-fold, P < 0.05, Supplementary Fig. 5C).
Blockade of both DLL and JAGGED is required for progression of liver metastases

We have recently developed Notch1-decoy variants (Fig. 6A): N11-24 decoy containing EGF repeats 1 to 24 blocks both DLL and JAGGED ligands; N11-13 decoy containing EGF repeats 1 to 13, specific for DLL ligands; and N110-24 decoy containing EGF repeats 10 to 24 blocks JAGGED ligands. Blockade of both DLL and JAGGED ligands increases sprouting of mSECs, whereas the ability of NGP-N11-24 to block JAGGED inhibition promoted development of early liver metastases (Fig. 6D). This suggests that DLL1/4 blockade promotes initial vascularization with ICAM-1(+) SECs. NGP-N11-24 metastases, however, displayed a 3.1-fold increase in αSMA(+) vasculature (Fig. 6D). Double-label IHC for desmin and αSMA demonstrated colocalization (arrows); inset (magnification, ×2); bar, 50 μm.

Histologic examination showed scant metastasis for NGP-Fc (74-fold, P < 0.001), whereas multiple metastases were seen with NGP-N11-24 decoy (296 vs. 246 μm, P < 0.05), with an increase in very large metastases (≈600 μm, P = 0.03, Fisher exact test).

IHC demonstrated no difference in ICAM-1(+) vasculature between NGP-N11-24 and NGP-N11-13 (Fig. 6D). This suggests that DLL1/4 blockade promotes initial vascularization with ICAM-1(+) SECs. NGP-N11-24 metastases, however, displayed a 3.1-fold increase in αSMA(+) vasculature (Fig. 6D). Double-label IHC for desmin and αSMA demonstrated colocalization in NGP-N11-24 metastases, but not in NGP-N11-13 metastases. Thus, DLL inhibition promoted development of early liver metastases by SECs, whereas the ability of NGP-N11-24 to block JAGGED increased activation and recruitment of HSC to the metastatic vasculature.

Blockade of DLL ligands increases sprouting of mSECs

Our in vivo results indicate that blockade of DLL, but not JAGGED, induced initial sprouting of vessels into hepatic metastases. To examine DLL specificity for sprouting, we performed in vitro assays with mSECs. Dextran beads were coated with SECs expressing N1-decoy variants, and assessed for sprout formation. On day 3, SECs expressing N1-124 or N11-13 decoy markedly increased sprouting per bead and sprout length (Fig. 7A–C), compared with either Fc or N11-24 decoy (similar results day 6, not shown). This DLL-specific effect on sprouting is not due to a...
selective effect on SEC proliferation, as both N11–13 and N110–24 increase proliferation (Supplementary Fig. S11C). Thus, SEC sprouting is mediated by inhibition of DLL-mediated signaling.

JAG1 blockade increases HSC migration

Our in vivo results suggest that blockade of JAGGED ligands promotes HSC recruitment, raising the possibility that JAG1 expressed on NGP cells may interact with Notch receptors on HSCs (28). GFP-labeled HSCs (which express Notch2/3, data not shown), were cocultured with NGP cells expressing Notch1 decoys and migration assessed in a Boyden chamber assay (Fig. 7D). Only decoys that blocked JAGGED, N11–24 decoy and N110–24 decoy, but not DLL, increased HSC migration (Fig. 7E). FACS isolation of HSC–GFP cells, after coculture with NGP cells, demonstrated that blockade of JAGGED, inhibited Notch signaling, as shown by decreased Hes1, Hey1, αSMA, and β-actin of FACS-sorted HSCs. Migration of HSC–GFP, at 48 hours, cocultured with NGP or NGP-siJag1; mean ± SD; *, P < 0.05; **, P < 0.01; ***, P < 0.001. H, model of Notch blockade effects on hepatic microenvironment. Quiescent, SECs are maintained in quiescence by Notch1 signaling. HSCs in the space of Disse are maintained in quiescence by Notch2/3 signaling from JAG1-expressing hepatocytes (28). SEC proliferation and sprouting, soluble NID inhibits DLL4 activation of Notch1 in SECs, leading to expression of ICAM-1 and proliferation of SECs, and subsequent sprouting into tumor micrometastases (<300 μm); HSC Recruitment, NID blocks tumor JAG1, promoting HSC activation [indicated by αSMA(+) expression], migration, and recruitment to the vasculature of macrometastases (>300 μm).

Figure 7.

DLL Blockade promotes SEC sprouting, whereas JAGGED blockade promotes HSC migration. A, fibrin bead assay with mSEC transfected with Fc, N11–24 decoy, N11–13 decoy, and N110–24 decoy. Arrows, sprouts; bar, 200 μm. B, sprouts per bead. C, sprout length; mean ± SD; *, P < 0.05; **, P < 0.01; ***, P < 0.001. D, in the top chamber of a Boyden chamber, HSC–GFP cells were cocultured with NGP cells expressing Notch1 decoys. E, migration of HSC–GFP measured at 48 hours; mean ± SD; *, P < 0.05; **, P < 0.01. F, immunoblot analysis of Hes1, Hey1, αSMA, and β-actin of FACS-sorted HSCs. G, migration of HSC–GFP, at 48 hours, cocultured with NGP or NGP-siJag1; mean ± SD; ***, P < 0.001. H, model of Notch blockade effects on hepatic microenvironment. Quiescent, SECs are maintained in quiescence by Notch1 signaling. HSCs in the space of Disse are maintained in quiescence by Notch2/3 signaling from JAG1-expressing hepatocytes (28). SEC proliferation and sprouting, soluble NID inhibits DLL4 activation of Notch1 in SECs, leading to expression of ICAM-1 and proliferation of SECs, and subsequent sprouting into tumor micrometastases (<300 μm); HSC Recruitment, NID blocks tumor JAG1, promoting HSC activation [indicated by αSMA(+) expression], migration, and recruitment to the vasculature of macrometastases (>300 μm).

