Ribosome biogenesis and control of cell proliferation: p53 is not alone.

Authors: Giulio Donati¹*, Lorenzo Montanaro¹ and Massimo Derenzini²§.

Running Title: p53-independent control of cell proliferation

Key words: ribosome biogenesis, cell proliferation, p53, cell cycle control, cancer

Affiliations: ¹ Department of Experimental Pathology, Alma Mater Studiorum - Bologna University, Bologna Italy. ²Clinical Department of Radiological and Histocytopathological Sciences, Alma Mater Studiorum - Bologna University.
* Current address: Bellvitge Biomedical Research Institute (IDIBELL)-Hospital Duran i Reynals, Barcelona, Spain
§ corresponding author: ²Clinical Department of Radiological and Histocytopathological Sciences, Alma Mater Studiorum - Bologna University, Via Massarenti, 9 40138 Bologna Italy. E-mail: massimo.derenzini@unibo.it

Word Count: 2681
Figure number: 2
Abstract

Cell growth is a pre-requisite for cell proliferation, and ribosome biogenesis is a limiting factor for cell growth. In mammalian cells, the tumour suppressor p53 has been shown to induce cell cycle arrest in response to impaired ribosome biogenesis. Recently, p53-independent mechanisms of cell cycle arrest in response to alterations of ribosome biogenesis have been described. These findings provide a rational basis for the use of drugs that specifically impact ribosome biogenesis for the treatment of cancers lacking active p53, and extend the scenario of mechanisms involved in the relationship between cell growth and cell proliferation.
Introduction

Cell growth (increase in cell mass), and cell proliferation (increase in cell number), are two tightly linked phenomena. In cells stimulated to proliferate, a progressive increase in cell constituents occurs before division ensuring appropriately sized daughter cells (1). Cell growth is the consequence of an enhanced stimulation of protein synthesis that characterizes proliferating cells. The increased demand for protein synthesis is accomplished by changes in the rate of ribosome biogenesis, and ribosome biogenesis is the major metabolic effort in a proliferating cell (2). Ribosome biogenesis is the result of a series of coordinated steps occurring in the nucleolus. These include the transcription of ribosomal genes by RNA polymerase I (Pol I) to produce the 47S rRNA precursor, the modification and processing of this transcript to generate the mature 18S, 5.8S, and 28S rRNA, the import to the nucleolus of 5S rRNA and ribosomal proteins (RPs), the assembly of the rRNAs with the RPs to form the large 60S and the small 40S subunits of the mature ribosome, and the export of ribosomal subunits to the cytoplasm (3). Transcription of ribosomal genes requires the assembly of a specific multiprotein complex at the rDNA promoter containing Pol I and, in mammals, at least three other basal factors: the Transcription Initiation Factor I (TIF-I) A, the Selectivity Factor I (SL1), and the Upstream Binding Factor (UBF) (reviewed in 4) (See Fig.1).

It is well established that at the end of G1-phase, the so-called restriction point defines a limit beyond which the cell is committed to divide, independent of growth (5). Therefore, it is within the G1 phase that a proliferating cell has to perform its effort of supplying the components necessary for the transit from G1 to S phase, and it is at the end of G1 phase that the cell must determine whether this effort has been successfully accomplished. Data indicate that it is the amount of ribosomes produced that controls the G1-S phase transition thus regulating the cell cycle progression. In partially hepatectomized mice with induced conditional deletion of 40S ribosomal protein S6, hepatocytes with hindered ribosome production but not hindered protein synthesis failed to enter the S phase (6). Drug-induced stimulation or inhibition of rRNA synthesis caused an accelerated or delayed G1/S-phase progression, respectively, in rat hepatoma cells, due to an accelerated or delayed achievement of the appropriate amount of ribosomes during the G1 phase (7). Regarding the mechanism that regulates the cross-talk between ribosome biogenesis and cell cycle progression there is a general consensus on the key role played by the tumour suppressor p53. In fact, as described more extensively below, it has been clearly demonstrated that altered ribosome biogenesis is responsible for p53 stabilization and, therefore, for the arrest of cell cycle progression.
(reviewed in 8-10). On the other hand, very recent data indicate that p53-independent mechanisms are also activated that hinder proliferation as a consequence of altered ribosome biogenesis (11-15).

