miR-214 and miR-148b Targeting Inhibits Dissemination of Melanoma and Breast Cancer

Francesca Orso1,2,3, Lorena Quirico1,2, Federico Virga1,2, Elisa Penna1,2, Daniela Dettori1,2, Daniela Cimino1,2,3, Roberto Coppo1,2, Elena Grassi1,2, Angela Rita Elia2, Davide Brusa4,5, Silvia Deaglio4,5, Maria Felice Brizzi6, Michael B. Stadler7, Paolo Provero1,2,8, Michele Caselle3,9, and Daniela Taverna1,2,3

Abstract

miR-214 and miR-148b have been proposed to antagonize the effects of each other in enabling or blocking metastasis, respectively. In this study, we provide evidence deepening their role and interrelationship in the process of metastatic dissemination. Depleting miR-214 or elevating miR-148b blocked the dissemination of melanoma or breast cancer cells, an effect that could be accentuated by dual alteration. Mechanistic investigations indicated that dual alteration suppressed passage of malignant cells through the blood vessel endothelium by reducing expression of the cell adhesion molecules ITGA5 and ALCAM. Notably, trans-endothelial migration in vitro and extravasation in vivo impaired by singly altering miR-214 or miR-148b could be overridden by overexpression of ITGA5 or ALCAM in the same tumor cells. In clinical specimens of primary breast cancer or metastatic melanoma, we found a positive correlation between miR-214 and ITGA5 or ALCAM along with an inverse correlation of miR-214 and miR-148b in the same specimens. Our findings define an antagonistic relationship of miR-214 and miR-148b in determining the dissemination of cancer cells via tumor–endothelial cell interactions, with possible implications for microRNA-mediated therapeutic interventions aimed at blocking cancer extravasation.

Introduction

Dissemination of primary tumor cells and the consequent formation of metastasis at distant organs are the main cause of cancer-related mortality. The currently available treatments (surgery, radio-, chemo-, and targeted therapy) mainly control primary tumors, but they only exert a mild effect on metastases, mostly because of resistance mechanisms activated by tumor cells. Therefore, it is crucial to identify key regulators of metastatic dissemination and to develop new targeted therapy approaches.

MicroRNAs (miRNAs) are small noncoding, single-stranded RNAs that act as negative regulators of gene expression. Increasing evidence shows that deregulation of miRNAs is a crucial event in tumor progression. Several miRNAs, including miR-137, miR-221/222, miR-182, miR-34a, and miR-214, have been found to be involved in melanoma malignancy by regulating key genes such as c-KIT, MITF, FOXO3, ITGB3, CCND1, p27Kip1, TFAP2, and ALCAM (1–3). Instead, let-7, miR-9, miR-10b, miR-21, miR-31, miR-146, miR-148b, miR-155, miR-200, and miR-221/222 are the main players in breast cancer dissemination (4–6). Therefore, it is essential to identify new miRNAs and understand how these small noncoding RNAs control the various steps of tumor malignancy and how they interact with one another to attempt miRNA-targeted therapies that can affect the molecular pathways involved in cancer progression. Several clinical trials based on the use of miRNA modulators in tumorigenesis have already been established (7).

We previously demonstrated that miR-214 promotes melanoma metastasis dissemination by increasing migration, invasion, extravasation, and survival of melanoma cells via a novel pathway involving the metastasis suppressor, TFAP2 transcription modulators (3). On the other hand, we found that miR-148b opposes breast cancer progression, acting directly on the integrin signaling players ITGA5, ROCK1, and PIK3CA/p110α (6). More recently, we showed that miR-214 downregulates miR-148b in tumor cells via TFAP2C with the consequent upregulation of miR-148b direct targets (2). In order to explore the relevance of miR-214 and miR-148b for miRNA-based therapeutic interventions in melanoma and breast cancer, we analyzed the dissemination of miR-214-depleted and miR-148b-overexpressing cells in mice, investigated how these small-RNAs influence cell metastatic traits and looked for the main molecular players involved in this process. We demonstrated that single or combined modulations of miR-214

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F. Orso and L. Quirico contributed equally to this article.

