A Novel Tankyrase Inhibitor Decreases Canonical Wnt Signaling in Colon Carcinoma Cells and Reduces Tumor Growth in Conditional APC Mutant Mice

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Introduction

The Wnt/β-catenin signaling pathway is a key regulator in numerous cellular processes including stem cell maintenance, fate decision, and cell-cycle control (1, 2). The main denominator of canonical Wnt signaling, β-catenin, has several cellular functions. At the cell membrane, it is associated with E-cadherin and participates in the formation of the adherens junctions (3). In the cytoplasm, β-catenin can form complexes with a multitude of partners, including the β-catenin destruction complex. Inhibition of Wnt/β-catenin signaling therefore is an attractive strategy for anticancer drugs. In this study, we have identified a novel small molecule inhibitor of the β-catenin signaling pathway, JW55, that functions via inhibition of the PARP domain of tankyrase 1 and tankyrase 2 (TNKS1/2), regulators of the β-catenin destruction complex. Inhibition of TNKS1/2 poly(ADP-ribosyl)ation activity by JW55 led to stabilization of AXIN2, a member of the β-catenin destruction complex, followed by increased degradation of β-catenin. In a dose-dependent manner, JW55 inhibited canonical Wnt signaling in colon carcinoma cells that contained mutations in either the APC (adenomatous polyposis coli) locus or in an allele of β-catenin. In addition, JW55 reduced XWnt8-induced axis duplication in Xenopus embryos and tamoxifen-induced polyposis formation in conditional APC mutant mice. Together, our findings provide a novel chemotype for targeting canonical Wnt/β-catenin signaling through inhibiting the PARP domain of TNKS1/2. Cancer Res; 1–11. ©2012 AACR.
In the nucleus, β-catenin exerts a number of functions, such as binding to the transcription factor TCF/LEF by replacing Groucho and activation of downstream transcription of genes, including c-Myc, Cyclin D1, and AXIN2 (2). Nuclear β-catenin levels are not only determined by the rate of β-catenin import to the nucleus but also by “nuclear trapping” of β-catenin through factors such as TCF/LEF (17).

In general, mutations in genes encoding central components of the Wnt/β-catenin pathway and the destruction complex lead to the accumulation of nuclear β-catenin and contribute to tumor initiation and progression (18, 19). Wnt activating mutations are found in a broad range of solid tumors including colon cancer, gastric cancer, hepatocellular carcinoma, breast cancer, medulloblastoma, melanoma, non–small cell lung cancer, pancreas adenocarcinoma, and prostate cancer (20). It has been well established that the majority of intestinal neoplasia with deregulated Wnt signaling harbor truncating mutations in both alleles of the APC tumor suppressor gene (21).

Considerable efforts have been made to identify drugs that inhibit Wnt/β-catenin signaling (18, 22–25). Published inhibitory drugs that interact with biotargets directly associated with canonical Wnt signaling include WAY-316606 for SFRP (26), niclosamide for frizzled (27), NCS668036 for Dsh (28), pyrvinium for CK1 (24), XAV939 and IWR-1 for TNSK1/2 (4, 22), 2,4-diamino-quinazoline, quercetin, PKF115-584 and ICG-001 for TNKS1/2, in particular, is a promising biotarget for pharmaceutical reagents (4, 6). TNSK1/2 is not only involved in controlling canonical Wnt signaling but has been associated with further cellular processes through either protein poly(ADP-ribosyl)ation (PARsylation) or by protein complex formation. These are (i) telomere maintenance by interacting with TRFI (33), (ii) spindle formation and stabilization by binding to NuMA (33), and (iii) glucose metabolism by regulating GLUT4 transport through IRAP (33).

In this work, we have identified a novel TNKS inhibitor, JW55, which inhibits the PARP domain of TNSK1/2, leading to the stabilization of AXIN2 followed by increased degradation of β-catenin. JW55 efficiently decreases canonical Wnt signaling in colon carcinoma cell lines, in a Xenopus axis duplication assay, and in tamoxifen-induced tumors in ApcKO/CKO/Lgr5-CreERT2 mice. The identified chemotype JW55 may serve as an attractive start point for the development of novel cancer therapeutics.

Materials and Methods

Cell lines and luciferase assays

All cell lines were purchased from American Type Culture Collection and maintained and treated according to the supplier’s recommendations. Reporter assays were carried out as previously described (34).