**Ribosome biogenesis and the control of cell cycle progression: the p53 paradigm.**

Defects in ribosome production cause p53 stabilization and induce a marked p53-dependent inhibition of cell proliferation (10). It is worth recalling that in proliferating mammalian cells the G1-S phase restriction point can be overwhelmed by the activation of the E2F transcription factor family (16), which regulate the expression of genes whose products are necessary for the synthesis of DNA, and therefore for the progression from G1 to S phase. The retinoblastoma protein (pRB), the product of the tumour-suppressor *RB1* gene, in its active hypophosphorylated form, is bound to E2Fs and prevents them from inducing E2F target genes. In the hyperphosphorylated form, pRB no longer binds to E2Fs, which can then activate their target genes. Phosphorylation of pRB is triggered in the early G1 phase by cyclin D-Cdk-4 and -6 complexes and is completed at the end of the G1 phase by cyclin E-Cdk-2 complexes. pRB phosphorylation is hindered by Cdk inhibitors: Cdk-4 and Cdk-6 are inhibited mainly by p16Ink4a whereas Cdk-2 is negatively regulated by p21(Cip1) and p27(Kip1) (Fig. 2a). p53 stabilization, occurring in response to a variety of cellular stresses, leads to induction of the Cdk-2 inhibitor p21, inhibition of cyclin E/Cdk2 complexes, and pRB-dependent cell cycle arrest at the G1/S phase check-point (16).

The mechanism of p53 stabilisation after perturbation of ribosome biogenesis is apparently the consequence of changes in functional and physical interactions of the tumour suppressor with MDM2. MDM2 negatively controls p53 activity in two ways: by binding to the protein and interfering with its transactivation activity, and by facilitating p53 proteasomal degradation thereby acting as an E3 ubiquitin ligase (17). As a consequence of altered ribosome biogenesis, several ribosomal proteins no longer used for ribosome construction bind to MDM2 and relieve its inhibitory activity toward p53. In fact, it was shown that the ribosomal proteins RPL5 (18), RPL11 (19-21), RPL23 (22,23) and RPS7 (24), bind MDM2, thus inducing p53 stabilisation by inhibiting MDM2’s E3 ubiquitin ligase function (also reviewed in 10). The inhibition of rRNA transcription, induced either by drugs such as actinomycin D, 5-fluorouracil, mycophenolic acid, or by depletion of essential Pol I complex components (10, 15, 25), stabilizes p53 by causing RP-induced inactivation of MDM2. These data are consistent with a mechanism by which altered ribosome biogenesis inhibits cell proliferation in a strictly p53-dependent manner through the activation of the ribosomal proteins-MDM2-p53 pathway.
p53-independent mechanisms

The regulation of cell cycle in response to defects in ribosome biogenesis, far from being exclusive to mammalian cells, also occurs in organisms where p53 does not regulate the cell cycle, or is not even present. In the metazoan Drosophila Melanogaster, p53 does not regulate p21 transcription, nor the cell cycle (26). In this organism, haploinsufficiency of several RP genes gives rise to the Minute phenotype. Cells from Minute mutants are the same size as wild type, but their proliferation rate is reduced (27), implying the existence of a mechanism controlling cell cycle progression in response to hampered ribosome biogenesis. Depletion of the RNA polymerase I co-factor TIF-IA induces a p53-independent proliferation arrest in Drosophila cells (28). Furthermore, in the yeast S. cerevisiae, where no p53 homologue has been found, defective ribosome biogenesis delays the cell cycle progression to S phase before any variation in protein synthesis capacity ensues (29).