Discussion

Given its important role in tumorigenesis, the Notch pathway has aroused great interest as a therapeutic target. Using intrarenal and intracardiac metastasis models with different Notch inhibitors, used in preclinical and clinical studies, we have shown that inhibition of Notch signaling in liver stromal microenvironment promotes metastatic growth. Notch blockade produced significantly larger and more highly vascularized hepatic metastatic lesions in neuroblastoma cell lines NGP and SH-SY5Y, and the breast cancer cell line MDA-MD-231. In these studies, we uncover a novel function of Notch, and show that inhibition of Notch signaling increases metastatic angiogenesis to support growth and enlargement.

Our results demonstrate that Notch inhibition is sufficient to promote liver metastases and increased vascularization, and is not limited by concurrent VEGF blockade. As bevacizumab binds selectively to human but not murine VEGF, angiogenesis in liver metastases is not dependent on tumor-derived VEGF. VEGF blockade, however, by slowing growth of the primary tumor and prolonging survival, may lead to an apparent increase in liver metastatic burden by extending the time period in which metastases can grow.
Notch1 inhibition is not known. However, the unique characteristics of liver SEC as well as specific liver metastasis in our mice models. Our data also show that knockdown of Notch1, in vitro, induces ICAM-1 in SECs but not in HUVECs. The reason for the selectivity of Notch1 inhibition is not known. However, the unique characteristics of liver SEC as well as specific anatomical and environmental factors may render them more proliferative upon Notch1 inhibition.

Liver metastasis progression is a multistep process that begins with the arrest of tumor cells in sinusoids, with ICAM-1 promoting the adhesion of tumor cells to liver sinusoidal endothelium (26). Our in vivo tumor cell liver adhesion assays showed that pretreatment with N1D adenosine elevated liver ICAM-1, but did not increase the retention of tumor cells in the liver. Therefore, attenuation of Notch signaling appears to affect another step of early survival or growth of tumor cells, rather than adhesion.

A role for Notch signaling in maintaining HSCs in a quiescent state has also been proposed (28). Cancer cells by releasing paracrine factors can create a prometastatic microenvironment within the liver, promoting the activation of HSCs (26, 31). Our results indicate that tumor-derived JAG1 has the opposite effect, suppressing the activation of HSCs. Blockade of both JAGGED and DLL (N1 1/3 decoy), produced larger metastases with increased HSC recruitment and activation, compared with DLL blockade only (N1 1/3 decoy). Consistent with these results, our in vitro studies demonstrated that HSCs, in which tumor-derived JAGGED was blocked, displayed increased migration and evidence of an activated state. Similarly, increased HSC migration was observed when HSCs were cocultured with NGP cells in which JAG1 was silenced. Our studies, thus, suggest that inhibition of tumor-derived JAG1 signaling disrupts Notch-mediated quiescence, promoting HSC activation and migration, and subsequent vascular maturation and enlargement of micrometastases (Fig. 7G).

In some settings, DLL4 and JAG1 can exert opposing effects on angiogenesis (32). DLL4-mediated Notch signaling inhibits the sprouting of endothelial tip cells in growing blood vessels. In contrast, JAG1 overexpression inhibits DLL4 signaling in ECs and increases sprouting angiogenesis (32). In our studies, however, we found that the Notch ligands in the liver have distinct rather than opposing effects, with both acting to inhibit metastatic progression.

Our findings have potentially serious implications for Notch inhibition therapy. Because the critical role of this pathway in regulating tumor angiogenesis, Notch components have emerged as an attractive potential target, and Notch inhibitors are entering clinical cancer trials. Our study shows that treatment with N1-decoys or the GSI PF-03084014 increase hepatic metastasis. These findings raise the concern that Notch signaling blockade could disrupt normal liver homeostasis and quiescence by pathologic activation of SECs and HSCs, creating a host microenvironment favorable for the metastatic growth of cancer cells.

Disclosure of Potential Conflicts of Interest
C.J. Shawber and J. Kitajewski report receiving commercial research support from Eisai, Ltd. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): D. Banerjee, A. Garcia, T. Kangsamaksin, J. Andrews, D.J. Yamashiro
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): D. Banerjee, S.L. Hernandez, A. Garcia, T. Kangsamaksin, D.J. Yamashiro
Writing, review, and/or revision of the manuscript: D. Banerjee, S.L. Hernandez, T. Kangsamaksin, A. Kadenhe-Chiweshe, C.J. Shawber, J.K. Kitajewski, J.J. Kandel, D.J. Yamashiro
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): S.L. Hernandez, E. Shihori, L.A. Foster, N. Wei, D.J. Yamashiro
Study supervision: C.J. Shawber, D.J. Yamashiro

Acknowledgments
The authors thank Arul Thirumoorthi, Minji Kim, Shuobo Zhang, and Jason Mitchell for technical assistance, and Robert Schware for assistance with HSC experiments.

Grant Support
These studies were supported by NIH SB01CA124644 (D.J. Yamashiro), 3RO1CA124644-03S1 (A. Kadenhe-Chiweshe), R01CA136673 (C.J. Shawber and J.K. Kitajewski), Pediatric Cancer Foundation (J.J. Kandel and D.J. Yamashiro).
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Received May 21, 2014; revised January 24, 2015; accepted January 29, 2015; published OnlineFirst March 5, 2015.

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**References**


Notch Suppresses Angiogenesis and Progression of Hepatic Metastases


Cancer Res 2015;75:1592-1602. Published OnlineFirst March 5, 2015.

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