Corresponding Author: Daniela Taverna, MBC and Department of Molecular Biotechnology and Health Sciences, University of Torino, Via Nizza, 52, 10126 Torino, Italy. Phone: 39-011-670-6497; Fax: 39-011-670-6432; E-mail: daniela.taverna@unito.it

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(inhibition) and miR-148b (overexpression) significantly inhibit metastatization when tumor cells cross the vessel endothelium by decreasing the expression of ITGA5 and ALCAM, two adhesion molecules respectively involved in tumor-Extra-Cellular Matrix (ECM) and cell–cell interactions.

**Materials and Methods**

**Cell culture**

MA-2 and MC-1 cells were kindly provided by L. Xu and R.O. Hynes (8) and maintained as described in refs. 2 and 3. Human HBL-100 were from ATCC and 4175-TGL and SK-MEL-28 were kindly provided, respectively, by J. Massagué (9) and L. Poliseno and maintained in standard conditions. Human umbilical vein endothelial cells (HUVEC) were kindly provided by L. Primo (with GFP) or generated by M.F. Brizzi and maintained as described in refs. 2 and 3. All used cell lines were authenticated in the last 6 months by BMR Genomics, using the CELL ID System (Promega).

**Reagents and antibodies**

miR *precursors and inhibitors*. Pre-miR miRNA Precursor Negative Control #1, Pre-miR miRNA Precursor Hsa-miR-214 (PM12124), Pre-miR miRNA Precursor hsa-miR-148b (PM10264), Anti-miR miRNA Inhibitor Negative Control #1, Anti-miR miRNA Inhibitor Hsa-miR-214 (AM12124), Anti-miR miRNA Inhibitor hsa-miR-148b (AM10264) (all from Applied Biosystems). TaqMan MicroRNA assays for miRNA detection: Hsa-miR-214 ID 002306, Hsa-miR-148b ID 000471, U6 snRNA ID001973, U44 snRNA ID001904 (all from Applied Biosystems). Primary antibodies: anti-PIK3CA #4255 (Cell Signaling Technology), anti-GFP ab290 (Abcam), anti-ITGA5 pAb RM10 kindly provided by G. Tarone, anti-CD166/ALCAM mAb MOG/07 (Novoceastra Laboratories), anti-ROCK-1 pAb H-85, anti-hsp90 mAb F-8, anti-GAPDH pAb V-18, anti-ACTIN I-19 pAb (all from Santa Cruz Biotechnology), anti-tubulin mAb B5-1-2 (Sigma), anti-CD31 pAb (Becton Dickinson). Secondary antibodies: HRP-conjugated goat anti-mouse IgG, goat anti-rabbit IgG, donkey anti-goat IgG (all from Santa Cruz Biotechnology), goat anti-rat IgG Alexa-Fluor-568 and goat anti-rat IgG Alexa-Fluor-488 (Molecular Probes, Invitrogen Life Technologies). siRNAs: si-ITGA5 (Hs_ITGA5_5 siRNA), si-ALCAM (Hs_ALCAM_5 siRNA), and All Stars Negative Control siRNA were purchased from Qiagen.

**Sponge design and recombinant vector preparations**

miR-214 sponges were described in ref. 3. miR-148b-specific sponge sequences containing eight miRNA binding sites interrupted by 15-nts spacers were designed to be perfectly complementary to the miR-148b seed region, with a bulge position 9–12 to prevent undesired cleavage of the sponge RNA. Sponges were synthesized by DNA 2.0, cloned into pLenti vector plasmids, excised using flanking HindIII sites, blunted and subcloned into blunted BamHI and SalI sites, downstream of EGFP into plentiCMV-GFP-Puro (6585-S) vector (Addgene), giving rise to pLenti148-sponge/B. Nucleotides (in Supplementary Table S1) were verified by sequencing. pLentiR-empty, pLentiR-214, and pLentiR-148b expression vectors were described in refs. 3 and 6. The same miR-148b expression cassette was also subcloned in a pLein148-V5 expression vector (kindly provided by C.M. DiPersio, Albany Medical College, Albany, NY).