The presence of a p53 independent pathway regulating the relationship between ribosome biogenesis and cell proliferation in mammalian cells was first suggested by the observation that selective inhibition of rRNA synthesis by actinomycin D was able to induce a perturbation of cell cycle progression also in cells silenced for p53 expression, although milder than that occurring in the presence of p53 (30). In the past two years some light has been shed on how altered ribosome biogenesis can hinder cell proliferation in a p53-independent way. A good example is constituted by pescadillo depletion. The pescadillo nucleolar protein plays a role in the processing of pre-rRNA molecules during assembly of 60S ribosomal subunits, through formation of the PeBoW complex with Bop1 and WDR12 proteins (11). Depletion of pescadillo inhibits ribosome biogenesis, and, similar to other mechanisms that hinder ribosome production, stabilises p53 leading to cell cycle arrest (11). However, pescadillo gene knockdown impaired cell proliferation in cells harbouring mutated p53, indicating that in these cells, the observed cell cycle control mediated by pescadillo protein was p53-independent (13). In fact, pescadillo depletion resulted in decreased expression of cell cycle protein cyclin D1 and up-regulation of the CDK inhibitor p27, with the consequent marked reduction of pRB phosphorylation (13).

Another p53-independent mechanism of cell proliferation arrest induced by impaired ribosome biogenesis was demonstrated by the down-regulation of PIM1 expression caused by ribosomal protein deficiency and various other ribosomal stressors (14). PIM1 is a constitutively active serine/threonine kinase regulated by cytokines, growth factors and hormones that has been shown to reduce the activity of p27Kip1 and increase its degradation (31). Furthermore, PIM1 kinase has been shown to interact with the ribosomal protein RPS19 and it has been demonstrated that depletion of RPS19 or other ribosomal proteins caused increased PIM1 kinase proteasome
degradation (14). The down-regulation of PIM1 kinase stabilized and activated p27Kip1, thus causing a block in cell cycle progression regardless of p53 status (14).

The data reported above indicate that changes in the biogenesis of single ribosome subunit hinder cell proliferation in cells with activated or inactivated p53. Moreover, data have recently been reported showing that specific inhibition of rRNA transcription by depletion of POLR1A hindered cell cycle progression in cancer cell lines with inactivated p53 (15). This was due to the fact that the inhibition of rRNA synthesis decreased the expression of E2F-1. Normally, E2F-1 is protected from proteasome-mediated degradation by the interaction with MDM2, which prevents the binding of other E3 ligases responsible for E2F-1 ubiquitination (32). The inhibition of rRNA synthesis releases the ribosomal protein L11, which, by binding to MDM2, prevents its stabilising function on the E2F-1 protein (15). The down-regulation of the E2F-1 protein caused a reduction in the expression of the E2Fs target genes that are necessary for the entry and progression through the S phase. RB1 silencing rescued the expression of these genes completely and prevented the effect of the rRNA synthesis inhibition on cell proliferation. The inhibition of rRNA synthesis is not always associated with a down-regulation of E2F-1. In fact, actinomycin D, cisplatin and etoposide – drugs that inhibit rRNA transcription – caused an accumulation of E2F-1 protein (33). However, these drugs are also DNA-damaging agents, and it has been demonstrated that these agents increase E2F-1 half-life and its transcriptional activity (33). Importantly, reducing the synthesis of rRNA by TIF-IA silencing, i.e. a non DNA-damaging procedure to inhibit rRNA transcription, reduced the expression of the E2F-1 protein (15).

Finally, another mechanism defining the relationship between ribosome biogenesis and cell proliferation in cells with inactive p53 is the down-regulation of c-Myc in response to ribosome stress. c-Myc, in addition to stimulating cell proliferation, controls all the steps of ribosome biogenesis: it increases Pol I activity by facilitating the recruitment of SL1 to promoters, stimulates ribosomal protein synthesis by increasing Pol II transcription, and enhances Pol III transcription by activating TFIIB (34). It was found that, in response to ribosome biogenesis inhibition, RPL11 binds to c-Myc reducing its transcriptional activity, and to c-Myc mRNA promoting its degradation (12). The RPL11-mediated down-regulation of c-Myc reduced cell proliferation, and this effect was also observed in p53-null cells, thus demonstrating another p53-independent mechanism of proliferation control (12).

