Ovarian Cancer Cell Detachment and Multicellular Aggregate Formation Are Regulated by Membrane Type 1 Matrix Metalloproteinase: A Potential Role in I.p. Metastatic Dissemination

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Abstract

An early event in the metastasis of epithelial ovarian carcinoma is shedding of cells from the primary tumor into the peritoneal cavity followed by diffuse i.p. seeding of secondary lesions. Anchorage-independent metastatic cells are present as both single cells and multicellular aggregates (MCA), the latter of which adhere to and disaggregate on human mesothelial cell monolayers, subsequently forming invasive foci. Although this unique metastatic mechanism presents a distinct set of therapeutic challenges, factors that regulate MCA formation and dissemination have not been extensively evaluated. Proteolytic activity is important at multiple stages in i.p. metastasis, catalyzing migration through the mesothelial monolayer and invasion of the collagen-rich submesothelial matrix to anchor secondary lesions, and acquisition of membrane type 1 matrix metalloproteinase (MT1-MMP; MMP-14) expression promotes a collagen-invasive phenotype in ovarian carcinoma. MT1-MMP is regulated posttranslationally through multiple mechanisms including phosphorylation of its cytoplasmic tail, and the current data using ovarian cancer cells expressing wild-type, phosphomimetic (T567E-MT1-MMP), and phosphodefective (T567A-MT1-MMP) MT1-MMP show that MT1-MMP promotes MCA formation. Confluent T567E-MT1-MMP–expressing cells exhibit rapid detachment kinetics, spontaneous release as cell–cell adherent sheets concomitant with MT1-MMP–catalyzed α5 integrin ectodomain shedding, and robust MCA formation. Expansive growth within three-dimensional collagen gels is also MT1-MMP dependent, with T567E-MT1-MMP–expressing cells exhibiting multiple collagen invasive foci. Analysis of human ovarian tumors shows elevated MT1-MMP in metastases relative to paired primary tumors. These data suggest that MT1-MMP activity may be key to ovarian carcinoma metastatic success by promoting both formation and dissemination of MCAs. [Cancer Res 2009;69(17):7121–9]

Introduction

Ovarian cancer is the leading cause of death from gynecologic malignancy due primarily to complications of metastasis (1). Unlike most carcinomas that rely on the vasculature for metastasis, an early event in ovarian cancer dissemination is shedding of cells from the primary tumor into the peritoneal cavity followed by diffuse "seeding" of the peritoneal cavity (2). This unique metastatic mechanism presents a distinct set of therapeutic challenges. Accumulation of malignant ascites is widely associated with advanced ovarian carcinoma (3) and it is hypothesized that ascites augment progression by facilitating the spread of cancer cells throughout the peritoneal cavity (4). Anchorage-independent metastatic cells are present as both single cells and multicellular aggregates (MCA) that survive in suspension (5). The factors that regulate MCA formation have not been extensively evaluated; however, evidence suggests that MCAs have a functional role in metastatic progression. MCAs adhere to and disaggregate on human mesothelial cell monolayers, subsequently forming invasive foci (6, 7). Transplanted ovarian cancer MCAs form murine xenografts with the same histopathology as the primary tumor and can be serially propagated in vivo (8, 9). Further, sphere-forming ovarian cancer-initiating cells are significantly more tumorigenic in xenograft models, further showing that the MCA population is a key target for antimetastatic therapy (10).

Proteolytic activity is important at multiple stages in i.p. metastasis, including localized proteinase-driven migration through the mesothelial monolayer and invasion of the collagen-rich submesothelial matrix to anchor secondary lesions (11, 12). Invasion of collagenous matrices by ovarian cancer cells requires membrane type 1 matrix metalloproteinase (MT1-MMP; MMP-14; refs. 13–15), a transmembrane collagenase that is not detected in normal ovarian surface epithelium or in benign ovarian tumors, however, evidence suggests that MCAs have a functional role in metastatic progression. MCAs adhere to and disaggregate on human mesothelial cell monolayers, subsequently forming invasive foci (6, 7). Transplanted ovarian cancer MCAs form murine xenografts with the same histopathology as the primary tumor and can be serially propagated in vivo (8, 9). Further, sphere-forming ovarian cancer-initiating cells are significantly more tumorigenic in xenograft models, further showing that the MCA population is a key target for antimetastatic therapy (10).

