Ibrutinib Exerts Potent Antifibrotic and Antitumor Activities in Mouse Models of Pancreatic Adenocarcinoma

Daniel Massó-Vallès,1 Toni Jauset,1 Erika Serrano1, Nicole M. Sodir2, Kim Pedersen1, Nesrine I. Affara3, Jonathan R. Whitfield1, Marie-Eve Beaulieu1, Gerard I. Evan2,3, Laurence Elias4, Joaquín Arribas1,5, and Laura Soucek1,5

Abstract

Pancreatic ductal adenocarcinoma (PDAC) is characterized by a dense stromal fibroinflammatory reaction that is a major obstacle to effective therapy. The desmoplastic stroma comprises many inflammatory cells, in particular mast cells as key components of the PDAC microenvironment, and such infiltration correlates with poor patient outcome. Indeed, it has been hypothesized that stromal ablation is critical to improve clinical response in patients with PDAC. Ibrutinib is a clinically approved Bruton’s tyrosine kinase inhibitor that inhibits mast cells and tumor progression in a mouse model of β-cell tumorigenesis. Here, we show that ibrutinib is highly effective at limiting the growth of PDAC in both transgenic mouse and patient-derived xenograft models of the disease. In these various experimental settings, ibrutinib effectively diminished fibrosis, extended survival, and improved the response to clinical standard-of-care therapy. Our results offer a preclinical rationale to immediately evaluate the clinical efficacy of ibrutinib in patients with PDAC. Cancer Res; 75(8); 1675–81. © 2015 AACR.

Introduction

Pancreatic cancer is the fourth most common cause of cancer death in the Western world and, although cancer mortality has in general been recently improving, it has a uniquely negative mortality trend in both sexes (1, 2). It is usually diagnosed at an advanced stage and characterized by poor prognosis. According to the American Cancer Society, the overall survival rates at 1 and 5 years are only 25% and 6%, respectively. The 5-year survival for localized disease is approximately 20%, and the median survival is 10 and 6 months for advanced and metastatic disease (which affects 80% of individuals), respectively (www.cancernetwork.com). Hence, there is an urgent need for new and effective therapies. Pancreatic adenocarcinoma is characterized by dense desmoplasia, composed of extracellular matrix, endothelial cells, immune cells, fibroblasts, and stellate cells. This epithelial and stromal compartment appears to enhance the aggressive nature of the disease and its resistance to therapy. Indeed, the dense stromal fibroinflammatory reaction results in decreased blood supply, poor drug delivery, and hypoxia (4). Hence, therapies targeting the stroma may be key to improving the clinical outcomes for patients with pancreatic cancer.

Analysis of human pancreatic ductal adenocarcinoma (PDAC) samples reported that mast cell infiltration correlated with higher tumor grade and worse survival (5), and high mast cell numbers at the intratumoral border correlated with the presence of lymphatic and microvascular invasion, as well as with lymph node metastasis (6). We have previously utilized a mouse model of pancreatic β-cell tumorigenesis to show that mast cells are required for angiogenesis and the macroscopic expansion and maintenance of tumors (7), thereby proposing mast cell targeting as a therapeutic strategy in such tumors. Since then, other groups have shown that this role for mast cells might be conserved in PDAC. For example, an influx of mast cells was observed in a mouse model of pancreatic cancer expressing high levels of KRASG12V (8) and, importantly, pancreatic tumors orthotopically transplanted in mast cell-deficient mice grew more slowly than in control mice (8), conferring a survival advantage compared with tumors grown in mast cell proficient animals.