**Transient transfections, vectors, and generation of stable cell lines**

To obtain transient anti-miR or pre-miR or siRNA expression, cells were plated at 50% confluency and immediately transfected using HiPerFect Transfection Reagent (Qiagen), with 100 nmol/L anti-miR, 75 nmol/L pre-miR, or 100 nmol/L siRNA. For transient cDNA overexpression, cells were plated at 90% confluency and transfected 24 hours later using Lipofectamine2000 reagent (Invitrogen Life Technologies). For ITGA5 or ALCAM overexpression pEGFP-N3-ITGA5 (6) or pLVX-ALCAM (2) expression vectors were used. All stable cell lines were generated via lentiviral infection. Sponge vectors were obtained as described above. The plKO.1-shALCAM lentiviral vector was from Open Biosystems (RH53979). Lentiviruses were produced by calcium phosphate transfection of 15-μg vector plasmid together with 15-μg packaging (pCMVdR8.74) and 6-μg envelope (pMD2.G-VSVG) plasmids in 293T cells, according to Trono’s lab protocol (http://tronolab.epfl.ch), and supernatants were harvested 48 hours after transfection.

**Protein or RNA isolation, immunoblotting, qRT-PCRs for miRNA detection, and proliferation assays**

Total protein or RNA extracts, immunoblotting, qRT-PCRs, and proliferation assays were performed as described in refs. 2 and 3.

**Migration, invasion, and transendothelial migration**

**Transwell assays**

To measure migration and matrigel invasion, 7.5 × 104 MA-2 or MC-1, 3 × 104 HBL-100, and 5 × 104 4175-TGL or SK-MEL-28 were seeded in serum-free medium in the upper chambers of cell culture transwells with 8.0-μm pore size membrane (BD Biosciences), precoated or not with 4 μg/well growth factor-reduced matrigel (BD Biosciences) or in BioCoat Matrigel Invasion Chambers (Becton Dickinson). The lower chambers were filled with complete growth medium. After 18 hours, the migrated cells on the lower side of the membrane were fixed in 2.5% glutaraldehyde, stained with 0.1% crystal violet and photographed using an Olympus IX70 microscope. For transendothelial migration assay, 105 HUVECs were seeded in complete medium in the upper part of Transwell inserts with 5.0-μm pore size membrane (Costar, Corning Inc.) coated by bronectin at 5 μg/cm2 or 0.1% gelatin, and grown till confluence. Then, 5 × 104 cells labeled with CellTracker Orange CMRA or Green CMFDA (Molecular Probes, Invitrogen Life Technologies) were seeded onto the HUVECs monolayer. Twelve hours later, HUVECs and nonmigrating cells were removed and the red or green fluorescent cells that migrated on the lower side of the membrane were fixed in 4% paraformaldehyde and photographed using Zeiss Axiosvert200M microscope. Migration, invasion, and transendothelial migration were evaluated by measuring the area occupied by migrated cells using the ImageJ software (http://rsbweb.nih.gov/ij/).

**ALCAM localization imaging**

A total of 2 × 105 HUVEC-GFP were seeded on coverslips coated with fibronectin at 5 μg/cm2, and grown till confluency. Then, 5 × 105 ALCAM-overexpressing MA-2 cells (stably transduced with pLVX-ALCAM expressing vector) were seeded on the HUVEC-GFP monolayer. Twenty-four hours later, the cocultures were fixed in cold methanol and immunostained for ALCAM protein. Briefly, samples were blocked with 5% BSA,
incubated with anti-CD166/ALCAM mAb M0G/07 (Novocasta Laboratories, 1:100 dilution) for 2 hours, then with anti-mouse IgG Alexa-Fluor-568 for 45 minutes and finally mounted on microscope slides for analyses and photos, performed with Confocal Leica SP5 microscope.