Because MT1-MMP is central to a variety of biological processes, proteolytic activity is stringently controlled. MT1-MMP is internalized from the cell surface through a mechanism involving the cytoplasmic domain (24, 25) and cytoplasmic tail truncation restricts MT1-MMP to the plasma membrane. The cytoplasmic domain of MT1-MMP has three potential phosphorylation sites: T567, Y573, and S577. Recent work indicates that MT1-MMP can be phosphorylated at T567 and Y573 (26–28). T567 is localized within the sequence RPRLLYCQRSLLDKV582 that has homology with the consensus sequence for protein kinase C (TXR) and

Note: Supplementary data for this article are available at Cancer Research Online (http://cancerres.aacrjournals.org/). Requests for reprints: M. Sharon Stack, Department of Pathology and Anatomical Sciences, University of Missouri School of Medicine, M214E Medical Sciences Building, 1 Hospital Drive, Columbia, MO 65212. Phone: 573-884-7301; Fax: 573-884-8104; E-mail: stackm@missouri.edu. ©2009 American Association for Cancer Research. doi:10.1158/0008-5472.CAN-08-4151
extracellular signal-regulated kinase 1/2 (XTP, ref. 29). To examine the potential effect of T567 phosphorylation in the unique metastatic mechanism of ovarian carcinoma, the properties of cells expressing wild-type MT1-MMP, a phosphomimetic mutant (T567E-MT1-MMP), or a phosphodefective mutant (T567A-MT1-MMP) were evaluated. Acquisition of MT1-MMP catalytic activity promotes rapid cell-matrix detachment kinetics concomitant with α3 integrin ectodomain shedding, enhanced MCA formation, and expansive growth in three-dimensional collagen. This prometa-
static phenotype was intensified in the phosphomimetic mutant T567E-MT1-MMP, suggesting that phosphorylation of the MT1-
MMP cytoplasmic tail may regulate i.p. metastatic dissemination.

Materials and Methods

Materials. DOV13 and OVCA433 cells were provided by Dr. R. Bast, Anti-
FLAG M2, anti-MT1-MMP (M3927), peroxidase-conjugated secondary
antibodies, and protein G-Sepharose beads were from Sigma. SuperSignal enhanced chemiluminescence reagents were purchased from Pierce. Tissue
inhibitor of metalloproteinase-2 (TIMP-2) was provided by Dr. R. Fridman. Rat
tail collagen type I, human type IV collagen, and human fibronectin
were purchased from BD Biosciences. Mouse anti-human integrin α3
(AMB1952Z and MARB2056) was purchased from Chemicon. Centriprep
was purchased from Millipore.

DNA constructs and generation of stable cell lines. The human MT1-
MMP cDNA with COOH-terminal FLAG tag (DYKDDDDK) was provided by
Dr. D. Pei. Subsequently, the T567A, T567E, and E240A point mutations were
generated using QuikChange (Stratagene). Inserts were sequenced to verify
mutation. Transfection of cells was done using FuGENE 6 (Roche) and
peroxidase-conjugated anti-rabbit IgG (1:4,000 in 3% BSA/TBST) and
300 mmol/L NaCl, and 0.2% Tween 20 (TBST). Membranes were incubated
(1 h, room temperature) with FLAG M2 monoclonal (1:1,000) or anti-MMP-
14 in 3% BSA/TBST. Immunoreactive bands were visualized with an
enhanced peroxidase-conjugated anti-rabbit IgG (1:4,000 in 3% BSA/TBST) and
enhanced chemiluminescence.