PCI-32765 (also known as ibrutinib) is a novel inhibitor of Bruton’s tyrosine kinase (BTK) approved for the treatment of mantle cell lymphoma and chronic lymphocytic leukemia (CLL) patients who have received at least one previous therapy, and CLL patients with 17p deletion (9). It is known to be capable of inhibiting mast cell or basophil degranulation (10). Recently, we validated ibrutinib as a potent systemic mast cell blocker in our mouse model of insulinoma, triggering collapse of tumor vasculature and tumor regression (11). Hence, in the present study, we sought to investigate the potential therapeutic impact of ibrutinib in a preclinical model of PDAC.
Materials and Methods

IHC and collagen staining

For IHC analyses, tissue samples were fixed overnight in neutral pH–buffered formalin, embedded in paraffin, and sectioned (5 μm). Sections were then deparaffinized, rehydrated, and microwaved for 1 minute in 0.01 mol/L citrate buffer (pH 6.0) for antigen retrieval. Primary antibodies [rat monoclonal anti–CD11b (clone M1/70), eBioscience; rabbit polyclonal anti–CD11b, Novus Biologicals; rat monoclonal anti-F4/80 (clone C13-1), rabbit monoclonal anti-Ki-67 (clone SP6), Neomarkers] were applied for 2 hours in blocking buffer (2.5% BSA, 5% goat serum, and 0.3% Triton X-100 in PBS), followed by Vectastain ABC kit and DAB reagents (Vector Laboratories). We identified mast cells by using 1% toluidine blue dissolved in ethanol. Picrosirius Red Stain Kit (Polysciences, Inc.) was used to stain for collagen types I and III according to manufacturer instructions. Images were obtained with an Axiovert 100 TV inverted fluorescence microscope (Zeiss) and Open Lab 3.5.1 software, with an Axiovert 100 inverted microscope (Zeiss) equipped with a Hamamatsu Orca digital camera or with an Olympus FFX100 microscope and FFX-BISW software.

Flow cytometry

Following resection, PDAC tumors isolated from PBS-perfused mice were immediately placed in ice-cold PBS, followed by manual mincing using scissors and a 20-minute enzymatic digestion with 1.25 mg/mL collagenase type IV (Roche), 0.1% trypsin inhibitor, and 50 U/mL DNase I (Roche) in serum-free DMEM (Life Technologies) at 37°C with continuous stirring. Single cell suspensions were then prepared by passing tissue through 70 μm nylon strainers (BD Biosciences). Cells were incubated for 30 minutes at 4°C with rat anti-mouse CD16/CD32 mAb (1:250, clone 2.4G2, BD Bioscience), in PBS, which also contained Live/Dead Aqua stain (1:250, Invitrogen) to differentiate between viable and dead cells. Cells were then incubated for 30 minutes in PBS containing 1.0% BSA (Sigma) and 2 mmol/L EDTA with 100 μL of fluorophore-conjugated anti-mouse antibodies (dilution; clone): PE-Cy7-CD45 (1:800; 30-F11), PerCP-Cy5.5-CD3ε (1:400; 145-2C11), PerCP-Cy5.5-CD19 (1:200; 6D5), PerCP-Cy5.5-CD49b (1:400; DX5), Alexa 700-CD11b (1:400; M1/70), APC-C70-CD11c (1:200; N418), eFluor450-MHCII (1:800; M5/14.15.2), APC-Ly6C (1:800; HK1.4), PE-Ly6G (1:400; 1A8), and PE-Cy5-F4/80 (1:400; BMB; eBioscience or Biogendes). Cells were then washed once in PBS containing 1.0% BSA (Sigma) and 2 mmol/L EDTA, followed by fixation with BD Cytofix for 30 minutes on ice. Following a final wash, the cells were stored at 4°C until data acquisition using a LSRII using FACS Diva software (BD Biosciences). Analysis was performed using Flowjo software program (Tree Star Inc.).

Animal studies

All the animal studies were performed in accordance with the ARRIVE guidelines and the 3 Rs rule of Replacement, Reduction and Refinement principles. Mice were housed and treated following the protocols approved by the Institutional Animal Care and Use Committee at the University of California, San Francisco (San Francisco, CA) and by the CEEA (Ethical Committee for the Use of Experimental Animals) at the Vall d’Hebron Institute of Oncology, Barcelona, Spain. Mouse weights were recorded for every experiment (Supplementary Fig. S1).