**In vivo tumor growth and metastasis assays**

All experiments performed with live animals complied with ethical care. For tumor growth, 5 \times 10^6 MC-1 cells (in PBS) were subcutaneously injected into the flanks of 8- to 12-week-old NOD/SCID/IL2R_null (NSG) immunocompromised mice, animals dissected 4 weeks later and proteins extracted from tumors. For experimental metastasis assays, 5 \times 10^5 MA-2 or MC-1 or SK-MEL-28 or 3 \times 10^5 4175-TGL cells were injected into the tail vein of 8- to 12-week-old SCID or NOD/SCID/IL2R_null (NSG) immunocompromised mice and the animals were dissected 7 or 4 weeks later, respectively. Green or red fluorescent lung metastases were evaluated and photographed in fresh lungs in toto using a Leica MZ16F fluorescence stereomicroscope. The number of metastases was measured on photographs using the ImageJ software (http://rsweb.nih.gov/j/). Micrometastases were evaluated on paraffin-embedded and haematoxylin and eosin (H&E)-stained slides, scanned with Panoramic Desk (3DHistech, Euroclone).

**In vivo extravasation assay**

A total of 1.5 \times 10^5 MC-1 or MA-2 or SK-MEL-28 cells, previously labeled with CellTracker Orange CMRA (Molecular Probes, Invitrogen Life Technologies), were injected into the tail vein of 4- to 6-week-old female NSG mice (Charles River Laboratories). Two or 48 hours later, mice were sacrificed, and 4% paraformaldehyde was injected into the trachea. Lungs were dissected and photographed in toto using a Leica MZ16F fluorescence stereomicroscope and red fluorescence was quantified 48 hours following injections using the ImageJ software (http://rsweb.nih.gov/j/). Lungs were embedded in OCT (Kifik, BioOptica), frozen, cryostat-cut in 6-µm-thick sections. Localization of tumor cells, inside/outside the vessels, was evaluated on sections at a Zeiss AxioObserver microscope with the ApoTome Module (3), following blood vessels staining with an anti-CD31 primary antibody in immunofluorescence.

**Human tumor correlation analyses**

Normalized expression values for mRNA and miRNA in breast cancer were downloaded from the European Genome-phenome Archive (EGAS00000000122, EGAD00001000434, and EGAD00001000438; refs. 10, 11), and 1302 samples were used. Instead, for melanomas, expression values were downloaded from The Cancer Genome Atlas (TCGA, https://tcga-data.nci.nih.gov/tcga/) and 352 samples were used. All analyses were performed with R using the packages stats (lm) and ggplot2 (12). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria (URL https://www.R-project.org/).

**Statistical analysis**

The results are shown as mean ± SD or as mean ± SEM, as indicated, and a two-tailed Student t test was used for comparison. ‘*’, P < 0.05; ‘**’, P < 0.01; ‘***’, P < 0.001 were considered to be statistically significant. ns indicates a nonstatistically significant P value.

**Results**

miR-214 depletion and miR-148b overexpression inhibit melanoma and breast cancer metastasis formation in mice

High levels of miR-214 are found in malignant melanomas and breast tumors and promote metastatization in mice by regulating a complex network of players, in a negative or positive manner, including the downregulation of the antimetastatic miR-148b. Here, we evaluated the potential therapeutic value of miR-214 depletion and miR-148b overexpression in tumor progression in mice.

Specific miR-214 or miR-148b sponges A or B were bio-informatically designed, cloned at the 3’ end of a Green Fluorescent Protein (GFP) expression cassette in lentivirus vectors as in Supplementary Table S1, Supplementary Fig. S1A, and ref. 3 and tested in MC-1 or MA-2 melanoma cells. Specifically, cells were transfected with pLenti-214-sponge/A or B or pLenti-148b-sponge/A or B or pLenti-empty (control) vectors together with precursors for miR-214 or miR-148b or controls (pre-miR-214 or pre-miR-148b or pre-control), and GFP levels were evaluated in cells at the microscope (Supplementary Fig. S1B and S1C), by Western Blot-WB (Supplementary Fig. S1D and S1E), or FACS analysis (Supplementary Fig. S1F and S1G). As shown, all sponges were able to inhibit GFP expression, suggesting miR-214 or miR-148b binding to their complementary sequences.