To detect integrin α3 in conditioned medium, 10^6 cells were serum
starved overnight ± the broad-spectrum MMP inhibitor GM6001. Conditioned
medium was collected, concentrated, then electrophoresed on 9% SDS-PAGE, transferred to polyvinylidene difluoro,
and blocked with 3% bovine serum albumin (BSA) in 50 mmol/L Trizma,
300 mmol/L NaCl, and 0.2% Tween 20 (TBST). Membranes were incubated
(1 h, room temperature) with anti-FLAG M2 monoclonal (1:1,000) or anti-MMP-
14 in 3% BSA/TBST. Immunoreactive bands were visualized with
peroxidase-conjugated anti-rabbit IgG (1:4,000 in 3% BSA/TBST) and
enhanced chemiluminescence.

Results

Expression of MT1-MMP in ovarian cancer cells. Based on
previous studies showing that phosphorylation of cytoplasmic tail
residues in MT1-MMP can modify cell behavior (26–28), MT1-MMP
mutants were generated in which T567 was mutated to glutamic
acid (T567E) to mimic constitutive phosphorylation or alanine
(T567A) to represent a phosphodefective mutation. Constructs
were transfected into OVCA433 cells chosen because of the lack
of endogenous MT1-MMP expression. Analysis of MT1-MMP
in whole-cell lysates obtained from wild-type and phosphomutant
cell lines showed equivalent expression of wild-type and mutant
MT1-MMP (Supplementary Fig. S1A). Relative to cells expressing
wild-type MT1-MMP, gelatin zymography indicated no significant
change in pro-MMP-2 activation by T567A-MT1-MMP or T567E-
MT1-MMP mutant cell lines (refs. 16, 21, 32–35; Supplementary
Fig. S1B). The cytoplasmic tail alters the surface presentation of
active MT1-MMP by regulating its internalization from the cell

MCA formation. MCAs were generated using a modification of the
hanging drop method as described previously (30). MCA formation
was monitored after 24 to 48 h.

DNA extraction, cDNA synthesis, and real-time reverse transcription-
PCR. RNA was extracted from 10^6 cells using the SV Total RNA
Isolation System (Promega). cDNA was synthesized from 5 to 10 μg total
RNA using iScript cDNA Synthesis Kit (Bio-Rad). Real-time PCR used SYBR
Green chemistry and the 7500 ABI Prism (Applied Biosystems). Reaction
setup, normalization, and primers were used as described previously and
fold changes were quantified as 2^(-ΔΔCT sample - ΔΔCT control) (15).

Quantification of cell-matrix detachment. Cells (10^3) were seeded onto
60 mm dishes and allowed to adhere overnight. Adherent cells were
subjected to trypsinization (0.25% trypsin/2.21 mmol/L EDTA in HBSS
without sodium bicarbonate, calcium, and magnesium) and both
adherent and detached cells were quantified. Experiments were repeated
three to five times.

Adhesion assays. Chambers were coated with 10 μg/ml type I collagen,
type IV collagen, or fibronectin in PBS for 4 h at 37°C, blocked with 3% BSA
in MEM for 1 h at 37°C, washed with PBS, and air-dried. Cells were seeded at
10^5 per well, allowed to adhere for 75 min (determined empirically based
on a time course of 0-2 h), washed with PBS to remove nonadherent cells,
fixed, stained using DiffQuick, and enumerated. Assays were done in triplicate.

Three-dimensional collagen culture. Three-dimensional cultures were
prepared as described (31) by diluting type I rat tail collagen with 10× MEM
to 1.5 mg/ml. Cells (10^4) were added to the collagen mixture before
solidification ± TIMP-2 (5 μg/mL). In additional controls, cells were added
atop solidified gels.