Statistical analysis

Statistical analysis was carried out by two-tailed Mann–Whitney test (for IHC counts), two-tailed unpaired t test (for flow-cytometric data), χ² test of homogeneity (for collagen staining), and log-rank test (for Kaplan–Meier survival curves). SE (±SEM) and SD (±SD) are either represented in the graphs or following the means of all measures, as stated in figure legends. Statistical analysis was performed using GraphPad Prism 6.
Results
Ibrutinib strongly affects the tumor microenvironment in PDAC

We made use of a p53ER/ER;LSLKRasG12D;Pdx1-cre mouse model that harbors a Cre-activated KRasG12D allele inserted into the endogenous KRas locus, combined with pancreas-specific Pdx1-driven Cre recombinase activity (12, 16). The disruption of p53 signaling in combination with KRAS mutation leads to rapid tumorigenesis and histopathological features typical of human pancreatic adenocarcinoma within 8 weeks. At this stage, we treated the animals with ibrutinib, added at a dose of 35 mg/kg/d to their drinking water, or with...
vehicle control. No detectable side effects of the treatment were observed and the animals did not lose weight throughout (Supplementary Fig. S1). The animals were euthanized 4 weeks later and their pancreata harvested for analysis. As expected, mast cells were still recruited to the tumor stroma, but their degranulation was efficiently inhibited by ibrutinib treatment (Supplementary Fig. S2A). Strikingly, tumors treated with ibrutinib displayed a significant reduction in proliferation rate (Fig. 1A; 67.8 ± 29.5% Ki-67 positive in untreated vs. 14.9 ± 7.0% treated animals). CD11b-positive cells, as well as F4/80-positive macrophages were both reduced as expected, mast cells were still recruited to the tumor stroma, rather than a direct effect on pancreatic cell survival—rather than a direct effect on pancreatic cell survival—as the best candidate for ibrutinib’s anti-tumorigenic effect in PDAC (Supplementary Fig. S3). With this line of reasoning in mind and to verify the hypothesis that mast cell interference played a role in the antifibrotic effect displayed by ibrutinib, we performed two independent control experiments. In the first one, we made use of sodium cromoglycate (cromolyn), a well-characterized blocker of mast cell degranulation and inflammmogen release (7, 17). We treated the p53ER/ER;LSKRasG12D;Pdx1-cre mice intraperitoneally (i.p.) with a daily injection of 10 mg per kg (bodyweight) of cromolyn, starting at 8 weeks of age. The animals were euthanized 4 weeks later and their pancreata analyzed. We verified that cromolyn treatment blocked degranulation (Supplementary Fig. S2B). Strikingly, treatment with cromolyn recapitulated the antifibrotic effect displayed by ibrutinib (Fig. 2A and B). Once again, this effect was accompanied by a reduction in F4/80+ cells and a significant decrease in CD11b+ cells in the tumor stroma (Fig. 2C and D). Although cromolyn was also described to display some cytotoxic effect on pancreatic cancer cells (18), the overlapping effect of ibrutinib and cromolyn on collagen deposition strongly suggests

The antifibrotic effect of ibrutinib is mast cell dependent

To verify that this effect on fibrosis was specifically due to mast cell interference, we first asked whether ibrutinib could have any direct activity on tumor cells per se. To address this question, we made use of four different pancreatic cancer cell lines and treated them with increasing amounts of ibrutinib in vitro, following their number over time. We chose four cell lines presenting a similar mutational profile to the one displayed by our mouse model (i.e., KRAS and p53 mutation): MIA PaCa-2, YAPC, PSN-1, and PA-TU898T. Importantly, none responded significantly to the treatment, pointing to the activity of ibrutinib on tumor stroma—rather than a direct effect on pancreatic cell survival—as the best candidate for ibrutinib’s anti-tumorigenic effect in PDAC (Supplementary Fig. S3).