MC-1 or MA-2 melanoma or 4175-TGL breast cancer cells were then transduced with lentiviruses expressing miR-214-sponges (pLenti-214-spongeA/B) or miR-148b (pLenti4/V5-148b, pLenti-miR-148b) or with empty controls (pLenti-empty, pLenti4/V5-empty, pLemiR-empty). Alternatively, MA-2 or SK-MEL-28 melanoma or 4175-TGL breast cancer cells were transduced with lentiviruses expressing miR-148b-sponges (pLenti-148b-spongeA/B) or miR-214 (pLemir-214) or with empty controls (pLenti-empty, pLemiR-empty) to verify the mechanism in the opposite directions. miR-214 and miR-148b levels were evaluated by qRT-PCR analyses as shown in Supplementary Figs. S2 to S5. Cells with the expected miR-214 or miR-148b modulations were injected in the tail vein of immunocompromised mice, and metastasis dissemination was evaluated 4 to 7 weeks later by measuring the number/area of lung metastasis in H&E-stained lung or liver sections or the fluorescent (green or red) tumor cells present in the whole organs (Fig. 1A–C, Supplementary Figs. S6A–S6D and S7A–S7D). Single miR-214 downmodulation or miR-148b overexpression significantly blocked metastasis dissemination for MC-1 or MA-2 melanoma or 4175-TGL breast cancer cells compared with controls. Relevantly, simultaneous depletion of miR-214 and increased levels of miR-148b further blocked tumor spreading of melanoma and breast tumor cells compared with single modulations, suggesting a combined action. In parallel, single miR-214 overexpression or miR-148b downmodulation, or simultaneous double targeting, favored melanoma or breast cancer cell dissemination compared with controls.

In conclusion, we can consider miR-214 and miR-148b as promising targets for miRNA-based therapeutic interventions in tumor progression.

miR-214 inhibition and miR-148b overexpression control-specific tumor cell metastatic traits

To understand which metastatic traits of melanoma (MA-2, MC-1, and SK-MEL-28) and normal or tumor breast (HBL-100 and 4175-TGL) cells were affected by miR-214 and miR-148b
miR-214 depletion and miR-148b overexpression inhibit metastasis formation in mice. Lung or liver colony formation in immunodeficient mice, 7 (A) or 4 (B and C) weeks after tail-vein injection of MC-1 (A) or 4175-TGL (B and C) cells. Cells were transduced with controls (pLenti-empty, pLenti4/V5-empty) or miR-214-sponge (pLenti-214-spongeB) or miR-148b-overexpressing (pLenti4/V5-148b) vectors, single or in combination, as indicated in the panels. Representative pictures of H&E-stained sections (A, B, a–d, bar, 500 μm; C, a–d, bar, 100 μm) are shown. Graphs (bottom of each figure) represent quantitated results as mean ± SEM H&E-stained colonies number (A) or as a percentage of metastatic/total areas (B) in lungs or as mean metastases/field in liver, referring to the indicated number of mice (n). Two independent experiments were performed and representative results are shown.

Figure 1.
modulations, we evaluated proliferation, migration, invasion through matrigel and transendothelial migration on a HUVEC monolayer in vitro. Cells were either transduced with lentiviruses for the depletion (pLenti-214-spongeB, pLenti-148b-spongeB) or overexpression (pLemiR-214, pLenti4/V5-148b) of miR-214 or miR-148b or with empty controls (pLemiR-empty, pLenti-empty, pLenti4/V5-empty) or transiently transfected with miRNA precursors or inhibitors (pre-miR-214, anti-miR-148b, pre/anti-control). miR-214 and miR-148b modulations were evaluated by qRT-PCR assays as shown in Supplementary Figs. S2 to S5. Single or double alterations of miR-214 and miR-148b did not significantly affect proliferation compared with controls in any tested cell line (Fig. 2A and B; and Supplementary Fig. S8). Instead, modulations of miR-214 or miR-148b significantly affected migration, cell movement across a HUVEC monolayer (transendothelial migration) and invasion in matrigel compared with controls (Fig. 3A–F; Supplementary Fig. S9A–F). Precisely, in all assays, cell movement was impaired in miR-214-depleted and miR-148b-overexpressing cells. Opposite results were observed when miR-214 was highly expressed in cells while miR-148b expression was reduced. Relevantly, in transendothelial migration assays, but generally not in migration or invasion analyses, simultaneous miR-214 inhibition and miR-148b overexpression almost always led to combined effects, like for in vivo metastasis (Fig. 1), suggesting a combinatorial, specific effect of miR-214 and miR-148b at the level of tumor–endothelial cell interactions.