Immunohistochemistry. Immunohistochemical analysis was done
retrospectively on tumor tissue microarrays prepared with institutional
review board approval by the Pathology Core Facility of Northwestern
University. Tissue microarrays included 17 paired primary and metastatic
ovarian cancer tissues obtained during the same surgical procedure from
patients who were not treated against stage III and IV ovarian cancer before
the operation (15 serous and 2 endometroid). Immunohistochemical
staining used antibody to MT1-MMP (clone RB1544B; 1:100 dilution;
Neomarkers; ref. 22) with breast carcinoma tissue as a positive control.
Scoring was assigned according to the average overall intensity of the
staining: 0, no staining; 1, fine granular staining; 2, somewhat coarse
staining but less than positive control tissue (breast carcinoma); and 3, very
coarse staining, similar to or greater than positive control tissue. Staining
<10% of tumor cells, regardless of intensity, was considered negative.
Staining of between 10% and 75% of tumor cells was considered focal
positive. Staining of >75% of tumor cells was considered diffuse positive.
surface (24, 25), the functional consequences of which may affect net proteolytic activity. Because the specific sequence(s) in the cytoplasmic tail responsible for modulating internalization have yet to be delineated, the effect of T567 mutation on MT1-MMP surface presentation was evaluated. Relative to cells expressing wild-type, T567A-MT1-MMP–, and T567E-MT1-MMP–expressing cells exhibit similar basal levels of surface expression (Supplementary Fig. S1C). Further, expression levels of exogenous MT1-MMP in OVCA433 stable transfectants are similar to endogenous MT1-MMP levels in DOV13 cells (Supplementary Fig. S1D and E). Potential changes in the relative distribution of MT1-MMP mutants within specific membrane microdomains or in rates of internalization in response to specific stimuli were not evaluated.

**MT1-MMP promotes spontaneous detachment and MCA formation.** Initial dissemination of ovarian cancer involves exfoliation of cells from the primary ovarian tumor into the peritoneal cavity as matrix-detached single cells and MCAs (5, 6). Malignant ascites accumulates in advanced ovarian carcinoma and contains a population of nonadherent MCAs ranging in size from 30 to 200 μm (36). These cells survive anoikis (37) and proliferate as a free-floating population of highly neoplastic cells (38); however, molecular mechanisms regulating the genesis of MCAs have not been extensively evaluated. Exploratory cDNA microarray analysis of changes in gene expression induced by MCA culture confirmed these results, showing a 2.1-fold increase in MT1-MMP in MCAs relative to two-dimensional cultures (Fig. 1A and B). To evaluate the effect of acquisition of MT1-MMP expression on MCA formation, OVCA433 cells expressing wild-type or mutant MT1-MMP were evaluated relative to DOV13 using the hanging drop method (30) to generate MCAs in vitro (Fig. 1C). Cells expressing MT1-MMP formed larger diameter MCAs relative to untransfected control cells (Fig. 1D). Interestingly, smaller MCAs were formed by cells expressing the phosphodefective T567A-MT1-MMP mutant relative to those expressing wild-type or T567E phosphomimetic mutant MT1-MMP, suggesting that phosphorylation at this site may modulate spheroid size and overall growth.

Although MCAs are prevalent in ascites obtained from women with ovarian carcinoma (6, 7, 36, 39), it is not known whether cellular aggregation occurs before or following detachment of metastatic cells from the primary tumor. Microscopic examination of cell cultures expressing T567E-MT1-MMP revealed a striking phenotype, showing spontaneous detachment as cell-cell adherent MCAs (Fig. 2A). To further characterize this phenotype, relative adhesive strength was evaluated using a controlled trypsinization