Incucyte cell growth measurement assay

A total of 1,000 SW480 or RKO cells were seeded in 96-well plates. The day after, the cell culture medium was exchanged to solutions that contained 0.1% dimethyl sulfoxide (DMSO) or 10 μmol/L JW55 for RKO cells and 0.1% DMSO or 10, 5, or 1 μmol/L JW55 for SW480 cells. All samples consisted of a minimum of 6 replicates. The plate was incubated in an IncuCyte (Essen BioScience) inside a cell culture incubator. Images were captured every second hour to monitor proliferation.

Colony assay

A total of 400 SW480, RKO, or HeLa cells were seeded in 6-cm plates and 1% PBS. The day after, the medium was exchanged to 0.05% DMSO or 5 μmol/L JW55 for RKO and HeLa cells, and 0.1% DMSO or 5, 1, or 0.1 μmol/L JW55 for SW480 cells (minimum 3 replicates). The cells were grown until colonies appeared in the control plates and medium was changed twice a week. The plates were stained and fixed with 0.2% methylene blue in methanol and washed with PBS. The colonies were counted manually with a colony counter (Scienceware). Representative pictures of colonies were captured with a Leica MZ Apo Stereomicroscope connected to a Leica DC200 camera and Adobe Photoshop.

Tumor initiation and treatment in ApcKO/CKO/Lgr5-CreERT2 mice

Seven 12-week old female ApcKO/CKO/Lgr5-CreERT2 mice were injected intraperitoneally with 25 mg/kg of tamoxifen (Sigma) diluted in an ethanol and corn oil (ratio 1:4). The mice were randomized into 2 groups and treated with either JW55 (100 mg/kg) or vehicle (DMSO). Daily per oral applications started the day after and continued for 3 weeks. The mouse body weight was measured twice a week. The mice were sacrificed and the intestines were dissected, washed in PBS, and fixed in formaldehyde [10% solution (v/v) in PBS]. The small intestines were stained using 1% methylene blue and the entire length of the intestine was stained with hematoxylin and eosin (H&E). Fixed and stained intestines were sectioned and stained with hematoxylin and eosin (H&E). Fixed colon sections were embedded in paraffin sectioned and stained with hematoxylin and eosin (H&E). Fixed colons were embedded in paraffin, sectioned and stained with an anti–β-catenin antibody (610153; BD Transduction Laboratories; 1:200 dilution). The number and size of the intestinal lesions were quantified by the Eclipse program (ViDiTo).

For other materials and methods, please see previous descriptions (34) and Supplementary Materials and Methods.

Results

JW55 is a potent and selective inhibitor of the canonical Wnt pathway

To identify chemical compounds that inhibit the canonical Wnt signaling pathway, a library consisting of 37,000 small molecules (ChemBioNet) was screened using HEK293 cells that were stably transfected with a SuperTOP-d1EGFP reporter (34). The vector contains a synthetic TCF-responsive promoter (SuperTOP: 7 x TCF binding sites) that initiates the expression of destabilized d1EGFP upon activation of canonical Wnt signaling (34). Canonical Wnt signaling was activated using conditioned media (CM) from L Wnt3a expressing cells. The screen resulted in 77 primary hits including the previously described compounds JW67 and JW74 (34) and JW55 (Fig. 1A, 1B).
JW55 inhibits Wnt signaling and colorectal cancer cell growth

Figure 1. JW55 specifically reduces canonical Wnt signaling in reporter cells and in Xenopus embryos. Reporter controls in 0.1% DMSO. A, left, luciferase reporter activity in HEK293 cells, transiently transfected with ST-Luc and Renilla plasmids and treated with JW55 at 0.1 to 10 μmol/L. +, 30% Wnt3a-CM; –, without Wnt3a-CM. The spotted line shows the IC50 value level. Right, chemical structure of JW55. B, effect of 1 and 10 μmol/L JW55 in NIH/3T3 Shh Light II cells (Gli1-Luc reporter) that were activated with 50% Shh-CM were not inhibited by 10 or 1 μmol/L JW55. Also, 10 or 1 μmol/L of JW55 in NIH/3T3 Shh Light II cells (Gli1-Luc reporter) that were activated with 50% Shh-CM were not inhibited by 10 or 1 μmol/L JW55.