**miR-214 depletion and miR-148b overexpression impair tumor cell extravasation in mice**

The effect of single or combined sponge-induced miR-214 depletion and miR-148b overexpression was investigated on in vivo cell extravasation of tumor cells. CMRA-labeled MC-1 or SK-MEL-28 cells stably expressing miR-214-sponge (pLenti-214-spongeB) or miR-148b (pLenti4/V5-148b) or control vectors (pLenti-empty or pLenti4/V5-empty) were injected in the tail vein of immunocompromised mice (Fig. 4A–L and graph, Supplementary Figs. S10 a–S10h and graph). Lodging to the lung vasculature was evaluated 2 hours after injection (Fig. 4A–D; Supplementary Fig. S10a–S10b), and no difference was observed among modified cells. Instead, a strong decrease in early (48 hours after injection) lung colonization was observed following single or combined sponge-driven miR-214 downmodulation and miR-148b overexpression compared with controls in MC-1 (Fig. 4E–H) or SK-MEL-28 (Supplementary Fig. S10c–S10d) cells. Note that cells were localized inside blood vessels or associated with them at 2 hours, as shown in Supplementary Figs. S10e–S10f. Instead, cells were found in the lung parenchyma at 48 hours (Fig. 4I–L; Supplementary Fig. S10g–S10h). As for metastasis dissemination (Fig. 1) and transendothelial migration (Fig. 3), simultaneous miR-214 inhibition and miR-148b overexpression led to combined effects, suggesting, once more, a combinatorial, specific effect of miR-214 and miR-148b at the level of tumor–endothelial cell interactions.

**Depletion of miR-214 and overexpression of miR-148b affect the adhesion molecules ITGA5 and ALCAM**

To identify the molecular players involved in reduced cancer dissemination/extravasation by miR-214 depletion and miR-148b overexpression in tumor cells, expression of ITGA5 and ALCAM, two validated miR-148b direct targets, known to be highly relevant for cancer cell dissemination, was analyzed in cell cultures or in mouse subcutaneous tumors. Single or combined miR-214 depletion and miR-148b overexpression in melanoma (MA-2, MC-1, SK-MEL-28) and breast cancer (4175-TGL) cells were obtained following stable transduction of lentivirus vectors for the expression of miR-214 sponges (pLenti-214-spongeB) or miR-148b (pLenti4/V5-148b) or empty controls (pLenti-empty, pLenti4/V5-empty). miR-214 and miR-148b alterations were evaluated by qRT-PCR analyses as in Supplementary Figs. S2 to S5. Important reduction of ITGA5 or ALCAM expression was observed for single or combined miRNA-modulations. Combined inhibitions of ITGA5 and ALCAM by dual miR-214/ miR-148b interventions were rarely observed compared with single alterations (Fig. 5A–D; and Supplementary Fig. S11A–E).

Considering the relevance of ITGA5 and ALCAM expression impairment for the inhibition of transendothelial migration following simultaneous miR-214/miR148b alterations, as presented below (see the next paragraph), we can speculate that the effects of combined miR-214/miR-148b changes occur specifically when tumor cells get in contact with endothelial cells. Alternatively, supplementary alterations of gene expression could be involved in the passage of tumor cells through the blood vessels. Opposite stable or transient modulations were obtained in some of the cells (MA-2, 4175-TGL) listed above and in HBL-100
miR-214 depletion and miR-148b overexpression inhibit transendothelial migration. Transwell migration assays were used to evaluate migration (through a porous membrane) or transendothelial migration (through an HUVEC monolayer on top of a porous membrane) for MC-1 (A, B) or 4175-TGL (C and D) or SK-MEL-28 (E and F) cells stably transduced with control (pLenti-empty, pLenti4/V5-empty) or miR-214-sponge (pLenti-214-spongeB) or miR-148b-overexpressing (pLenti4/V5-148b) vectors, single or in combination, as indicated in the panels. For migration, results are indicated as a ratio of mean ± SEM of the area covered by migrated versus plated tumor cells; for transendothelial migration, results are shown as mean ± SEM of the area covered by tumor migrated cells. At least two independent experiments (with triplicates) were performed, and representative results are shown.