In conclusion, there is now evidence that ribosomal stress hinders cell proliferation in mammalian cells with and without p53. The p53-dependent and –independent mechanisms by which ribosome biogenesis controls cell cycle progression are schematically summarised in Fig. 2.
Ribosome biogenesis inhibition and chemotherapy of p53-deficient cancer

The disruption of p53 function occurs in more than 50% of human tumours, thus representing the most frequent gene alteration in cancers (35). Commonly used chemotherapeutic agents in human cancer induce some cell stress, which activates the p53-mediated cell cycle blockage and/or apoptosis. Therefore, tumours without functioning p53 are less sensitive to cytostatic and cytotoxic drug treatment than those harbouring wild type p53 (36). The p53-independent control of ribosome biogenesis on cell proliferation may explain some recent data demonstrating an anti-proliferative effect of Pol I complex inhibitors on cancer lacking functional p53. Indeed, a study conducted by Drygin and colleagues (37) showed that targeting factors of the rDNA transcription complex represents a promising strategy for chemotherapeutic treatment of cancers, both p53 proficient and deficient. During a screening assay for agents that selectively inhibit Pol I transcription relative to Pol II transcription, CX-5461 (Cylene Pharmaceuticals Inc.) a potent small-molecule that selectively inhibits Pol I–driven transcription was identified (37). It was shown that CX-5461 disrupts the binding of the SL1 transcription factor to the rDNA promoter, thus preventing the initiation of rRNA synthesis by the Pol I multiprotein complex. In fact, SL1 mediates specific interactions between the rDNA promoter region and the Pol I enzyme complex by recruiting Pol I, together with a series of Pol I–associated factors, to rDNA. CX-5461 exhibited anti-proliferative activity in numerous cancer cells in vitro, and, importantly, this occurred in wild-type and mutant p53 cell lines. In this study, the authors found that the mechanisms leading to cell death after inhibition of rRNA synthesis involved autophagy and senescence. Unfortunately, potential mechanisms affecting the control of cell cycle progression were not investigated. However, accordingly to the results obtained by Pol I and TIF-IA depletion, it is likely that the inhibitory effect on proliferation of cells lacking active p53 observed after CX-5461 treatment might be mediated by a down-regulation of E2F-1.

Perspectives

Studies on p53-independent mechanisms regulating the relationship between cell growth and cell cycle progression are expected to not only increase our knowledge of the homeostatic control of cell proliferation, but also reveal new aspects of the process of tumourigenesis and potential chemotherapeutic therapies for p53-negative cancers. The stabilisation of p53 following ribosomal stress plays an important role against neoplastic transformation (38). An altered ribosome biogenesis may be responsible for a failure in the p53 activation pathway, and this may facilitate tumour onset (39). Similarly, functional inhibition of p53-independent mechanisms for opposing cell cycle progression after ribosomal stress could
down-regulate the tumour suppressor potential of the cell. Experimental studies of the effect of alterations of factors involved in these mechanisms (such as Pim1, pescadillo, Bop1) on tumourigenesis, together with the analysis of the integrity of their function in human tumours could possibly point out their involvement in the process of neoplastic transformation and identify new targets for therapeutic applications.

The results reported in this review strongly encourage studies to develop inhibitors of ribosome biogenesis that down-regulate E2F-1 expression. Furthermore, considering the role of pRB in controlling E2F-1 function, it might be of interest to combine the ribosome biogenesis inhibitors with drugs that hinder the activity of the CDKs. The resulting reduction of E2F-1 expression, together with the increased E2F-1 binding to hypophosphorylated pRB can be reasonably expected to have a very strong impact on cell proliferation in cancers lacking active p53. A similar chemotherapeutic approach should be highly effective on those cancers in which this transcription factor is known to exert a major role in the biology of the transformed cells. For example, aberrant E2F-1 activity is indeed a critical factor for the initiation and progression of B-cell Burkitt’s lymphoma (40) and for the emergence of castration-resistant prostate cancer (41).