The selectivity and specificity of JW55 was further tested in an in vivo Xenopus axis duplication assay, a highly stringent test for examining inhibitors of canonical Wnt signaling. When injected into the ventral blastomeres of 4-cell stage Xenopus laevis embryos, XWnt8 mRNA induced Wnt signaling resulting in the formation of a second body axis. By using selective Wnt inhibitors, the axis duplication can be inhibited and a normal phenotype restored (34). Coinjection of 10 pg XWnt8 and 2 pmol of JW55 resulted in a significant 51% reduction of the axis duplication incidence in developing frog embryos when compared with the DMSO vehicle control (z-test, P = 0.001; Fig. 1B, left). This result provided evidence that JW55 acts as a specific and effective inhibitor of canonical Wnt signaling.

JW55 regulates β-catenin stability at the level of the destruction complex

To identify the interference point of JW55 with canonical Wnt signaling, the effect of JW55 was monitored after LiCl-induced activation of the pathway. The activity of GSK3β, which is part of the β-catenin destruction complex, is reduced in the presence of LiCl (35), rendering the N-terminal phosphorylation of β-catenin by GSK3β ineffective. This causes nuclear accumulation of β-catenin and active Wnt signaling. HEK293 cells, transiently transfected with ST-Luc and Renilla, were incubated with 25 mmol/L LiCl and different doses of JW55. The LiCl-activating effect was counteracted by JW55 in a dose-dependent manner with an IC50 value of 360 nmol/L, indicating that JW55 acts at the level or downstream of the destruction complex (Fig. 2A, left and Supplementary Fig. S1B).

Next, the effect of JW55 on regulating the activity of β-catenin was tested. We used (i) wild-type β-catenin and (ii) β-catenin with point mutations in the N-terminal phosphorylation sites (S33, S37, T41, and S45) that resists degradation and functions as dominant active (da-Cat). Plasmids containing full-length β-catenin or da-Cat, along with ST-Luc and Renilla, were transiently transfected in HEK293 cells. The expression of wild-type β-catenin led to an increased ST-Luc reporter activity that could be reduced by 55% when the transfected cells were exposed to 10 μmol/L of JW55 (normality test failed, rank sum test: P = 0.001; Fig. 2A, right). In contrast, the activation of the pathway by da-Cat could not be inhibited by 10 μmol/L JW55, further indicating that JW55 acts at the level or downstream of the destruction complex (normality test failed, rank sum test: P = 0.457; Fig. 2A, right).

JW55 inhibits canonical Wnt signaling in colorectal cancer cell lines in vitro

Mutations in the APC gene, which occur in nearly all colorectal cancers (CRC; ref. 18), lead to ineffective degradation of β-catenin and aberrant upregulation of Wnt signaling. The cell lines SW480 and HCT-15 (mutated in codon 1338 and 1417 of the APC gene, respectively) were stably transfected with ST-
Figure 2. JW55 inhibits canonical Wnt signaling in CRC cells. A, the control bars are shown in gray and JW55-treated samples are shown in black. Left, inhibition of GSK3β [25 mmol/L LiCl]) in HEK293 cells, transiently transfected with ST-Luc and Renilla, is counteracted by 0.1 to 10 μmol/L JW55. +, 25 mmol/L LiCl; −, no LiCl. Controls in 0.1% DMSO. The mean values of 3 independent assays and SDs are shown. Right, HEK293 cells transiently transfected with ST-Luc and Renilla plus full-length β-catenin or da-Cat. +, with β-catenin plasmids; −, without β-catenin. Controls in 0.1% DMSO. Ten μmol/L JW55 inhibited luciferase activity induced by ectopic wild-type β-catenin (P < 0.05), whereas da-Cat induction was unaffected. n = total number of measurements from multiple independent assays. The SDs are shown as error bars. B, JW55-mediated reduction of luciferase activity in different CRC cell lines stably transfected with ST-Luc and Renilla. Left, a dose-dependent reduction of ST-Luc activation was detected in HCT-15 and SW480 cells harboring mutant APC. Right, JW55-mediated reduction in HCT116 cells containing a single allele mutation in S45 of β-catenin. The mean values and the SE of several independent assays are shown. C, 25 or 10 μmol/L JW55 reduced the relative expression of AXIN2, SP5, and NKD1 mRNA in the CRC cells SW480 and DLD-1 as shown by real-time RT-PCR analysis. The means of 3 independent experiments are shown along with error bars depicting SDs.