Figure 3.
normal breast cells to further evaluate this mechanism. Here, cells were transduced with pLenti-148b-spongeA/B or pLemiR-214 or the relative empty controls (pLenti-empty, pLenti4/V5-empty) or miR-214-depletion or miR-148b-overexpressing (pLenti4/V5-148b) vectors, single or in combination, as indicated in the panels. Representative pictures of whole red fluorescent lungs at 2 hours or 48 hours after injection (A–H) and representative fields of murine lung sections, 48 hours after injection (I–L), stained for CD31 and counterstained with DAPI are shown; bar, 800 μm. Results are indicated in the graphs (bottom) as mean ± SEM of the number of extravasated cells at 48 hours for n = 4 mice per group. White arrows, extravasated cells. Two independent experiments were performed, and representative results are shown.

Figure 4.

miR-214 depletion and miR-148b overexpression inhibit extravasation in mice. In vivo extravasation 2 hours (A–D) or 48 hours (E–H) following tail-vein injections in immunodeficient mice of CMRA-labeled MC-1 cells transduced with control (pLenti-empty, pLenti4/V5-empty) or miR-214-depletion (pLenti-214-spongeB) or miR-148b-overexpressing (pLenti4/V5-148b) vectors, single or in combination, as indicated in the panels. Representative pictures of whole red fluorescent lungs at 2 hours or 48 hours after injection (A–H) and representative fields of murine lung sections, 48 hours after injection (I–L), stained for CD31 and counterstained with DAPI are shown; bar, 800 μm. Results are indicated in the graphs (bottom) as mean ± SEM of the number of extravasated cells at 48 hours for n = 4 mice per group. White arrows, extravasated cells. Two independent experiments were performed, and representative results are shown.
Taken together, our results prove that the negative targeting of miR-214 and/or the positive modulation of miR-148b impairs dissemination by acting on ITGA5 or ALCAM, two main players for tumor cell extravasation.

miR-214 expression correlates with ITGA5 and ALCAM levels, while it anticorrelates with miR-148b, in melanoma metastases and in primary breast tumors. Breast cancer (n = 1302; refs. 10, 11) and melanoma metastasis (TCGA, https://tcga-data.nci.nih.gov/tcga/; n = 352) datasets were used to evaluate miR-214, miR-148b, ITGA5, and ALCAM expression and to look for possible correlations or anticorrelations with one another (Fig. 7A and B). Relevantly, we found that miR-214 and miR-148b anticorrelate in melanoma metastases (P = 7.91e−05) and in primary breast tumors (P = 5.43e−08), while miR-214 significantly correlates with ITGA5 and ALCAM in both datasets. P values were the following: for melanoma metastases: ITGA5 (P = 3.09e−24) and ALCAM (P = 2.69e−09); for primary breast tumors: ITGA5 (P = 1.68e−23) and ALCAM (P = 9.27e−07). These results are therefore in line with our above-presented investigations, thus strengthening the link between miR-214, miR-148b, ITGA5, and ALCAM and its relevance in human tumor progression.