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*Figure 1. MT1-MMP expression in MCAs. A, expression of MT1-MMP RNA in DOV13 MCAs or 2D cultures. Real time RT-PCR was performed to detect RNA expression. Shown is the average of three independent experiments ± standard deviation. (*, P < 0.05). B, expression of MT1-MMP protein. Western blot was performed to detect relative MT1-MMP protein levels with β-tubulin as a loading control. Arrow, 55 kDa active MT1-MMP; arrowhead, 43 kDa MT1-MMP autolysis product. C, representative MCA formed by OVCA 433 or DOV13 cells. D, expression of MT1-MMP augments MCA formation. Cells were cultured for MCA formation using the hanging drop method and measured using a Zeiss Axiosvert Imaging Software. Data shown are expressed in μm, and represent the mean diameter measurement of n = 10 MCAs. +, P < 0.05, relative to vector-transfected cells; *, P < 0.05, relative to cells expressing wild type MT1-MMP. Untransfected DOV13 cells, that express similar levels of endogenous MT1-MMP, are shown for comparison.*

**MT1-MMP in I.p. Metastatic Dissemination**

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6 M.V. Barbolina and M.S. Stack, unpublished results.

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assay to monitor the kinetics of cell-matrix dissociation. In subconfluent cultures, detachment kinetics were similar in cells expressing wild-type, T567A-MT1-MMP, or T567E-MT1-MMP (Fig. 2B). On confluence, cells expressing T567E-MT1-MMP exhibit a more complete and accelerated detachment relative to cells expressing wild-type or T567A-MT1-MMP (Fig. 2C). Cells lacking MT1-MMP expression (vector controls) are strongly adherent and detach with extended kinetics (>20 min) under these conditions (data not shown). DOV13 cells do not exhibit spontaneous detachment; however, it should be noted that these cells also express high levels of the endogenous MMP inhibitor TIMP-2 (13, 15). To determine whether MT1-MMP catalytic activity was required for aggregate detachment, the active site mutation E240A was introduced into the T567E background to generate T567E/E240A-MT1-MMP–expressing cells. Detachment kinetics of T567E/E240A-MT1-MMP cells were significantly attenuated relative to cells expressing the catalytically active phosphomimetic mutant (Fig. 2D). Together, these results suggest that cell-cell communication regulates cell-matrix adhesion via the activity of MT1-MMP.

**MT1-MMP Catalyzes α3 Integrin Ectodomain Shedding.** Reciprocal interplay between integrin and cadherin function has been reported (40, 41) and the results described above suggest that cadherin-engaged, confluent cells lose integrin function. Relative to phosphodeficient T567A cells, expression of both wild-type and T567E-MT1-MMP significantly impaired cell adhesion to both type I collagen (Fig. 3A) and type IV collagen (Supplementary Fig. S2A). In contrast, adhesion to fibronectin was unaffected by MT1-MMP expression (Supplementary Fig. S2B), suggesting a specific defect in the expression and/or function of collagen-binding integrins. As detachment of cell-cell adherent sheets of T567E-MT1-MMP–expressing cells occurs on confluence (Fig. 2A), integrin profiles were compared under these conditions. A significant reduction in levels of cell surface α3 integrin was observed in confluent cultures of T567E-MT1-MMP–expressing cells (Fig. 3B, inset, red trace) relative to cells expressing wild-type (Fig. 3B, inset, black trace) and T567A-MT1-MMP (Fig. 3B, inset, blue trace). Vector control cells maintain α3 expression at levels seen in subconfluent cells (data not shown). To evaluate a potential role for MT1-MMP activity in this process, cells were cultured with GM6001 to block MT1-MMP catalytic activity. Inhibition of MMP catalytic activity rescues α3 integrin surface expression in confluent cells expressing wild-type or T567E-MT1-MMP (Fig. 3B). In control experiments, surface levels of α3 integrin were similar in subconfluent cells expressing wild-type, T567A-MT1-MMP, or T567E-MT1-MMP (Supplementary Fig. S3A). Additional controls show no loss of surface levels of α2 integrin (Supplementary Fig. S3B), β1 integrin (Supplementary Fig. S3C), E-cadherin (Supplementary Fig. S3D), and transferrin receptor or integrins αv, α5, and α6 (data not shown) in subconfluent or confluent cells in the presence or absence of GM6001, suggesting that loss of α3 integrin expression does not represent nonspecific loss of cell surface protein.