JW55 destabilizes β-catenin by increasing cytoplasmic AXIN2 levels

Previous studies have shown that an increase in AXIN2 steady-state levels induced by TNKS1/2 or CK1α inhibitors is accompanied by a decrease in β-catenin concentrations (4, 22, 24, 34). Increased levels of AXIN2 protein promote degradation of β-catenin even in cells with truncated APC (4, 22, 38). Western blot analysis of SW480 cells lysates revealed a dose-dependent increase of cytoplasmic AXIN2 after JW55 treatment (range: 10 μmol/L–100 nmol/L; Fig. 3A and Supplementary Fig. S2). Furthermore, an antibody against the active and nonphosphorylated form of β-catenin (active β-catenin, ABC) identified reduced levels of β-catenin in the cytoplasm of JW55-treated cells (Fig. 3A and Supplementary Fig. S2). In addition, a modest reduction of total β-catenin and a substantial decrease of nuclear ABC were observed, and an increase in phosphorylated β-catenin (p9-catenin) levels was seen indicating ongoing β-catenin degradation (Fig. 3A and Supplementary Fig. S2).

To gain further insight into the changes in cellular distribution of AXIN2 and β-catenin, JW55-treated SW480 cells were analyzed by immunofluorescence. A general reduction of total β-catenin, both in the cytoplasmic and nuclear compartments, was detected at the doses of 5 and 1 μmol/L (equal shutter
Supplementary Fig. S3). Confocal microscopy (equal shutter speeds) revealed, in accordance with the Western blot analysis, that the levels of cytoplasmic AXIN2 were significantly increased (Fig. 3B) and large protein foci, probably representing accumulated destruction complexes, were observed (Fig. 3B, arrows). Clusters of colocalized cytoplasmic β-catenin and AXIN2 have previously been detected in SW480 cells after treatment with the Wnt antagonist JW74 (34). Similar to JW74, enhanced phosphorylation and resulting degradation of β-catenin after JW55 exposure seemed to be orchestrated by stabilization of AXIN2 in the destruction complex (Fig. 3 and 5).

JW55 specifically inhibits the PARsylatation activity of TNKS1 and TNKS2

By inhibiting the PARP domain of TNKS1/2, XAV939, IWR-1, and JW74 prevent auto-PARsylatation of TNKS1/2 and PARsylatation of AXIN2 (4, 22, 39, 40). This leads to a stabilization of AXIN2, an accumulation of proteins in the destruction complex followed ultimately by an increased degradation of β-catenin (4, 22). To test whether JW55 decreased canonical Wnt signaling by directly inhibiting the PARP domain of TNKS1/2, we carried out biochemical assays for monitoring the activity of TNKS1/2 and PARP. JW55 decreased auto-PARsylatation of TNKS1/2 in vitro with IC50 values of 1.9 μmol/L and 830 nmol/L, respectively (Fig. 4A and 4B, XAV939, IWR-1).

Figure 3. JW55 mediates increased AXIN2 stability and induces β-catenin degradation. A, Western blot analysis of lysates from SW480 cells after incubations with JW55 (24 hours). Controls in 0.1% DMSO). Antibodies against AXIN2, ABC (active β-catenin), and pβ-catenin (N-terminal phosphorylated β-catenin) were used. ACTIN (cytoplasmic) or LAMIN B1 (nuclear extracts) documented equal protein loading. Full-length and uncropped blots are shown in Supplementary Fig. S2. The blots show representative data derived from multiple experiments. B, cellular redistribution of β-catenin and stabilization of AXIN2 in SW480 cells incubated with 5 μmol/L JW55 (48 hours). The arrows show clustered AXIN2 and β-catenin. The confocal microscopy images are representative examples from one of several independent experiments.