The use of drugs targeting the CDKs in combination with ribosome biogenesis inhibitors that down-regulate E2F-1 expression may be also very useful for treatment of cancers characterised by Myc overexpression, which is frequently observed in human tumours of different origin. In fact, these drugs may exert a p53-independent anti-proliferative activity by both down-regulating c-Myc mRNA expression as a consequence of an increased availability of RPL11 after the inhibition of rRNA synthesis, and by reducing E2F-1 expression and activity.

Acknowledgements: We thank Dr. Angela Drew for critically editing the manuscript.

Grant Support: This work was funded by the Pallotti legacy for cancer research and RFO funds from Bologna University to LM and MD. GD was supported by a grant fellowship by the Vanini Cavagnino Legacy (Centro Interdipartimentale per le Ricerche sul Cancro “G.Prodi”, Bologna University.
References


**Figure legends**

**Fig. 1.** Schematic representation of the main processes involved in ribosome biogenesis. One ribosomal gene unit is repeated hundreds of times in the genome and includes the promoter and the transcribed region. The major basal factors binding to the promoter region and required for ribosomal genes transcription are shown in the inset. After transcription, the 47S pre-rRNA is processed. The mature rRNAs are assembled with ribosomal proteins (RPs) to generate mature ribosomal sub-units. In particular 18S is assembled with small subunit RPs to generate the 40S small ribosomal subunit, while 5.8S and 28S rRNA are assembled with 5S rRNA, (which is transcribed by PolIII in the nucleus), and with large subunit RPs to form the 60S large ribosomal subunit. Mature 40S and 60S subunits are then exported to the cytoplasm to constitute the ribosomes.

**Fig. 2.** a) Schematic and simplified representation of the mechanisms leading to the cell cycle progression from G1 to S phase. After cell cycle entry cyclin D-Cdk-4 and -6 and cyclin E-Cdk-2 complexes phosphorylate pRB during G1 phase. Phosphorylation of pRB leaves E2F1 free to activate the E2F1 target genes whose products are necessary for entry and progression through S phase.


c) p53-independent cell cycle arrest after ribosomal stress. Alterations in ribosomal assembly and ribosomal processing increase the level of the cyclin-dependent kinase inhibitor p27 thus reducing pRB phosphorylation. This should lead to a decreased activation of E2F-1. Changes of ribosomal biogenesis caused by inhibition of factors constituting the complex of rDNA gene promoter increases RP availability. RP binding to MDM2 lowers its protective effect on E2F-1. The amount of E2F-1 is therefore reduced by proteasome-mediated digestion with a consequent reduced activation of the E2F-1 target genes.
Normal cell cycle progression

Mitogens/growth factors → cyclin D,E → CDK 4/6,2 → p16

Ribosomal stress: p53 dependent mechanism of cell cycle arrest

Ribosomal assembly → free RP → MDM2 → p21 → cyclin D,E → CDK 4/6,2

Ribosomal stress: p53 independent mechanisms of cell cycle arrest

Ribosomal assembly (RPS19) → PIM1 → p27 → cyclin D,E → CDK 4/6,2 → E2F target genes

Downloaded from cancerres.aacrjournals.org
Ribosome biogenesis and control of cell proliferation: p53 is not alone.

Giulio Donati, Lorenzo Montanaro and Massimo Derenzini

*Cancer Res* Published OnlineFirst January 26, 2012.

**Updated version**
Access the most recent version of this article at:
doi:10.1158/0008-5472.CAN-11-3992

**Author Manuscript**
Author manuscripts have been peer reviewed and accepted for publication but have not yet been edited.

**E-mail alerts**
Sign up to receive free email-alerts related to this article or journal.

**Reprints and Subscriptions**
To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

**Permissions**
To request permission to re-use all or part of this article, use this link http://cancerres.aacrjournals.org/content/early/2012/01/24/0008-5472.CAN-11-3992. Click on "Request Permissions" which will take you to the Copyright Clearance Center's (CCC) Rightslink site.