The ability of GM6001 to rescue surface expression of integrin α3 indicates that the observed reduction in surface presentation may

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**Figure 2.** MT1-MMP expression promotes detachment of cell-cell adherent monolayers and MCA formation. A, confluence initiates detachment of T567E-MT1-MMP–expressing cells. Image shows adherent and detached portions of confluent monolayer with floating aggregate. B–D, quantitation of detachment using controlled trypsinization. Kinetics of cell detachment were evaluated by using (B) subconfluent or (C, D) confluent cultures of cells. Experiments were performed in triplicate.
be the result of a MT1-MMP–catalyzed cleavage event. To explore this possibility, conditioned medium from confluent T567E-MT1-MMP–expressing cells was concentrated and analyzed for the presence of polypeptides that cross-react with anti–α3 integrin antibodies. Western blot analysis of concentrated conditioned medium identified a protein of 70.8 kDa and this species was undetectable in the medium of cells cultured overnight with GM6001 (Fig. 3C, compare lanes 2 and 3). Similar results were obtained following immunoprecipitation of nonconcentrated conditioned medium with anti–α3 integrin (Fig. 3C, lanes 4 and 5). Impaired collagen adhesion, spontaneous detachment, decreased surface α3 integrin levels, and α3 integrin ectodomain shedding were not observed in cells expressing the catalytically inactive T567E/E240A-MT1-MMP (Fig. 3D; data not shown). Together, these data support a mechanism for loss of adhesion subsequent to MT1-MMP–catalyzed α3 integrin ectodomain shedding, particularly evident in phosphomimetic T567E-MT1-MMP–expressing cells.

MT1-MMP enables matrix-embedded proliferative growth. Successful i.p. metastasis requires proliferation within the confines of the interstitial collagen-rich submesothelial matrix to establish collagen-anchored i.p. secondary lesions (11, 42, 43). MT1-MMP is necessary for cell proliferation in a three-dimensional collagen network, functioning to remove cytoskeletal constraints necessary to drive a proliferative response (23). To examine the contribution of MT1-MMP activity to growth of ovarian cancer cells in three-dimensional collagen, cells were seeded at single-cell density in three-dimensional collagen gels and qualitatively assessed for growth after 6 days (31). Large MCA structures were generated by all MT1-MMP–expressing cells (Fig. 4A; Supplementary Table S1). Proliferation was significantly attenuated in vector-transfected control cells (data not shown) or by copolymerization of TIMP-2 within collagen gels (Fig. 4B), verifying the dependence of growth in three-dimensional collagen on MT1-MMP activity. DOV13 cells were also highly proliferative within three-dimensional collagen gels, forming clusters of cells averaging 2.43 ± 0.65 × 10^5 per cluster. Although overall structures generated by all MT1-MMP–expressing cells were similar in size (Supplementary Table S1), MCAs generated by T567A-MT1-MMP–expressing cells were more spherical (length/width ratio of 1.47) and lacked distinct foci of collagen invasion (Fig. 4A, left; Supplementary Table S1). Cells expressing wild-type MT1-MMP produced more elongated structures with small invasive foci (Fig. 4A, middle; Supplementary Table S1). In contrast, cells expressing the phosphomimetic mutant
T567E-MT1-MMP grew as multicellular prolate ellipsoid aggregates (length/width ratio of 2.16) with multiple large invasive foci (Fig. 4A, right; Supplementary Table S1). Quantitative analysis of invasive projections indicates that both projection number and projection length were significantly increased relative to cells expressing wild-type MT1-MMP (Supplementary Table S1). These results support the hypothesis that the phosphorylation status of MT1-MMP cytoplasmic residue T567 may regulate invasive growth within the collagen-rich microenvironment of the submesothelial matrix. Furthermore, the data indicate that the ability of ovarian cancer cells to survive long-term and proliferate in three-dimensional collagen is enhanced by MT1-MMP activity, as proliferation is blocked by inclusion of TIMP-2 within the three-dimensional collagen gels (Fig. 4B). This is supported by an immunohistochemical analysis of MT1-MMP expression in sets of paired primary ovarian tumors and peritoneal metastatic lesions derived from the same patient (n = 17; Fig. 5A–C; Supplementary Table S2). Although the data set is not sufficiently powered for rigorous statistical analysis, the data support a trend wherein 76% (13 of 17) show sustained or increased high-level (2+ and 3+) MT1-MMP expression in metastases. A further 12% (2 of 17) maintain low level (1+) expression in metastases, whereas only 12% (2 of 17) show decreased expression relative to the primary tumor. It is interesting to note that MT1-MMP is expressed in 100% of the metastatic lesions overall (Supplementary Table S2).