Figure 4. JW55 specifically inhibits TNKS1, TNKS2 but not PARP1 in biochemical assays. A, JW55 B, XAV939 (logarithmic scale). C, the calculated IC50 values are displayed in the table. The mean values represent 2 independent experiments and the error bars show SDs. D, immunoblotting of TNKS1/2 after exposures of SW480 cells to JW55, XAV939, or IWR-1.
Supplementary Fig. S1C). However, in contrast to XAV939 (ref. 4; Fig. 4A and Supplementary Fig. S1D), but similar to IWR-1 (4), JW55 exhibited no inhibition of PARP1 at doses up to 20 μmol/L (Fig. 4A). Although further details may emerge, present evidence suggests that JW55 decreases canonical Wnt signaling by specifically inhibiting the PARP domain of TNKS1/2 (Fig. 5), although leaving the activity of the PARP domain of at least PARP1 unaffected. Molecular docking of JW55 into the adenosine site of the human PARP domain of TNKS2 using the structure of the PARP domain in complex with IWR-1 (X-ray, PDB code 3UA9) reveals that the binding site and position of JW55 in the model is similar to IWR-1 (40), but different from XAV939 (Supplementary Fig. S4). Binding of JW55 to recombinant TNKS2 protein was also observed by using a fluorescence polarization competition assay, with a structurally similar but inactive analog as a negative control (ref. 41; Supplementary Fig. S5). Interestingly, opposite to the effect of XAV939 (4), a decrease in endogenous TNKS1/2 levels was detected after exposure to JW55 and IWR-1 in SW480 cells.

JW55 reduces growth of SW480 colon cancer cells

CRC cells can enter cell-cycle arrest as a result of antagonized canonical Wnt signaling (42–44). Various proliferation assays were carried out to see whether JW55-mediated inhibition of canonical Wnt signaling would affect CRC cell growth. First, SW480 cells were incubated with 10 μmol/L JW55 and labeled with bromodeoxyuridine (BrdUrd) and propidium iodide (PI). The subsequent flow cytometry cell-cycle analysis revealed that JW55 treatment lowered the proportion of cells in the S phase (from 28.4% in DMSO-treated controls to 22.2%), raised moderately the cell fraction in the G1 phase (from 37.3%–38.8%) and increased the number of cells in G2–M phase (from 34.3%–39%; Fig. 6A).

Next, we monitored the cell proliferation kinetics using IncuCyte. SW480 cells were exposed to 10, 5, or 1 μmol/L of JW55 for 9 days, and the confluency was measured every 2 hours. All doses of JW55 reduced SW480 cell growth and the most robust effect was detected at a dose of 10 μmol/L JW55. The confluency was reduced to 55% relative to the DMSO control at the end of experiment (Fig. 6B). In parallel, the CRC cell line RKO, which contains wild-type APC and β-catenin and exerts Wnt-independent cell growth, was used as a control. RKO cells reached 100% confluency within 7 days and were not affected by treatment with 10 μmol/L JW55 (Fig. 6B).

Furthermore, proliferation of SW480 cells was examined by consecutive passages in the presence of 5, 2.5, or 1 μmol/L JW55. A dose-dependent decrease of cell numbers over 3 passages was noted in SW480 cells in the presence of JW55, whereas HeLa cells (Wnt-independent cervical cancer cells) remained unaffected (Fig. 6C). Finally, SW480 cells were grown under low serum conditions (1% FBS) along with various concentrations of JW55 (5, 1, and 0.1 μmol/L) and the formation of colonies was quantified. We observed a concentration-dependent reduction of colony numbers in SW480 cells and no reduction in colony numbers in the control cell lines HeLa and RKO when cultured in 5 μmol/L JW55 (Fig. 6D, left). All SW480 colonies, which formed in the presence of JW55, were substantially smaller (Fig. 6D, right).

Taken together, these data showed that JW55-mediated inhibition of canonical Wnt signaling resulted in reduced cell-cycle progression, proliferation, and colony formation in the CRC cell line SW480 in vitro.
Recently, the leucine-rich repeat containing G-protein–coupled receptor 5 (Lgr5) was established as a specific marker for intestinal epithelium stem cells (ISC). The knockin mouse Lgr5-EGFP-IRES-CreERT2, further referred to as Lgr5-CreERT2, expresses a tamoxifen-regulated variant of Cre recombinase that is under the control of the Lgr5 locus (45). The Cre-mediated excision of the floxed exon 14 in mice with conditional Apc alleles (CKO) changes the reading frame downstream of the deletion. This results in the production of a truncated and nonfunctional 580 amino acid (aa) Apc polypeptide (46) that relates to the homozygous mutations seen in the intestinal cancer model mouse ApcMin (multiple intestinal neoplasia, Min; ref. 47).