Figure 2.
Mast cell function inhibition by cromolyn diminishes tissue fibrosis. A, 8-week-old p53ER/ER;LSKRasG12D;Pdx1-cre mice were treated with intraperitoneal sodium cromoglycate (n = 6) or vehicle control (n = 5) for 4 weeks. Histologic analysis of pancreata by Picrosirius Red staining shows reduced collagen deposition in cromolyn-treated mice compared with vehicle-treated control animals. Five animals per condition and at least five sections per animal were analyzed. Red staining shows collagen in the tumor stroma. B, scoring of treated (n = 30) and untreated (n = 25) tumors according to their collagen content shows a significant difference between groups (**, P = 0.0088). Sections stained with Picrosirius Red staining were scored blindly and assigned to three categories (+, ++, or +++). The percentages of tumors classified into each category are shown. A χ² test of homogeneity was utilized for statistical analysis of the data. C, quantification of F4/80-positive cells per microscopic field in the tumor area shows a nonsignificant reduction in cromolyn-treated versus vehicle-treated mice. Results shown represent mean ± SEM. Five animals per condition and four sections per animal were analyzed. Statistical significance was determined via two-tailed Mann-Whitney test. D, quantification of CD11b-positive cells per microscopic field in the tumor area shows a significant reduction in cromolyn-treated versus vehicle-treated mice (**, P = 0.0021, two-tailed Mann-Whitney test). Results shown represent mean ± SEM. Five animals per condition and four sections per animal were analyzed.
that mast cell inhibition is the most likely explanation for this phenomenon. This finding is consistent with the recently published work showing that mast cells could exacerbate the cellular and extracellular dynamics of the tumor microenvironment found in PDAC (19).

In a second set of experiments, we made use of subcutaneous patient-derived xenografts (PDX) of a patient-derived tumor in NOD/SCID mice (Fig. 3A). These mice are defective for both B- and T-cell function. Once again, ibrutinib prevented mast cell degranulation (Supplementary Fig. S2C) and led to a striking reduction in tissue fibrosis detected by Picosirius Red staining (Fig. 3A and B), excluding B- and T-cell signaling modulation as ibrutinib’s main mechanism of action in this context. The animals also showed a significant survival advantage when treated with ibrutinib compared with untreated littermates (Fig. 3C). Interestingly, IHC analysis of F4/80 and CD11b-positive cells showed a slight but nonsignificant reduction of both cell types (Fig. 3D and E).

These experimental models combined point to mast cells as crucial players in stimulating collagen deposition, in line with previously published data showing that mast cell tryptase might sustain liver fibrosis by promoting stellate cell proliferation and collagen synthesis (20) and with the concept that mast cells could be the culprit in various fibrotic diseases (21, 22).

Ibrutinib is an effective therapy and improves the outcome of standard care in a transgenic mouse model of PDAC

In PDAC, dense stromal fibrosis is a considerable obstacle to therapeutic intervention. Consequently, we hypothesized that the antifibrotic effect of ibrutinib represented a promising therapeutic opportunity. We therefore performed two independent survival experiments in the p53^R172H;LSL-Ras^G12D;Pdx1-cre mouse model:
Advantage to the treated animals (survival) of ibrutinib plus gemcitabine compared with untreated controls (Fig. 4A). In the second experiment, gemcitabine alone resulted in slight toxicity, although ibrutinib ameliorated the toxicity and significantly extended survival compared with gemcitabine alone (Fig. 4B), confirming our hypothesis that the standard-of-care outcome can be improved by the addition of ibrutinib.

Discussion

Our work provides preclinical evidence for the use of ibrutinib, which is already approved for clinical use, in the treatment of PDAC, based on the observed systemic mast cell inhibitory effects of ibrutinib as a potentially new and potent antifibrotic drug. In line with our results, it has been shown that ibrutinib specifically inhibits the release of IL8, MPC-1, and TNFα from mast cells (10). All these factors have a known role in fibrosis: IL8 is elevated in the serum of idiopathic pulmonary fibrosis patients (23) and correlates with the degree of fibrosis in infants with chronic liver disease (24); MCP-1 is a key determinant in the development of skin fibrosis, it could influence collagen fiber formation in mice (25) and is involved in the development of interstitial fibrosis in a mouse model of crescentic nephritis (26); finally, TNFα, which has a controversial role in fibrotic disease (27), promotes survival of activated hepatic stellate cells in vitro and in vivo, increases liver fibrosis in mice (28), mediates the transition from pulmonary inflammation to fibrosis (29) and its inhibition reduces fibrosis in a mouse model of autoimmune thyroiditis (30). Therefore, in addition to its beneficial role in reducing tumor fibrosis, the use of ibrutinib could potentially be extended to the treatment of various fibrotic diseases, such as liver fibrosis (31) or chronic pancreatitis (32).