**Discussion**

Reversible phosphorylation is widely recognized as a key post-translational modification that regulates protein function and a growing body of work suggests that MT1-MMP action may be altered through phosphorylation of cytoplasmic tail residues (26–28). For example, src-dependent or epidermal growth factor–dependent phosphorylation of cytoplasmic residue T567 has been shown to modulate cell migration and invasion (26, 28), suggesting that the intracellular domain of MT1-MMP may be vital to “inside-out” signaling processes. The observation that several cell surface proteins undergo phosphorylation at multiple sites underscores our observations implicating T567 as a second site for post-translational modification (27). Although regulatory mechanisms for the control of T567 phosphorylation have yet to be elucidated, results in the present study highlight a pivotal role for the cytoplasmic tail in regulating MT1-MMP function and associated cellular phenotypes relevant to metastasis of ovarian carcinoma.

The majority of women with ovarian cancer are initially diagnosed with disseminated intra-abdominal disease (2, 3), indicating that a more detailed understanding of the cellular and biological factors that promote successful metastatic dissemination can ultimately improve patient survival. Unlike other solid tumors, hematogenous dissemination of ovarian cancer cells is uncommon, as metastasis proceeds primarily through exfoliation of cells from the primary tumor into the peritoneal cavity as free-floating cells.
and MCAs. This imposes challenges to tumor cell survival because these metastatic cells must escape anoikis, and it has been speculated that MCA formation may function to promote anchorage-independent growth (5). Although originally considered a nonadhesive subset of ovarian tumor cells, recent data show that human ovarian cancer ascites-derived MCAs adhere to and invade mesothelial monolayers and can thereby contribute to i.p. implantation and metastasis (6, 7, 12). Further, MCAs that survive in ascites may generate a subpopulation of highly neoplastic cells (38). This is supported by recent data showing that cells isolated from murine ascites are more aggressive than parental cells when reinjected in vivo (8). Gene expression profiling shows clustering of MCA expression profiles with tumor xenograft patterns, rather than with monolayer cells (9, 44), suggesting that MCAs represent a more advanced stage of malignancy. Self-renewing spheroid-forming cells isolated from ovarian primary tumors show increased tumorigenicity and are associated with metastasis to omentum and colon (10). Together, these data support the hypothesis that the MCA population in human ovarian ascites may be a primary source of i.p. metastases and thereby represents a key target for antimetastatic therapy.