Lgr5-CreERT2 mice were intercrossed with ApcCKO/CKO animals and multiple tumors were observed after tamoxifen injections in the small intestine. To evaluate the JW55-mediated decrease of intestinal tumor development in vivo, ApcCKO/CKO Lgr5-CreERT2 mice were injected intraperitoneally with a 25 mg/kg single dose of tamoxifen. A day after, daily per oral applications of JW55 (100 mg/kg; 3 females) or vehicle (DMSO; 4 females) were initiated. The dose of 100 mg/kg was chosen to counteract the rapid liver metabolism of JW55 as indicated by the human liver microsome stability analysis (t1/2 = 10.1 minutes; Supplementary Fig. S7A).

No measurable effects on mouse body weight were noticed throughout the experiment period (Supplementary Fig. S7B). After 21 days, the mice were sacrificed and the dissected intestines were embedded in paraffin and sectioned. Immunohistochemical staining revealed that the neoplastic lesions expressed β-catenin and the mouse ISC marker Ephrin type-B receptor 2 (EphB2), indicating aberrant activation of canonical Wnt signaling in the tumor tissue (Fig. 7B; refs. 44, 48–50). The β-catenin–stained colon adenomas contrasted with the surrounding healthy mucosa, and image analysis software (Ellipse) was used to quantify the number and areas of β-catenin–positive regions in the tumors. 

**Figure 6.** Inhibition of canonical Wnt signaling by JW55 promotes cell-cycle arrest and specifically reduces proliferation in SW480 CRC cells. A, left, fluorescence-activated cell sorting scatter plots showing SW480 cells labeled with BrdUrd and PI after incubation in 10 μmol/L or 0.1% DMSO. The different cell-cycle phases, G1, S, and G2/M, are gated in 3 different compartments. Right, table shows a representative percent cell-cycle phase distribution (mean ± SEM) of cells under different conditions. B, cell growth curve as measured by IncuCyte shows a concentration-dependent decrease of proliferation in SW480 cells compared with the Wnt-independent CRC control cell line RKO. SW480 and RKO cells were incubated with 0.1% DMSO or exposed to JW55. The plot shows the mean value of 2 independent experiments and all relative SDs are below 10%. C, relative cell count (%) of SW480 and RKO controls in 0.05% DMSO. The graphs depict the mean values of several experiments and the error bars show the SDs. D, a colony assay that shows a specific decrease in colony formation of SW480 cells cultured in various doses of JW55 and 1% DMSO. Left, the bar chart shows a dose-dependent reduction of SW480 colonies and unaffected growth of the control cell lines HeLa and RKO. Controls in 0.05% DMSO. The graphs show the mean values of several measurements from multiple experiments along with error bars depicting SDs. Right, the images from a representative assay show that the general SW480 colony sizes are smaller in increased doses of JW55.
**A**

DMSO  
JW55  
Wt

**B**

DMSO  
JW55  
DMSO  
JW55

**C**

Ileum (small intestine)

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<th>DMSO (Vehicle), n = 4</th>
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<td>Mean total tumor area (mm²)</td>
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**D**

Wt  
JW55

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lesions in the colon (Supplementary Fig. S7C). As it was impossible to distinguish the borders of individual tumors in the small intestine (ileum), only the total tumor area per mouse was recorded. In the ileum, a significant reduction of the total tumor area was observed after JW55 injections (mean: 2.93 mm² and median: 2.95 mm²) when compared with the control group (mean: 8.84 mm² and median: 9.51 mm²; normality test failed, rank sum test: P = 0.003; Fig. 7A and C, top panel). In the colon, the tumor count was substantially reduced in JW55-treated mice (mean: 9.7 and median: 6.0) when evaluated against the control group (mean: 33.8 and median: 26.0; normality test failed, rank sum test: P = 0.057; Fig. 7C, bottom panel). Furthermore, a significant decrease of the total tumor area was noticed after injections with JW55 (mean: 0.22 mm²) when compared with the control group (mean: 0.154 mm²; normality test failed, rank sum test: P = 0.0025 mm²) when compared with the untreated group (mean: 0.0049 mm²; Students t test: P = 0.043; Fig. 7C, bottom panel). Interestingly, the proportion of cells that expressed Ki67, a marker of proliferating and ISC-like cells (50), was substantially decreased in adenomas exposed to JW55 when compared with tumors that developed in the control mice (Fig. 7B, panel h and i).