The decrease in the leukocytic component of the tumor microenvironment (CD11b-positive cells and F4/80-positive macrophages) after treatment with ibrutinib and cromolyn could be explained by the fact that several chemokines produced by mast cells, such as IL6, are known to be potent stimulants for monocytic cell migration and macrophage activation (33), so that inhibiting mast cells could prevent their recruitment.

Intriguingly, recent work suggests that ibrutinib could be redeployed to treat lung cancer owing to inhibitory activity against the EGFR (34). However, EGFR mutations are not frequent in PDAC and none of our experimental systems presented EGFR alterations (data not shown).

It remains possible that additional anti-inflammatory effects of ibrutinib (specifically inhibition of B-cell signaling) might contribute to its therapeutic impact and modulation of other inflammatory components, even though we have demonstrated that such signaling is not required for decreased fibrosis. Further study of the effect of ibrutinib on other members of the complex PDAC stroma will be required to fully characterize its precise mechanism of action.

Disclosure of Potential Conflicts of Interest

L. Elias is the senior director of Pharmacyclics Inc., has ownership interest (including patents) in Pharmacyclics Inc. (Stockholder), and is a consultant/advisory board member for Pharmacyclics Consultant (Contractor). L. Soucek is a consultant/advisory board member for Pharmacyclics Inc. No potential conflicts of interest were disclosed by the other authors.

Authors’ Contributions

Conception and design: D. Massó-Vallés, T. Jaquet, N.M. Sodir, L. Elias, J. Arribas, L. Soucek

Development of methodology: D. Massó-Vallés, K. Pedersen, G. Evan, L. Soucek

Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): D. Massó-Vallés, T. Jaquet, E. Serrano, N.M. Sodir, K. Pedersen, N.I. Affara, L. Soucek

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): D. Massó-Vallés, T. Jaquet, E. Serrano, N.I. Affara, M.-E. Beauclerc, L. Elias, L. Soucek

Writing, review, and/or revision of the manuscript: D. Massó-Vallés, T. Jaquet, J.R. Whitfield, L. Elias, J. Arribas, L. Soucek

Administrative, technical, or material support (i.e., acquiring data, constructing databases): D. Massó-Vallés, E. Serrano, N.M. Sodir, G. Evan, L. Soucek

Study supervision: D. Massó-Vallés, J.R. Whitfield, L. Soucek

Acknowledgments

The authors thank Lisa Coussens for her valuable feedback and for useful reagents. The authors also acknowledge Dr. Joaquin Balcells and Dr. Maria Salcedo for their help in the collection and analysis of the patient derived tumor samples.

Grant Support

This work was supported by grants from the Miguel Servet Program (L. Soucek), the FERO Foundation (L. Soucek and D. Massó-Vallés), the Bear...
Ibrutinib Reduces Fibrosis in PDAC

Necessities Pediatric Cancer Foundation (L. Soucek), Secretaria d’Universitats i Recerca del Departament d’Economia i Coneixement de la Generalitat de Catalunya (D. Massó-Valles), the European Social Fund (D. Massó-Valles), and Asociacion Española Contra el Cáncer (J. Arribas).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received September 29, 2014; revised January 9, 2015; accepted January 28, 2015, published OnlineFirst April 15, 2015.

References

Ibrutinib Exerts Potent Antifibrotic and Antitumor Activities in Mouse Models of Pancreatic Adenocarcinoma


Updated version
Access the most recent version of this article at:
http://cancerres.aacrjournals.org/content/75/8/1675

Supplementary Material
Access the most recent supplemental material at:
http://cancerres.aacrjournals.org/content/suppl/2015/04/24/75.8.1675.DC1

Cited articles
This article cites 33 articles, 9 of which you can access for free at:
http://cancerres.aacrjournals.org/content/75/8/1675.full#ref-list-1

Citing articles
This article has been cited by 5 HighWire-hosted articles. Access the articles at:
http://cancerres.aacrjournals.org/content/75/8/1675.full#related-urls

E-mail alerts
Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions
To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.