Although recent studies highlight the importance of MCAs in ovarian cancer pathobiology, the process of self-assembly of ovarian tumor cells into microtissues such as MCAs has not been extensively evaluated. Further, it is unknown whether the temporal sequence of events in MCA formation involves shedding of individual tumor cells, which then aggregate in ascites versus exfoliation of multicellular tumor cell sheets that subsequently reorganize into MCAs. The current data suggest that acquisition of MT1-MMP activity would promote MCA formation by either mechanism. Cells expressing endogenous MT1-MMP (DOV13) readily aggregate from single cells into MCAs, and quantitative PCR and Western blotting analyses confirm elevated expression of MT1-MMP in MCA cultures. Alternatively, OVCA433 cells transfected with wild-type or T567E-MT1-MMP exhibit rapid detachment kinetics and sheet-like exfoliation as cell-cell adherent aggregates. Interestingly, this striking detachment phenotype is not manifested in subconfluent cultures, suggesting the potential contribution of cell-cell adhesion molecules to the regulation of cell-matrix adhesion. It is interesting to speculate that acquisition of MT1-MMP expression by primary tumor cells may promote metastasis via enhanced tumor cell shedding. In support of this hypothesis, previous immunohistochemical and in situ hybridization analyses show MT1-MMP expression in 78% to 100% of primary ovarian tumors (15, 18). MT1-MMP expression is also detected in cancer cells obtained from malignant effusions (17) and in 88% to 98% of peritoneal metastases, wherein expression is correlated with poor outcome (current study; ref. 17).

Detachment of cell-cell adherent sheets bears similarities to cohort migration (45). The ability of protease inhibitors to impede this process reinforces the role for MMP activity in establishing this phenotype. This is further supported by results from the current study showing that the catalytically inactive mutant T567E/E240A-MT1-MMP did not induce cell detachment and sheet-like exfoliation. Our data suggest that MT1-MMP catalytic activity contributes mechanistically to cell detachment via catalysis of α₃ integrin ectodomain shedding, as cell adhesion was restored and soluble integrin ectodomain was not detectable in the presence of a broad-spectrum MMP inhibitor or with the catalytically inactive mutant. It is plausible that shedding of cells as multicellular masses may represent a more efficient means of dissemination. In addition, the ability of cells to function collectively may allow the mass to produce high levels of matrix proteases that promote migration and invasion at secondary sites (45).

Metastasizing ovarian cancer cells encounter an interstitial collagen-rich microenvironment, as the submesothelial matrix is composed primarily of types I and III collagen (12, 46–50). Acquisition of MT1-MMP collagenolytic activity may be key to

Figure 5. Immunohistochemical analysis of MT1-MMP expression in paired primary ovarian tumors and peritoneal metastatic lesions. A–C, examples of paired primary tumor (top) and corresponding metastatic lesion (bottom) from the same patient stained with MT1-MMP-specific antibodies (1:100 dilution) and peroxidase-conjugated secondary antibody. Inset, positive control breast carcinoma tissue. Numbers in top right corners, immunohistological score of each tumor.
metastatic success, as MT1-MMP activity is required for invasion of three-dimensional collagen gels by ovarian cancer cells (19). Furthermore, MT1-MMP collagenolysis is necessary to remove matrix barriers to allow for the cytосkeletal reorganization necessary to drive cellular proliferation (23). This is supported by results of the current study showing lack of proliferation within three-dimensional collagen gels in the absence of MT1-MMP expression or in gels containing TIMP-2. Although all MT1-MMP–expressing cultures formed proliferative colonies in three-dimensional collagen gels, cells expressing the phosphomimetic T567E-MT1-MMP construct exhibited more invasive patterns of growth. Thus, it is interesting to speculate that both the presence and the phosphorylation status of MT1-MMP may control metastatic success. Metastatic disease is the main cause of death for women with epithelial ovarian carcinoma, as disseminated tumor cells attach to abdominal surfaces, anchor, and grow multiple secondary lesions that disrupt the function of essential organs (1, 2). A molecular-level understanding of metastasis is necessary for the development of therapies to inhibit i.p. spread and thereby improve the long-term survival of thousands of women with ovarian cancer. To this end, a more detailed understanding of what regulates the transition from primary tumor to free-floating MCA to life-threatening peritoneally anchored metastatic lesion may provide novel insight necessary to target i.p. therapies to appropriate multicellular populations.

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

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