Importantly, when examining the healthy mucosa with antibodies against various epithelial cell populations, such as EphB2 for ISCs and Krt20 for terminally differentiated epithelial cells (50), we did not observe changes in its morphology, proliferation, or differentiation after exposure to JW55 (Fig. 7D). In summary, the results obtained from JW55-treated conditional Apc knockout mice indicated that in vivo small-molecular inhibition of TNKS1/2 leads to decreased canonical Wnt signaling followed by reduced adenoma induction and possibly progression of adenoma carcinoma.

Discussion

Altered properties of Wnt/β-catenin signaling are enabling factors for a multitude of diseases, including cancer. Among known interference points in the canonical Wnt signaling pathway, the PARP domains of TNKS1/2 seem to be a particular suitable target for inhibiting the pathway. Blocking the PARP domain of TNKS1/2 with low molecular inhibitors, such as XAV939, IWR-1, and JW74 (4, 34), leads to the stabilization of the destruction complex that triggers increased degradation of β-catenin (19). As predicted for a TNKS inhibitor, we observed a massive cytoplasmic accumulation of AXIN2 after treatment with 500 nmol/L JW55, followed by a significant reduction of β-catenin levels in vitro (Fig. 5). A similar stabilization of AXIN2 was also seen with JW74, XAV939, and IWR-1 (4, 22, 34). However, despite the substantial increased specificity of JW55 to the PARP domain of TNKS1/2 when compared with XAV939, we cannot rule out that JW55 may affect additional mechanisms that contribute to the observed effects in colon cancer cell lines and in Apc<sup>Cre</sup>/Lgr5-CreERT2<sup>+</sup> mice. For instance pyrvinium decreases Wnt signaling both by stabilizing AXIN2 and through inhibition of CK1ε (24). Furthermore, in complementation to altering canonical Wnt/β-catenin signaling, a JW55-mediated inhibition of TNKS1/2 may induce additional intracellular effects including altered telomere maintenance or spindle formation.

The in vivo treatment of tamoxifen-induced polyposis in Apc<sup>Cre</sup>/Lgr5-CreERT2<sup>+</sup> mice JW55 showed a profound effect on tumor progression. In comparison with the well-established Apc<sup>Mie</sup> mice (47), this mouse model provides a clearer assay for scoring adenoma formation because the tumor induction and treatment are time controlled. However, as adenoma induction is rather strong, it is difficult to assess later stages of CRC development as mouse mortality does not allow long-term tracking of the tumors. Thus, the presented data further solidify the functional implication of PARsylation by TNKS1/2 in a malignant transformation of the colon and the small intestine.

Interestingly, in our setting, the exposure of progenitor cells in a normal crypt to JW55 did not exert visible alterations in the intact mucosa as measured with the markers EphB2, Ki67, and Krt20. However, we cannot exclude that a longer treatment may cause reduction in cell turnover in the intestine epithelium (45). The efficient in vivo reduction of adenomas in mice and CRC growth in vitro by a tankyrase-specific inhibitor further strengthens the importance of TNKS and β-catenin as potential therapeutic biotargets.

TNKS1/2 is a member of a family of 5 PARP proteins in humans, including PARP1, which is an extensively explored...
target for drug development (33). Particularly in the BRCA1/2 mutant background, PARP1 inhibitors exhibit therapeutic effects in vitro and in vivo. Because PARP proteins are involved in a range of biologic functions, including genome maintenance, the selectivity of PARP inhibitors may be an issue in the therapeutic setting. Similar to XAV939, JW55 inhibits auto-PARsylation of TNKS1 and TNKS2 in a biochemical assay. However, in contrast to XAV939, we see no evidence for the inhibition of PARP1 by JW55, making this chemical assay. However, in contrast to XAV939, we see no evidence for the inhibition of PARP1 by JW55, making this possible for further development toward a TNKS1/2 selective inhibitor. It remains to be determined in which settings a preferred PARP1 inhibitor such as for example olaparib, a broad PARP inhibitor such as XAV939, or a selective TNKS inhibitor such as JW55, JW74, and IWR-1 may be the favorable choice.

Disclosure of Potential Conflicts of Interest
The described chemical compound may have commercial value if further investigated.

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