Discussion

We previously demonstrated the prometastatic role of miR-214 (3) and its link with the antimetastatic miR-148b (2). Here, we show that single or combined miR-214 inhibition and miR-148b overexpression in tumor cells strongly modulate metastasis formation by acting mainly during the passage across the vessel endothelium (transendothelial migration/extravasation), a metastatic trait that involves two direct miR-148b targets, the adhesion receptors ITGA5 and ALCAM, in a cell–fibronectin and cell–cell dependent manner. Our data suggest...
Figure 6.
Transendothelial migration depends on ITGA5 and ALCAM expression in miR-214-depleted and/or miR-148b-overexpressing tumor cells. A–F, transendothelial migration was evaluated in MC-1 (A, C, E) or 4175-TGL (B, D, F) cells stably transduced with miR-214-sponge (pLenti-214-spongeB) or miR-148b-overexpressing (pLenti4/V5-148b) vectors, single or in combination, and, in addition, transiently transfected with recombinant vectors for the overexpression of ITGA5 or ALCAM or empty controls, as indicated in the panels. ITGA5 and ALCAM levels were evaluated by Western blot analysis, 24 hours or 48 hours following transfections. Protein modulations were calculated relative to negative controls, normalized on loading controls (GAPDH or actin) and expressed as percentages (%). Transmigration results are indicated as mean ± SEM of the area covered by tumor-migrated cells. At least two independent experiments (with triplicates) were performed, and representative results are shown.
that miR-214 and miR-148b are valuable candidates for miRNA-based targeted therapy.

miR-214 is highly expressed in malignant cutaneous and ocular melanomas (3, 13, 14) as well as in breast, osteosarcoma, ovary, pancreas, prostate, and gastric cancers (15–20). In line with these findings, upregulation of miR-214 in various tumor cells increases metastasis formation (2, 3, 21–23). On the other hand, miR-148b is poorly expressed in melanomas and in breast, pancreatic, and hepatocellular carcinomas (1, 6, 24, 25) and its modulation in tumor cell lines reveals its antimetastatic function (6, 26–28). Here, by modulating miR-214 and miR-148b, respectively, in a negative and positive manner, we show that miR-214 and miR-148b are part of a miR-ON-miR regulatory axis where miR-214 favors tumor dissemination following the downregulation of miR-148b and the consequent upregulation of miR-148b direct targets, ITGA5 and ALCAM as well as some of their downstream targets.

Figure 7. miR-214 anticorrelates with miR-148b, while it correlates with miR-148b targets ITGA5 and ALCAM, in melanoma metastases or human primary breast tumors. The indicated datasets were used to evaluate miR-214, miR-148b, ITGA5, and ALCAM expression in melanoma metastases (A) and in primary breast tumors (B). Statistically significant negative or positive correlations are shown for miR-214 and miR-148b (anticorrelations) and for miR-214 and ITGA5 or ALCAM (correlations). Correlations of normalized expression are represented with a dot plot superimposing the regression line. The shaded area represents the 0.95 standard error confidence interval of the model predictions. Statistically significant R² and P values and number of samples (n) are indicated.
in mice, gives hope for an miRNA-based therapy. Due to the relevance of ALCAM and ITGA5 in the pathway presented here, one could even speculate to target these two adhesion molecules with specific antibodies, in addition to miRNA targeting, to further affect metastasis formation. Considering that the main issue of the miRNA-based targeted therapy is the in vivo delivery, it is essential to identify safe, selective, and efficient compound systemic deliveries. For this purpose, we are currently developing new tools to administer miR-214 inhibitors and miR-148b precursors to animals and test their efficacy on metastasis formation.

In conclusion, our data demonstrate that the cascade of events, including miR-214, miR-148b, ALCAM, and ITGA5, is controlling melanoma and breast cancer progression and it can be exploited for combinatorial therapeutic interventions.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Conception and design: F. Orso, L. Quirico, D. Taverna
Development of methodology: M.F. Brizzi, M.B. Stadler
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): F. Vinga, E. Penna, D. Cimino, D. Brusa, S. Deaglio
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): F. Orso, L. Quirico, F. Vinga, D. Detorri, D. Cimino, E. Grassi, A.R. Elia, M.B. Stadler, P. Provero, M. Caselle
Writing, review, and/or revision of the manuscript: F. Orso, L. Quirico, M.B. Stadler, D. Taverna
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Francesca Orso, Lorena Quirico, Federico Virga, et al.


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